

Synthesis of (+) and (-)-Streptomyces coelicolor Butanolide 5 (SCB-5)

Jonas Donges,^[a] Sandra Hofmann,^[b] Moritz Brüggemann,^[c] Andrea Frank,^[a] Dieter Schollmeyer,^[a] and Udo Nubbemeyer^{*[a]}

Various 1-(1-hydroxyalkyl) paraconyl alcohols are important signaling molecules within antibiotics production in Streptomyces sp. Intending developing a flexible convergent chemical synthesis of such butanolides, a zwitterionic aza-Claisen rearrangement was chosen as reliable strategy generating the central stereotriad. Reaction of enantiopure *N*-allyl pyrrolidines and 4-phenylbutenoic acid fluoride delivered defined configured amides displaying the 2,3,1' stereotriads. The configuration

Introduction

Small molecule hormones are known to trigger the antibiotic production in various bacteria.^[1] Focusing on Streptomyces sp., defined 2,3 disubstituted γ -butyrolactones had been identified as a family of compounds characterized by prominent bioactivity.^[2] Systematic investigation and structure elucidation had been initiated by the pioneering work of Khokhlov et al. in 1976 isolating and characterizing the so-termed A-factor 1g from Streptomyces griseus (S. griseus) displaying the only 2acyl-3-hydroxymethyl γ -butyrolactone natural product found so far (Figure 1).^[3] Since then, three series of trans 2-(1'-hydroxyalkyl)-3-hydroxymethyl y-butyrolactones had been isolated from Streptomyces successively sp. such as S. viridochromogenes,^[4a] S. bikiniensis,^[5a] S. cyaneofuscatus,^[5a] S. coelicolor,^[4a-d] S. virginiae,^[5c,d] S. lavendulae,^[4e] S. badius.^[6a] Despite of the fact, that the relative and absolute stereochemistry of several γ -butyrolactones (GBL's) remain to be validated,^[4a,5a,7] three substitution pattern types predominate. The Streptomyces coelicolor butanolide/IM-2 type 2 (SCB) comprises (2R, 3R, 1'R)-trans-2-(1'-hydroxyalkyl)-3-(hydroxymethyl)- γ -butyrolactones SCB-1 (Gräfe SCB-Factor 1) to SCB-8 and

r - 1	1 Dan and A. Frank, Dr. D. Cakallarawan, Draf, Dr. 11 Nichhamanan
[a]	J. Donges, A. Frank, Dr. D. Schollmeyer, Prof. Dr. U. Nubberneyer
	Organische Chemie
	Johannes Gutenberg-Universität Mainz
	Duesbergweg 10–14, 55128 Mainz, Germany
	E-mail: nubbemey@uni-mainz.de
[b]	S. Hofmann
	Konrad-Adenauer-Gymnasium
	Wörthstr. 16, 56457 Westerburg, Germany
[c]	Dr. M. Brüggemann
	Shimadzu Deutschland GmbH
	Im Leuschnerpark 4, 64347 Griesheim, Germany
	Supporting information for this article is available on the WWW under https://doi.org/10.1002/ejoc.202100497
raf (© 2021 The Authors. European Journal of Organic Chemistry published by

© 2021 The Authors. European Journal of Organic Chemistry published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. was determined by the allyl alcohol moiety indicating a complete remote stereo control. Amide removal by iodolactonization and proceeding reductions, halocyclization and elimination gave key alkylidene tetrahydrofuran derivatives. Stepwise degradation of the olefins through ozonolysis, reductive workup and protecting group removal delivered both enantiomers of the target Streptomyces coelicolor butanolide 5.



Figure 1. γ-Butyrolactones from natural sources/bioreactors: (C₇)**1g** (–)-A-factor (R=3-Me–C₄H₈ [*i*C₅H₁₁]). **2** (SCB-type, increasing C2 chain length) (C₄) **2a** (–)-IM-2 (R=Me). (C₆) **2c** (–)-SCB-8 (R=nC₃H₇). **2d** (–)-SCB-5 (R=2-Me–C₃H₆ [*i*C₄H₉]). **2e** (+)-SCB-4 (R=1-Me–C₃H₆ [*s*C₄H₉], 2 diastereomers), (C₇) **2f** (–)-SCB-6 (R=nC₄H₉). **2g** (–)-SCB-1, Gräfe SCB-Factor 1 (R=3-Me–C₄H₈ [*i*C₅H₁₁]). (C₆) **2h** (–)-SCB-2 (R=nC₅H₁₁), **2j** SCB-3 (R=3-Me–C₅H₁₀ [*s*C₆H₁₃], 2 diastereomers). (C₉) **2k** SCB-7 (R=nC₆H₁₃). 2-(1,n-dihydroxylalkyl) SCB-series: (C₄) **2l** (R=3-hydroxy-3-Me–C₄H₇). (C₅) **2m** (R=3-hydroxy-4-Me–C₅H₉, 2 diastereomers). (C6): **2n** (R=4-hydroxy-5-Me–C₆H₁₁, 2 diastereomers). **2o** (R=5-hydroxy-5-Me–C₆H₁₁). **3** (*W*B-type, increasing C2 chain length) (C₅) **3b** (–)-VB–E (R=1-Me–C₅C₄ [*i*C₄H₇]). (C₆) **3c** (–)-VB–C (R=nC₃H₇). **3d** (–)-VB–A, (–)-Gräfe VB-factor 1, (R=2-Me–C₃H₆ [*i*C₄H₉]). **3e** (–)-VB–B (R=1-Me–C₃H₆ [*s*C₄H₉]), **2d** iastereomers). (C₇) **3f** (–)-VB–D (R=nC₄H₉), **3g** (–)-Gräfe VB-factor III (R=3-Me–C₄H₈ [*i*C₅H₁₁]). (C₆) **3i** (–)-Gräfe VB-factor III (R=4-Me–C₅H₁₀ [*i*C₆H₁₃]).

IM-2 2a-2k.^[4] In contrast, the Virginiae butanolide (VB) type 3 displays the (2*R*, 3*R*, 1'S) diastereomers VB–A (Gräfe VB-Factor I) to VB–E and Gräfe VB-Factors II and III 3a-3i.^[5] The SCB-type

Chemistry Europe European Chemical Societies Publishing

GBL's incorporating 2-(1',n-dihydroxyalkyl) side chains should be attributed to a third family 2I-2o.^[6]

Various defined GBL's have been isolated from different streptomyces strains.^[4,5] Obviously, a particular GBL influences biosynthesis pathways depending on the individual bacterium.^[2e,4d,8] Analyzing the SCB and the VB diastereomers and enantiomers of the SCB-1 type, Takano et al. found, that the bioactivity of the all (*R*) configured (–)-SCB-1 showed by far the highest activity within a Kanamycin bioassay in comparison to the three analogues pointing out the importance of the defined substituted and configured compound in connection with the bioactivity of a special organism.^[4c] Furthermore, investigation of a series of GBL's gave an optimal C2 chain length of 7 to 9 carbon atoms, an additional methyl group at C6 (favored) and C7 (less favored) and a 1' (*R*) OH group. Generally, the corresponding 1'(*S*) VB diastereomers were described to be less active.^[4c]

In contrast, VB-bioessays gave different results in respect to bioactivity. Investigations by Y. Yamada et al. gave optimal C2 chain lengths of 6 to 8 carbon atoms,^[2e,8] an additional methyl group at C5 (VB–A) and the standard 1' (S) configured OH group.^[9] In contrast, additional methyl groups at C4 caused a drastic loss of bioactivity.^[Sd] The corresponding SCB-type diastereomers were described to be about 10-fold less active.^[9a] Investigation of the VB–A hyperproducing strain S. antibioticus NF-18 improved the biosynthesis of (–)-VB–A (major) and both enantiomers of SCB-5 (minor).^[9d] However, the bioactivity regulating role of the GBL's remains unclear.

Despite of the fact, that modified hyperproducing Streptomyces strains deliver (semi)preparative amounts of suitable GBL derivatives, chemical syntheses of A-factor,^[10a-e] SCB-type^[4b,10f-i] and VB-type^[10j-k] products promised a higher flexibility upon generating defined diastereomers and enantiomers.^[4b,5c,10]

Chemical syntheses of GBL's **A** known so far can be subdivided in two strategies (Figure 2). Most attempts described the generation of optically active protected paraconyl



Figure 2. Chemical syntheses of butanolides (X=O-alkyl, NH-alkyl, N-acyl/ alkyl; PG=OH, TBS, Bn, Ph, acyl, etc.). For details see Ref 10.

alcohol **C** from paraconic acid **B-1** and open chain reactants **B-1/B-2**, respectively. The final steps incorporate the butanolide C2 side chain introduction via aldol type reaction (using suitable aldehydes, direct formation of **A**)^[10f] or via Claisen condensation (using acid derivatives) affording 2-acyl butanolides **C** and subsequent reduction of the ketone moiety (\rightarrow **A**).^[10f-i] Jørgensen et al.^[10k] started with a Michael addition of a suitable β -ketoester (X=OR)/amide (X=NR₂) derivative **D** and nitroalkene **E** to generate a protected 3-nitroallyl alcohol **F**. Ring closure generating the γ -lactone **G**, reduction of the ketone moiety and Nef reaction of the nitro group and a final reduction delivered the target molecules **A**.

Overall, all attempts incorporated a late-stage introduction of almost two stereogenic centers requiring challenging structure elucidations. Until 1990, the VB-type GBL's had been described as 2,3-*cis* butanolides displaying 2(*S*), 3(*R*), 1'(*R*) configurations (see K. Mori et al. 1990).^[10h] In 1991, Y. Yamada et al. published a corrected assignment of the stereocenters of the VB-type GBL's as 2(*R*), 3(*R*), 1'(*S*).^[10h] Obviously, the so termed "*trans*-VB-type" diastereomers effectively displayed the SCBtype 2(*R*), 3(*R*), 1'(*R*) configuration.^[10h,i] Consequently, synthesis strategies separating the generation of the stereogenic centers and the complete assembly of the β hydroxy lactone moiety occurred advantageous meeting the stereoselectivity challenges.

Results and Discussion

Re-analyzing the substitution patterns of the Streptomyces butanolides, the sequence of three continuous stereogenic centers (2, 3, 1') can be defined as the central segment of all targets. The defined generation of the optically active stereotriad independent of final ring closure and functional group transformations promised developing flexible and reliable syntheses of selected natural products. The retrosynthesis concentrated on assembling the backbone including the stereogenic centers (1', 2, 3) using a kit of building blocks and final functional group variations enabling completion of the syntheses (Scheme 1).

Starting from target butanolide I, lactone and hydroxymethyl function should originate from the corresponding olefins in II via ozonolytic cleavage and reductive work-up without endangering epimerization of the adjacent C2 and C3 positions. The vinyl ether in II must be obtained from unsaturated carbinol III via haloether formation and elimination. Carbinol III represents the reduction product of amide IV, which should be build-up by a zwitterionic aza-Claisen rearrangement from allylamine V and styryl acetic acid fluoride VI.^[11] Acyl fluoride VI can be generated in two steps from malonic acid and phenyl acetaldehyde via Knoevenagel condensation and acid fluoride formation.^[12] Allylamine V should be obtained from aldehyde VII and a suitable N-propargyl prolinol derivative VIII via alkyne addition reduction and carbinol protection.^[13] Prolinol derivative VIII can be generated from L-(–)-proline via reduction/OH protection and N propargylation adapting literature procedures.^[13]

Full Papers doi.org/10.1002/ejoc.202100497





Scheme 1. Retrosynthesis of defined γ -butyrolactones. Streptomyces coelicolor butanolide 5 (SCB-5) 2d and Virginiae butanolide A (VB–A) 3d substitution pattern: R=4-Me–C₅H₁₀/iC₆H₁₃.

For a first total synthesis, the SCB-5/VB–A 2 d/3 d substitution pattern was chosen. (–)-VB–A is described as the most bioactive compound of the VB family. Including the corrections of Yamada et al.^[10i] Mori et al. already published the syntheses of (–)-SCB-5 2 d and (–)-VB–A 3 d in 1990 as a mixture of diastereomers.^[10h,14] Therefore, diastereoselective syntheses aiming for assembling complete data sets of these compounds have been developed in our laboratory.

The synthesis started synthesizing the key *N*-allyl prolinol diastereomers **10** and **11** (retrosynthesis compound **V**) as well as acid fluoride **12** (retrosynthesis compound **VI**) according literature procedures.^[15]

For optically active pyrrolidine derivative **7** and **8** syntheses, *L*-(–)-proline methyl ester **4** was used as starting material (Scheme 2). A five-step sequence (N-benzylation, LiAlH₄ reduction, Buchwald phenyl ether formation, benzylamine hydrogenolysis and N-propargylation) afforded *N*-propargyl prolinol **5** via five steps with 74% yield overall.^[13] Alkynyl anion coupling with aldehyde **6** (two steps from 5-methyl hexanoic acid: LiAlH₄ reduction, TEMPO oxidation, 96%), HPLC resolution of the



Scheme 2. Syntheses of optically active *N*-allylpyrrolidine diastereomers (4*S*)-7 and (4*R*)-8 according ref. 13.

diastereomers of the corresponding *O*-acetyl mandelic esters, LiAlH₄ reduction and allylalcohol TBS protection delivered both, *N*-allyl prolinol derivative (4*R*)-**8** with 34% yield and diastereomer (4*S*)-**7** with 37% yield. Alternatively, a five-step sequence of alkynyl anion (from **5**) coupling with aldehyde **6**, MnO₂ oxidation and subsequent Midland's reagent controlled diastereoselective ketone reduction, LiAlH₄ reduction and TBS protection allylalcohol moiety selectively afforded *N*-allyl prolinol derivative (4*R*)-**8** with 42% yield.^[16]

Acid fluoride **9** was obtained from phenylacetaldehyde and malonic acid via two steps (Scheme 3). Literature known Knoevenagel condensation of phenylacetaldehyde and malonic acid delivered 4-phenyl-3-butenoic acid (78% yield),^[17] which then was activated with cyanuric fluoride and pyridine to give acid fluoride **9** with quantitative yield.^[12c]

The aza-Claisen rearrangement using allylamine diastereomers (4S)-7 and (4R)-8, respectively, and acid fluoride 9 served as the key step upon generating the stereotriad displaying amides 10 and 11 (Scheme 4).^[11] Predictably, the reaction of TBS-ether derived allylamine (45)-7 and acid fluoride 9 gave product amide 10 with 93% yield as a single diastereomer (matched combination of the stereodirecting factors, see Figure 5 below). First attempts intending for direct reduction of the amide to the corresponding primary alcohol failed despite of wide variation of the reaction conditions.^[18] Because of unconvincing results running direct tertiary amide cleavage processes,^[19] a reliable two-step sequence was applied. Removal of the pyrrolidine auxiliary moiety succeeded via iodo lactonization.[11g,19,20] Treatment of amide 10 with iodine in DME/H₂O delivered a mixture of C4- α iodolactone *ent*- α -11 and C4- β iodolactone *ent*- β -11 as a 0.85:1 mixture of C4-epimers with 91% yield. The relative configuration of the new stereogenic centers (C2/C3 anti) was elucidated by means of NOESY analyses. Furthermore, iodolactone ent- β -11 crystallized and the X-ray analysis improved absolute and relative configurations of all stereogenic centers (Figure 3).^[21] Reductive opening of the lactones ent-11 with Zn/ HOAc gave the intermediate acid (85% yield, not shown), which was treated with diazomethane solution to afford corresponding ester ent-15 with 88% yield (75% yield over 2 steps). Furthermore, reaction of the intermediate acid with SOCl₂ in MeOH caused TBS group removal and subsequent cyclization to give lactone ent-13 with 90% yield (77% yield over 2 steps). Careful structure elucidation via NOESY analysis gave the cis arrangement of all γ -butyrolactone side chains.

Surprisingly, reaction of allylamine (4*R*)-**8** and acid fluoride **9** delivered amide **14** with 90% yield as a single diastereomer (Scheme 4). No minor compounds had been found after careful analyses (HPLC, NMR). Despite of the fact, that the mismatched combination of the stereodirecting ether and pyrrolidine



Scheme 3. Synthesis of acid fluoride 9. Conditions and yields: iv) (a) *N*-methyl morpholine, 95 °C, 14 h, 78%. Ref 17. (b) $C_3F_3N_3$, CH_2CI_2 , pyridine, 0 °C to 23 °C, 3 h, > 99%. According Ref. 12c.

Full Papers doi.org/10.1002/ejoc.202100497



Scheme 4. Aza-Claisen rearrangement of allylamines (45)-7 and (4*R*)-8, structure elucidation and auxiliary removal. Conditions and yields: i) 9, Me₃Al, Na₂CO₃, CH₂Cl₂, 0 °C, 4 d, yields: 93 % (10), 90 % (14). (ii) I₂, DME, H₂O, 23 °C, 4 d, yields: 38 % β -11 + 38 % α -11 (from 13); yields: 49 % *ent*- β -11 + 42 % *ent*- α -11 (from 10), 21 h. (iii) Zn, HOAc, Et₂O, 23 °C, 2 d, (iv) SOCl₂, MeOH, 23 °C, 3 d, yields: 55 % 13 (from 11); 77 % *ent*-13 from *ent*-11 (two steps each). (v) CH₂N₂, Et₂O, 0 °C, 5 h, yields: 75 % 12 (from 11); 75 % *ent*-12 from *ent*-11 (two steps each). Remark: Since α and β characterize the relative arrangement of the iodomethyl group within the present drawing, α -11/*ent*- β -11 as well as β -11/*ent*- α -11 represent enantiomers.

moieties had been involved, one of these factors completely predominated the diastereoselectivity of the process (see Figure 5 below). Again, removal of the pyrrolidine via iodocyclization delivered lactones α -11 and β -11 as a 1:1 mixture of C4-epimers (76% yield). The relative configuration of the new stereogenic centers (C2/C3 *anti*) could be elucidated by means of NOESY analyses. In analogy to the enantiomer series mentioned above, iodolactone α -11 crystallized and the X-ray analysis improved absolute and relative configurations of all stereogenic centers (Figure 4).^[21] Finally, reductive opening of the lactones 11 with Zn/HOAc gave the intermediate acid (81% yield, not shown), which was treated with diazomethane solution to afford corresponding ester 12 with 92% yield (75% yield over 2 steps). Furthermore, reaction of the intermediate



Figure 3. X-ray structure of iodolactone ent- β -11 (only selected hydrogens are shown).



Figure 4. X-ray structure of iodolactone α -11 (only selected hydrogens are shown).

acid with SOCl₂ in MeOH caused TBS group removal and subsequent cyclization to give lactone **13** with 68% yield (55% yield over 2 steps). Careful structure elucidation via NOESY analysis gave the *cis* arrangement of all γ -butyrolactone side chains.

Methyl esters **12** and *ent*-**12** displaying both enantiomeric stereotriades served as starting key intermediates for SCB/IM-2 butenolide syntheses (2R,3R,1'R) (-)-SCB-5 **2d** (Scheme 6, Scheme 7, Scheme 9) and (2S,3S,1'S) (+)-SCB-5 *ent*-**2d** (Scheme 5, Scheme 8).^[22] Reduction of the ester functions with DIBAL–H afforded the carbinols **15** and *ent*-**15** with high yields (99% of **15** and 85% of *ent*-**15**, respectively).

A most straight forward process heading for the target butanolide *ent-***2d** formation was the oxidative cleavage of both double bonds of carbinol *ent-***15** (Scheme 4). Ozonolysis



Scheme 5. Synthesis of iodo tetrahydrofuranes *ent*-20 and *ent*-21: Conditions and yields: (i) DIBAL–H, CH_2CI_{2r} –78 °C, 5 h, yield: 85% *ent*-15; (ii) SnCI₂, PhSeBr, CH_2CI_2 , 23 °C, 15 min, yield *ent*-17 77% overall. HPLC separation gave β (CH_2SePh) *ent*-17: 32% (OTBS + 9% (OH), α (CH_2SePh) *ent*-17: 26% (OTBS) + 10% (OH). (iii) from *ent*-15 via crude *ent*-17: mCPBA, pyridine, DME, -78 °C, 30 min, 23 °C, 1.5 h, yield *ent*-18 (mixture of diastereomers) 97% (from *ent*-15). (iv) pyridine, DME, 80 °C 4 h, yield *ent*-19: 19% (from *ent*-15). (v) I_2 , NaHCO₃, Et₂O, H₂O, 23 °C, 9 h yields: 36% *ent*-20 and 48% *ent*-21 (d.r. 43:57). (vi) Zn, HOAc, Et₂O, 23 °C, 22 h, yield: >99%. (vii) tBuOK, THF, 60 °C, 6 d, yield: >99% *ent*-22.

had to generate two aldehyde functions, five-membered ring lactol formation should protect the C1 position. The remaining aldehyde moiety had to be reduced selectively^[20] to deliver hydroxymethyl lactol *ent*-**16**. However, various attempts varying reagents and conditions failed because of the rapid destruction of the material under the oxidizing conditions or the competing reduction of the lactol delivering triol side products (stereo-chemical information of C3 is lost).

The failure of the twofold oxidative cleavage of the double bonds in combination with proceeding chemoselective aldehyde reduction (hydroxymethyl moiety) and oxidation (lactol moiety) motivated to generate an enolether function first (Scheme 5). Now, ozonolysis of olefin and enolether type *ent-*22 would have generated a lactono aldehyde intermediate, the so formed aldehyde function had to be selectively reduced to build-up hydroxy lactones type *ent-*23.

For tetrahydrofuran formation and subsequent β elimination to generate exo enol ethers of type ent-22, a selenocyclization,^[24a-c] oxidation, *syn*-elimination cascade^[24d,e] had been tested (Scheme 5). Treatment of carbinol ent-15 with PhSeBr and SnCl₂ in CH₂Cl₂ smoothly delivered 2-(phenylselanylmethyl) tetrahydrofuran ent-17 with 77% yield as a 41:36 mixture of C1 β and α -epimers.^[25] Subsequent oxidation with mCPBA afforded the corresponding selenoxide ent-18 diastereomers with nearly quantitative yields (> 97%). However, in contrast to literature precedence describing tetrahydropyran enolether formation from phenylselenyl methyl ехо precursors,^[24d,e] no β elimination generating enolether *ent*-22 could be enforced. Heating of the material induced decomposition and competing product formation (e.g. furofuran derivative ent-19 after silyl group removal and cyclization). Furthermore, no exo enolether ent-22 and hydroxylactones type ent-23 compounds were found upon running selenium oxidation, oxidative cleavage of the olefins and aldehyde reduction as a one pot process.

Ring closure by means of iodocyclization required careful optimization.^[26] In contrast to the iodolactonizations using amides **10/14** and the selenocyclizations described above,

treatment of carbinols 15 and ent-15 with iodine in Et₂O/aq. NaHCO₃ afforded two regioisomer tetrahydrofurans 20/21 (63% yield, ratio 48:52) and ent-20/ent-21 (84% yield, ratio 43:57) as single diastereomers, respectively (Scheme 5, Scheme 6). After careful column chromatography and HPLC separation 2-(iodomethyl) tetrahydrofurans 21/ent-21 displayed the standard 5-exo-trig substitution pattern in favor of the trans C1/C2 arrangement of the side chains.[26a-f] The formation of the regioisomers 20/ent-20 indicated a competing 5-endo-trig reaction.^[26g,h] Obviously, the attack of the iodo cation at the styryl double bond generated an intermediate benzyl cation, which finally underwent intramolecular five membered ring ether formation affording the 2-phenyl-tetrahydrofurans 20/ent-20 with trans arrangement of all side chain substituents. Wide variation of iodo cation donor and reaction conditions resulted no higher regioselectivity in favor of regioisomers 21/ent-21. However, starting from iodo tetrahydrofuran ent-20, reductive elimination using Zn and HOAc enabled regeneration of carbinol ent-15 with nearly quantitative yield (Scheme 5).

Then, installation of the exo enolether moiety had been investigated (\rightarrow *ent*-22) (Scheme 5).^[27] Reactions of iodomethyl derivative *ent*-21 at room temperature induced substitution



Scheme 6. Synthesis of iodo tetrahydrofuranes 20 and 21: Conditions and yields: (i) DIBAL–H, CH₂Cl₂, -78 °C, 5 h, yield: >99% 15; (ii) I₂, NaHCO₃, Et₂O, H₂O, 23 °C, 17 h yields: 32% 21 and 31% 20 (d.r. 52:48).

0990690 20 22. Downloaded from https://chemistry-europe.onlinelibrary.wiley.com/doi/10.1002/ejoc.202100497 by Cochrane Germany, Wiley Online Library on [17/10/2022]. See the Terms and Conditions (https://onlinelibrary.wiley.com/termsand-conditions) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons

License

processes. AgOTf/tBuOK, pyridine replacement of iodide only generated carbinols and pyridinium salts, respectively.^[28] In contrast, long lasting subjecting of iodomethyl tetrahydrofuran *ent*-**21** to potassium *tert*-butoxide in refluxing THF gave *exo* enolether *ent*-**22** with > 99% yield after 9 days of reaction time. Again, simultaneous oxidative cleavage of both double bonds failed (\rightarrow *ent*-**23**). The intermediately formed lactonoaldeyde might have suffered from proceeding condensation (β -eliminaton of the lactone carboxyl group) and further oxidation of the nascent acrolein moiety.

Overall, the simultaneous oxidative cleavage of both olefines failed, enforcing development of a stepwise process. In this connection, the iodoethers **20**/*ent*-**20** and **21**/*ent*-**21** served as appropriate starting materials.

2-Phenyl tetrahydrofuran 20 was chosen to install the carboxyl function first.^[29] Ozonolysis of iodoether 20 in MeOH and oxidative work-up with aqueous H_2O_2 afforded acid 24 (Scheme 7). Because of work-up and purification problems, a disappointing yield of 21 % was found. For facilitating chromatographic purification and characterization, the crude acid 24 was immediately treated with a solution of freshly prepared diazomethane. Methyl ester 25 a was isolated with moderate 26% yield (3 steps). Remaining aldehyde 25b (21% yield) indicated an incomplete second H₂O₂ oxidation step. Reductive ring opening with Zn in AcOH/Et₂O induced regeneration of the styrene moiety, the intermediate hydroxy acid underwent ylactonization delivering butanolide 26 with 49% yield from acid 24 and 87% yield from ester 25a. Best results were achieved running a cascade of oxidative degradation of the vinyl group and reductive ring opening of the iodo tetrahydrofuran segment with final lactonization. Stating from iodoether 20, such a sequence enabled generation of butanolide ${\bf 26}$ with 36% yield overall. $^{\scriptscriptstyle [30]}$

Ozonolytic cleavage of the styryl double bond of lactone **26** in MeOH/CH₂Cl₂ and subsequent reductive work-up using NaBH₄ afforded 3-(hydroxymethyl) lactone **23** with 71% yield.^[31,11d] Finally, protecting group removal with buffered TBAF- solution in THF/HOAc enabled to complete a first total synthesis of (–) SCB-5 **2 d** with 98% yield.^[32]

lodomethyl tetrahydrofuran *ent*-**21** served as key intermediate for a synthesis of (+)-SCB-5 *ent*-**2d**. The starting sequence focused on introduction of the 3-(hydroxymethyl) group first (Scheme 8). Treatment of reactant *ent*-**21** with O_3/O_2 in MeOH, subsequent reduction with NaBH₄ and final protection of the carbinol delivered TBS ether *ent*-**27** with >76% yield (2 steps). Reductive ring opening using Zn/HOAc afforded alkenol *ent*-**28** with 79% yield.

For introduction of the lactone carboxyl function, two alternative reaction paths were tested (Scheme 8). In analogy to the oxidative degradation of the vinyl group of tetrahydrofuran **21** delivering lactone **26** (see above, Scheme 7), hydroxyalkene *ent*-**28** was subjected to O_3/O_2 in CH_2CI_2 and subsequent oxidative work-up with 35% aq. H_2O_2 . Surprisingly, a lactol only (82% yield, not shown) was found indicating the failure of the final oxidation. No lactone *ent*-**30** was detected. Consequently, introduction of the carboxyl function required an additional transformation. Reaction of the intermediate lactol with PCC in dry CH_2CI_2 gave the desired TBS protected SCB-5 derivative *ent*-



Scheme 7. Synthesis of (–)-SCB-5 2 d. Conditions and yields: (i) O_3/O_2 , MeOH, -78 °C, 20 min, then: 35% aq. H₂O₂, MeOH, -78 °C to 23 °C, 15 h. Yield: 21% 24. (ii) CH₂N₂, Et₂O, 0 °C, 3 h, yield 26% 25 a (from 20) + 21% of the aldehyde 25 b. (iii) Zn, HOAc, Et₂O, 23 °C, 36 h, yields of 26: 49% (from 24), 87% (from 25 a), 36% (from 20). (iv) O_3/O_2 , MeOH/CH₂Cl₂, -78 °C, 1 h, then: MeOH/ CH₂Cl₂, NaBH₄, -78 °C to 23 °C, 23 h, Yield 71% of 23. (v) TBAF, HOAc, THF, 23 °C, 19 h, yield: 98% 2 d, (–)-SCB-5.



Scheme 8. Synthesis of (+)-SCB-5 *ent*-2d. Conditions and yields: (i) O_3/O_2 , CH_2Cl_2 , -78 °C, 15 min, then: MeOH, NaBH₄, -78 °C to 23 °C, 22 h, Yield 99% of carbinol. (ii) TBSOTf, Et₃N, CH₂Cl₂, 23 °C, 20 h, yield 78% silylether *ent*-27. (iii) Zn, HOAc, Et₂O, 23 °C, 24 h, yield 79% olefin *ent*-28. (iv) (a) O_3/O_2 , CH_2Cl_2 , -78 °C, 15 min, then: 35% aq. H_2O_2 , CH_2Cl_2 , 23 °C to reflux, 2 h, yield: 82% of lactol. (b) PCC, 3 Å molsieves, CH_2Cl_2 , 23 °C, 1 h, Yield: 34% lactone *ent*-30 (+ 43% of ketoformiate, see supporting information). (v) tBuOK, THF, 50 °C, 3 d, Yield 98% of enolether *ent*-29. (vi) O_3/O_2 , CH_2Cl_2 , -78 °C, 1 h, then: Me₂S, CH_2Cl_2 , -78 °C, 15 h, yield: 99% of target (+)-SCB-5 *ent*-20. (vii) TBAF, HOAc, THF, 23 °C, 15 h, yield: 99% of target (+)-SCB-5

30 with moderate 34% yield. Competing degradation products could be isolated in up to 43% yield.^[33]

The cumbersome oxidative cleavage of alkenol *ent*-**28** advised reactivation of the enol ether pathway (Scheme 8, in analogy to the consideration to generate lactone moiety in *ent*-**23**, Scheme 5). Starting from TBS ether *ent*-**27**, H–I β -elimination succeeded using KOtBu in THF at 50 °C. After 3 d of reaction time *exo* enol ether *ent*-**29** could be isolated with 98% yield. Oxidative cleavage of the double bond with O₃/O₂ in CH₂Cl₂ and standard work-up with Me₂S delivered the desired TBS protected SCB-5 *ent*-**30** with 75% yield. Finally, global protective group removal with TBAF/HOAc in THF gave the target (+)-SCB-5 derivative *ent*-**2 d** with <99% yield.^[32]

In addition, the synthesis of (-)-SCB-5 2d had been run via the optimized route developed for (+)-SCB-5 ent-2d applying slightly varied conditions (Scheme 9). The oxidative cleavage of the styrene subunit starting from tetrahydrofuran 21 with O₃/O₂ in CH₂Cl₂, subsequent reduction with NaBH₄ and final protection of the carbinol delivered TBS ether 27 with 70% yield (2 steps). Too, reductive ring opening using Zn/HOAc enabled to generate alkenol 28 with 81% yield. Again, the enol ether route proved successful. Starting from TBS ether 27, H–I β -elimination was achieved using KOtBu in THF at 50 °C. After 3 d of reaction time exo-enol ether 29 could be isolated with nearly quantitative yield. Oxidative cleavage of the double bond with O_3/O_2 in CH₂Cl₂ and standard work-up with Me₂S delivered the desired TBS protected SCB-5 derivative 30 with 63% yield. Finally, global protective group removal with TBAF/HOAc in THF gave the target (–)-SCB-5 **2 d** with < 99% yield.^[32]



 $\begin{array}{l} \label{eq:scheme 9. Synthesis of (-)-SCB-5 2 d. Conditions and yields: (i) O_3/O_2, CH_2Cl_2, $-78 °C, 1 h, then: MeOH, NaBH_4$, $-78 °C to 23 °C, 21 h, Yield 82% of carbinol. (ii) TBSOTf, $Et_3N, CH_2Cl_2, $23 °C, $20 h, yield 86% silylether 27. (iii) $Zn, HOAc$, $Et_2O, $23 °C, $21 h, yield 81% olefin 28. (iv) tBuOK, $THF, $50 °C, $3 d, Yield 99% of enolether 29. (v) O_3/O_2, CH_2Cl_2, $-78 °C, $1 h, then: Me_2$, CH_2Cl_2, $-78 °C to $23 °C, $3 h, Yield 63% of lactone 30. (vi) TBAF$, $HOAc$, THF, $23 °C, $20 h, yield: $>99% of target (-)-SCB-5 2 d. \\ \end{array}$

Comparison of the ¹H NMR data of our (–)-SCB-5 with these published by Mori et al. (corrected structure)^[10h,i] gave a convincing agreement of both sets. Especially, the lactone protons H-5 and H-5' were described as two dd systems at 4.41 and 3.97 ppm. The proton attached to C-1' was found as a m between 4.05 and 4.00 ppm. In contrast, the VB–A protons are found as dd systems at 4.42 and 4.1 ppm, respectively, the proton attached to C-1' was described as a m between 4.15 and 4.10 ppm. Comparison of the data of further SCB/VB type substitution pattern pairs of several compounds **2/3** displayed the same tendency in connection with the protons mentioned above. Unfortunately, no ¹³C NMR data of SCB-5 have been published in the literature so far.^[34]

Generally, the aza-Claisen rearrangement as the central key step offers two options in respect of the stereocontrol building up stereotriads (Figure 5) involving *N*-allylpyrrolidines (4*S*)-**7** and (4*R*)-**8**, respectively and activated intermediate ketene **9a** (from acid fluoride **9** and Me₃Al, evolution of methane).^[11] The addition of the lone pair of the nitrogen at the carbonyl center of the ketene **9a** was the initiating step in both cases, delivering the central zwitterionic intermediate with high diastereoselectivity. Then, Claisen rearrangement required the passing of a chair-like transition state. In this connection, the stereodirecting factors need to be examined.

1) Starting from N-allyl pyrrolidines 7 and 8 bearing a defined ether function in position 4, 1,2 asymmetric induction enables to generate stereotriads with anti/syn substitution pattern with high diastereoselectivity (minimized repulsive interactions because of anti-arrangement of enolate C and bulky TBS ether).^[35] 2) Starting from optically active N-allyl (S)prolinol derivatives 7 and 8, auxiliary induced rearrangements deliver 2,3 anti-amides with 2 (R) configurations (quasi equatorial arrangement of the bulky branched pyrrolidine moiety). The combination of both independently directing stereoinductors in a single molecule should affect matched and mismatched results concerning the amide formation. The matched case is found in reactant (4S)-7/(S)-prolinol derivative delivering the syn-anti amide 10 as an intermediate of a (+)-SCB-5 ent-2d synthesis. In contrast, the mismatched combination of reactant (4R)-8/(S)-prolinol derivative resulted syn-anti amide 14 (via mismatched-s, predominating substrate control, transition state suffering from quasi axial arrangement of the bulky branched pyrrolidine moiety) as a (-)-SCB-5 2d precursor. However, formation of anti-anti amide 14a (via mismatched-a, predominating auxiliary control, transition state suffering from repulsive interactions because of syn-arrangement of enolate C and bulky TBS ether) inducing a (+)-VB-A 3d synthesis was not observed.

Conclusion

The stereoselective synthesis of enantiopure Streptomyces coelicolor butanolide hormones required efficient access to the optically active (2,3,1') stereotriads characterizing the key fragments of this series of natural products. Zwitterionic aza-Claisen rearrangements developed earlier proved to serve as reliable tools to generate such stereotriads with high selectivity.

Full Papers doi.org/10.1002/ejoc.202100497



Figure 5. Stereochemical outcome of the zwitterionic aza-Claisen rearrangement key-step: axial N lone pair in (4*S*)-7 and (4*R*)-8, attack at the carbonyl C of ketene **9a**, passing of chair-like transition states with minimized repulsive interactions. "matched" gives *syn anti* **10** (\rightarrow (+)-SCB-5 *ent*-**2d**), mismatched-a auxiliary control gives *anti anti* **14a** (\rightarrow (+)-VB-A *ent*-**3d**), mismatched-s substrate control gives *syn anti* **14** (\rightarrow (-)-SCB-5 **2d**).

Starting from suitably substituted *N*-allylpyrrolidine (4*S*)-**7**, reaction with 4-phenylbutenoyl fluoride **9** delivered the $\gamma_i \delta$ unsaturated amide **10** as a single diastereomer with high yield (93%, matched combination of stereochemistry inducing factors). Despite of involving differently directing stereochemistry inducing factors as present in pyrrolidine (4*R*)-**8**, 1,2 asymmetric induction (remote stereocontrol) of the (4*R*)-**TBS** ether moiety predominated delivering $\gamma_i \delta$ -unsaturated amide **14** only with 90% yield. Two step removal of the amide via iodocyclization, reductive ring opening and subsequent reduction of the carboxyl function afforded carbinols **15** (*R*,*R*,*R* stereotriad, 55% yield) and *ent*-**15** (*S*,*S*,*S* stereotriad, 58% yield). Structure elucidation succeeded by means of NOESY and X-ray analyses of appropriate intermediates in both series.

Chemoselective conversion of the olefins into carboxyl and hydroxymethyl groups required careful elaborated reaction sequences. Despite of several attempts, the simultaneous degradation of both double bonds failed. lodocyclizaton of carbinols 15/*ent*-15 delivered two regioisomeric iodoethers 20/ *ent*-20 (protected styryl double bond) and 21/*ent*-21 (protected vinyl group).^[36]

Starting from iodoether **20** allowed introduction of the carboxyl function first. Best results were achieved via ozonolysis/oxidative work-up and reductive iodoether ring opening generating γ lactone **26** with 36% yield. A second ozonolysis/ reductive work-up gave protected (–)-SCB-5, a final silyl ether removal enabled completion of the total synthesis of target (–)-**2d** (25% overall from iodoether **20**).

Starting from iodoether *ent*-**21** enabled introduction of the hydroxymethyl function first. Ozonolysis/reductive work-up and silyl ether protection delivered tetrahydrofuran *ent*-**27** (77% yield). Since the carboxyl group introduction with regeneration of the vinyl group (via *ent*-**28**) and subsequent oxidative degradation gave moderate results upon generating protected

(+)-SCB-5 *ent*-**30** (6 steps from *ent*-**21**, 17% yield), potassium *tert*-butoxide induced HI elimination gave enol ether *ent*-**29**. Subsequent ozonolysis/standard work-up of the double bond afforded lactone *ent*-**30** with 74% yield (2 steps). Final protecting group removal gave (+)-SCB-5 *ent*-**2d** with nearly quantitative yield (overall: 5 steps from *ent*-**21**, 56% yield). In addition, starting from iodoether **21** enabled completion of the total synthesis of (-)-SCB-5 **2d** with 44% yield (5 steps from **21** applying the enol ether **28** cascade).

The data of the SCB-5 enantiomers synthesized here match with these published by Mori.^[10h] In contrast, a VB–A synthesis requires a late-stage inversion of the C1' OH group.^[37] Alternatively, more attractive auxiliary dominated zwitterionic aza-Claisen rearrangement would require a sterically less encumbered C4 carbinol protection group and a suitable auxiliary substituent. Further investigations in our laboratory aim for such Virginiae butanolide total syntheses.

Experimental Section

General Remarks: Thin film IR spectra were recorded with a Jasco FT/IR-4100 spectrometer with single reflection horizontal ATR (ZnSe window). Melting points were determined with an IA 9100 apparatus, manufactured by Electrothermal Engineering Ltd. Optical rotation was recorded with MC 241 polarimeter (Perkin-Elmer). ¹H NMR, ¹³C NMR and 2D NMR (COSY, HSQC, HMBC, NOESY) spectra were recorded at room temperature with a Bruker Avance III HD 300, Avance II 400 or Avance III HD 400 spectrometer in C₆D₆ or CDCl₃, e.g. using the signal of residual CHCl₃ (¹H: 7.26 ppm; ¹³C: 77.16 ppm) as internal standard. FD mass spectra were obtained using a Finnigan MAT 95, ESI spectra were measured using a Waters Micromass QTOF Ultima 3. HRMS ESI spectra were recorded using an Agilent G6545A Q-TOF spectrometer. Column chromatography was performed on MN silica gel 60 M from Macherey-Nagel GmbH & Co. KG with a grain size of 0.040–0.063 nm. The analytical HPLC

10990690,

system was used to analyze the products: Knauer HPLC Pump 64 connected to a HPLC column (see table below), a Knauer Variable Wavelength Monitor at $\lambda = 254$ nm or 220 nm and Knauer Differential Refractometer. Preparative HPLC system: Knauer WellChrom Preparative Pump K-1800 connected to the HPLC column, a Knauer Variable Wavelength Monitor at $\lambda = 254$ nm or 220 nm and Bischoff RI-detector 8100. A selection of experimental procedures is given in the following. For all procedures and the numbering used in the ¹H and ¹³C NMR spectra, see supporting information.

(2R,3S,4S)-4-((tert-Butyldimethylsilyloxy)-8-methyl-1-((S)-2-

(phenoxymethyl)-pyrrolidin-1-yl)-2-((E)-styryl)-3-vinylnonan-1one 10: Sodium carbonate (1.446 g, 13.644 mmol, 6.0 eq) was dried and N-allyl pyrrolidine (4S)-7 (1.007 g, 2.259 mmol, 1.0 eq.) in DCM (30 mL) was added. Acyl fluoride 9 (0.904 g, 5.507 mmol, 2.4 eq) in DCM (10 mL) was added at 0 °C. Then, trimethylaluminum (2.75 mL, 0.40 g, 5.5 mmol, 2.4 eq, 2 m in toluene) was added dropwise over 30 min. After stirring for 4 d at 0°C (cryostat), the mixture was warmed-up room temperature. Water (2 mL) and saturated aqueous KNa tartrate (15 mL) were added dropwise and the resulting mixture was stirred for 3 h. The organic layer was washed with saturated aqueous K,Na tartrate (100 mL). The aqueous layer was extracted with DCM (3×100 mL) and the combined organic layers were dried (MgSO₄). The solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:10 to 1:4) to afford amide 10 (1.234 g, 2.091 mmol, 93%) as a yellow oil. $R_f = 0.29$ (EtOAc/petroleum ether 1:15, ninhydrin, UV). $[\alpha]_{D}^{22} = -16.87^{\circ}$ (CH₂Cl₂, c = 1.00 g/100 mL). IR (thin film): 2953 (s), 2927 (s), 2853 (m), 1727 (w), 1637 (s), 1600 (m), 1497 (s), 1469 (m), 1415 (s), 1245 (s), 1088 (m), 1038 (w), 1004 (m), 964 (w), 914 (m), 836 (s), 775 (s), 752 (vs), 691 (s), 617 (m). ¹H NMR (CDCl₃, 400 MHz): δ [ppm] = 7.41–7.16 (m, 7H, *H*-17, *H*-22, *H*-23, *H*-24), 7.00– 6.83 (m, 3H, H-16, H-18), 6.59 (d, ³J=16.1 Hz, 0.2H, H-20B), 6.49 (d, ³J=16.0 Hz, 0.8H, H-20A), 6.26 (dd, ³J=16.0 Hz, ³J=9.2 Hz, 0.8H, H-19A), 6.22 (dd, ${}^{3}J=16.1$ Hz, ${}^{3}J=9.3$ Hz, 0.2H, H-19B), 5.91–5.78 (m, 1H, H-25), 5.30-5.20 (m, 0.4H, H-26B), 5.21-5.10 (m, 1.6H, H-26A), 4.42-4.33 (m, 1H, H-10), 4.22 (dd, ²J=9.7 Hz, ³J=3.2 Hz, 0.8H, H-14Aa), 4.02 (dd, ²J=9.0 Hz, ³J=3.8 Hz, 0.2H, H-14Ba), 3.89 (dd, ²J= 9.7 Hz, ³J=7.6 Hz, 0.8H, H-14Ab), 3.86-3.79 (m, 1H, H-4), 3.78-3.70 (m, 0.2H, H-14Bb), 3.70-3.62 (m, 1H, H-2), 3.61-3.43 (m, 2H, H-13), 2.85-2.75 (m, 0.2H, H-3B), 2.71-2.63 (m, 0.8H, H-3A), 2.12-2.04 (m, 2H, H-11a, H-12a), 1.98-1.88 (m, 2H, H-11b, H-12b), 1.54-1.43 (m, 2H, H-5a, H-8), 1.43-1.32 (m, 1H, H-5b), 1.30-1.16 (m, 1H, H-6a), 1.15–1.04 (m, 3H, H-6b, H-7), 0.96 (s, 7.2H, H-29A), 0.83 (d, ${}^{3}J=6.6$ Hz, 6H, H-9, H-9'), 0.74 (s, 1.8H, H-29B), 0.06 (s, 2.4H, H-27A), 0.05 (s, 2.4H, H-27A'), -0.02 (s, 1.2H, H-27B, H-27B'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm] = 172.5 [171.6] (C-1), 158.8 [158.6] (C-15), 137.0 [136.9] (C-21), 134.7 [134.7] (C-25), 132.8 [133.0] (C-20), 129.5 [129.6] (C-17), 128.7 [128.7] (C-23), 128.4 [128.2] (C-19), 127.6 [127.7] (C-24), 126.5 [126.4] (C-22), 120.7 [121.2] (C-18), 119.1 [119.6] (C-26), 114.8 [114.4] (C-16), 71.6 [71.7] (C-4), 67.1 [68.5] (C-14), 56.0 [56.5] (C-10), 50.5 [50.3] (C-3), 50.1 [49.7] (C-2), 47.5 [46.2] (C-13), 39.2 [39.3] (C-7), 36.1 [36.2] (C-5), 28.0 [28.0] (C-8), 27.6 [29.0] (C-11), 26.3 [26.0] (C-29), 24.2 [21.8] (C-12), 23.1 [23.1] (C-6), 22.8 [22.9] (C-9), 22.6 (C-9'), 18.4 [18.2] (C-28), -3.0 [-3.1] (C-27), -4.3 [-4.5] (C-27'). HRMS (ESI): $m/z [M+H]^+$ calcd for $C_{37}H_{56}NO_3Si^+$: 590.4024; found: 590.4003. Because of the slowly reconverting amide rotamers, ¹H and ¹³C NMR spectra display a double set of signals (ratio 4:1). The absolute configuration of new stereogenic centers was elucidated via X-ray analysis of iodolactones.

lodolactonization: lodine (3.607 g, 14.231 mmol, 3 eq) was added to amide 14 (2.789 g, 4.727 mmol, 1.0 eq) in dimethoxy ethane (40 mL) and water (10 mL). The reaction mixture was stirred at room temperature for 21 h with exclusion of light. DCM (100 mL) and an aqueous $Na_2S_2O_3$ (30 mL, 10%) were added. The layers were separated and the aqueous layer was extracted with DCM (3×

100 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:30) to afford the iodolactones *ent*- β -11 and *ent*- α -11 (2.391 g, 4.296 mmol, 91%) as a yellow oil, 1:0.85 mixture of diastereomers. For characterization, an analytical amount was separated via column chromatography (EtOAc/petroleum ether 1:30).

Analytical data of (3*R*,4*R*,5*S*)-4-((*S*)-1-(*tert*-butyldimethylsilyloxy)-5methylhexyl)-5-(iodo-methyl)-3-((*E*)-phenethenyl)-dihydrofuran-

2(3H)-one ent- α -11: R_f=0.18 (EtOAc/petroleum ether 1:30, cerium reagent, UV). HPLC: $t_0 = 1.28 \text{ min}$, k = 4.50 (Nucleosil 50–5; EtOAc/ hexane 5:95, 2 mL/min, 102 bar), $t_0 = 1.60$ min, k = 15.60 (S,S-Whelk-O1; EtOAc/hexane 8:92, 2 mL/min, 50 bar. $[a]_{D}^{29} = +124.0^{\circ}$ (CH₂Cl₂, c=0.99 g/100 mL). IR (thin film): 2952 (m), 2931 (m), 2856 (w), 1779 (vs), 1496 (w), 1469 (m), 1385 (w), 1327 (w), 1254 (m), 1169 (w), 1132 (m), 1079 (s), 1006 (m), 962 (m), 888 (w), 835 (s), 805 (m), 775 (s), 745 (s), 692 (s), 614 (w), 602 (w), 586 (w). ¹H NMR (CDCl₃, 400 MHz): δ [ppm] = 7.38–7.30 (m, 4H, H-15, H-16), 7.29–7.24 (m, 1H, H-17), 6.61 (d, ${}^{3}J = 15.8$ Hz, 1H, H-13), 6.00 (dd, ${}^{3}J = 15.9$ Hz, ³J=8.0 Hz, 1H, H-12), 4.87–4.77 (m, 1H, H-5), 4.01–3.92 (m, 1H, H-6), 3.66–3.52 (m, 2H, H-3, H-18a), 3.31 (dd, ²J=11.3 Hz, ³J=9.7 Hz, 1H, H-18b), 2.75-2.65 (m, 1H, H-4), 1.65-1.43 (m, 3H, H-7, H-10), 1.36-1.22 (m, 2H, H-8), 1.20-1.04 (m, 2H, H-9), 0.91 (s, 9H, H-21), 0.85 (d, ³J=6.6 Hz, 3H, H-11), 0.84 (d, ³J=6.6 Hz, 3H, H-11'), 0.11 (s, 3H, H-19), 0.10 (s, 3H, H-19'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm] = 175.7 (C-2), 136.1 (C-14), 135.8 (C-13), 128.8 (C-15), 128.3 (C-17), 126.6 (C-16), 122.9 (C-12), 80.3 (C-5), 69.8 (C-6), 49.7 (C-4), 44.4 (C-3), 39.0 (C-9), 35.5 (C-7), 28.1 (C-10), 26.0 (C-21), 23.3 (C-8), 22.7 (C-11), 22.6 (C-11'), 18.1 (C-20), 4.3 (C-18), -3.7 (C-19), -4.4 (C-19'). HRMS (ESI): m/z $[M+H]^+$ calcd for C₂₆H₄₁INaO₃Si⁺: 579.1759; found: 579.1762.

Analytical data of (3*R*,4*R*,5*R*)-4-((*S*)-1-(*tert*-butyldimethylsilyloxy)-5methylhexyl)-5-(iodomethyl)-3-((*E*)-phenethenyl)-dihydrofuran-

2(3H)-one ent-β-11: R_f=0.26 (EtOAc/petroleum ether 1:30, cerium reagent, UV). HPLC: $t_0 = 1.70$ min, k = 3.25 (Nucleosil 50–5; EtOAc/ hexane 5:95, 2 mL/min, 102 bar), $t_0 = 1.49$ min, k = 20.42 (S,S-Whelk-O1; EtOAc/hexane 1:9, 2 mL/min, 50 bar). $[a]_{D}^{29} = +19.0^{\circ}$ (CH₂Cl₂, c=1.00 g/100 mL). IR (thin film): 2952 (s), 2926 (s), 2854 (m), 1778 (vs), 1496 (w), 1463 (m), 1412 (w), 1362 (w), 1256 (m), 1163 (s), 1063 (m), 963 (m), 901 (w), 835 (s), 805 (m), 775 (s), 745 (s), 692 (s), 624 (w), 613 (w), 587 (w). ¹H NMR (CDCl₃, 400 MHz): δ [ppm]=7.41-7.36 (m, 2H, H-16), 7.36-7.30 (m, 2H, H-15), 7.31-7.22 (m, 1H, H-17), 6.59 (d, ${}^{3}J = 15.8$ Hz, 1H, H-13), 6.17 (dd, ${}^{3}J = 15.8$ Hz, ³J=8.1 Hz, 1H, H-12), 4.37–4.30 (m, 1H, H-5), 3.81–3.77 (m, 1H, H-6), 3.65 (dd, ²J=11.1 Hz, ³J=4.0 Hz, 1H, H-18a), 3.42-3.33 (m, 2H, H-3, H-18b), 2.54-2.46 (m, 1H, H-4), 1.64-1.27 (m, 5H, H-15, H-16, H-17), 1.16-1.07 (m, 2H, H-9), 0.91 (s, 9H, H-21), 0.84 (d, ³J=6.6 Hz, 3H, H-11), 0.83 (d, ³J=6.6 Hz, 3H, H-11'), 0.13 (s, 3H, H-19), 0.11 (s, 3H, H-19'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm] = 175.3 (C-2), 136.2 (C-14), 135.1 (C-13), 128.7 (C-15), 128.2 (C-17), 126.7 (C-16), 123.9 (C-12), 77.3 (C-5), 72.2 (C-6), 51.0 (C-4), 47.5 (C-3), 39.1 (C-9), 35.6 (C-7), 28.0 (C-10), 26.0 (C-21), 22.7 (C-11), 22.6 (C-11'), 22.6 (C-8), 18.2 (C-20), 11.1 (C-18), -3.8 (C-19), -4.2 (C-19'). HRMS (ESI): m/z [M+H]⁺ calcd for C₂₆H₄₁INaO₃Si⁺: 579.1762; found: 579.1745.

(2*R*,3*S*,4*S*)-4-((*tert*-Butyldimethylsilyloxy)-8-methyl-2-((*E*)-styryl)-3vinyl-nonanoic acid: Zinc powder (1.658 g) and acetic acid (10 drops) were added to iodolactone *ent*-11 (43.3 mg, 0.078 mmol, 1.0 eq, mixture of diastereomers) in Et₂O (5 mL). The reaction mixture was stirred for 2 d. After filtration through a silica gel pad and careful rinsing with DCM, the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/ petroleum ether 1:20 to 1:10+0.1% AcOH) to afford the acid (28.5 mg, 66.2 µmol, 85%) as yellow oil. R_f=0.25 (EtOAc/petroleum ether 1:10+0,1% AcOH, ninhydrin, UV). For data see supporting information.

0990690,

Methyl (2R,3S,4S)-4-((tert-butyldimethylsilyloxy)-8-methyl-2-((E)-2-phenylethenyl)-3-vinyl-nonanoate ent-12: N-Methyl-N-nitroso urea (139.6 mg, 1.354 mmol, 20.5 eq) was dissolved in Et₂O (20 mL) and aqueous KOH (20 mL, 40%) was added in a beaker at 0°C. After stirring for 10 min the bright yellow color of diazomethane within the organic layer occurred. In a second beaker the crude acid (28.5 mg, 0.066 mmol, 1.0 eq) in Et₂O (5 mL) was treated with diazomethane in ether until the yellow color of surplus diazomethane remained. Stirring was continued at $0\,^{\circ}\text{C}$ for 5 h. Then acetic acid (5 mL, 2 M) in Et₂O was added dropwise to quench the excess of diazomethane and stirring was continued for further 30 min. The reaction mixture was washed with saturated aqueous NaHCO₃ (50 mL) and the aqueous layer was extracted with Et_2O (3 × 30 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:50) to afford ester ent-12 (25.8 mg, 0.058 mmol, 88%) as a colorless oil. R_f=0.25 (EtOAc/petroleum ether 1:50, cerium reagent, UV). $[\alpha]_{D}^{23} = +28.4^{\circ}$ (CH₂Cl₂, c=1.01 g/100 mL). IR (thin film): 2954 (s), 2931 (s), 2858 (m), 1740 (vs), 1471 (m), 1434 (m), 1363 (w), 1256 (s), 1155 (s), 1090 (s), 1002 (w), 969 (m), 919 (m), 836 (vs), 774 (s), 751 (m), 690 (m), 605 (w). ¹H NMR (CDCl₃, 400 MHz): δ [ppm]=7.39-7.29 (m, 4H, H-13, H-14), 7.27–7.22 (m, 1H, H-15), 6.56 (d, ³J=15.9 Hz, 1H, H-11), 6.12 (dd, ³J=15.9 Hz, ³J=9.6 Hz, 1H, H-10), 5.85 (ddd, ³J=17.3 Hz, ³J=10.3 Hz, ³J=10.1 Hz 1H, H-16), 5.13 (dd, ³J=10.3 Hz, ²J=2.2 Hz, 1H, H-17a), 5.07 (dd, ³J=17.3 Hz, ²J=2.2 Hz, 1H, H-17b), 3.76 (ddd, ${}^{3}J = 9.5$ Hz, ${}^{3}J = 4.6$ Hz, ${}^{3}J = 1.7$ Hz, 1H, H-4), 3.62 (s, 3H, H-21), 3.49– 3.40 (m, 1H, H-2), 2.49 (ddd, ${}^{3}J = 11.0$ Hz, ${}^{3}J = 10.1$ Hz, ${}^{3}J = 1.7$ Hz, 1H, H-3), 1.55-1.47 (m, 1H, H-8), 1.49-1.25 (m, 2H, H-5), 1.25-1.16 (m, 1H, H-6a), 1.19-1.05 (m, 3H, H-6b, H-7), 0.93 (s, 9H, H-20), 0.85 (d, ³J=6.6 Hz, 6H, H-9, H-9'), 0.03 (s, 3H, H-18), 0.02 (s, 3H, H-18'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm] = 173.8 (C-1), 136.8 (C-12), 135.3 (C-16), 134.0 (C-11), 128.7 (C-14), 127.8 (C-15), 126.5 (C-13), 126.4 (C-10), 118.4 (C-17), 71.2 (C-4), 52.1 (C-2), 51.7 (C-21), 50.8 (C-3), 39.2 (C-7), 36.0 (C-5), 28.0 (C-8), 26.2 (C-20), 23.1 (C-6), 22.9 (C-9), 22.6 (C-9'), 18.4 (C-19), -3.2 (C-18), -4.2 (C-18'). HRMS (ESI): m/z [M+H]⁺ calcd for C₂₇H₄₅O₃Si⁺: 445.3132; found: 445.3135.

(2R,3S,4S)-4-((tert-Butyldimethylsilyloxy)-8-methyl-2-((E)-2-phe-

nylethenyl)-3-vinylnonan-1-ol ent-15: DIBAL-H (0.82 mL 0.82 mmol, 2.7 eq, 1.0 m in toluene) was added dropwise to ester ent-12 (133.1 mg, 0.299 mmol, 1.0 eq) in DCM (2 mL) at -78 °C. After stirring at this temperature for 3 h, the reaction mixture was warmed to 0 °C. Saturated aqueous KNa tartrate (15 mL) was added dropwise and the resulting mixture was stirred for 1.5 h at room temperature. The layers were separated and the aqueous phase was extracted with DCM (5×10 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:20) to afford alcohol ent-15 (106.0 mg, 0.254 mmol, 85%) as a colorless oil. $R_f = 0.29$ (EtOAc/petroleum ether 1:10, vanillin, UV). $[\alpha]_{D}^{25} = +0.6^{\circ}$ (CH₂Cl₂, c = 0.99 g/100 mL). IR (thin film): 3393 (br, m), 2954 (s), 2929 (s), 2857 (m), 1638 (w), 1494 (m), 1471 (m), 1386 (m), 1365 (m), 1254 (m), 1089 (s), 1057 (s), 1004 (m), 970 (m), 913 (m), 834 (vs), 807 (m), 773 (s), 748 (s), 692 (s), 665 (m), 614 (w), 601 (w), 592 (w). ¹H NMR (CDCl₃, 400 MHz): δ [ppm]=7.40–7.30 (m, 4H, H-13, H-14), 7.27–7.22 (m, 1H, H-15), 6.55 (d, ³J=16.0 Hz, 1H, H-11), 6.00 (dd, ${}^{3}J = 16.0$ Hz, ${}^{3}J = 9.3$ Hz, 1H, H-10), 5.85 (ddd, ${}^{3}J = 17.4$ Hz, ${}^{3}J = 10.1$ Hz, ${}^{3}J = 10.1$ Hz, 1H, H-16), 5.19 (dd, ${}^{3}J = 10.1$ Hz, ${}^{2}J = 2.2$ Hz, 1H, H-17a), 5.08 (dd, ${}^{3}J=17.4$ Hz, ${}^{2}J=2.2$ Hz, 1H, H-17b), 3.77 (ddd, ³*J*=9.4 Hz, ³*J*=4.7 Hz, ³*J*=1.6 Hz, 1H, *H*-4), 3.74–3.68 (m, 1H, H-1a), 3.45 (dd, ²J=11.0 Hz, ³J=8.0 Hz, 1H, H-1b), 2.69-2.58 (m, 1H, H-2), 2.14 (ddd, ³J=10.2 Hz, ³J=10.1 Hz, ³J=1.6 Hz, 1H, H-3), 1.60 (s, 1H, H-21), 1.56-1.43 (m, 2H, H-5a, H-8), 1.40-1.31 (m, 1H, H-5b), 1.29-1.16 (m, 1H, H-6a), 1.15-1.06 (m, 3H, H-6b, H-7), 0.91 (s, 9H, H-20), 0.85 (d, ³J=6.6 Hz, 6H, H-9, H-9'), 0.03 (s, 3H, H-18), 0.01 (s, 3H, *H*-18'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm]=137.1 (C-12), 136.9 (C- 16), 133.8 (C-11), 130.9 (C-10), 128.7 (C-14), 127.6 (C-15), 126.3 (C-13), 117.8 (C-17), 72.8 (C-4), 65.1 (C-1), 50.2 (C-3), 46.3 (C-2), 39.2 (C-7), 35.9 (C-5), 28.0 (C-8), 26.2 (C-20), 23.3 (C-6), 22.9 (C-9), 22.6 (C-9'), 18.4 (C-19), -3.3 (C-18), -4.2 (C-18'). HRMS (ESI): m/z [M+H]⁺ calcd for $C_{26}H_{45}O_2Si^+$: 417.3184; found: 417.3184.

lodocyclization: lodine (997.4 mg, 3.93 mmol, 3.03 eq) was added to alcohol *ent*-**15** (540.9 mg, 1.298 mmol, 1.0 eq) in Et₂O (10 mL) and saturated aqueous NaHCO₃ (18 mL). The reaction mixture was stirred at room temperature for 9 h with exclusion of light. Et₂O (20 mL) and an aqueous Na₂S₂O₃ (20 mL, 10%) were added. The layers were separated and the aqueous layer was extracted with Et₂O (3×20 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was filtered through a silica gel pad (careful rinsing with DCM). The solvent was removed in vacuo and the residue was purified via HPLC (Gemini NX C18, MeCN, 40 mL/min, 37 bar) to afford iodoether *ent*-**21** (334.9 mg, 0.617 mmol, 48%) and iodoether *ent*-**20** (250.5 mg, 0.462 mmol, 36%) as colorless oils. For data of minor *cis* alkene *ent*-**21 a** see supporting information.

Analytical data of (2R,3R,4R)-3-(((1S)-tert-butyldimethylsilyloxy)-5methylhexyl)-2-(iodomethyl)-4-((E)-styryl)-tetrahydrofuran ent-21: $R_f = 0.19$ (EtOAc/ petroleum ether 1:100, cerium reagent, UV). HPLC: $t_0 = 1.33 \text{ min}, k = 2.09$ (Gemini NX C18, MeCN, 2 mL/min, 73 bar). $[\alpha]_{\mathsf{D}}^{2}$ $c^{3} = +12.2^{\circ}$ (CH₂Cl₂, c = 0.99 g/100 mL). IR (thin film): 3027 (w), 2954 (s), 2929 (s), 2899 (m), 2857 (s), 1740 (m), 1600 (w), 1493 (w), 1471 (m), 1411 (w), 1385 (w), 1364 (w), 1255 (s), 1075 (s), 1051 (s), 1006 (w), 967 (s), 938 (w), 835 (vs), 807 (m), 774 (vs), 747 (s), 693 (s), 615 (w), 597 (w), 585 (w). $^1\mathrm{H}$ NMR (CDCl_3, 400 MHz): δ [ppm] =7.38– 7.26 (m, 4H, H-15, H-16), 7.27–7.18 (m, 1H, H-17), 6.42 (d, ³J=15.8 Hz, 1H, H-13), 6.13 (dd, ${}^{3}J = 15.8$ Hz, ${}^{3}J = 9.1$ Hz, 1H, H-12), 3.99 (dd, ²J=8.8 Hz, ³J=7.5 Hz, 1H, H-5a), 3.94 (ddd, ³J=6.6 Hz, ³J=6.4 Hz, ³J=3.1 Hz, 1H, H-2), 3.85–3.75 (m, 2H, H-5b, H-6), 3.56 (dd, ²J=10.5 Hz, ³J=3.1 Hz, 1H, H-21a), 3.32 (dd, ²J=10.5 Hz, ³J=6.4 Hz, 1H, H-21b), 3.00-2.88 (m, 1H, H-4), 2.08-1.99 (m, 1H, H-3), 1.63-1.42 (m, 3H, H-7, H-10), 1.42-1.16 (m, 2H, H-8), 1.13-1.01 (m, 2H, H-9), 0.91 (s, 9H, H-20), 0.81 (d, ³J=6.6 Hz, 3H, H-11), 0.79 (d, ³J=6.6 Hz, 3H, H-11'), 0.11 (s, 3H, H-18), 0.08 (s, 3H, H-18'). ¹H NMR (C_6D_6 , 600 MHz): δ [ppm] = 7.26-7.22 (m, 2H, H-16), 7.16-7.10 (m, 2H, H-15), 7.08-7.02 (m, 1H, H-17), 6.34 (d, ${}^{3}J = 15.8$ Hz, 1H, H-13), 6.06 (dd, ${}^{3}J = 15.8$ Hz, ³J=9.2 Hz, 1H, H-12), 4.00–3.94 (m, 1H, H-2), 3.89 (dd, ²J=8.7 Hz, ³J=7.5 Hz, 1H, H-5a), 3.78-3.73 (m, 1H, H-5b), 3.73-3.67 (m, 1H, H-6), 3.47 (dd, ²J=10.4 Hz, ³J=3.4 Hz, 1H, H-21a), 3.18 (dd, ²J=10.4 Hz, ³J=5.9 Hz, 1H, H-21b), 2.80–2.72 (m, 1H, H-4), 2.08 (ddd, ³J=7.9 Hz, ³J=6.3 Hz, ³J=6.3 Hz, 1H, H-3), 1.55-1.45 (m, 1H, H-7a), 1.47-1.39 (m, 1H, H-10), 1.41-1.31 (m, 2H, H-7b, H-8a), 1.33-1.23 (m, 1H, H-8b), 1.10-1.01 (m, 2H, H-9), 0.97 (s, 9H, H-20), 0.83 (d, ³J=6.6 Hz, 3H, H-11), 0.81 (d, ³J=6.6 Hz, 3H, H-11'), 0.10 (s, 3H, H-18), 0.06 (s, 3H, H-18').¹³C NMR (CDCl₃, 101 MHz): δ [ppm]=137.1 (C-14), 131.6 (C-13), 129.9 (C-12), 128.7 (C-15), 127.6 (C-17), 126.3 (C-16), 80.9 (C-2), 72.8 (C-6), 72.8 (C-5), 55.3 (C-3), 47.9 (C-4), 39.2 (C-9), 36.0 (C-7), 28.1 (C-10), 26.1 (C-20), 22.7 (C-11), 22.7 (C-11'), 22.0 (C-8), 18.2 (C-19), 13.0 (C-21), -3.9 (C-18), -4.1 (C-18'). ¹³C NMR (C₆D₆, 151 MHz): δ [ppm] = 137.4 (C-14), 131.9 (C-13), 130.3 (C-12), 128.9 (C-16), 127.7(C-17), 126.5 (C-15), 81.2 (C-2), 73.4 (C-6), 72.8 (C-5), 55.7 (C-3), 48.1 (C-4), 39.5 (C-9), 35.9 (C-7), 28.3 (C-10), 26.2 (C-20), 22.8 (C-11), 22.7 (C-11'), 22.2 (C-8), 18.3 (C-19), 13.1 (C-21), -3.9 (C-18), -4.1 (C-18'). HRMS (ESI): m/z $[M + Na]^+$ calcd for $C_{26}H_{43}INaO_2Si^+$: 565.1969; found: 565.1970.

Analytical data of (2*R*,3*S*,4*R*)-4-(((2*S*)-*tert*-butyldimethylsilyloxy)-(1*S*)-ethenyl-6-methylheptyl)-3-iodo-2-phenyl-tetrahydrofuran *ent*-20: R_{*i*}=0.15 (EtOAc/ petroleum ether 1:100, cerium reagent, UV). HPLC: $t_0 = 1.33$ min, k = 3.01 (Gemini NX C18, MeCN, 2 mL/min, 73 bar). $[\alpha]_D^{23} = -43.3^{\circ}$ (CH₂Cl₂, c = 0.98 g/100 mL). IR (thin film): 3068 (w), 2954 (s), 2929 (s), 2895 (m), 2860 (s), 1741 (m), 1636 (w), 1492 (w), 1471 (m), 1382 (w), 1365 (w), 1255 (s), 1086 (s), 1050 (s),



1027 (s), 1005 (m), 922 (m), 874 (w), 836 (vs), 807 (m), 774 (vs), 759 (s), 733 (w), 698 (s), 61w (w), 597 (w), 586 (w). ¹H NMR (CDCl₃, 400 MHz): δ [ppm]=7.43-7.39 (m, 2H, H-15), 7.37-7.30 (m, 3H, H-14, H-16), 5.79 (ddd, ${}^{3}J=17.2$ Hz, ${}^{3}J=10.2$ Hz, ${}^{3}J=9.9$ Hz, 1H, H-17), 5.26 (dd, ³*J*=10.2 Hz, ²*J*=2.1 Hz, 1H, *H*-18a), 5.14 (dd, ³*J*=17.2 Hz, ²J=2.1 Hz, 1H, H-18b), 4.96 (d, ³J=9.0 Hz, 1H, H-2), 4.07 (dd, ²J=8.8 Hz, ³J=8.7 Hz, 1H, H-5a), 4.00 (dd, ²J=8.8 Hz, ³J=7.5 Hz, 1H, H-5b), 3.78-3.67 (m, 2H, H-3, H-7), 2.81-2.71 (m, 1H, H-4), 2.55 (ddd, ³J= 9.9 Hz, ³J=3.8 Hz, ³J=3.8 Hz, 1H, H-6), 1.59-1.52 (m, 1H, H-11), 1.51-1.42 (m, 1H, H-8a), 1.43-1.30 (m, 1H, H-8b), 1.29-1.18 (m, 2H, *H*-9), 1.18–1.10 (m, 2H, *H*-10), 0.91 (s, 9H, *H*-21), 0.88 (d, ${}^{3}J$ =6.6 Hz, 3H, H-12), 0.88 (d, ${}^{3}J$ = 6.6 Hz, 3H, H-12'), 0.11 (s, 3H, H-19), 0.08 (s, 3H, H-19'). ¹H NMR ($C_6 D_{67}$ 600 MHz): δ [ppm] = 7.58–7.54 (m, 2H, H-14), 7.19-7.15 (m, 2H, H-15), 7.11-7.08 (m, 1H, H-16), 5.74-5.66 (m, 1H, H-17), 5.07 (d, ³J=8.8 Hz, 1H, H-2), 5.06–4.99 (m, 2H, H-18), 4.05-3.98 (m, 2H, H-5), 3.89 (dd, ³J=8.8 Hz, ³J=8.8 Hz, 1H, H-3), 3.68 (ddd, ³*J*=6.1 Hz, ³*J*=6.1 Hz, ³*J*=3.8 Hz, 1H, *H*-7), 2.86–2.80 (m, 1H, H-4), 2.63 (ddd, ³J=9.8 Hz, ³J=4.0 Hz, ³J=3.8 Hz, 1H, H-6), 1.54– 1.44 (m, 2H, H-8a, H-11), 1.41-1.20 (m, 3H, H-8b, H-9), 1.17-1.10 (m, 2H, H-10), 1.00 (s, 9H, H-21), 0.90 (d, ³J=6.7 Hz, 3H, H-12), 0.90 (d, ³J=6.7 Hz, 3H, H-12'), 0.14 (s, 3H, H-19), 0.08 (s, 3H, H-19'). ¹³C NMR (CDCl₃, 101 MHz): δ [ppm] = 138.9 (C-13), 133.9 (C-17), 128.6 (C-14), 128.5 (C-16), 126.9 (C-15), 120.1 (C-18), 88.0 (C-2), 75.0 (C-7), 69.0 (C-5), 51.8 (C-4), 48.6 (C-6), 39.2 (C-10), 35.4 (C-8), 34.8 (C-3), 28.0 (C-11), 26.2 (C-21), 23.1 (C-9), 22.8 (C-12), 22.7 (C-12'), 18.3 (C-20), -3.8 (C-19), -3.8 (C-19'). ¹³C NMR (C₆D₆, 151 MHz): δ [ppm] = 139.6 (C-13), 134.2 (C-17), 128.6 (C-15), 128.3 (C-16), 126.9 (C-14), 119.7 (C-18), 88.2 (C-2), 74.9 (C-7), 68.7 (C-5), 51.8 (C-4), 49.2 (C-6), 39.2 (C-10), 35.4 (C-8), 35.3 (C-3), 28.1 (C-11), 26.1 (C-21), 23.2 (C-9), 22.7 (C-12), 22.6 (C-12'), 18.2 (C-20), -3.9 (C-19), -4.0 (C-19'). HRMS (ESI): m/z $[M+H]^+$ calcd for C₂₆H₄₄IO₃Si⁺: 543.2150; found: 543.2156.

((2R,3R,4S)-3-((1S)-((tert-Butyldimethylsilyloxy)-5-methylhexyl)-4-(hydroxymethyl)-2-(iodomethyl)-tetrahydrofuran: Alkene ent-21 (85.1 mg, 0.157 mmol, 1.0 eq) in DCM (10 mL) was cooled to -78°C. Oxygen was bubbled through the solution for 10 min. Then, ozone was bubbled through the solution for 1 h until the blue color of unreacted ozone occurred. The ozone was displaced by oxygen (10 min) and argon (15 min). Then, sodium borohydride (68.6 mg, 1.81 mmol, 11.6 eq) in MeOH (4 mL, cooled to -78 °C) was added dropwise. The mixture was stirred for 21 h, the temperature was allowed to warm-up to 23. After washing with semi-saturated aqueous NH₄Cl (20 mL), the aqueous layer was extracted with DCM (4×10 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:6) to afford the ent-alcohol (72.8 mg, 0.155 mmol, 99%) as a colorless oil. $R_f = 0.25$ (EtOAc/petroleum ether 1:5, vanillin, UV). For analytical data see supporting information.

(2R,3R,4R)-3-((1S)-1-(tert-Butyldimethylsilyloxy)-5-methylhexyl)-4tert-butyldimethylsilyloxymethyl-1-(iodomethyl)-tetrahydrofuran ent-27: Triethylamine (0.12 mL, 0.87 mmol, 7.4 eq) and TBSOTf (0.14 mL, 0.61 mmol, 5.2 eq) were added dropwise to ent-alcohol (55.3 mg, 0.118 mmol, 1.0 eq) in DCM (3 mL) at 0°C. The mixture was stirred at room temperature for 20 h. After dilution with DCM (10 mL) and washing with saturated aqueous NaHCO₃ (8 mL), the resulting aqueous layer was extracted with DCM (4×6 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:100) to afford silyl ether ent-27 (53.8 mg, 0.092 mmol, 78%) as a colorless oil. R_f=0.20 (EtOAc/petroleum ether 1:50, cerium reagent). $[\alpha]_D^{23} = -1.2^\circ$ (CH₂Cl₂, c = 1.00 g/100 mL). IR (thin film): 2954 (s), 2929 (s), 2857 (s), 1471 (m), 1411 (w), 1386 (m), 1362 (m), 1255 (s), 1176 (w), 1093 (s), 1073 (s), 1006 (m), 938 (w), 915 (w), 835 (vs), 774 (vs), 726 (w), 666 (m), 630 (w), 617 (w), 607 (w), 588 (w). ¹H NMR ($C_6 D_6$, 400 MHz): δ [ppm] = 3.89 (dd, ${}^{2}J$ = 8.9 Hz, ${}^{3}J$ = 4.9 Hz, 1H, H-5a), 3.86–3.77 (m, 2H, H-2, H-5b), 3.75–3.69 (m, 1H, H-6), 3.64–3.58 (m, 2H, H-12), 3.42 (dd, ${}^{2}J$ = 10.2 Hz, ${}^{3}J$ = 4.6 Hz, 1H, H-19a), 3.14 (dd, ${}^{2}J$ = 10.2 Hz, ${}^{3}J$ = 5.3 Hz, 1H, H-19b), 2.36–2.26 (m, 1H, H-4), 2.01–1.94 (m, 1H, H-3), 1.61–1.48 (m, 1H, H-10), 1.48–1.32 (m, 4H, H-7, H-8), 1.19–1.10 (m, 2H, H-9), 0.97 (s, 9H, H-18), 0.96 (s, 9H, H-15), 0.92 (d, ${}^{3}J$ = 6.6 Hz, 6H, H-11, H-11'), 0.11 (s, 3H, H-16), 0.08 (s, 3H, H-16'), 0.07 (s, 6H, H-13, H-13'). ¹³C NMR (C₆D₆, 101 MHz): δ [ppm] = 80.4 (C-2), 74.0 (C-6), 70.3 (C-5), 64.9 (C-12), 52.1 (C-3), 46.0 (C-4), 39.6 (C-9), 35.6 (C-7), 28.4 (C-10), 26.2 (C-15), 26.2 (C-18), 23.3 (C-8), 22.9 (C-11), 22.8 (C-11'), 18.5 (C-17), 18.3 (C-14), 12.2 (C-19), -3.9 (C-16), -4.0 (C-16'), -5.2 (C-13), -5.2 (C-13'). HRMS (ESI): m/z [M+H]⁺ calcd for C₂₅H₅₄IO₃Si₂⁺: 585.2651; found: 585.2654.

(3S,4R)-3-((1S)-tert-butyldimethylsilyloxy-5-methylhexyl)-4-(tertbutyldimethylsilyloxymethyl)-2-(methylene)-tetrahydrofuran ent-29: Potassium tert-butoxide (50.0 mg, 0.446 mmol, 14.6 eq) was added to iodoether ent-27 (17.9 mg, 30.6 µmol, 1.0 eq) in THF (10 mL). The reaction mixture was heated for 3 d to 50 °C. Then, the solvent was removed in vacuo. The residue was dissolved in Et₂O (10 mL) and washed with water (10 mL). The aqueous layer was extracted with Et_2O (3×10 mL). The combined organic layers were dried (MgSO₄) and the solvent was removed in vacuo to afford olefine ent-29 (13.7 mg, 30.0 μ mol, 98%) as a yellow oil. R_f=0.19 (EtOAc/petroleum ether 1:20, cerium reagent). $[\alpha]_{D}^{25} = -1.4^{\circ}$ (CH₂Cl₂, c=1.00 g/100 mL). IR (thin film): 2954 (s), 2929 (s), 2896 (m), 2857 (m), 1732 (w), 1667 (m), 1602 (w), 1471 (m), 1386 (m), 1363 (m), 1256 (s), 1092 (s), 1066 (s), 1006 (m), 939 (w), 889 (w), 837 (vs), 798 (m), 775 (vs), 713 (w), 666 (w), 625 (w), 613 (w), 605 (w), 594 (w). ¹H NMR ($C_6 D_{6t}$ 400 MHz): δ [ppm] = 4.63 (dd, ⁴J = 1.6 Hz, ²J = 1.5 Hz, 1H, H-19a), 4.15 (dd, ²J=8.8 Hz, ³J=6.9 Hz, 1H, H-5a), 4.07 (dd, ^{2}J =8.8 Hz, ^{3}J =4.2 Hz, 1H, H-5b), 3.90 (dd, ^{2}J =1.5 Hz, ^{4}J =1.4 Hz, 1H, H-19b), 3.89–3.85 (m, 1H, H-6), 3.71 (dd, ²J=9.8 Hz, ³J=5.1 Hz, 1H, H-12a), 3.45 (dd, ²J=9.8 Hz, ³J=8.8 Hz, 1H, H-12b), 2.73–2.69 (m, 1H, H-3), 2.68-2.59 (m, 1H, H-4), 1.68-1.60 (m, 1H, H-7a), 1.60-1.54 (m, 1H, H-8a), 1.54-1.46 (m, 1H, H-10), 1.45-1.34 (m, 1H, H-7b), 1.33-1.24 (m, 1H, H-8b), 1.24-1.05 (m, 2H, H-9), 0.99 (s, 9H, H-18), 0.94 (s, 9H, H-15), 0.90 (d, ³J=6.7 Hz, 3H, H-11), 0.90 (d, ³J=6.7 Hz, 3H, H-11'), 0.09 (s, 3H, H-16), 0.07 (s, 3H, H-16'), 0.04 (s, 6H, H-13, H-13'). ¹³C NMR (C₆D₆, 101 MHz): δ [ppm]=164.3 (C-2), 80.9 (C-19), 75.1 (C-6), 73.0 (C-5), 65.7 (C-12), 51.1 (C-3), 41.9 (C-4), 39.2 (C-9), 33.2 (C-7), 28.4 (C-10), 26.1 (C-18), 26.1 (C-15), 24.9 (C-8), 22.8 (C-11), 22.8 (C-11'), 18.5 (C-14), 18.3 (C-17), -4.0 (C-16), -4.5 (C-16'), -5.3 (C-13), -5.3 (C-13'). HRMS (ESI): m/z $[M+Na]^+$ calcd for C₂₅H₅₂O₃Si₂Na⁺: 479.3347; found: 479.3359.

(3S,4R)-3-((1S)-1-((tert-Butyldimethylsilyloxy)-5-methylhexyl)-4-

((tert-butyldimethylsilyloxy)-methyl)-dihydrofuran-2(3H)-one ent-30: Alkene ent-29 (13.7 mg, 30 µmol, 1.0 eq) was dissolved in DCM (10 mL) and cooled to -78 °C. Then, ozone was bubbled through the solution for 1 h until the blue color of unreacted ozone occurred. The ozone was displaced by oxygen (10 min) and argon (15 min). Dimethyl sulfide (0.33 mL, 280.5 mg, 4.73 mmol, 145.8 eq) was added dropwise and the reaction mixture was stirred for 2 h at -78°C and 1 h at room temperature. The solvent and the excess dimethyl sulfide were removed in vacuo. The residue was purified via column chromatography (EtOAc/petroleum ether 1:50) to afford lactone ent-30 (10.3 mg, 22.5 µmol, 75%) as a colorless oil. $R_{f}{=}\,0.13$ (EtOAc/petroleum ether 1:50, cerium reagent). $[\alpha]_{n}^{23}{=}\,+$ 30.5° (CH₂Cl₂, c=0.21 g/100 mL). IR (thin film): 2954 (s), 2929 (s), 2902 (m), 2857 (s), 1775 (s), 1736 (m), 1605 (w), 1514 (w), 1471 (m), 1389 (m), 1363 (m), 1255 (s), 1172 (m), 1094 (s), 1071 (s), 1006 (m), 938 (w), 837 (vs), 776 (vs), 713 (w), 667 (w), 627 (w), 598 (w). ¹H NMR (CDCl₃, 600 MHz): δ [ppm]=4.34 (dd, ²J=9.0 Hz, ³J=8.2 Hz, 1H, H-5a), 4.09 (dd, ²J=9.0 Hz, ³J=5.9 Hz, 1H, H-5b), 4.05 (ddd, ³J=8.4 Hz, ³J=5.2 Hz, ³J=3.5 Hz, 1H, *H*-6), 3.73 (dd, ²J=10.0 Hz, ³J=4.6 Hz, 1H, H-12a), 3.58 (dd, ²J=10.0 Hz, ³J=6.5 Hz, 1H, H-12b), 2.75-2.69 (m, 1H, H-4), 2.61 (dd, ³J=6.4 Hz, ³J=3.5 Hz, 1H, H-3), 1.60–1.53 (m, 1H, H-7a), 1.55-1.49 (m, 1H, H-10), 1.49-1.42 (m, 1H, H-7b), 1.42-1.33 (m, 1H, H-8a), 1.32-1.26 (m, 1H, H-8b), 1.24-1.17 (m, 1H, H-9a), 1.17-1.09 (m, 1H, H-9b), 0.88 (s, 9H, H-18), 0.88 (s, 9H, H-15), 0.86 (d, ${}^{3}J =$ 6.6 Hz, 3H, H-11), 0.86 (d, ³J=6.6 Hz, 3H, H-11'), 0.09 (s, 3H, H-16), 0.07 (s, 3H, H-16'), 0.05 (s, 3H, H-13), 0.05 (s, 3H, H-13'). ¹H NMR $(C_6D_{6'} 600 \text{ MHz}): \delta \text{ [ppm]} = 4.20 \text{ (ddd, } {}^3J = 7.9 \text{ Hz}, {}^3J = 4.8 \text{ Hz}, {}^3J = 2.8,$ 1H, H-6), 4.08–4.03 (m, 1H, H-5a), 3.77 (dd, ${}^{2}J=9.1$ Hz, ${}^{3}J=5.6$ Hz, 1H, H-5b), 3.52-3.48 (m, 1H, H-12a), 3.33-3.28 (m, 1H, H-12b), 2.59-2.52 (m, 2H, H-3, H-4), 1.77-1.69 (m, 1H, H-7a), 1.54-1.42 (m, 3H, H-7b, H-8a, H-10), 1.39–1.25 (m, 1H, H-8b), 1.21–1.14 (m, 1H, H-9a), 1.14-1.06 (m, 1H, H-9b), 0.94 (s, 9H, H-18), 0.92 (s, 9H, H-15), 0.90 (d, ³J=6.3 Hz, 3H, H-11), 0.89 (d, ³J=6.3 Hz, 3H, H-11'), 0.05 (s, 3H, H-16), 0.02 (s, 3H, H-16'), 0.00 (s, 3H, H-13), -0.01 (s, 3H, H-13'). ¹³C NMR (CDCl₃, 151 MHz): δ [ppm] = 177.0 (C-2), 72.5 (C-6), 69.9 (C-5), 64.3 (C-12), 47.8 (C-3), 39.5 (C-4), 39.0 (C-9), 34.2 (C-7), 28.1 (C-10), 25.9 (C-15, C-18), 24.2 (C-8), 22.7 (C-11), 22.6 (C-11'), 18.3 (C-14), 18.1 (C-17), -4.2 (C-16), -4.4 (C-16'), -5.4 (C-13), -5.4 (C-13'). ¹³C NMR $(C_6 D_{61} 151 \text{ MHz}): \delta \text{ [ppm]} = 175.7 (C-2), 72.8 (C-6), 69.5 (C-5), 64.8 (C-6))$ 12), 48.3 (C-3), 39.5 (C-4), 39.1 (C-9), 34.4 (C-7), 28.3 (C-10), 26.0 (C-18), 26.0 (C-15), 24.6 (C-8), 22.8 (C-11), 22.7 (C-11'), 18.4 (C-14), 18.2 (C-17), -4.3 (C-16), -4.5 (C-16'), -5.4 (C-13), -5.4 (C-13'). HRMS (ESI): $m/z [M + Na]^+$ calcd for $C_{24}H_{50}O_4Si_2Na^+$: 481.3140; found: 481.3140.

(3S,4S)-3-((S)-1-Hydroxy-5-methylhexyl)-4-(hydroxymethyl)-dihydrofuran-2(3H)-one ent-2d (+)-SCB-5: Acetic acid (16.8 mg, 0.280 mmol, 12.5 eq) and TBAF (0.23 mL, 0.23 mmol, 10.3 eq, 1.0 M in THF) were added dropwise to a solution of silvl ether ent-30 (10.3 mg, 22.5 µmol, 1.0 eq) in THF (4 mL). The reaction mixture was stirred at room temperature for 20 h. After dilution with Et₂O (10 mL) and washing with an aqueous NaHCO₃ (10 mL), the aqueous layer was extracted with Et_2O (3×10 mL). The combined organic layers were dried (MgSO₄), the solvent was removed in vacuo and the residue was purified via column chromatography (EtOAc/petroleum ether 1:10 to 1:1) to afford (+)-SCB-5 ent-2d (5.1 mg, 22 μ mol, 99%) as a colorless oil. R_f=0.20 (EtOAc/petroleum ether 1:1, vanillin). $[\alpha]_{D}^{19} = +13.5^{\circ}$ (CH₂Cl₂, c = 0.50 g/100 mL). IR (thin film): 3421 (s, br), 2953 (s), 2923 (s), 2869 (s), 1767 (s), 1666 (w), 1467 (m), 1388 (m), 1260 (m), 1222 (w), 1179 (s), 1132 (m), 1027 (vs), 906 (w), 846 (w), 804 (w), 680 (w), 651 (w), 625 (w), 613 (w), 597 (w), 583 (w). ¹H NMR (C_6D_6 , 600 MHz): δ [ppm]=3.80–3.74 (m, 1H, H-6), 3.73 (dd, ²J=8.6 Hz, ³J=7.8 Hz, H-5a), 3.28 (dd, ²J=8.6 Hz, ³J=8.6 Hz, 1H, H-5b), 3.00 (dd, ²J=10.6 Hz, ³J=4.8 Hz, 1H, H-12a), 2.90 (dd, ²J=10.6 Hz, ³J=6.2 Hz, 1H, H-12b), 2.81 (s, 1H, H-13), 2.16-2.07 (m, 2H, H-3, H-4), 1.95 (s, 1H, H-14), 1.53-1.45 (m, 2H, H-8a, H-10), 1.45-1.39 (m, 1H, H-7a), 1.38-1.24 (m, 2H, H-7b, H-8b), 1.18-1.05 (m, 2H, H-9), 0.90 (d, ³J=6.7 Hz, 3H, H-11), 0.90 (d, ³J=6.7 Hz, 3H, H-11'). ¹³C NMR (C_6D_6 , 151 MHz): δ [ppm] = 176.9 (C-2), 70.9 (C-6), 67.9 (C-5), 62.9 (C-12), 49.4 (C-3), 40.1 (C-4), 39.1 (C-9), 34.4 (C-7), 28.3 (C-10), 24.1 (C-8), 22.8 (C-11), 22.8 (C-11'). HRMS (ESI): m/z [M+H]⁺ calcd for C₁₂H₂₃O₄⁺: 231.1592; found: 231.1591.

Acknowledgement

This work was supported by the "Naturstoffzentrum Rheinland-Pfalz". The authors are grateful for helpful discussions and financial aid. Open access funding enabled and organized by Projekt DEAL.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: Bioactive γ -butyrolactone \cdot Diastereoselective synthesis \cdot lodocyclization \cdot Ozonolysis \cdot Zwitterionic aza-Claisen rearrangement

- Small molecule hormones triggering antibiotics production in Streptomyces: a) N. B. Thao, S. Kitani, H. Nitta, T. Tomioka, T. Nihira, J. Antibiot. 2017, 70, 1004–1008; b) C. Corre, L. Son, S. O'Rouke, K. F. Chater, G. L. Challis, Proc. Natl. Acad. Sci. USA 2008, 105, 17510–17515; c) E. Takano, Curr. Opin. Microbiol. 2006, 9, 287–294; d) M. J. Bibb, Curr. Opin. Microbiol. 2005, 8, 208–215.
- [2] GBL's act as signaling molecules: a) Z. Zou, D. Du, Y. Zhang, J. Zhang, G. Niu, H. Tan, *Mol. Microbiol.* 2014, *94*, 490–505; b) J. M. Willey, A. A. Gaskell, *Chem. Rev.* 2011, *111*, 174–187; c) S. Kitani, M. Doi, T. Shimizu, A. Maeda, T. Nihira, *Arch. Microbiol.* 2010, *192*, 211–220; d) S. Horinouchi, T. Beppu, *Proc. Jpn. Acad. Ser. B* 2007, *83*, 277–295; e) N. Shikura, J. Yamamura, T. Nihira, *J. Bacteriol.* 2002, *184*, 5151–5157; f) R. Kawachi, T. Akashi, Y. Kamitani, A. Sy, U. Wangchaisoonthorn, T. Nihira, Y. Yamada, *Mol. Microbiol.* 2000, *36*, 302–313; g) M. Waki, T. Nihira, Y. Yamada, *J. Bacteriol.* 1997, *179*, 5131–5137; h) M. Kawabuchi, Y. Hara, T. Nihira, Y. Yamada, *FEMS Microbiol. Lett.* 1997, *157*, 81–85.
- [3] A-Factor from S. griseus: a) E. M. Kleiner, S. A. Pliner, V. S. Soifer, V. V. Onoprienko, T. A. Balasheva, B. V. Rozynov, A. S. Khokhlov, *Bioorg. Khim.* 1976, 2, 1142–1147; b) N. L. Anisova, I. N. Blinova, O. V. Efremenkova, L. P. Koz'min, V. V. Onoprienko, *Izv. Akad, Nauk. SSSR Biol.* 1984, 1, 98–108.
- [4] SCB-1/Gräfe SCB-Factor 1: from S. viridochromogenes a) U. Gräfe, W. Schade, I. Eritt, W. F. Fleck, L. Radies, J. Antibiot. 1982, 35, 1722–1723; from S. coelicolor A3(2) (incl. syntheses SCB-1, Gräfe VB-II): b) E. Takano, T. Nihira, Y. Hara, J. J. Jones, C. J. L. Gershater, Y. Yamada, M. Bibb, J. Biol. Chem. 2000, 275, 11010–11016; SCB-2-3 from S. coelicolor M145: c) N.-H. Hsiao, S. Nakayama, M. E. Merlo, M. de Vries, R. Bunet, S. Kitani, T. Nihira, E. Takano, Chem. Biol. 2009, 16, 951–960; SCB-1-SCB-8 from S. M1152: d) J. D. Sidda, V. Poon, L. Song, W. Wang, K. Yang, C. Corre, Org. Biomol. Chem. 2016, 14, 6390–6393; IM-2 from S. sp. FRI-5: e) K. Sato, T. Nihira, S. Sakuda, M. Yanagimoto, Y. Yamada, J. Ferment. Bioeng. 1989, 68, 170–173.
- [5] Gräfe-VB-Factor I–III from S. bikiniensis and S. cyaneofuscatus (III only) a) U. Gräfe, G. Reinhardt, W. Schade, I. Eritt, W. F. Fleck, L. Radics, *Biotechnol. Lett.* 1983, *5*, 591–596; VB-A from S. antibioticus: b) H. Y.-H. Ohashi, Zheng, Y. Yamada, *J. Antibiot.* 1989, *42*, 1191–1195; VB-A-C from S. virginiae (incl. synthesis VB-C): c) Y. Yamada, K. Sugamura, K. Kondo, M. Yanagimoto, H. Okada, *J. Antibiot.* 1987, *40*, 496–504; VB-A-E from S. virginiae: d) K. Kondo, Y. Higuchi, S. Sakuda, T. Nihira, Y. Yamada, *J. Antibiot.* 1989, *42*, 1873–1876; VB-C, VB-E from S. sp. CA-12953: e) K. Georgousaki, N. Tsafantakis, S. Gumeni, I. Gonzalez, T. A. Mackenzie, F. Reyes, C. Lambert, I. P. Trougakos, O. Genilloud, N. Fokialakis, *Bioorg. Med. Chem. Lett.* 2020, *30*, Art.-No. 126952.
- [6] SCB-1 derivatives displaying additional hydroxyl groups within the side chain see from S. badius: a) Y. Mu, X. Yu, Z. Zheng, W. Liu, G. Li, J. Liu, Y. Jiang, L. Han, X. Huang, *Magn. Res. Chem.* 2019, *57*, 1150–1157; From S. sp. W5: b) G. Wei, N. Zhu, Y. Zeng, Y. Shen, P. Zhao, *Ann. Microbiol.* 2010, *60*, 249–253.
- [7] Generally, GBL's isolated from various (modified) streptomyces strains had been characterized via several spectroscopic methods. Until now, the ¹H NMR data of the GBL's enable to subdivide into SCB and VB type species. However, the determination of the absolute configuration based on chemical synthesis of suitable enantiomers and diastereomers. Then, activity tests elucidated the bioactivity connected with the individual structure of the GBL's. Actually, it cannot be excluded, that the relative and absolute configurations of the compounds given in Figure 1 need further correction and addenda. Further investigations are required.
- [8] SAR Ref. 2e, a) S.-U. Choi, C.-K. Lee, Y.-I. Hwang, H. Kinoshita, T. Nihira, Arch. Microbiol. 2003, 180, 303–307; b) S. Kitani, Y. Yamada, T. Nihira, J. Bacteriol. 2001, 183, 4357–4363; c) S. Okamoto, K. Nakamura, T. Nihira, Y. Yamada, J. Biol. Chem. 1995, 270, 12319–12326; d) S. Okamoto, T. Nihira, H. Kataoka, A. Suzuki, Y. Yamada, J. Biol. Chem. 1992, 267, 1093– 1098; e) H. S. Kim, H. Tada, T. Nihira, Y. Yamada, J. Antibiot. 1990, 43, 692–706.
- [9] a) T. Nihira, Y. Simizu, H. S. Kim, Y. Yamada, J. Antibiot. 1988, 41, 1828– 1837; b) K. Hashimoto, T. Nihira, Y. Yamada, J. Ferment. Bioeng. 1992, 73, 61–65; c) M. Kawabuchi, Y. Hara, T. Nihira, Y. Yamada, FEMS Microbiol.

Lett. **1997**, *157*, 81–85; d) N. Shikura, T. Nihira, Y. Yamada, *Biochim. Biophys. Acta* **2000**, *1475*, 329–336.

- [10] Syntheses A-Factor: a) J. B. Morin, K. L. Adams, J. K. Sello, Org. Biomol. Chem. 2012, 10, 1517–1520 (biosynthesis); b) J. M. Crawforth, J. Fawcett, B. J. Rawlings, J. Chem. Soc. Perkin Trans. 1 1998, 1721–1725; c) P. J. Parsons, P. Lacrouts, A. D. Buss, J. Chem. Soc. Chem. Commun. 1995, 437–438; d) G. H. Posner, M. Weitzberg, S.-S. Jew, Synth. Commun. 1987, 17, 611–620; e) K. Mori, Tetrahedron 1983, 39, 3107–3109; f) K. Mori, K. Yamane, Tetrahedron 1982, 38, 2919–2921; SCB-1: Ref. 4b; SCB-2: g) A. M. Sarkale, A. Kumar, C. Appayee, J. Org. Chem. 2018, 83, 4167– 4172; IM-2: h) K. Mizuno, S. Sakuda, T. Nihira, Y. Yamada, Tetrahedron 1994, 50, 10849–10858; VB-A-D: i) K. Mori, N. Chiba, Liebigs Ann. Chem. 1990, 31–37; structure correction: j) S. Sakuda, Y. Yamada, Tetrahedron Lett. 1991, 32, 1817–1820; VB-C: k) K. Takabe, N. Mase, H. Matsumura, T. Hasegawa, Y. Iida, H. Kuribayashi, K. Adachi, H. Yoda, M. Ao, Bioorg. Med. Chem. Lett. 2002, 12, 2295–1197; VB-D: I) P. Elsner, H. Jiang, J. B. Nielsen, F. Pasi, K. A. Jørgensen, Chem. Commun. 2008, 5827–5829.
- [11] for a Review see: a) U. Nubbemeyer, Synthesis 2003, 961–1008; Substrate control: b) U. Nubbemeyer, J. Org. Chem. 1995, 60, 3773– 3780; c) U. Nubbemeyer, J. Org. Chem. 1996, 61, 3677–3686; d) C. Heescher, D. Schollmeyer, U. Nubbemeyer, Eur. J. Org. Chem. 2013, 4399–4404; Auxiliary control: e) S. Laabs, W. Münch, U. Nubbemeyer, J.-W. Bats, Tetrahedron 2002, 58, 1317–1334; f) N. Zhang, U. Nubbemeyer, Synthesis 2002, 242–252; g) N. M. Friedemann, A. Härter, S. Brandes, S. Groß, D. Gerlach, W. Münch, D. Schollmeyer, U. Nubbemeyer, Eur. J. Org. Chem. 2012, 2346–2358.
- [12] a) X. Wang, F. Wang, F. Huang, C. Ni, J. Hu, Org. Lett. 2021, 23, 1764–1768; b) T. Scattolin, K. Deckers, F. Schoenebeck, Org. Lett. 2017, 19, 5740–5743; c) S. Groß, S. Laabs, A. Scherrmann, A. Sudau, U. Nubbemeyer, J. Prakt. Chem. 2000, 342, 711–714.
- [13] J. Donges, S. Hofmann, J. C. Walter, J. Reichertz, M. Brüggemann, A. Frank, U. Nubbemeyer, *Synthesis* 2021, published online doi.org/ 10.1055/s-0037-1610770.
- [14] In 1990, Mori et al. (ref. 10 h) published ¹H NMR, IR and [α] data of both compounds **2d/3d**. For VB–A **3d** additional ¹³C NMR and MS data are published (ref. 5b, 5c). Focusing on SCB-5 **2d**, only incomplete data (no ¹³C-NMR, bioactivity data) are reported in the literature. Nihira et al. (ref. 9d) found, that a cell free extract of a VB–A overproducer induced a NADPH-dependent hydrogenation of racemic 1'-dehydro VB–A **1d** to give enantiopure (-)-VB–A (*2R*,*3R*,1'S) **3d** as well as the minor SCB-5 analogues (-)-(*2R*,*3R*,1'R) **2d** and (+)-(2*S*,*3S*,1'S) *ent*-**2d** separable via chiral HPLC. In 2016, Corre et al. (ref 4d) reported on the SCB-5 diastereomer **2d** biosynthesis, but MS and HPLC data only had been published. The bioactivity of both SCB-5 enantiomers **2d** remains to be investigated.
- [15] For testing purpose, a racemic sequence had been conducted. Starting from racemic *N*-(4-benzyloxy-8-methyl-2-nonenyl)-pyrrolidine (ref. 13, supporting information), aza-Claisen rearrangement, amide cleavage and reduction delivered racemic key intermediate *rac*-15 analog with high diastereoselectivity. For procedures and data see supporting information.
- [16] For preparative details and data see ref. 13 and supporting information connected with this paper. For the reagent controlled diastereoselective reduction using Midland's reagents (both enantiomers available) see a) M. M. Midland, A. Tramontano, A. Kazubski, R. S. Graham, D. J. S. Tsai, D. B. Cardin, *Tetrahedron* 1984, 40, 1371–1380; b) F. Liron, P. Knochel, *Chem. Commun.* 2004, 304–305; c) R. Lunkwitz, K. Zab, C. Tschierske, J. Prakt. Chem. 1998, 340, 662–668.
- [17] a) S. E. Denmark, M. G. Edwards, J. Org. Chem. 2006, 71, 7293–7306; b) I.
 Iriarte, O. Olaziola, S. Vera, I. Gamboa, M. Oierbide, C. Palomo, Angew.
 Chem. 2017, 129, 8986–8990; Angew. Chem. Int. Ed. 2017, 56, 8860–8864.
- [18] Amide reduction a) H. C. Brown, S. C. Kim, S. Krishnamurthy, J. Org. Chem. **1980**, 45, 1–12; b) G. B. Fisher, J. C. Fuller, J. Harrison, S. G. Alvarez, E. R. Burkhardt, C. T. Goralski, B. Singaram, J. Org. Chem. **1994**, 59, 6378–6385; c) A. G. Myers, B. H. Yang, L. McKinstry, S. J. Kopecky, J. L. Gleason, J. Am. Chem. Soc. **1997**, *119*, 6496–6511.
- [19] N. M. Kaluza, D. Schollmeyer, U. Nubbemeyer, *Eur. J. Org. Chem.* **2016**, 357–366.
- [20] a) M. S. Oberinde, H. N. Hunter, S. W. Bremner, M. G. Organ, *Eur. J. Org. Chem.* 2012, 175–182; b) Z. Liu, H. Qu, X. Gu, K.-S. Lee, B. V. Grossmann, K. Kumirov, V. J. Hruby, *Tetrahedron Lett.* 2010, *51*, 3518–3520; c) H. Nonaka, N. Ogawa, N. Maeda, Y.-G. Wang, Y. Kobayashi, *Org. Biomol. Chem.* 2010, *8*, 5212–5223; d) V. Blot, V. Reboul, P. Metzner, *Eur. J. Org. Chem.* 2006, 1934–1939; e) N. Noguchi, M. Nakada, *Org. Lett.* 2006, *8*,

2039–2042; f) V. Blot, V. Reboul, P. Metzner, J. Org. Chem. 2004, 69, 1196–1201; g) P. Metz, B. Hungerhoff, J. Org. Chem. 1997, 62, 4442–4448; h) Y. Tamaru, M. Mizutani, Y. Furukawa, S. Kawamura, Z. Yoshida, K. Yanagi, M. Minobe, J. Am. Chem. Soc. 1984, 106, 1079–1085.

- [21] Deposition Numbers 2076145 (for ent- β -11), and 2076144 (for α -11) contain the supplementary crystallographic data for this paper. These data are provided free of charge by the joint Cambridge Crystallographic Data Centre and Fachinformationszentrum Karlsruhe Access Structures service www.ccdc.cam.ac.uk/structures.
- [22] The zwitterionic aza-Claisen rearrangement led to the syntheses of both enantiomers of ester 12 and carbinol 15, respectively. For completion of both, (-)-SCB-5 and (+)-SCB-5 total synthesis, several strategies had been tested involving only one of the enantiomer series. For clarification, all intermediates heading for (-)-SCB-5 2d, the simple molecule numbers are used. All intermediates leading to (+)-SCB-5 ent-2d are presented using the prefix "ent".
- [23] a) K. Tatsuta, H. Takahashi, Y. Amemiya, M. Kinoshita, J. Am. Chem. Soc.
 1983, 105, 4096–4097; b) M. F. Jones, S. A. Noble, C. A. Robertson, R. Storer, *Tetrahedron Lett.* 1991, 32, 247–250; c) H. Takahashi, Y. Iwai, Y. Hitomi, S. Ikegami, Org. Lett. 2002, 4, 2401–2403.
- [24] Selenocyclization a) Z. M. Burgacic, M. P. Gavrilovic, V. M. Divac, *Monatsh. Chem.* 2007, *138*, 149–151; b) M. Tiecco, L. Testaferri, L. Bagnolli, V. Purgatorio, A. Temperini, F. Marini, C. Santi, *Tetrahedron Asymmetry* 2004, *15*, 405–412; c) A. Stojanovic, P. Renaud, K. Schenk, *Helv. Chim. Acta* 1998, *81*, 268–284; Oxidation and *syn-elimination: d*) H. J. Kim, Y.-n. Liu, R. M. McCarty, H.-w. Liu, *J. Am. Chem. Soc.* 2017, *139*, 16084–16087; e) A. G. Myers, D. Y. Gin, D. H. Rogers, *J. Am. Chem. Soc.* 1994, *116*, 4697–4718.
- [25] Work-up and purification via column chromatography of selanides ent-17 for analytical purposes was connected with a partial cleavage of the TBS group delivering minor amounts of the corresponding carbinols ent-17a (Scheme 5). However, such silylether cleavage was not found after running the selenocyclization and the subsequent oxidation. Selenoxides ent-18 were isolated with 97% yield.
- [26] Iodocyclization: 5-exo-trig a) A. Ramdular, K. A. Woerpel, Org. Lett. 2020, 22, 4113–4117; b) J. Halder, D. Das, S. Nanda, Org. Biomol. Chem. 2018, 16, 2549–2575; c) N.-C. Hsueh, Y.-T. Hsiao, M.-Y. Chang, Tetrahedron 2017, 73, 4398–4406; d) S. Konda, M. Khatravath, N. K. Mallurwar, P. Rao, S. Sripelly, J. Iqbal, P. Arya, Synthesis 2016, 48, 1663–1683; e) J. A. Goodwin, C. F. Ballesteros, A. Aponick, Org. Lett. 2015, 17, 5574–5577; f) A. S.-Y. Lee, K.-W. Tsao, Y.-T. Chang, S.-F. Chu, Tetrahedron Lett. 2007, 48, 6790–6793; 5-endo-trig: g) J. I. Nallasivam, R. A. Fernandes, J. Am. Chem. Soc. 2016, 138, 13238–13245; h) W. Yang, Z. Wang, J. Sun, Angew. Chem. 2016, 128, 7068–7072; Angew. Chem. Int. Ed. 2016, 55, 6954–6958.
- [27] Elimination: a) M. Bege, I. Bereczki, M. Herczeg, M. Kicsák, D. Eszenyi, P. Herczegh, A. Borbás, Org. Biomol. Chem. 2017, 15, 9226–9233; b) S. Nie, X. Chen, Y. Ma, W. Li, B. Yu, Carbohydr. Res. 2016, 432, 36–40; c) W. Li, Y. Niu, D.-C. Xiong, X. Cao, X.-S. Ye, J. Med. Chem. 2015, 58, 7972–7990; d) B. Ramakrishna, P. R. Sridhar, Org. Lett. 2013, 15, 4474–4477; e) A. Neogi, T. P. Majhi, N. Goshal, P. Chattopadhyay, Tetrahedron 2005, 61, 9368–9374.
- [28] For procedures and data delivering several iodide substitution products see supporting information.
- [29] Ozonolysis vinyl group in Ester a) P. Wipf, M. D. Manojlovic, *Beilstein J. Org. Chem.* 2011, *7*, 824–830; b) Y. Kobayashi, Y. Kitano, Y. Takeda, F. Sato, *Tetrahedron* 1986, *42*, 2937–2944; RuCl₃, NalO₄: c) P. R. McGuirk, D. B. Collum, *J. Am. Chem. Soc.* 1982, *104*, 4496–4497.
- [30] Sequence tetrahydrofurane 20 to lactone 26: via isolation/purification of acid 24: 10% yield, via isolation/purification of ester 25a: 23% yield, sequence without purification of the intermediates. 36% yield. Higher yields might be achievable optimizing the intermediate aldehyde 25b oxidation. However, such oxidation hampered from several competing processes in analogy to the formation of *ent*-30 (see below).
- [31] Ozonolysis of the vinyl group in carbinol: Ref. 11d, a) Z. Lu, X. Zhang, Z. Guo, Y. Chen, T. Mu, A. Li, J. Am. Chem. Soc. 2018, 140, 9211–9218; b) Y. Sahara, J. Cui, M. Furutachi, J. Chen, T. Watanabe, M. Shibasaki, Synthesis 2017, 49, 69–75; c) M. Körner, M. Hiersemann, Synthesis 2016, 48, 2466–2482; d) C. Lv, X. Yan, Q. Tu, Y. Di, C. Yuan, X. Fang, Y. Ben-David, L. Xia, J. Gong, Y. Shen, Z. Yang, X. Hao, Angew. Chem. 2016, 128, 7665–7669; Angew. Chem. Int. Ed. 2016, 55, 7539–7543; e) R. Tsutsumi, S. Hong, M. Krische, Chem. Eur. J. 2015, 21, 12903–12907.
- [32] For a final removal of the TBS ether strong basic conditions should be avoided. Any intramolecular transesterification incorporating the C3

hydroxymethyl group would have caused a C3 epimerization potentially leading to competing 2,3 *cis* configured GBL side products.

- [33] Furthermore, a ketoformate was isolated indicating some over oxidation within this last Cr(VI) reaction. Presumably, the lactol suffered from H_2O elimination and subsequent dihydrofuran oxidative cleavage. For details and data see supporting information.
- [34] For details and data lists see supporting information (NMR data measured in CDCl₃ and C₆D₆, respectively. Mori: NMR data in CDCl₃ published, ref. 10 h).
- [35] Furthermore, orientation of the electron rich enolate terminus in the proximity of the empty *anti* bonding orbital of the C–OTBS bond, might be rated as an attractive interaction, too.
- [36] Always, starting from both iodoethers 20/21, reductive ring opening allowed regeneration of carbinols 15 with nearly quantitative yield.

Thus, the likewise generation of iodoethers **20** and **21**, respectively, could be optimized by recycling the undesired isomer.

[37] First experiments introducing an ester group at the 1' OH function gave small amounts of the ester. The major product displayed an *exo* double bond indicating a rapid condensation destroying the stereogenic centers at C2 an C1'. However, OH group inversion requires the absence of the lactone carbonyl function. For details see supporting information.

Manuscript received: April 23, 2021 Revised manuscript received: May 17, 2021 Accepted manuscript online: May 18, 2021