

# DISSERTATION

## **The Protective Role and Underlying Mechanism of Hydrogen Sulfide against glaucomatous injuries *in vitro* and *in vivo***

Submitted in fulfillment of the requirements for the academic degree of

**Medical Doctor (MD)**

at the University Medical Center  
of the Johannes Gutenberg-University Mainz

by

**Hanhan Liu**

**Born 16<sup>th</sup> of December 1989**

**in Shanghai, China**

**Mainz, September 2020**



---

***„Science is a wonderful thing.”***

Albert Einstein

## Declaration I

Parts of this dissertation have been published in international journals and presented at scientific conferences:

### Published papers/manuscripts in revision

- 1) **Proteomics reveals the potential protective mechanism of hydrogen sulfide on retinal ganglion cells in an ischemia/reperfusion injury animal model**  
Liu H, Perumal N, Manicam C, Mercieca K, Prokosch V  
Pharmaceuticals (Basel) 13, no. 9, 2020, PMID: 32867129
- 2) **Hydrogen Sulfide and  $\beta$ -Synuclein Are Involved and Interlinked in the Aging Glaucomatous Retina**  
Liu H, Mercieca K, Anders F, Prokosch V  
Journal of Ophthalmology, 2020, PMID: 32351728
- 3) **Mitochondrial Markers in Aging and Primary Open-Angle Glaucoma**  
Liu H, Mercieca K, Prokosch V  
Journal of Glaucoma, 2020, PMID: 31977544
- 4) **Proteome alterations in aqueous humour of primary open angle glaucoma patients**  
Liu H, Anders F, Funke S, Mercieca K, Grus F, Prokosch V  
International Journal of Ophthalmology, 2020, PMID: 31956586
- 5) **Hydrogen Sulfide Protects Retinal Ganglion Cells Against Glaucomatous Injury In Vitro and In Vivo**  
Liu H, Anders F, Thanos S, Mann C, Liu A, Grus FH, Pfeiffer N, Prokosch V  
Investigative Ophthalmology & Visual Science, 2017, PMID: 28986598
- 6) **Alterations in Tight- and Adherens-Junction Proteins Related to Glaucoma Mimicked in the Organotypically Cultivated Mouse Retina Under Elevated Pressure.**  
Brockhaus K, Melkonyan H, Prokosch-Willing V, Liu H, and Thanos S.  
Investigative Ophthalmology & Visual Science, 2020, PMID: 32207812
- 7) **Elevated intraocular pressure induces neuron-specific beta-III-tubulin expression in non-neuronal vascular cells**  
Prokosch V, Brockhaus K, Anders F, Liu H, Mercieca K, Gericke A, Melkonyan H, Thanos S

Acta Ophthalmologica, 2019, PMID: 31885180

**8) Neuroprotective and neuroregenerative effects of CRMP-5 on retinal ganglion cells in an experimental in vivo and in vitro model of glaucoma**

Lauzi J, Anders F, **Liu H**, Pfeiffer N, Grus F, Thanos S, Arnhold S, Prokosch V  
PLoS One, 2019, PMID: 30673694

**9) Relaxin 2 fails to lower intraocular pressure and to dilate retinal vessels in rats**

Hampel U, Träger K, **Liu H**, Teister J, Grus F, Prokosch-Willing V  
International Ophthalmology, 2018, PMID: 29536410

**10) Morphological and Quantitative Changes in Retinal and Optic Nerve Vessels in Experimental Glaucoma Model with Elevated IOP for 7 Weeks**

Mann C, Anders F, **Liu H**, Brockhaus K, Liu A, Grus FH, Pfeiffer N, Thanos S, Prokosch V

Klinische Monatsblätter für Augenheilkunde, 2018, PMID: 29590684

**11) Femtosecond laser in refractive and cataract surgeries**

**Liu H**, Hu Y, Cui H

International Journal of Ophthalmology, 2015, PMID: 25938066

**Poster presentations at scientific conferences:**

- 1) Proteomics reveals the potential mechanisms hydrogen sulfide involve in to protect retinal ganglion cells in a glaucomatous animal model

**Liu H**, Perumal N, Manicam C, Prokosch V<sup>1</sup>

Deutsche Ophthalmologische Gesellschaft Kongress 2019 in Berlin

- 2) Analysis of mitochondrial proteomic profile to protect retinal ganglion cells from glaucomatous injury

**Liu H**, Perumal N, Anders F, Grus FH, Pfeiffer N, Prokosch V

Deutsche Ophthalmologische Gesellschaft Kongress 2017 in Berlin

- 3) Hydrogen sulfide protects retinal ganglion cells from cell death due to elevated hydrostatic pressure in vitro and in vivo

**Liu H**, Anders F, Thanos S, Mann C, Liu A, Grus FH, Pfeiffer N, Prokosch V

Deutsche Ophthalmologische Gesellschaft Kongress 2016 in Berlin

## Declaration II

The present thesis was written under the supervision of Prof. Dr. Prokosch at the Department of Ophthalmology, University Medical Care Center, Johannes Gutenberg-University in Mainz. The doctoral thesis is designed as a cumulative dissertation and is based on the following publications and manuscripts:

**1) Hydrogen Sulfide Protects Retinal Ganglion Cells Against Glaucomatous Injury In Vitro and In Vivo**

Liu H, Anders F, Thanos S, Mann C, Liu A, Grus FH, Pfeiffer N, Prokosch V  
Investigative Ophthalmology & Visual Science, 2017, PMID: 28986598

**2) Hydrogen Sulfide and  $\beta$ -Synuclein Are Involved and Interlinked in the Aging Glaucomatous Retina**

Liu H, Mercieca K, Anders F, Prokosch V  
Journal of Ophthalmology, 2020, PMID: 32351728

**3) Proteomics reveals the potential protective mechanism of hydrogen sulfide on retinal ganglion cells in an ischemia/reperfusion injury animal model**

Liu H, Perumal N, Manicam C, Mercieca K, Prokosch V  
Pharmaceuticals (Basel) 13, no. 9, 2020, PMID: 32867129

---

**Table of Contents**

<b>DECLARATION I</b> .....	<b>IV</b>
<b>DECLARATION II</b> .....	<b>VI</b>
<b>TABLE OF CONTENTS</b> .....	<b>VII</b>
<b>ABSTRACT</b> .....	<b>1</b>
<b>ZUSAMMENFASSUNG</b> .....	<b>3</b>
<b>1 INTRODUCTION</b> .....	<b>5</b>
1.1 GLAUCOMA – PATHOLOGY AND TREATMENT.....	5
1.2 GASEOUS SIGNALING MOLECULE IN THE EYE .....	5
1.3 HYDROGEN SULFIDE- A NOVEL ENDOGENOUS GASEOUS SIGNALING MOLECULE.....	6
1.4 NEUROPROTECTIVE PROPERTIES OF H <sub>2</sub> S.....	6
1.5 SYNUCLEINS –STRUCTURE, CLASSES AND FUNCTIONS .....	7
1.6 AIMS OF THE THESIS.....	8
<b>2 MANUSCRIPTS</b> .....	<b>9</b>
2.1 PUBLICATION I.....	9
2.2 PUBLICATION II .....	10
2.2 PUBLICATION III .....	11
<b>3 SUMMERAY OF THE RESULTS AND DISCUSSION</b> .....	<b>12</b>
3.1 HYDROGEN SULFIDE PROTECTS RETINAL GANGLION CELLS AGAINST GLAUCOMATOUS INJURY IN VITRO AND IN VIVO .....	14
3.2 HYDROGEN SULFIDE AND B-SYNUCLEIN ARE INVOLVED AND INTERLINKED IN THE AGEING GLAUCOMATOUS RETINA .....	16
3.3 PROTEOMICS REVEALS THE POTENTIAL PROTECTIVE MECHANISM OF HYDROGEN SULFIDE ON RETINAL GANGLION CELLS IN AN ISCHEMIA/REPERFUSION INJURY ANIMAL MODEL .....	20
3.3.1 <i>Changes in Iron Homeostasis and ROS Regulation</i> .....	21
3.3.2 <i>Changes in Retinal Metabolism, Mitochondrial Homeostasis and Function</i> .....	23
3.3.3 <i>Changes in Retinal Vascular Function</i> .....	26
3.3.4 <i>Changes in GABA Receptor Signaling</i> .....	29
3.3.5 <i>Changes in DNA Repair</i> .....	30
3.4 CRITICAL DISCUSSION .....	31
<b>4 CONCLUSION AND OUTLOOK</b> .....	<b>33</b>
<b>5. REFERENCE</b> .....	<b>36</b>
<b>6 APPENDIX</b> .....	<b>V</b>
6.1 CONTRIBUTIONS TO THE MANUSCRIPTS.....	V
6.2 LIST OF ABBREVIATIONS .....	VI
6.3 LIST OF FIGURES.....	VII
6.4 CURRICULUM VITAE .....	VIII
6.5 DECLARATION OF ACADEMIC HONESTY .....	IX

**Abstract**

Glaucoma, one of the leading causes of irreversible blindness worldwide and is characterized by progressive optic nerve and retinal ganglion cell (RGC) degeneration. Hydrogen sulfide (H<sub>2</sub>S) plays a role as a potent neurotransmitter and has been proven to protect RGCs against glaucomatous injury in vitro and in vivo, although the exact mechanism is unclear.

The main objective of this doctoral thesis was to reveal the potential roles of H<sub>2</sub>S in glaucoma pathophysiology and to better understand the mechanism of H<sub>2</sub>S in neuroprotection.

In the process of aging, RGCs and  $\beta$ -synuclein (SNCB) are significantly changed in old animals. Under chronic IOP elevation there is a significant RGC loss in old animals, whereas no significant change in young animals; SNCB is significantly downregulated and 3-mercaptosulfurtransferase, a H<sub>2</sub>S producing enzyme, showed a 3-fold up-regulation within the retina in young animals after IOP elevation, while no significant changes in old ones are notable.

GY4137, a H<sub>2</sub>S slow-releasing donor, was observed to protect RGC against elevated pressure and oxidative stress in vitro depending on the concentration used ( $p < 0.005$ ). In vivo, intravitreal administration of GY4137 preserved RGCs from acute ischemic injury and optic nerve crush ( $p < 0.0001$ ). Under acute ischemic injury, exogenous H<sub>2</sub>S also significantly downregulated SNCB levels. Furthermore, retinal vessel diameters enlarged after intravitreal GY4137 injection ( $p < 0.0001$ ).

As final part of this doctoral thesis, we examined the potential protective mechanisms activated by H<sub>2</sub>S in a glaucoma animal model. We used mass spectrometry-based proteomics to elucidate how protein expression changes at the cellular level. In total 1115 proteins were identified, 48 proteins were significantly differentially expressed due to I/R. Abundance of 18 key proteins were restored by H<sub>2</sub>S. Another 11 proteins were differentially expressed following H<sub>2</sub>S. Proteomic and ingenuity pathway analysis (IPA) revealed a significant H<sub>2</sub>S-mediated activation of pathways related to mitochondrial function, iron homeostasis and vasodilation.

In conclusion, the present doctoral thesis is the first study to analyse the effect of H<sub>2</sub>S in the pathogenesis of experimental glaucoma and provide an overall insight into the

mainstay retinal proteins and pivotal signaling pathways that interact with H<sub>2</sub>S to maintain retinal homeostasis against glaucomatous injury. These results form the basis for further research of H<sub>2</sub>S as an innovative treatment strategy for glaucoma.

## Zusammenfassung

Das Glaukom, eine der weltweit führenden Ursachen für irreversible Blindheit, ist durch eine fortschreitende Degeneration von Sehnerven und retinalen Ganglienzellen (RGC) gekennzeichnet. Schwefelwasserstoff ( $H_2S$ ) spielt eine Rolle als potenter Neurotransmitter und schützt nachweislich RGCs in vitro und in vivo vor glaukomatösen Verletzungen, obwohl der genaue Mechanismus unklar ist.

Das Hauptziel dieser Doktorarbeit war es, die möglichen Rollen von  $H_2S$  in der Pathophysiologie des Glaukoms aufzudecken und den Mechanismus von  $H_2S$  bei der Neuroprotektion besser zu verstehen.

Im Alterungsprozess verändern sich RGCs und  $\beta$ -Synuclein (SNCB) bei alten Tieren signifikant. Bei chronischer Erhöhung des Augeninnendrucks gibt es bei alten Tieren einen signifikanten RGC-Verlust, während bei jungen Tieren keine signifikante Veränderung auftritt. SNCB ist signifikant herunterreguliert und 3-Mercaptosulfurtransferase, die  $H_2S$ -produzierendes Enzyme, zeigte bei jungen Tieren nach Erhöhung des Augeninnendrucks eine dreifache Hochregulation innerhalb der Netzhaut, während bei alten Tieren keine signifikanten Veränderungen bemerkenswert sind.

Es wurde beobachtet, dass GYY4137, ein langsam freisetzender  $H_2S$ -Donor, RGC in vitro in Abhängigkeit von der verwendeten Konzentration vor erhöhtem Druck und oxidativem Stress schützt ( $p < 0,005$ ). In vivo bewahrte die intravitreale Verabreichung von GYY4137 RGCs vor akuten ischämischen Verletzungen und Sehnervenquetschungen ( $p < 0,0001$ ). Bei akuten ischämischen Verletzungen regulierte exogenes  $H_2S$  auch die SNCB-Spiegel signifikant herunter. Darüber hinaus vergrößerten sich die Durchmesser der Netzhautgefäße nach intravitrealer GYY4137-Injektion ( $p < 0,0001$ ).

Als letzten Teil dieser Doktorarbeit untersuchten wir die möglichen Schutzmechanismen, die durch  $H_2S$  in einem Glaukom-Tiermodell aktiviert werden. Wir verwendeten massenspektrometrische Proteomik, um zu untersuchen, wie sich die Proteinexpression auf zellulärer Ebene ändert. Insgesamt wurden 1115 Proteine identifiziert, 48 Proteine wurden aufgrund von I / R signifikant unterschiedlich exprimiert. Die Häufigkeit von 18 Schlüsselproteinen wurde durch  $H_2S$  wiederhergestellt. Weitere 11 Proteine wurden nach  $H_2S$  unterschiedlich exprimiert.

Die Proteom- und Einfallsreichtumsanalyse (IPA) ergab eine signifikante H<sub>2</sub>S-vermittelte Aktivierung von Signalwegen im Zusammenhang mit Mitochondrienfunktion, Eisenhomöostase und Vasodilatation.

Zusammenfassend ist die vorliegende Doktorarbeit die erste Studie, die die Wirkung von H<sub>2</sub>S auf die Pathogenese des experimentellen Glaukoms analysiert und einen umfassenden Einblick in die Hauptproteine der Netzhaut und die zentralen Signalwege bietet, die mit H<sub>2</sub>S interagieren, um die Homöostase der Netzhaut gegen glaukomatöse Verletzungen aufrechtzuerhalten. Diese Ergebnisse bilden die Grundlage für die weitere Erforschung von H<sub>2</sub>S als innovative Behandlungsstrategie für das Glaukom.

## **1 Introduction**

### **1.1 Glaucoma – Pathology and treatment**

Glaucoma is one of the leading causes of irreversible blindness worldwide[1]. Glaucoma is a group of diseases characterized by the accelerated death of retinal ganglion cells (RGCs) and their axons. The death of RGCs ultimately leads to progressive visual field loss and irreversible blindness[2]. Elevated intraocular pressure (IOP) is a main risk factor for glaucoma and the mainstay of treatment, but simply lowering IOP may fail to halt the disease progression in many patients despite delaying RGC death. Novel strategies which halt RGC loss are desperately needed to prolong visual function in glaucoma[3]. Various other treatment options have been examined to delay or halt RGC loss in order to preserve visual function in glaucoma patients without combating results. In recent years the role other factors like glutamate excitotoxicity, disorganized metabolism of nitric oxide (NO), overproduction of reactive oxygen species (ROS), increased endothelin-2 levels and reduction of ocular blood flow has also been discussed to play a key role[4] .

### **1.2 Gaseous signaling molecule in the eye**

The importance of NO in biology and medicine was highlighted in 1998 when the Nobel Prize was awarded in Physiology and Medicine to Robert Furchgott, Louis Ignarro and Ferid Murad for their pioneering work on the role of NO in the nervous, cardiovascular and immune systems [5]. In the same time period, carbon monoxide (CO) has also been recognized as a putative neurotransmitter [6]. They are both involved in several important aspects of neuronal function. Under physiological conditions, NO formed in the eye is beneficial in preventing the genesis of glaucoma or halting its progress; however, excessive NO would become pathogenic in the progress of glaucomatous optic neuropathy [7]. In vitro and in vivo studies have shown evidence of NO's involvement in retinal degeneration, the regulation of IOP and the production of aqueous humor [8–10]. Drugs inhibiting NO formation showed protective effects in the retina against elevated IOP in glaucoma animal models [11–13]. On the other hand, The CO production system in the retina has been highlighted as a protective mechanism, its protecting role is demonstrated in the regulation of Ischemia-reperfusion (I/R) -induced damage, protecting retinal ganglion cells against oxidative damage, increases retinal and choroidal blood flows and downregulating inflammatory response in the anterior segment of the eye [14–16].

### 1.3 Hydrogen sulfide- a novel endogenous gaseous signaling molecule

Hydrogen sulfide ( $H_2S$ ), along with CO and NO, has been recognized as a third endogenous gaseous signaling molecule[5]. It is produced via cysteine catabolism by the cytoplasmic enzymes, cystathionine- $\beta$ -synthase (CBS) and cystathionine- $\gamma$ -lyase (CSE), and also via 3-mercaptopyruvate catabolism by 3-mercaptopyruvate sulfurtransferase (3-MST) [6].  $H_2S$  has recently been considered an endogenous gasotransmitter with protective potential at low concentrations. Alteration of endogenous  $H_2S$  levels in the retina is also linked to different pathological conditions, and its exogenous donors have been shown to exhibit potential in protecting retinal ganglion cells against insults such as diabetic retinopathy, ischemia–reperfusion injury and N-methyl-D-aspartic acid (NMDA)-induced excitatory neurotoxicity [11-13].

Recently cell protective properties within the cardiovascular system have been found and thus its action in the cardiovascular system excessively studied. It has been shown, that endogenously-generated and exogenously-administered  $H_2S$  exerts a wide range of actions in vascular and myocardial cells including vasodilator/vasoconstrictor effects via modification of the smooth muscle tone [7].

There are different  $H_2S$  donors available, the most widely used donors are inorganic sulfite salts, such as NaHS and  $Na_2S$ . The slow releasing  $H_2S$  donor GYY4137 is chosen for this study, it seems more beneficial to expose cells to low concentrations of the gas over a longer period of time. GYY4137 slowly releases low but consistent concentrations of  $H_2S$  over several hours in aqueous solutions at physiological pH and temperature. GYY4137 therewith mimics the time course of the physiological  $H_2S$  release *in vivo*. Additionally, as known so far GYY4137 does not cause any significant cytotoxic effects.

### 1.4 Neuroprotective properties of $H_2S$

Besides these vascular effects, neuroprotective properties have been assumed. In the central nervous system(CNS)  $H_2S$  has been found to facilitate long-term potentiation and regulate intracellular calcium concentration and pH level in brain cells. Antioxidant, anti-apoptotic, and anti-inflammatory effects of  $H_2S$  have been found e.g. in Alzheimer's disease(AD), Parkinson's disease(PD), and vascular dementia. Furthermore an abnormal generation and metabolism of  $H_2S$  has been seen involved in most of these neurodegenerative disorders [8]. Moreover,  $H_2S$  protects neurons against glutamate-mediated oxidative stress, or oxytosis, through the pleiotropic

---

effects of maintaining the activities of cystathionine- $\gamma$ -lyase and cystine transport, leading to an increase in glutathione levels [9]. The neuroprotective effect of H<sub>2</sub>S was partly attributed to its capability of vaso-relaxation, anti-oxidative stress, neuroendocrine regulation, and inflammation suppression [10-12], but our current understanding of the mechanism behind RGC apoptosis and protective properties of H<sub>2</sub>S is far from comprehensive.

### **1.5 Synucleins –Structure, classes and functions**

Synuclein is a family of small proteins including  $\alpha$  (SNCA),  $\beta$  (SNCB) and  $\gamma$  (SNCG) synucleins[13,14] and is involved in various neurodegenerations in the CNS. Specifically, SNCA is a major constituent of Lewy bodies, pathological neuronal inclusion bodies found in Parkinson's disease, Alzheimer's disease, and other neurodegenerative disorders [15,16]. Mutations of SNCA play a central role in PD pathology, and misfolding and aggregation of SNCA directly linked to microglial activation, followed by inflammation and oxidative stress resulting in neurodegeneration[17]. Synucleins are present in the retina and optic nerve[18] and are associated with glaucomatous alterations in the optic nerve[19]. SNCA autoantibody was found to be downregulated in serum and upregulated in aqueous humor of glaucoma patients[20] and in our previous study, intravitreal injection of SNCA antibodies is found to be neuroprotective in glaucoma animal model[21].

$\beta$  –synuclein shares a similar protein structure to SNCA [22], but lack the non-amyloid- $\beta$  component domain[23]. Its expression is documented to be increased in cerebrospinal fluid in patients with neurodegenerative diseases and in neuronal retina and visual cortex of rats and nonhuman primates with age and external stress [24-26]. SNCB is thought to function as a physiological inhibitor of SNCA in neurodegenerative diseases [26,27] , it retain anti-apoptotic ability in a dose-dependent manner [13],and  $\beta$ -synuclein-derived peptides behave as anti-aggregating agents [15].

While SNCA and SNCB are mainly associated with diseases in the CNS,  $\gamma$ -synuclein is first identified as breast-specific gene protein 1[28], but it is also involved in axonal spheroid-like lesions in Parkinson's disease, deposition in glial cells in glaucoma, and motor neuron dysfunction and death[19,29,30].

Because of the pivotal role of SNCA in neurodegeneration in CNS, it has been extensively studied in CNS. But its role in retina or glaucoma, as well as SNCB and

SNCG's roles and functions in glaucoma is still sparse and remain to be thoroughly explored.

### **1.6 Aims of the thesis**

In first part of the study, we analyse the expression changes of H<sub>2</sub>S in an experimental animal model of glaucoma and second H<sub>2</sub>S's potential neuroprotective effect on RGC towards elevated pressure in glaucoma models in vitro and in vivo by addition of the slow-releasing H<sub>2</sub>S donor GYY4137.

Studies have shown that H<sub>2</sub>S and synucleins are involved in several mutual pathophysiological processes, such as microglia activation, p53-mediated apoptosis, inflammatory response and free radical reactions [5,31-33]. As second part of the thesis, we first elucidate the potential roles and functions of synucleins in glaucomatous neuropathy, following this, to investigate the potential of H<sub>2</sub>S to regulate it, and to better understanding the mechanism of H<sub>2</sub>S in neuroprotection.

At the end of the project, a mass spectrometry-based proteomics approach was employed to analyze the retinal proteome, and bio-informatics to algorithmically generate protein connections, which allowed us to identify the most plausible signaling pathway alterations related to H<sub>2</sub>S's neuroprotective properties against ischemia-reperfusion injury, and to provide a more precise direction for further studies.

2 Manuscripts

## **2.1 Publication I**

Hydrogen Sulfide Protects Retinal Ganglion Cells Against Glaucomatous Injury In Vitro and In Vivo

**Liu H**, Anders F, Thanos S, Mann C, Liu A, Grus FH, Pfeiffer N, Prokosch V

Investigative Ophthalmology & Visual Science, 2017

PMID: 28986598, DOI: 10.1167/iovs.17-22200

## **2.2 Publication II**

Hydrogen Sulfide and  $\beta$ -Synuclein Are Involved and Interlinked in the Aging Glaucomatous Retina

**Liu H**, Mercieca K, Anders F, Prokosch V

Journal of Ophthalmology, 2020

PMID: 32351728, DOI: 10.1155/2020/8642135

### **2.3 Publication III**

Proteomics reveals the potential protective mechanism of hydrogen sulfide on retinal ganglion cells in an ischemia/reperfusion injury animal model

**Liu H**, Perumal N, Manicam C, Mercieca K, Prokosch V

Pharmaceuticals, 2020

PMID: 32867129, DOI: 10.3390/ph13090213

---

### 3 Summeray of the results and discussion

Glaucoma, one of the leading causes of irreversible blindness worldwide[34], is a group of disorders characterized by progressive retinal ganglion cell loss and axon atrophy, which leads to gradually visual field loss [35].

By far the only known modifiable risk factor of glaucoma is intraocular pressure, however, lowering IOP is not able to halt the deterioration of glaucoma in most patients in clinic practice, indicating again the multifactorial pathogenesis and the complexity of glaucoma [36]. The other main risk factor is age. Alternative approaches independent of IOP and probably combating ageing as well as focusing on the pathophysiological processes are in demand to ameliorate glaucoma neuropathy. Other pathophysiological processes including oxidative stress, inflammatory reaction, glial activation, vascular dysfunctions and abnormal protein accumulation are proven to be closely involved[37-40]. Various other treatment options have been examined to delay or halt RGC loss in order to preserve visual function in glaucoma patients without combating results. In recent years the role other factors like glutamate excitotoxicity, disorganized metabolism of nitric oxide (NO), overproduction of reactive oxygen species (ROS) and reduction of ocular blood flow has also been discussed to play a key role [4] .

An interesting molecule in this context is H<sub>2</sub>S. H<sub>2</sub>S is the third identified gaseous transmitter after NO and CO and it is endogenously generated in mammalian endothelial cells [41]. It is produced via cysteine catabolism by the cytoplasmic enzymes, cystathionine-β-synthase (CBS) and cystathionine-γ-lyase (CSE), and also via 3-mercaptopyruvate catabolism by 3-mercaptopyruvate sulfurtransferase (3-MST) [6].

Ever since H<sub>2</sub>S has been recognized as the third endogenous gaseous signaling molecule, its role in various physiological and pathological processes has been explored. As a potent reductant, H<sub>2</sub>S plays critical roles in multiple physiological and pathological processes, it works to alleviate inflammatory responses, oxidative stress, and restores energy shortage[31,42,43]. H<sub>2</sub>S has shown profound therapeutic efficiency potential in neurodegenerative diseases in CNS [44-47]. Researches focus on H<sub>2</sub>S in connection with glaucoma have increasingly emerged in last few years, our knowledge on this topic is still lacking and remains to be thoroughly expanded. Alteration of endogenous H<sub>2</sub>S level in retina is correlated with different pathological

situations, and its exogenous donors exhibited potential in protecting retinal ganglion cells against assaults, such as diabetic retinopathy, ischemia–reperfusion injury and N-methyl-D-aspartic acid (NMDA)-induced excitatory neurotoxicity[48-50].

In first part of study, covered in Publication I, endogenous H<sub>2</sub>S synthases are observed to increase in a glaucoma animal model after seven weeks of chronic elevated IOP. Furthermore, we demonstrated that administration of GYY4137, a slow-release H<sub>2</sub>S donor, significantly improved RGC survival under different glaucomatous injuries in vitro and in vivo[51].

The neuroprotective effect of H<sub>2</sub>S in the retina was partly attributed to its capability of vasorelaxation, anti-oxidative stress, neuroendocrine regulation, and inflammation suppression[52] but the underlying mechanism and the specific protein interaction networks and the mechanisms involved in protective properties of H<sub>2</sub>S remain to be elucidated.

Synuclein is a family of small proteins including  $\alpha$  (SNCA),  $\beta$  (SNCB) and  $\gamma$  (SNGG) synucleins[13,14] and is involved in various neurodegenerations in the CNS. Synucleins are also present in the retina and optic nerve[18] and are associated with glaucomatous alterations in the optic nerve[19]. SNCA autoantibody was found to be downregulated in serum and upregulated in aqueous humor of glaucoma patients [20].

In second part of the study, covered in Publication II, we investigated the level changes of synucleins in the retina in different glaucoma animal models, a chronic progressive model of glaucoma at different age stages and an acute IOP elevation induced by ischemia-reperfusion injury. Furthermore, as synucleins and H<sub>2</sub>S are involved in several mutual pathophysiological processes, we explored the correlation between H<sub>2</sub>S and synucleins in order to better understand the mechanism H<sub>2</sub>S's neuroprotective properties in experimental glaucoma.

Finally, building on previous results, we employed a mass spectrometry-based proteomics approach to analyze the retinal proteome, and bio-informatics to algorithmically generate protein connections, which allowed us to identify the most plausible signaling pathway alterations related to H<sub>2</sub>S's neuroprotective properties against ischemia-reperfusion injury in present study. Recognition of the intricate alterations is beneficial to the future treatment in glaucoma therapy with exogenous

H<sub>2</sub>S, and to show how these molecules play a pivotal role in restoring retinal homeostasis against I/R injury *in vivo*.

### **3.1 Hydrogen sulfide protects retinal ganglion cells against glaucomatous injury *in vitro* and *in vivo***

In recent years the role other factors like glutamate excitotoxicity, disorganized metabolism of NO, overproduction of ROS, increased endothelin-2 levels and reduction of ocular blood flow has also been discussed to play a key role in glaucoma pathology [4] .

In the central nervous system H<sub>2</sub>S has been found to facilitate long-term potentiation and regulate intracellular calcium concentration and pH level in brain cells. Antioxidant, anti-apoptotic, and anti-inflammatory effects of H<sub>2</sub>S have been found e.g. in Alzheimer's disease, Parkinson's disease, and vascular dementia. Furthermore an abnormal generation and metabolism of H<sub>2</sub>S has been seen involved in most of these neurodegenerative disorders [8]. However, the role of H<sub>2</sub>S in glaucoma remains obscure.

This part of the research focused on the expression changes of H<sub>2</sub>S in an experimental animal model of glaucoma and second its potential neuroprotective effect on retinal ganglion cells (RGC) in different glaucoma models *in vitro* and *in vivo* by addition of the slow-releasing H<sub>2</sub>S donor GYY4137.

Firstly, by mass spectrometry, we confirmed our hypothesis, the potential correlation between H<sub>2</sub>S and glaucoma. To do so, IOP was induced for 7 weeks in Sprague-Dawley rats by thermic occlusion of three episcleral veins, and pressure induced RGC loss was confirmed by anti-Brn3a immunostaining retinal wholemounts, which we repeatedly used to quantify RGCs in our further study. 3-mercaptopyruvate sulfurtransferase (3-MST) is clearly upregulated (3- fold), whilst house-keeping proteins remained unchanged in the proteomics approach, indicating some relation between experimental glaucomatous injury, neuronal impairment and the protein 3-MST. 3-MST is a mitochondrial protein,[53] which is known to synthesize H<sub>2</sub>S in mammalian tissue through its enzymatic action[54]. It has been reported that in brain injury, 3-MST can rapidly release H<sub>2</sub>S on stimulation[54]. We assume that in our study, 3-MST is upregulated due to the chronically elevated IOP, which suggesting H<sub>2</sub>S may also play an important role in pathology of glaucoma.

Meanwhile, as H<sub>2</sub>S has been excessively studied in cardiovascular system, and its vasodilating effect is widely recognized. We tested its potential to decrease IOP by dilating episcleral veins, by applying the solution of GYY4137, a slow-releasing H<sub>2</sub>S donor, as eye-drop in normotensive, untreated Sprague-Dawley rats. But it failed to decrease IOP in normotensive Sprague-Dawley rats.

Based on the mass spectrometry result, we proceeded with applying GYY4137 to retinal explants, under high-pressure condition or oxidative stress respectively. These in vitro studies proved H<sub>2</sub>S's dose-dependent protective property against glaucomatous retinal damage in vitro, it is well-known that the effect of H<sub>2</sub>S is depending upon the concentration. High concentrations (above 250 μmol/L) are supposed to exert toxic effects, while low ones have become regarded cell protective in recent years [55]. Furthermore, these studies also gave us optimal concentration to carry on in further in vivo studies.

With the positive results of in vitro studies, we moved on to test its function in vivo, in two different glaucomatous animal model, two different glaucomatous injuries were induced in experimental animals, by ischemia-reperfusion injury and optic nerve crush surgery. Considering the nature of two operations, GYY4137 was injected before inducing ischemia injury and immediately after optic nerve crush. Despite the differences in the mechanisms causing RGC loss in these two experimental in vivo models, both procedures caused significant RGC loss in operated eyes, but in those eyes with intravitreal injections of GYY4137, RGC survival was significantly higher than in untreated ones.

Endogenous H<sub>2</sub>S is produced in both vascular smooth muscle cells and endothelial cells, is involved in the regulation of many physiological processes, including the vascular tone. H<sub>2</sub>S produced in vascular smooth muscle cells can directly regulate the vascular tone in the autocrine manner. And H<sub>2</sub>S synthesized in endothelial cells can regulate independently of smooth muscle cells by mediators binding to endothelial cell receptors. Endothelial H<sub>2</sub>S signalling is up-regulated in some pathological conditions including ischemia-reperfusion injury [56]. As there is evidence suggesting that the major cause of blood flow reduction in glaucoma patients is rather a vascular dysregulation. H<sub>2</sub>S may have a specific protective potential by maintaining the regulation of the vascular tone.

As H<sub>2</sub>S has solid vasodilating effect, we tested it further on dilating fundus vessels, GYY4137 solution was injected intravitreally in experimental animals, which we inducted their IOP by thermic occlusion of three episcleral veins. GYY4137 was injected intravitreally after IOP was elevated for 6 weeks in animals, and fundus images were monitored by Optical Coherence Tomography (OCT) for over 3 hours after injection, and repeated again one week later (7 weeks after IOP elevated), fundus images obtained before thermic occlusion surgery served as baseline. H<sub>2</sub>S's action in the cardiovascular system has been excessively studied. Endogenously-generated and exogenously-administered H<sub>2</sub>S exerts a wide range of actions in vascular and myocardial cells including vasodilator/vasoconstrictor effects via modification of the smooth muscle tone. It has been reported that GYY4137 has protective effects against myocardial ischemia and reperfusion injury by attenuating oxidative stress and apoptosis [57].

Retinal blood flow in glaucomatous condition was restricted due to optic nerve injury and vasospasms caused by elevated IOP, enlarged vessel calibres will possibly improve the retinal perfusion and RGC survival under glaucomatous conditions in the long-term. These results present the possibility that H<sub>2</sub>S may act as a vasoregulator. H<sub>2</sub>S may exert its protective effect in glaucoma, by stabilizing ocular perfusion and easing ischemia-reperfusion injury. Whilst optic nerve injury has a direct impact on ganglion cell axons, this part of the mechanism by which H<sub>2</sub>S resulted in protection of RGCs against optic nerve injury remains unclear.

### **3.2 Hydrogen sulfide and $\beta$ -synuclein are involved and interlinked in the ageing glaucomatous retina**

Synuclein is a family of small proteins including  $\alpha$  (SNCA),  $\beta$  (SNCB) and  $\gamma$  (SNGG) synucleins[13,14] and is involved in various neurodegenerations in the CNS. Specifically, Mutations of SNCA play a central role in Parkinson's disease pathology, and misfolding and aggregation of SNCA directly linked to microglial activation, followed by inflammation and oxidative stress resulting in neurodegeneration[17]. Synucleins are also present in the retina and optic nerve[18] and are associated with glaucomatous alterations in the optic nerve[19]. SNCA autoantibody was found to be downregulated in serum and upregulated in aqueous humor of glaucoma patients [20] and intravitreal injection of SNCA antibodies is found to be neuroprotective in glaucoma animal model[21]. Because of the pivotal role of SNCA in neurodegeneration

in CNS. But its role in retina or glaucoma, as well as SNCB and SNCG's roles and functions in glaucoma is still sparse and remain to be thoroughly explored.

As synucleins and H<sub>2</sub>S are involved in several mutual pathophysiological processes, such as microglia activation, p53-mediated apoptosis, inflammatory response and free radical reactions. In second part of this doctoral thesis, we explore the correlation between H<sub>2</sub>S and synucleins in order to better understand the mechanism H<sub>2</sub>S's neuroprotective properties in experimental glaucoma.

In first part of the research, we chronically elevated IOP in adult female Sprague-Dawley rats for 7 weeks by cauterization of 3 episcleral veins. The IOP of all operated eyes increased significantly three weeks after the episcleral vein occlusion(EVO). Subsequent loss of RGCs is in agreement with thinning of the retinal nerve fiber layer, which suggests that EVO is a sufficient glaucoma model. As predominantly elder people are affected in glaucoma, older animals also showed a higher susceptibility to IOP elevation resulting in significant loss of RGCs and retinal nerve fiber layer (RNFL) thickness, while younger animals seemed to show resistance against mildly elevated IOP.

Secondly, we employed label-free quantification process following LC-ESI-LTQ-Orbitrap mass spectrometry to measure the abundance of synucleins in the retina. We found that SNCB has significantly more abundant in retina than other family members, furthermore, its level is significantly altered due to aging and elevated IOP, while the other two family members didn't show noticeable changes, which indicates that SNCB might have a more pivotal role to play in neurodegenerations in retina than other family members.

In physiological aging, the abundance of SNCB declines, which is correlated with the decreased RGC density and increased susceptibility to IOP. As under physiological conditions, SNCB is thought to be neuroprotective by functioning as a physiological inhibitor of SNCA and behaving as anti-aggregating agents. Studies in autopsy brains of PD, dementia with Lewy bodies and AD suggest that decreased amount of SNCB may lead to relative loss of protective functions of SNCB against neurotoxicity caused by SNCA[58]. Furthermore, downregulation of SNCB could occur not only in aggregation of SNCA, but also in other types of neurodegenerative disease[59].

SNCB's downregulation with aging increases RGC's susceptibility to glaucomatous assaults secondary to elevated IOP, such as elevated mechanic stress, insufficient retinal perfusion, and increased oxidative stress.

Under pathological conditions—chronic elevated IOP, SNCB in juvenile animals is downregulated, and the downregulation of SNCB is correlated with reduced RGC loss. While in aged animals, there is no significant alteration of SNCB in respond to assault, but more significant RGC loss.

According to mass spectrometry results, the amount of 3MST in rat retina is not significantly altered through the process of aging. In juvenile animals, 3MST was significantly upregulated in retina when exposed to elevated IOP over a period of 7 weeks, while no significant change is observed in old animals. Associating with the data from immunofluorescence staining of RGC, it suggests that upregulation of 3MST, a key H<sub>2</sub>S producing enzyme, is correlated with reduced RGC loss induced by elevated IOP. The self-regulation of H<sub>2</sub>S is decreased with aging. Therefore, we assume that downregulating SNCB and upregulating endogenous H<sub>2</sub>S level are neuroprotective against elevated IOP, and the function of regulating them is weakened with aging, which renders RGC's vulnerability.

Furthermore, acute IOP elevation is induced in juvenile animals, which led to significant RGC loss and SNCB downregulation. Although juvenile animals are more resilient than old animals to mildly elevated IOP (ca. 18mmHg), but when it reaches a threshold, acute IOP elevation (IOP ca. 55mmHg) led to a significant RGC loss. Downregulation of SNCB might be therefore a self-protective mechanism presenting from the beginning of the IOP elevation, but an exhaustion of the functional reserve eventually led to RGC loss.

Our data on SNCB abundance and RGC loss agrees with recent studies in retina, showing that the protective property of SNCB is exerted in a dose-dependent manner[13], which means overexpression and accumulation of SNCB increases oxidative stress and inflammatory responses, and furthermore promote the apoptosis, while lower concentrations of SNCB shows anti-apoptotic effect[26,60]. In various aspects of neurodegeneration, accumulation of SNCB is present, such as in dystrophic neurites in the hippocampal region in brains from PD and DLB patients, which suggest that accumulation of SNCB is involved in the axonal pathology[29]. SNCB was found to form toxic cytosolic inclusions in a similar manner to SNCA, and shares similar

toxicity mechanisms, including vesicular trafficking impairment and induction of oxidative stress[61]. Overexpression of SNCB in cultured primary cortical neurons led to cell loss and signs of metabolic impairment, in a similar manner to overexpressing SNCA neurons[62].

Treatments targeting SNCA to reduce its levels and toxicity have shown positive results in rescuing neuronal cells and halting the neurodegeneration process in preclinical studies[59,63]. For example, in our previous study, intravitreal injection of SNCA antibodies is found to be neuroprotective in glaucoma animal model [21].

Thus, it is reasonable to target the pathogenic SNCB and to decrease the intracellular SNCB as novel strategies for therapeutic intervention in neurodegeneration. Removal of pathogenic SNCB or reduce its abundancy may be effective to rescue neuron and halt the progression of glaucoma.

H<sub>2</sub>S has shown profound involvement in various retinal neuropathy processes, in previous studies by different groups including us, exogenous donors exhibited therapeutic potential in conditions of several retinal diseases[31,51]. The underlying mechanism, through which H<sub>2</sub>S exerts its neuroprotection, was partly attributed to its capability of vasorelaxation, anti-oxidative stress, neuroendocrine regulation, and inflammation suppression[10-12]. Nevertheless, H<sub>2</sub>S is involved in several mutual pathophysiological processes with SNCB[5,31-33].

Quantification of Brn3a positive RGCs showed that administration of exogenous H<sub>2</sub>S correlated positively with RGC survival improvement in acute IOP elevation. The mass spectrometric–assisted proteomics analysis of the retinal tissue demonstrated that administration of H<sub>2</sub>S also further downregulated SNCB.

Overall, the work described in this part of the dissertation suggested that downregulating SNCB partly contributes to the neuroprotection by H<sub>2</sub>S under glaucomatous condition. The extent to which internal mechanism and/or inflammatory factors, signalling pathways or the disruption of vascular function participate in the process is to be elucidated.

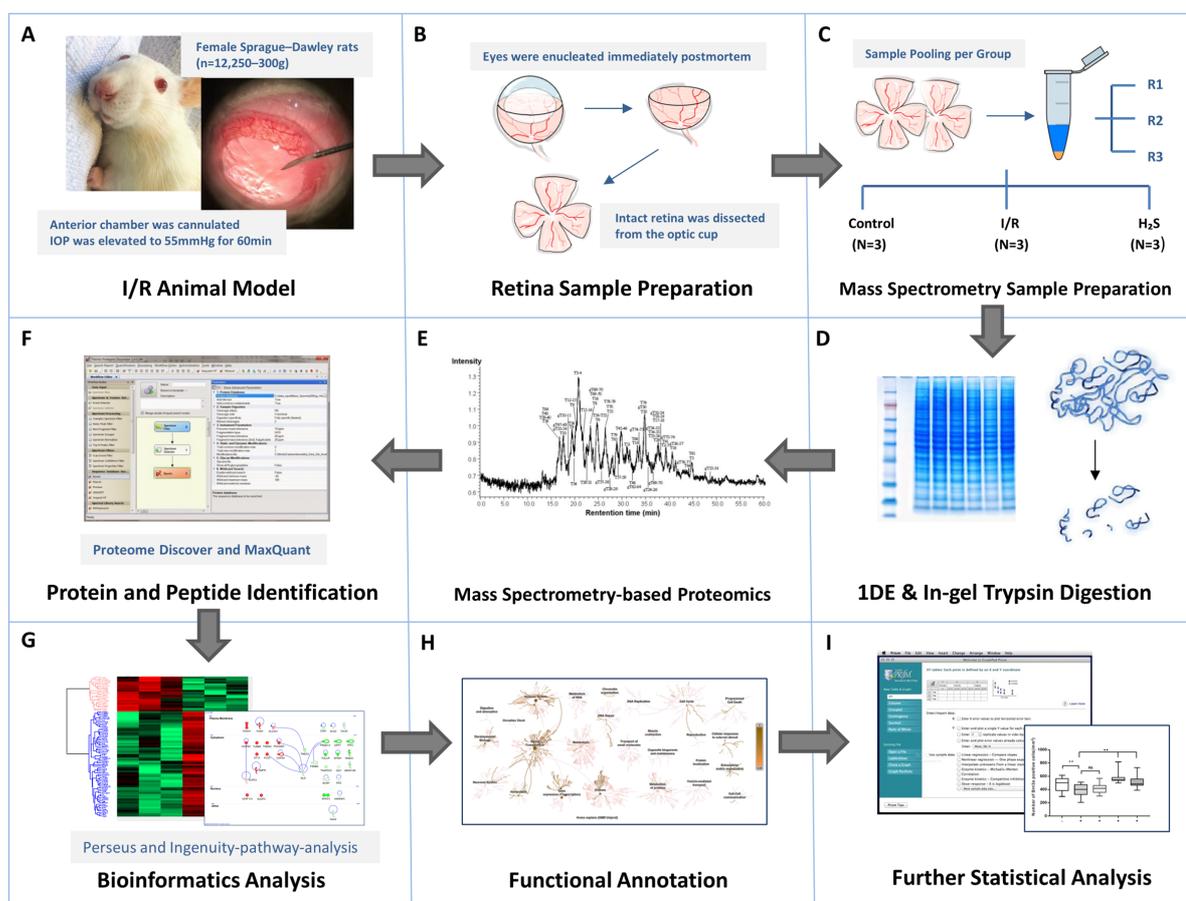
### **3.3 Proteomics reveals the potential protective mechanism of hydrogen sulfide on retinal ganglion cells in an ischemia/reperfusion injury animal model**

Ever since H<sub>2</sub>S has been recognized as the third endogenous gaseous signalling molecule and its role in various physiological and pathological processes has been explored. Building on previous results, we employed a mass spectrometry (MS)-based proteomics approach to analyze the retinal proteome, and bio-informatics to algorithmically generate protein connections, which allowed us to identify the most plausible signalling pathway alterations related to H<sub>2</sub>S's neuroprotective properties against ischemia-reperfusion injury in present study. Addressing the intricate alterations at the protein level can be attributed to the treatment with exogenous H<sub>2</sub>S, and to show how these molecules play a pivotal role in restoring retinal homeostasis against I/R injury in vivo.

We elevated the IOP of the Spargue-Dawley to 55mmHg for 60min to induce regional ischemic injury, followed by 24 hours of reperfusion. Compared to the contralateral control, I/R injury resulted in significant reduction in the number of RGC in the operated eyes, injection of GYY4137 prior to I/R injury significantly improved RGC survival.

MS-based discovery proteomics was utilized to identify and distinguish proteome profiles of rat retina undergoing I/R injury with or without pre-treatment of H<sub>2</sub>S. The overview of sampling and proteomics workflow used in this study is presented in Figure 1.

In total 1115 retinal proteins were identified with a false discovery rate (FDR) of 1%. Following I/R injury, the abundance of 48 proteins was significantly altered. Changes of the signaling pathways involved in mitochondrial homeostasis and function, calcium dysregulation, cytotoxicity regulation, ROS scavenging, neural transduction and vascular function were found. The proteins, which were regulated by H<sub>2</sub>S were categorized into two clusters: 1) a cluster 18 proteins, the abundance of which was significantly altered in I/R, and was then restored to normal or near normal level by H<sub>2</sub>S; 2) a cluster of 11 proteins, the abundance of which was significantly altered due to H<sub>2</sub>S administration compared to the CTRL group. Based on the results from the ingenuity pathway analysis, the signaling pathways regulated by H<sub>2</sub>S comprised iron homeostasis and ROS regulation, mitochondrial homeostasis and function, vasodilation and DNA repairing.



**Figure 1.** Workflow overview. I/R injury was induced in the left eyes of adult female Sprague-Dawley rats ( $n=12$ ), six of which received intravitreal injection of GYY4137, a H<sub>2</sub>S slow-releasing precursor, shortly before intervention. Animals were executed 24h after intervention, retinae were harvested immediately postmortem. Retinae from contralateral eyes were designated as controls. Retinal samples were immediately weighed and lysed by T-PER Tissue Protein Extraction Reagent and Bullet Blender Storm. Six samples of retinal protein from respective groups were pooled equally into three biological replicates after protein measurements, represented by R1, R2 and R3, and subsequently subjected to first dimensional gel electrophoresis. The protein bands were sliced and digested by trypsin prior to proteomic analysis by LC - ESI - MS/MS. The emerging datasets were subjected to robust bioinformatics analyses and functional annotations to identify the differential protein expressions and protein interaction networks.

### 3.3.1 Changes in Iron Homeostasis and ROS Regulation

Redox mechanisms are known to partially contribute to the protective properties of H<sub>2</sub>S in various tissues. Firstly, we observed that *Slc2a1* level is significantly downregulated in I/R and restored by H<sub>2</sub>S. *Slc2a1* facilitates docosahexaenoic acid (DHA), the oxidized form of vitamin C, transport across the inner blood-retina barrier and convert DHA to ascorbic acid and accumulates as ascorbic acid in the retina [64]. As *Slc2a1*

level is downregulated following I/R injury, Vitamin-C transport is downregulated accordingly. Vitamin C is the primary circulatory antioxidant to be quickly used and depleted when excessive ROS is generated by the pathophysiological pathways triggered by I/R injury, therefore sparing other endogenous antioxidants[65]. Oxidative stress promotes the oxidation of ascorbic acid to DHA, which is shown by studies to promote neuronal death under oxidative stress [66]. Downregulated Vitamin-C transport indicates that I/R injury impaired anti-oxidative property of the retina. H<sub>2</sub>S increased Vitamin-C transport, subsequently strengthening the anti-oxidative property of the retina as opposed to the ROS generated due to I/R.

Secondly, several proteins involved in iron homeostasis, such as biliverdin reductase A (*Bilvra*), *Cyb5r3* and *Glx3*, were differentially abundant due to I/R and/or H<sub>2</sub>S administration. Iron metabolism and regulation is crucial in mammals and is essential for physiological neuronal functions such as neural respiration and metabolic activities, myelin synthesis, neurotransmitter production and synaptic plasticity[67]. However its overload triggers axonal degeneration and neuronal cell death[68]. Excessive iron may catalyze the formation of highly reactive hydroxyl radicals, which eventually induce the accumulation of ROS [69,70]. Nevertheless, it is well known that the disproportionately increased brain Fe level is correlated with neurodegenerative diseases such as Parkinson's disease, Alzheimer's disease, Huntington's disease and neurodegeneration with brain Fe accumulation.

Most of the body Fe is contained within the protoporphyrin ring of heme [70]. Heme is essential for the survival of organisms, while being potentially toxic. The excessive release of heme from hemoproteins can lead to the generation of unfettered oxidative stress and programmed cell death [67]. Heme degradation is therefore important for maintaining iron homeostasis and preventing its cytotoxicity and oxidative stress [71].

The abundance of *Bilvra*, the main isoform of biliverdin reductase [72], is significantly increased due to I/R injury. Increased *Bilvra* level corresponds to upregulated heme degradation.

*Bilvra* is a pleiotropic enzyme primarily known for reducing heme-derived biliverdin into the powerful antioxidant and anti-nitrosative molecule bilirubin [73,74]. Bilirubin is able to inhibit free radical chain reactions and protects against oxidant-induced damage [75]. Studies reported that reducing *Bilvra* level in cell or knockout *Bilvra* gene in mice both led to increased oxidative stress [75,76]. On the other hand, increasing the levels

of *Blvra* is beneficial through an increase in intracellular bilirubin generation and direct signaling through cytoprotective pathways [72,77,78].

*Cyb5r3*, the principal reductase in mitochondria, is also significantly upregulated due to I/R injury, and can be restored by H<sub>2</sub>S. *Cyb5r3* is also a heme reductase and recycles oxidized heme (Fe<sup>3+</sup>) back to its reduced form (Fe<sup>2+</sup>) [79]. Upregulation of *Blvra* and *Cyb5r3* level following I/R injury is likely to confer self-protection against oxidative stress.

*Glrx3* level was the most significantly modulated by H<sub>2</sub>S when compared to the control group, while it was not impacted by I/R injury. Depletion of *Glrx3* in mammalian cells was associated with moderate deficiencies of cytosolic Fe-S cluster enzymes and evidence of altered iron homeostasis [80]. Overexpressed *Glrx3* can compensate for the lack of other reducing equivalents [81].

*Glrx3* belongs to the glutaredoxin family, it utilizes the reducing power of glutathione to maintain and regulate the cellular redox state and is essential for iron-sulfur (Fe-S) cluster assembly [82,83]. Iron-sulfur clusters are ancient, ubiquitous cofactors composed of iron and inorganic sulfur which participate in numerous biological processes [84-87].

In mammalian cells, *Glrx3* localizes to the cytosol and it has been proposed that it plays dual roles in iron trafficking and regulation [88,89]. *Glrx3* is a monothiol glutaredoxin, being able to form [2Fe-2S] cluster-bridged dimers and deliver Fe-S clusters to recipient proteins[90,91].

Detoxification of neural Fe overload is a potential therapeutic approach in the treatment of neurodegenerative disease. Based on the results of proteomic and IPA analysis, we assume that, when exposed to oxidative stress, cells increase their capability to reduce oxidized heme and to degrade it, which partially contributes to a self-protective mechanism that reduces the deleterious effects of free heme. H<sub>2</sub>S plays a similar reducing role as *Cyb5r3* in recycling oxidized heme to its reduced state; the abundance of *Cyb5r3* was thus not increased when H<sub>2</sub>S was present. Nevertheless, H<sub>2</sub>S has shown a strong association with *Glrx3*, which is able to regulate cellular iron homeostasis [92,93].

### 3.3.2 Changes in Retinal Metabolism, Mitochondrial Homeostasis and Function

As mentioned above, the abundance of *Slc2a1* is significantly downregulated in I/R and restored by H<sub>2</sub>S to near normal level, which also corresponds to increased HIF1 $\alpha$  signaling. *Slc2a1* abundance in HIF1 $\alpha$  signaling plays a key role in promoting neuronal survival by mediating the endogenous protective responses after hypoxia-ischemia. In glaucoma mice, decreased HIF1 $\alpha$  expression is observed to be correlated with RGC loss[94]. H<sub>2</sub>S regulates HIF1 factors in different patterns, depending on different cell type and experimental conditions. It increases expression of the HIF1 $\alpha$  in rat brain capillary endothelial cells and in mouse spinal-cord primary culture [95,96], and inhibits HIF1 activation in human hepatoma Hep3B cells, cervical carcinoma HeLa cells, and aortic smooth-muscle cells [97,98].

Our proteomic data indicate that H<sub>2</sub>S administration increased HIF1 $\alpha$  signaling. Upregulating expression of HIF1 $\alpha$  induces an increase of aerobic glycolysis, which transforms glucose to lactate and generates nicotinamide adenine dinucleotide (NAD<sup>+</sup>) [99]. HIF1 also directly induces pyruvate dehydrogenase kinase 1 [100], which phosphorylates the pyruvate dehydrogenase (PDH) complex, and consequently inhibits PDH complex from catalyzing pyruvate to form acetyl CoA. This fuels the mitochondrial tricarboxylic acid cycle (TCA) cycle, which provides the mitochondrial NADH needed to power electron transport. Hence, HIF1 $\alpha$  actively represses mitochondrial respiration.

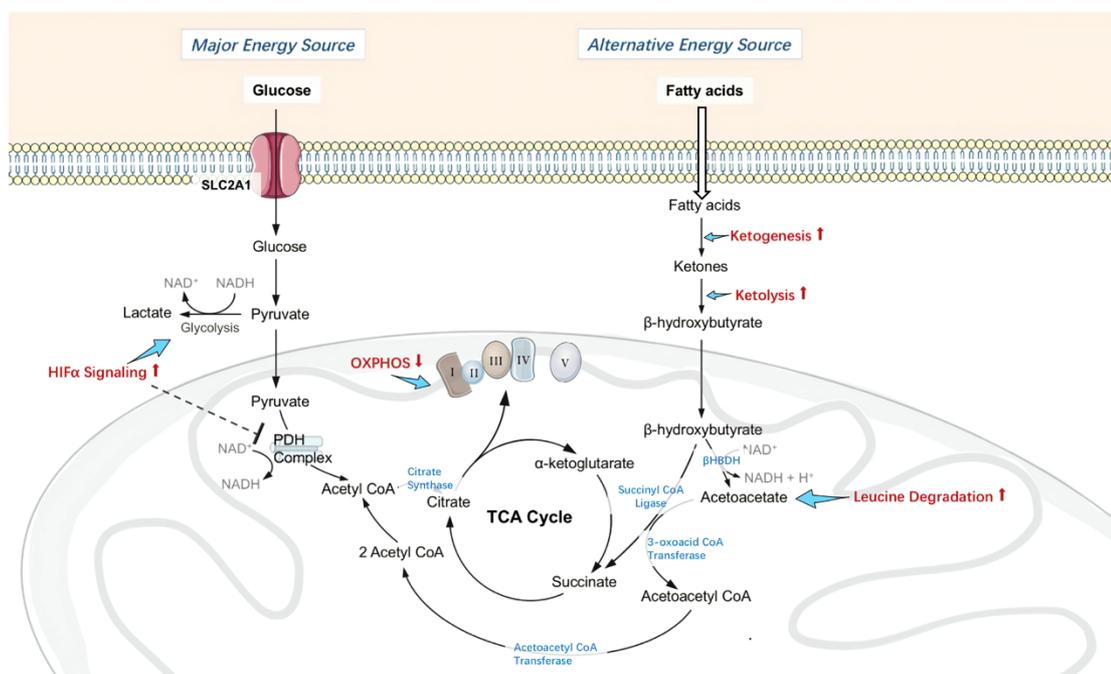
Furthermore, in this study, H<sub>2</sub>S reduced the abundance of NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 5 (*Ndufa5*), which subsequently also decreased mitochondrial respiration. *Ndufa5*, localizes to the inner mitochondrial membrane and is an accessory sub-unit of complex I [101]. Reducing *Ndufa5* suppresses complex I activity [102,103]. It is generally believed, that mitochondrial complex I deficiency contributes to the process of neurodegeneration because of reduced adenosine triphosphate (ATP) production, oxygen consumption and increased ROS production. However, complex I has been reported as a site of mitochondrial ROS generation [104,105], and actively suppressing its activity may actually be protective. Neuronal complex I deficiency is not correlated with the neurodegeneration, mitochondrial DNA damage in Parkinson's disease brain, nor in mice with a central nervous system knockout of *Ndufa5* [102,106]. It has been shown in neurodegenerative disorders that reversible inhibition of complex I can be protective against ischemia-reperfusion injury by modifying mitochondrial ROS production [107,108].

Studies strongly suggest that active suppression of the TCA cycle, mitochondrial respiration and ROS production, as well as augmentation of ATP levels, are crucial for the cell survival under I/R injury [100,109]. We assume that by suppressing mitochondrial respiration through upregulating HIF1- $\alpha$  and downregulating *Ndufa5* levels, H<sub>2</sub>S reduces mitochondrial oxidative phosphorylation, thereby repressing mitochondrial oxygen consumption and resulting in the relative increase of intracellular oxygen tension and limited ROS production under I/R.

As discussed above, mitochondrial PDH activity is attenuated by HIF1 $\alpha$  following H<sub>2</sub>S administration. Neurons usually prefer glucose for energy, but when mitochondrial metabolism of pyruvate is limited, ketone bodies can be oxidized and release the only alternative source of acetyl CoA for neurons [110]. Using ketone bodies as an alternative energy source has shown neuroprotective effects in Alzheimer's disease, Parkinson's disease and ischemic and traumatic brain injury [111-116]. The metabolic pathway that produces ketone bodies is ketogenesis. In this study, the signaling pathways most significantly downregulated by I/R injury and then restored by H<sub>2</sub>S were ketogenesis, ketolysis and leucine degradation.

In comparison to glucose, ketone bodies have a higher inherent energy[117]. As ketone bodies provide more energy per unit of oxygen than glucose [118,119], in states of metabolic stress such as I/R injury, ketone bodies are more energy-efficient fuel for neurons. They can also bypass the inhibited PDH complex and maintain the metabolites of TCA cycle, therefore permitted continued ATP production under ischemia.

Administration of H<sub>2</sub>S has shown evidence of actively suppressing oxidative phosphorylation and limiting the utilization of glucose as the energy source, therefore increasing intracellular oxygen tension under ischemia and limiting ROS production during reperfusion. Furthermore, H<sub>2</sub>S promotes the utilization of ketone bodies as an alternative energy source, which is more energy-efficient than glucose and maintains ATP production. H<sub>2</sub>S therefore enhanced ability of retinal neurons to withstand metabolic stress induced by I/R, which would normally deplete the resilience of the neurons and result in neuronal cell loss (see in Figure 2).



**Figure 2.** Changes in retinal metabolism, mitochondrial homeostasis and function. Administration of H<sub>2</sub>S actively inhibited pyruvate dehydrogenase (PDH) complex activity by upregulating HIF1 $\alpha$  signaling, therefore limited using glucose as energy source, and suppressed oxidative phosphorylation by inhibiting Complex I activity, consequently increased intracellular oxygen tension under ischemia and limited ROS production during reperfusion. Furthermore, H<sub>2</sub>S promotes the utilization of ketone bodies as an alternative energy source, which is more energy-efficient than glucose, thus permitted continuous adenosine triphosphate (ATP) production. H<sub>2</sub>S enhanced the ability of retinal neurons to withstand metabolic stress induced by I/R, therefore less neuronal cell loss.

### 3.3.3 Changes in Retinal Vascular Function

Decreased average blood flow in the retinal, optic nerve head and choroidal circulations is also demonstrated in glaucoma patients [120-122]. The “vascular theory” of glaucoma hypothesizes RGC loss as a consequence of insufficient blood supply [123]. Vasospasm and autoregulatory dysfunction have been postulated to reduce ocular blood flow [124]. Increased IOP has also been shown to reduce vascular caliber in rat retina while H<sub>2</sub>S acted as a vasodilator in our previous studies [51]. H<sub>2</sub>S can be endogenously generated in vascular smooth muscle cells. Its vasorelaxant potency is partially mediated by a functional endothelium [125].

However, I/R is known to cause endothelial dysfunction. In this study, eNOS signaling is downregulated following I/R injury. Endothelium-derived NO is a critical regulator of vascular homeostasis and tone [126]. NO continually regulates the diameter of blood vessels and maintains an anti-apoptotic environment in the vessel wall [127].

Downregulation of eNOS signaling is an indicator of impaired endothelial function, which results in diminished microcirculation and reduced response to endothelial-dependent vasodilators and vasoconstrictors. This causes dysregulated blood flow and loss of the endothelial barrier.

While eNOS signaling is downregulated, protein kinase A (PKA) signaling is significantly upregulated following I/R. PKA represents a signaling hub for a large variety of hormones, neurotransmitters and cytokines [128]. PKA has been shown to regulate different aspects of endothelial cell physiology [129-131], and to inhibit angiogenesis when activated [132,133]. Upregulation of PKA is likely a self-protective mechanism.

Furthermore, one of the most significantly upregulated proteins I/R, which can be restored by H<sub>2</sub>S to normal level, is *Cyb5r3*. It is the principal reductase involved in the mitochondrial amidoxime reducing component (mARC)-containing enzyme system [134]. In vascular smooth muscle, *Cyb5r3* functions as a soluble guanylyl cyclase (sGC) heme iron reductase, and is critical for vasodilation [135]. Through its reductase activity, *Cyb5r3* maintains NO-sGC-cGMP function [136]. The NO-sGC-cGMP pathway in aortic smooth muscle cell (SMC) is known to relax SMC and dilate vessels [136].

Hydrogen sulfide and the -SH anion reduce a variety of organic substrates [137]. It is reasonable to assume that H<sub>2</sub>S plays a similar reducing role as *Cyb5r3* in maintaining vascular relaxation and enable a less constricted vascular environment due to I/R.

Although the abundance of *DDX5* was not influenced by I/R injury, it is significantly more abundant following the administration of H<sub>2</sub>S. As *DDX5* regulates the expression of a number of key regulators upstream of the estrogen-receptor [138], estrogen-receptor signaling was also significantly upregulated in our PPI results.

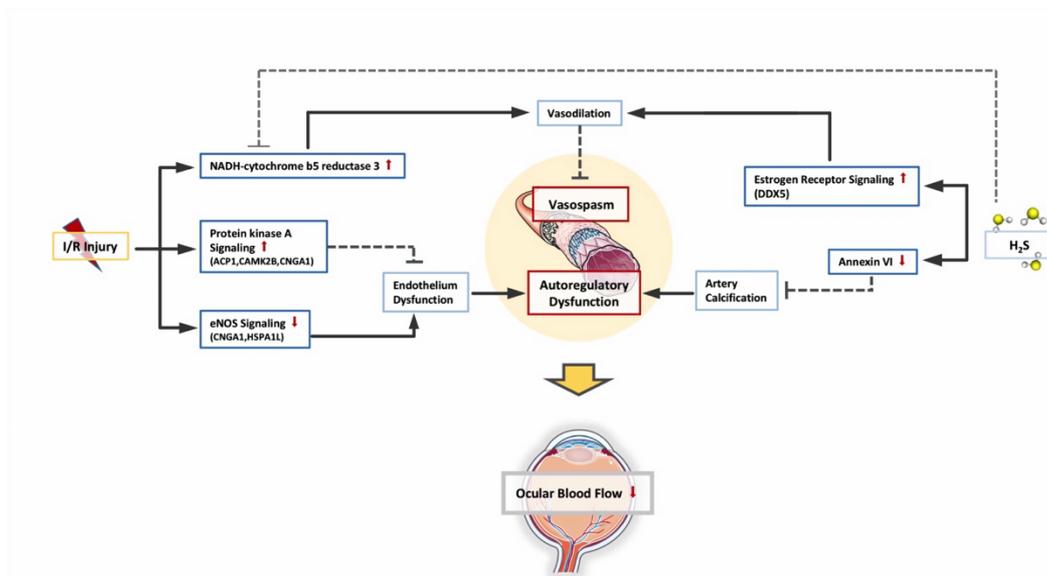
Increasing evidence suggests that estrogen exposure may have a neuroprotective effect on the progression of glaucoma and may alter its pathogenesis [139]. The vasodilation caused by estrogen and its effects on aqueous humor outflow may contribute [140]. These estrogen receptors are abundantly expressed throughout the eye, and in particular the retina and more specifically in the RGC [141,142]. In various CNS injuries and diseases, the activation of estrogen receptor signaling has shown neuroprotective effects via attenuation of neuro-inflammation and neurodegeneration

[143]. In different animal models of RGC degeneration, activation of estrogen receptor attenuated RGC apoptosis under acute elevated IOP [144]; impaired estrogen receptor expression is observed in a mouse model with early RGC apoptosis [145]; and in Leber's hereditary optic neuropathy cells activating estrogen receptors improve cell viability by reducing apoptosis [146]. Moreover, inhibition of estrogen synthesis is associated with IOP elevation and reduced RGC counts in female mice [147] and blockage of estrogen receptor signaling in rats intensifies impairment in visual function and retinal structure after ocular hypertension [148]. Although estrogen receptor signaling was not impacted by I/R injury in our study, activating estrogen receptor signaling via upregulating *DDX5* is still one of the possible pathways through which H<sub>2</sub>S exerts protective effects and partially contributes to its capability of vasorelaxation, anti-oxidative stress and inflammation suppression.

H<sub>2</sub>S also significantly downregulated annexin A6 compared to the control group, which is a calcium dependent phospholipid binding protein [149].

Annexins are important in Ca(2+)-induced neurotoxicity or neuroprotection[150]. Annexin A6 has been shown to play a central role in vascular remodelling processes[151]. It has been demonstrated that annexin A6 accumulates and converts exosomes in vascular smooth muscle cells into calcifications under calcium stress, which is known to be induced by I/R[151]. Vascular smooth muscle cells contribute significantly to physiological regulation of vascular tone and arterial blood pressure [152]. The presence of coronary artery calcification is significantly associated with raised IOP regardless of conventional cardiovascular risk factors [153]. By reducing the abundance of annexin A6, H<sub>2</sub>S potentially makes the calcification in vascular smooth muscle cells subside due to I/R, thus maintaining vascular autoregulation.

PKA signaling and *Cyb5r3* level is upregulated, likely serving as a self-protective mechanism to maintain endothelial function and vasodilation against I/R injury, while eNOS signaling is downregulated as an indication of endothelial dysfunction. Administration of H<sub>2</sub>S regulated the abundance of *Cyb5r3* and activated estrogen receptor signaling to facilitate vascular relaxation. H<sub>2</sub>S also reduced the abundance of annexin A6, which plays a central role in artery calcification. Combined together, H<sub>2</sub>S enabled a less constricted vascular environment in the retina, thereby resulting in better retinal perfusion to counteract I/R injury (see Figure 3).



**Figure 3.** Changes in retinal vascular function. Due to I/R, PKA (protein kinase A) signaling and NADH-cytochrome b5 reductase 3 (*Cyb5r3*) is upregulated as self-protective mechanism to maintain endothelial function and vasodilation, while eNOS signaling is downregulated as an indication of endothelial dysfunction. Administration of H<sub>2</sub>S activated estrogen receptor signaling to facilitate vascular relaxation. H<sub>2</sub>S also reduced the abundance of annexin A6, which plays a central role in artery calcification. Combined together, H<sub>2</sub>S protected the blood flow regulatory mechanisms and enabled a less constricted vascular environment in retina against I/R injury.

### 3.3.4 Changes in GABA Receptor Signaling

GABA receptor signaling is the most impacted signaling pathway by I/R and this can be restored by H<sub>2</sub>S. The *AP2M1* and *SLC32A1* levels are upregulated due to I/R, which correspond to upregulated GABA receptor signaling. Gama-Aminobutyric acid (GABA) is the main inhibitory neurotransmitter in the mammalian brain [154]. In the retina, GABA is similarly important in neural inhibition with approximately 40% of all retinal cells having been described as GABAergic [155]. GABA-containing vesicles release the neurotransmitter into the synaptic cleft to bind with GABA receptors present on the post-synaptic neuron [156].

*SLC32A1*, solute carrier family 32 (GABA vesicular transporter) member 1, is a transmembrane transporter which is responsible for the storage of GABA or glycine in synaptic vesicles [157]. *AP2M1* is highly expressed in the central nervous system [158,159], and encodes the  $\mu$ -subunit of the adaptor protein complex 2 (*AP-2*) [160]. *AP-2* regulates the neuronal surface levels of GABA and glutamate receptors [161,162]. An imbalance between excitatory and inhibitory synaptic transmission is

thought to contribute to excitotoxicity and neuronal cell death during ischemic insult. AP-2 was found to be crucial in reducing the loss of synaptic GABA receptors during simulated ischemia in the rat brain [163]. Increased abundance of *SLC32A1* and *AP2M1* suggests raised retinal GABA levels following I/R injury, which may occur in order to maintain excitatory/inhibitory balance, thereby improving the survival of RGCs, the latter being most susceptible to extracellular glutamate.

However, similar to glutamate, the functions of GABA are also controversial. There is evidence that the retina is extremely sensitive to abnormally accumulated GABA and its resulting toxicity, particularly in the presence of ischemia [164-166]. An antiepileptic drug, vigabatrin (Gama-vinyl GABA), exerts its effect by increasing GABA levels [167]; as the drug increases retinal GABA levels much more significantly than in the brain and retina has a lower tolerance to GABA toxicity than brain [165,166], one third of patients receiving vigabatrin present binasal visual field loss [168].

GABA also has a role in the regulation of vascular tone [165]. GABA receptors are known to interact with perivascular astrocytes which synthesise vasoactive materials [169,170]. In the rat retina, GABA is shown to be capable of eliciting both vasodilator and vasoconstrictor responses but endogenous GABA is unlikely to be an important regulator of resting vascular diameter and blood flow in the retina [171].

The exact role of GABA receptor signaling in this study is unclear but the results certainly raise intriguing questions about the involvement of GABA receptor signaling in neurodegeneration and the interaction between GABA signaling and H<sub>2</sub>S.

### 3.3.5 Changes in DNA Repair

The nucleotide excision repair (NER) pathway is at the top of the canonical pathways modulated by H<sub>2</sub>S, with a particularly significant upregulation of *COPS6* level. In combination with the ubiquitin (Ub) proteasome system (UPS) the COP9 signalosome controls the stability of cellular regulators [172] and regulates several important intracellular pathways, including DNA repair, cell cycle, developmental changes, and some aspects of immune responses [173-175].

*COPS6* has been shown to be cleaved by caspase 3 during apoptosis in vitro and in vivo [176] and this cleavage can be completely blocked by specific caspase 8 inhibitor [177]. Upregulation of *COPS6* led to upregulation of the NER pathway, one of the major

cellular DNA repair pathways for the removal of bulky helix lesions [178,179]. Enhancement of the NER pathway has shown protective effects in peripheral neurons both *in vitro* and *in vivo*[180]. The roles of DNA repair in neuronal cell survival and the response to aging and ROS such as that generated by mitochondrial respiration in glaucoma is of particular interest. The promotion of DNA repair by activation of the NER pathway through COP9 Signalosome Subunit upregulation could be an interesting aspect to explore in future studies.

Neurodegenerative diseases are characterized by decades of apparent normality, during which local deficits are compensated. Eventually the deficits become too accentuated and/or the compensatory mechanisms fail[181]. From the results we obtained in this study, I/R injury led to changes in ROS regulation, retinal metabolism, mitochondrial homeostasis and function, retinal vascular function and metal homeostasis, with over half of the altered pathways in the I/R group being self-protective mechanisms. H<sub>2</sub>S provided additional support where self-protective mechanisms fell short and demonstrated DNA repair properties, eventually providing neurons a better outcome against I/R injury.

### 3.4 Critical discussion

Admittedly there are limitations in this study. There are limitations of the study. Unfortunately, there is no proper glaucoma model, neither *in vitro* nor *in vivo*. Therefore, in the first part of this thesis, we used very different kind of glaucoma models to cover it all. Furthermore, the measurement of the vessels by means of OCT might be affected by not spotlessly clean measurements. But however within the framework of those limitations mentioned, we could see neuroprotective properties and vasodilatative effects, which are in accordance with previous findings.

Furthermore, in the second part of this doctoral thesis (manuscript II) we could confirm that downregulating SNCB partly contributes to the neuroprotection by H<sub>2</sub>S under glaucomatous condition. However, the extent to which internal mechanism and/or inflammatory factors, signaling pathways or the disruption of vascular function participate in the process is to be elucidated

The last part of this theses provides for the first time an overall insight into the mainstay retinal proteins and pivotal signaling pathways that interact with H<sub>2</sub>S to maintain retinal homeostasis against I/R injury.

Firstly, the results attained from the ischemia/reperfusion model are not necessarily valid in other glaucomatous models. There are various experimental models of glaucoma in the marketplace; each of them has specific pros and cons. It seems that degeneration caused by chronic elevated IOP mimics the characteristics of glaucoma better than I/R model. Clinical observations have demonstrated retinal vascular abnormalities and impaired blood flow at the optic nerve head which suggest ischemia plays a key role in the pathogenesis of glaucoma[182,183]. A pure ischemic lesion certainly cannot represent the glaucomatous damage, reperfusion injury is also present in glaucoma patients [184]. It is evident that IOP fluctuations are more damaging than a stabled increased IOP, and reduced circulation due to vascular dysregulation is more damaging than reduced circulation due to arteriosclerosis[185].

Furthermore, individual retinal samples were deliberately pooled into biological replicates to minimize inter-individual variations [186,187]. Due to the limitation of sample material and the variety of the signaling pathways H<sub>2</sub>S is involved in, the differentially expressed proteins could not be confirmed by a second technique in the present study design.

## **4 Conclusion and outlook**

The main objective of the present doctoral thesis was to evaluate the potential roles of H<sub>2</sub>S against glaucomatous injuries in retina and to unravel the underlying mechanisms of its neuroprotective effects. To answer these questions, we firstly employed a state-of-the-art LC-MS-based proteomic approach to analyse the expression changes of H<sub>2</sub>S in an experimental animal model of glaucoma and secondly H<sub>2</sub>S's potential neuroprotective effect on RGC towards elevated pressure in glaucoma models in vitro and in vivo by addition of the slow-releasing H<sub>2</sub>S donor GYY4137 (manuscript I). As second part of the thesis, we elucidate the potential roles and functions of synucleins in glaucomatous neuropathy, following this, to investigate the potential of H<sub>2</sub>S to regulate it, and to better understanding the mechanism of H<sub>2</sub>S in neuroprotection (manuscript II). As the final part of the thesis, a mass spectrometry-based proteomics approach was employed to analyze the retinal proteome, and bio-informatics to algorithmically generate protein connections, which allowed us to identify the most plausible signaling pathway alterations related to H<sub>2</sub>S's neuroprotective properties against ischemia-reperfusion injury (manuscript III).

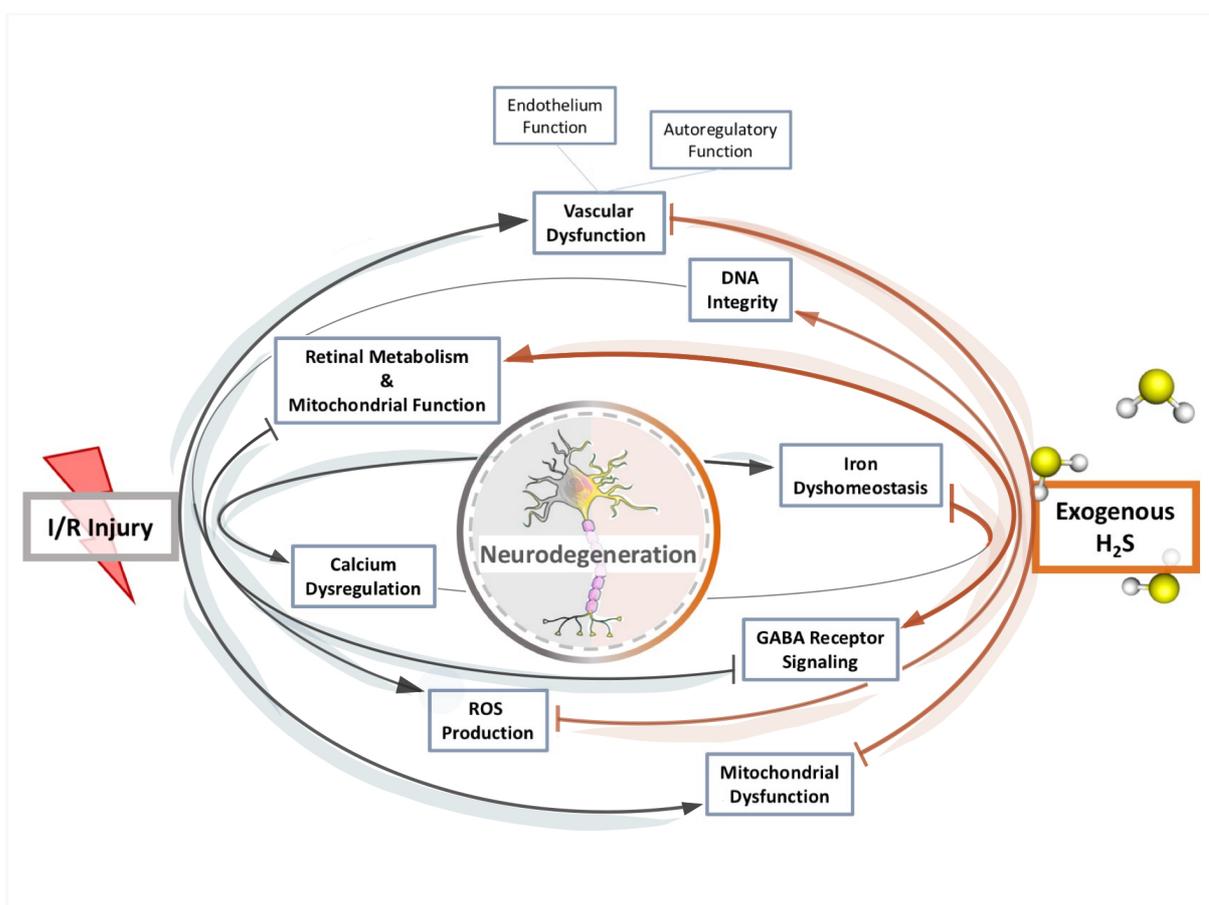
In the first part of the thesis, the results present the possibility that H<sub>2</sub>S plays a role in the pathology of glaucoma. And it exerts protective effect against glaucomatous injuries in different glaucoma animal models in vitro and in vivo. Furthermore, H<sub>2</sub>S may exert its protective effect in glaucoma, by stabilizing ocular perfusion and easing ischemia-reperfusion injury. Whilst optic nerve injury has a direct impact on ganglion cell axons, this part of the mechanism by which H<sub>2</sub>S resulted in protection of RGCs against optic nerve injury remains unclear.

Based on the previous results, in the second part of the doctoral thesis, we explore the underlying mechanism of H<sub>2</sub>S's neuroprotective property in relation with synucleins. Our data demonstrated that SNCB's downregulation with aging increases RGC's susceptibility to glaucomatous assaults secondary to elevated IOP, such as elevated mechanic stress, insufficient retinal perfusion, and increased oxidative stress.

Under pathological conditions—chronic elevated IOP, SNCB in juvenile animals is downregulated, and the downregulation of SNCB is correlated with reduced RGC loss. While in aged animals, there is no significant alteration of SNCB in respond to assault, but more significant RGC loss.

The self-regulation of H<sub>2</sub>S is decreased with aging. Therefore, we assume that downregulating SNCB and upregulating endogenous H<sub>2</sub>S level are neuroprotective against elevated IOP, and the function of regulating them is weakened with aging, which renders RGC's vulnerability.

Our data on SNCB abundance and RGC loss agrees with recent studies in retina, showing that the protective property of SNCB is exerted in a dose-dependent manner[13], which means overexpression and accumulation of SNCB increases oxidative stress and inflammatory responses, and furthermore promote the apoptosis, while lower concentrations of SNCB shows anti-apoptotic effect[26,60]. In various aspects of neurodegeneration, accumulation of SNCB is present.



**Figure 4.** Overview of altered signalling pathways due to I/R injury and H<sub>2</sub>S treatment.

It is reasonable to target the pathogenic SNCB and to decrease the intracellular SNCB as novel strategies for therapeutic intervention in neurodegeneration. Removal of pathogenic SNCB or reduce its abundancy may be effective to rescue neuron and halt the progression of glaucoma.

Overall, this part of the thesis suggested that downregulating SNCB partly contributes to the neuroprotection by H<sub>2</sub>S under glaucomatous condition. The extent to which internal mechanism and/or inflammatory factors, signalling pathways or the disruption of vascular function participate in the process is to be elucidated.

As final part of the present doctoral thesis, a state-of-the-art LC-MS-based proteomic approach is employed to provide a thorough overview of the retina proteome changes related to neuroprotective properties of H<sub>2</sub>S in retina, the main focus was to acquire a comprehensive perspective of the complex interaction between different proteins. But the findings presented here point to directions for further research, for example exploring the significance of these identified proteins and pathways and investigating related data from other glaucoma models. From the results we obtained from this part of the study, I/R injury led to changes in ROS regulation, retinal metabolism, mitochondrial homeostasis and function, retinal vascular function and metal homeostasis, with over half of the altered pathways in the I/R group being self-protective mechanisms. H<sub>2</sub>S provided additional support where self-protective mechanisms fell short and demonstrated DNA repair properties, eventually providing neurons a better outcome against I/R injury (Figure 4).

In conclusion, the present thesis provided important new insights into the complex neuroprotective effects of H<sub>2</sub>S in glaucoma and emphasized H<sub>2</sub>S as a promising and effective therapeutic tool for glaucoma.

## 5. Reference

1. Flaxman, S.R.; Bourne, R.R.A.; Resnikoff, S.; Ackland, P.; Braithwaite, T.; Cicinelli, M.V.; Das, A.; Jonas, J.B.; Keeffe, J.; Kempen, J.H., et al. Global causes of blindness and distance vision impairment 1990–2020: a systematic review and meta-analysis. *The Lancet Global Health* 2017, 5, e1221-e1234, doi:10.1016/s2214-109x(17)30393-5.
2. Almasieh, M.; Wilson, A.M.; Morquette, B.; Cueva Vargas, J.L.; Di Polo, A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res* 2012, 31, 152-181, doi:10.1016/j.preteyeres.2011.11.002.
3. Keating, D.J. Mitochondrial dysfunction, oxidative stress, regulation of exocytosis and their relevance to neurodegenerative diseases. *Journal of neurochemistry* 2008, 104, 298-305, doi:10.1111/j.1471-4159.2007.04997.x.
4. Doozandeh, A.; Yazdani, S. Neuroprotection in Glaucoma. *J Ophthalmic Vis Res* 2016, 11, 209-220, doi:10.4103/2008-322X.183923.
5. Tabassum, R.; Jeong, N.Y.; Jung, J. Therapeutic importance of hydrogen sulfide in age-associated neurodegenerative diseases. *Neural Regen Res* 2020, 15, 653-662, doi:10.4103/1673-5374.266911.
6. Kimura, H. Signaling molecules: hydrogen sulfide and polysulfide. *Antioxid Redox Signal* 2015, 22, 362-376, doi:10.1089/ars.2014.5869.
7. Elsey, D.J.; Fowkes, R.C.; Baxter, G.F. Regulation of cardiovascular cell function by hydrogen sulfide (H<sub>2</sub>S). *Cell Biochem Funct* 2010, 28, 95-106, doi:10.1002/cbf.1618.
8. Gong, Q.H.; Shi, X.R.; Hong, Z.Y.; Pan, L.L.; Liu, X.H.; Zhu, Y.Z. A new hope for neurodegeneration: possible role of hydrogen sulfide. *J Alzheimers Dis* 2011, 24 Suppl 2, 173-182, doi:10.3233/JAD-2011-110128.
9. Kimura, Y.; Kimura, H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 2004, 18, 1165-1167, doi:10.1096/fj.04-1815fje.
10. Yang, G.; Wu, L.; Jiang, B.; Yang, W.; Qi, J.; Cao, K.; Meng, Q.; Mustafa, A.K.; Mu, W.; Zhang, S., et al. H<sub>2</sub>S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 2008, 322, 587-590, doi:10.1126/science.1162667.
11. Kaneko, Y.; Kimura, Y.; Kimura, H.; Niki, I. L-cysteine inhibits insulin release from the pancreatic beta-cell: possible involvement of metabolic production of hydrogen sulfide, a novel gasotransmitter. *Diabetes* 2006, 55, 1391-1397, doi:10.2337/db05-1082.
12. Zhang, H.; Bhatia, M. Hydrogen sulfide: a novel mediator of leukocyte activation. *Immunopharmacol Immunotoxicol* 2008, 30, 631-645, doi:10.1080/08923970802278045.
13. da Costa, C.A.; Masliah, E.; Checler, F. Beta-synuclein displays an antiapoptotic p53-dependent phenotype and protects neurons from 6-hydroxydopamine-induced caspase 3

- activation: cross-talk with alpha-synuclein and implication for Parkinson's disease. *The Journal of biological chemistry* 2003, 278, 37330-37335, doi:10.1074/jbc.M306083200.
14. Surguchov, A. Intracellular Dynamics of Synucleins: "Here, There and Everywhere". *Int Rev Cell Mol Biol* 2015, 320, 103-169, doi:10.1016/bs.ircmb.2015.07.007.
15. Windisch, M.; Hutter-Paier, B.; Rockenstein, E.; Hashimoto, M.; Mallory, M.; Masliah, E. Development of a new treatment for Alzheimer's disease and Parkinson's disease using anti-aggregatory beta-synuclein-derived peptides. *J Mol Neurosci* 2002, 19, 63-69, doi:10.1007/s12031-002-0012-8.
16. Hattori, N.; Machida, Y.; Noda, K. [Pathogenesis of Parkinson's disease: a common pathway between alpha-synuclein and parkin and the mechanism of Lewy bodies formation]. *Rinsho Shinkeigaku* 2005, 45, 905-907.
17. Beraud, D.; Twomey, M.; Bloom, B.; Mittereder, A.; Ton, V.; Neitzke, K.; Chasovskikh, S.; Mhyre, T.R.; Maguire-Zeiss, K.A. alpha-Synuclein Alters Toll-Like Receptor Expression. *Front Neurosci* 2011, 5, 80, doi:10.3389/fnins.2011.00080.
18. Surguchov, A.; McMahan, B.; Masliah, E.; Surgucheva, I. Synucleins in ocular tissues. *J Neurosci Res* 2001, 65, 68-77, doi:10.1002/jnr.1129.
19. Surgucheva, I.; McMahan, B.; Ahmed, F.; Tomarev, S.; Wax, M.B.; Surguchov, A. Synucleins in glaucoma: implication of gamma-synuclein in glaucomatous alterations in the optic nerve. *J Neurosci Res* 2002, 68, 97-106, doi:10.1002/jnr.10198.
20. Boehm, N.; Wolters, D.; Thiel, U.; Lossbrand, U.; Wiegel, N.; Pfeiffer, N.; Grus, F.H. New insights into autoantibody profiles from immune privileged sites in the eye: a glaucoma study. *Brain Behav Immun* 2012, 26, 96-102, doi:10.1016/j.bbi.2011.07.241.
21. Teister, J.; Anders, F.; Beck, S.; Funke, S.; von Pein, H.; Prokosch, V.; Pfeiffer, N.; Grus, F. Decelerated neurodegeneration after intravitreal injection of alpha-synuclein antibodies in a glaucoma animal model. *Scientific reports* 2017, 7, 6260, doi:10.1038/s41598-017-06702-1.
22. Jensen, P.H.; Sorensen, E.S.; Petersen, T.E.; Gliemann, J.; Rasmussen, L.K. Residues in the synuclein consensus motif of the alpha-synuclein fragment, NAC, participate in transglutaminase-catalysed cross-linking to Alzheimer-disease amyloid beta A4 peptide. *Biochem J* 1995, 310 ( Pt 1), 91-94, doi:10.1042/bj3100091.
23. Tsigelny, I.F.; Bar-On, P.; Sharikov, Y.; Crews, L.; Hashimoto, M.; Miller, M.A.; Keller, S.H.; Platoshyn, O.; Yuan, J.X.; Masliah, E. Dynamics of alpha-synuclein aggregation and inhibition of pore-like oligomer development by beta-synuclein. *The FEBS journal* 2007, 274, 1862-1877, doi:10.1111/j.1742-4658.2007.05733.x.

24. Bohm, M.R.; Melkonyan, H.; Thanos, S. Life-time expression of the proteins peroxiredoxin, beta-synuclein, PARK7/DJ-1, and stathmin in the primary visual and primary somatosensory cortices in rats. *Front Neuroanat* 2015, 9, 16, doi:10.3389/fnana.2015.00016.
25. Bohm, M.R.; Mertsch, S.; Konig, S.; Spieker, T.; Thanos, S. Macula-less rat and macula-bearing monkey retinas exhibit common lifelong proteomic changes. *Neurobiol Aging* 2013, 34, 2659-2675, doi:10.1016/j.neurobiolaging.2013.04.020.
26. Hadrian, K.; Melkonyan, H.; Schlatt, S.; Wistuba, J.; Wasmuth, S.; Heiligenhaus, A.; Thanos, S.; Bohm, M.R.R. Age-related distribution and potential role of SNCB in topographically different retinal areas of the common marmoset *Callithrix jacchus*, including the macula. *Experimental eye research* 2019, 185, 107676, doi:10.1016/j.exer.2019.05.016.
27. Hashimoto, M.; Rockenstein, E.; Mante, M.; Mallory, M.; Masliah, E. beta-Synuclein inhibits alpha-synuclein aggregation: a possible role as an anti-parkinsonian factor. *Neuron* 2001, 32, 213-223, doi:10.1016/s0896-6273(01)00462-7.
28. Ji, H.; Liu, Y.E.; Jia, T.; Wang, M.; Liu, J.; Xiao, G.; Joseph, B.K.; Rosen, C.; Shi, Y.E. Identification of a breast cancer-specific gene, BCSG1, by direct differential cDNA sequencing. *Cancer Res* 1997, 57, 759-764.
29. Galvin, J.E.; Uryu, K.; Lee, V.M.; Trojanowski, J.Q. Axon pathology in Parkinson's disease and Lewy body dementia hippocampus contains alpha-, beta-, and gamma-synuclein. *Proceedings of the National Academy of Sciences of the United States of America* 1999, 96, 13450-13455, doi: 10.1073/pnas.96.23.13450.
30. Ninkina, N.; Peters, O.; Millership, S.; Salem, H.; van der Putten, H.; Buchman, V.L. Gamma-synucleinopathy: neurodegeneration associated with overexpression of the mouse protein. *Hum Mol Genet* 2009, 18, 1779-1794, doi:10.1093/hmg/ddp090.
31. Huang, S.; Huang, P.; Yu, H.; Lin, Z.; Liu, X.; Shen, X.; Guo, L.; Zhong, Y. Extracellular Signal-Regulated Kinase 1/2 Pathway Is Insufficiently Involved in the Neuroprotective Effect by Hydrogen Sulfide Supplement in Experimental Glaucoma. *Investigative ophthalmology & visual science* 2019, 60, 4346-4359, doi:10.1167/iovs.19-27507.
32. Kumar, M.; Sandhir, R. Hydrogen sulfide suppresses homocysteine-induced glial activation and inflammatory response. *Nitric Oxide* 2019, 90, 15-28, doi:10.1016/j.niox.2019.05.008.
33. Longhena, F.; Faustini, G.; Brembati, V.; Pizzi, M.; Bellucci, A. The good and bad of therapeutic strategies that directly target alpha-synuclein. *IUBMB Life* 2019, 10.1002/iub.2194, doi:10.1002/iub.2194.
34. Resnikoff, S.; Pascolini, D.; Mariotti, S.P.; Pokharel, G.P. Global magnitude of visual impairment caused by uncorrected refractive errors in 2004. *Bull World Health Organ* 2008, 86, 63-70, doi:10.2471/blt.07.041210.

35. Quigley, H.A. Number of people with glaucoma worldwide. *The British journal of ophthalmology* 1996, 80, 389-393, doi:10.1136/bjo.80.5.389.
36. Chang, E.E.; Goldberg, J.L. Glaucoma 2.0: neuroprotection, neuroregeneration, neuroenhancement. *Ophthalmology* 2012, 119, 979-986, doi:10.1016/j.ophtha.2011.11.003.
37. Johnson, E.C.; Morrison, J.C. Friend or foe? Resolving the impact of glial responses in glaucoma. *J Glaucoma* 2009, 18, 341-353, doi:10.1097/IJG.0b013e31818c6ef6.
38. Wilson, G.N.; Inman, D.M.; Dengler Crish, C.M.; Smith, M.A.; Crish, S.D. Early pro-inflammatory cytokine elevations in the DBA/2J mouse model of glaucoma. *J Neuroinflammation* 2015, 12, 176, doi:10.1186/s12974-015-0399-0.
39. Tezel, G. Oxidative stress in glaucomatous neurodegeneration: mechanisms and consequences. *Prog Retin Eye Res* 2006, 25, 490-513, doi:10.1016/j.preteyeres.2006.07.003.
40. Lansbury, P.T.; Lashuel, H.A. A century-old debate on protein aggregation and neurodegeneration enters the clinic. *Nature* 2006, 443, 774-779, doi:10.1038/nature05290.
41. Lowicka, E.; Beltowski, J. Hydrogen sulfide (H<sub>2</sub>S) - the third gas of interest for pharmacologists. *Pharmacological reports : PR* 2007, 59, 4-24.
42. Salloum, F.N. Hydrogen sulfide and cardioprotection--Mechanistic insights and clinical translatability. *Pharmacol Ther* 2015, 152, 11-17, doi:10.1016/j.pharmthera.2015.04.004.
43. Panthi, S.; Chung, H.J.; Jung, J.; Jeong, N.Y. Physiological Importance of Hydrogen Sulfide: Emerging Potent Neuroprotector and Neuromodulator. *Oxid Med Cell Longev* 2016, 2016, 9049782, doi:10.1155/2016/9049782.
44. Liu, Y.; Deng, Y.; Liu, H.; Yin, C.; Li, X.; Gong, Q. Corrigendum to: "Hydrogen sulfide ameliorates learning memory impairment in APP/PS1 transgenic mice: A novel mechanism mediated by the activation of Nrf2" [*Pharmacol. Biochem. Behav.* 150-151 (2016) 207-216]. *Pharmacol Biochem Behav* 2017, 153, 191, doi:10.1016/j.pbb.2016.12.004.
45. Liu, Y.; Deng, Y.; Liu, H.; Yin, C.; Li, X.; Gong, Q. Hydrogen sulfide ameliorates learning memory impairment in APP/PS1 transgenic mice: A novel mechanism mediated by the activation of Nrf2. *Pharmacol Biochem Behav* 2016, 150-151, 207-216, doi:10.1016/j.pbb.2016.11.002.
46. Xie, L.; Yu, S.; Yang, K.; Li, C.; Liang, Y. Hydrogen Sulfide Inhibits Autophagic Neuronal Cell Death by Reducing Oxidative Stress in Spinal Cord Ischemia Reperfusion Injury. *Oxid Med Cell Longev* 2017, 2017, 8640284, doi:10.1155/2017/8640284.
47. Sarookhani, M.R.; Haghdoost-Yazdi, H.; Sarbazi-Golezari, A.; Babayan-Tazehkand, A.; Rastgoo, N. Involvement of adenosine triphosphate-sensitive potassium channels in the neuroprotective activity of hydrogen sulfide in the 6-hydroxydopamine-induced animal

---

model of Parkinson's disease. *Behav Pharmacol* 2018, 29, 336-343, doi:10.1097/FBP.0000000000000358.

48. Sakamoto, K.; Suzuki, Y.; Kurauchi, Y.; Mori, A.; Nakahara, T.; Ishii, K. Hydrogen sulfide attenuates NMDA-induced neuronal injury via its anti-oxidative activity in the rat retina. *Experimental eye research* 2014, 120, 90-96, doi:10.1016/j.exer.2014.01.008.
49. Biermann, J.; Lagreze, W.A.; Schallner, N.; Schwer, C.I.; Goebel, U. Inhalative preconditioning with hydrogen sulfide attenuated apoptosis after retinal ischemia/reperfusion injury. *Mol Vis* 2011, 17, 1275-1286.
50. Si, Y.F.; Wang, J.; Guan, J.; Zhou, L.; Sheng, Y.; Zhao, J. Treatment with hydrogen sulfide alleviates streptozotocin-induced diabetic retinopathy in rats. *Br J Pharmacol* 2013, 169, 619-631, doi:10.1111/bph.12163.
51. Liu, H.; Anders, F.; Thanos, S.; Mann, C.; Liu, A.; Grus, F.H.; Pfeiffer, N.; Prokosch-Willing, V. Hydrogen Sulfide Protects Retinal Ganglion Cells Against Glaucomatous Injury In Vitro and In Vivo. *Investigative ophthalmology & visual science* 2017, 58, 5129-5141, doi:10.1167/iovs.17-22200.
52. Liu, H.; Mercieca, K.; Anders, F.; Prokosch, V. Hydrogen Sulfide and beta-Synuclein Are Involved and Interlinked in the Aging Glaucomatous Retina. *J Ophthalmol* 2020, 2020, 8642135, doi:10.1155/2020/8642135.
53. Predmore, B.L.; Lefer, D.J.; Gojon, G. Hydrogen sulfide in biochemistry and medicine. *Antioxid Redox Signal* 2012, 17, 119-140, doi:10.1089/ars.2012.4612.
54. Shibuya, N.; Tanaka, M.; Yoshida, M.; Ogasawara, Y.; Togawa, T.; Ishii, K.; Kimura, H. 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxidants & redox signaling* 2009, 11, 703-714, doi:10.1089/ARS.2008.2253.
55. Wallace, J.L. Hydrogen sulfide-releasing anti-inflammatory drugs. *Trends Pharmacol Sci* 2007, 28, 501-505, doi:10.1016/j.tips.2007.09.003.
56. Beltowski, J.; Jamroz-Wisniewska, A. Hydrogen sulfide and endothelium-dependent vasorelaxation. *Molecules* 2014, 19, 21183-21199, doi:10.3390/molecules191221183.
57. Meng, G.; Wang, J.; Xiao, Y.; Bai, W.; Xie, L.; Shan, L.; Moore, P.K.; Ji, Y. GYY4137 protects against myocardial ischemia and reperfusion injury by attenuating oxidative stress and apoptosis in rats. *J Biomed Res* 2015, 29, 203-213, doi:10.7555/JBR.28.20140037.
58. Rockenstein, E.; Hansen, L.A.; Mallory, M.; Trojanowski, J.Q.; Galasko, D.; Masliah, E. Altered expression of the synuclein family mRNA in Lewy body and Alzheimer's disease. *Brain Res* 2001, 914, 48-56, doi:10.1016/s0006-8993(01)02772-x.
59. Fujita, M.; Sekigawa, A.; Sekiyama, K.; Takamatsu, Y.; Hashimoto, M. Possible alterations in beta-Synuclein, the non-amyloidogenic homologue of alpha-Synuclein, during

- progression of sporadic alpha-synucleinopathies. *International journal of molecular sciences* 2012, 13, 11584-11592, doi:10.3390/ijms130911584.
60. Brockhaus, K.; Bohm, M.R.R.; Melkonyan, H.; Thanos, S. Age-related Beta-synuclein Alters the p53/Mdm2 Pathway and Induces the Apoptosis of Brain Microvascular Endothelial Cells In Vitro. *Cell Transplant* 2018, 27, 796-813, doi:10.1177/0963689718755706.
61. Tenreiro, S.; Rosado-Ramos, R.; Gerhardt, E.; Favretto, F.; Magalhaes, F.; Popova, B.; Becker, S.; Zweckstetter, M.; Braus, G.H.; Outeiro, T.F. Yeast reveals similar molecular mechanisms underlying alpha- and beta-synuclein toxicity. *Hum Mol Genet* 2016, 25, 275-290, doi:10.1093/hmg/ddv470.
62. Taschenberger, G.; Toloe, J.; Tereshchenko, J.; Akerboom, J.; Wales, P.; Benz, R.; Becker, S.; Outeiro, T.F.; Looger, L.L.; Bahr, M., et al. beta-synuclein aggregates and induces neurodegeneration in dopaminergic neurons. *Ann Neurol* 2013, 74, 109-118, doi:10.1002/ana.23905.
63. Tolmasov, M.; Djaldetti, R.; Lev, N.; Gilgun-Sherki, Y. Pathological and clinical aspects of alpha/beta synuclein in Parkinson's disease and related disorders. *Expert Rev Neurother* 2016, 16, 505-513, doi:10.1586/14737175.2016.1164600.
64. Hosoya, K.; Nakamura, G.; Akanuma, S.; Tomi, M.; Tachikawa, M. Dehydroascorbic acid uptake and intracellular ascorbic acid accumulation in cultured Muller glial cells (TR-MUL). *Neurochem Int* 2008, 52, 1351-1357, doi:10.1016/j.neuint.2008.02.001.
65. Spoelstra-de Man, A.M.E.; Elbers, P.W.G.; Oudemans-Van Straaten, H.M. Vitamin C: should we supplement? *Curr Opin Crit Care* 2018, 24, 248-255, doi:10.1097/MCC.0000000000000510.
66. Garcia-Krauss, A.; Ferrada, L.; Astuya, A.; Salazar, K.; Cisternas, P.; Martinez, F.; Ramirez, E.; Nualart, F. Dehydroascorbic Acid Promotes Cell Death in Neurons Under Oxidative Stress: a Protective Role for Astrocytes. *Mol Neurobiol* 2016, 53, 5847-5863, doi:10.1007/s12035-015-9497-3.
67. Gozzelino, R. The Pathophysiology of Heme in the Brain. *Curr Alzheimer Res* 2016, 13, 174-184, doi:10.2174/1567205012666150921103304.
68. Zhang, D.L.; Ghosh, M.C.; Rouault, T.A. The physiological functions of iron regulatory proteins in iron homeostasis - an update. *Front Pharmacol* 2014, 5, 124, doi:10.3389/fphar.2014.00124.
69. Gutteridge, J.M.; Halliwell, B. Free radicals and antioxidants in the year 2000. A historical look to the future. *Ann N Y Acad Sci* 2000, 899, 136-147, doi:10.1111/j.1749-6632.2000.tb06182.x.
70. Gozzelino, R.; Arosio, P. Iron Homeostasis in Health and Disease. *Int J Mol Sci* 2016, 17, doi:10.3390/ijms17010130.

- 
71. Alavi, F.S.; Gheidi, M.; Zahedi, M.; Safari, N.; Ryde, U. A novel mechanism of heme degradation to biliverdin studied by QM/MM and QM calculations. *Dalton Trans* 2018, 47, 8283-8291, doi:10.1039/c8dt00064f.
72. Kapitulnik, J.; Maines, M.D. Pleiotropic functions of biliverdin reductase: cellular signaling and generation of cytoprotective and cytotoxic bilirubin. *Trends Pharmacol Sci* 2009, 30, 129-137, doi:10.1016/j.tips.2008.12.003.
73. Stocker, R. Antioxidant activities of bile pigments. *Antioxid Redox Signal* 2004, 6, 841-849, doi:10.1089/ars.2004.6.841.
74. Barone, E.; Trombino, S.; Cassano, R.; Sgambato, A.; De Paola, B.; Di Stasio, E.; Picci, N.; Preziosi, P.; Mancuso, C. Characterization of the S-denitrosylating activity of bilirubin. *J Cell Mol Med* 2009, 13, 2365-2375, doi:10.1111/j.1582-4934.2009.00680.x.
75. Chen, W.; Maghzal, G.J.; Ayer, A.; Suarna, C.; Dunn, L.L.; Stocker, R. Absence of the biliverdin reductase-a gene is associated with increased endogenous oxidative stress. *Free Radic Biol Med* 2018, 115, 156-165, doi:10.1016/j.freeradbiomed.2017.11.020.
76. Sharma, N.; Tramutola, A.; Lanzillotta, C.; Arena, A.; Blarzino, C.; Cassano, T.; Butterfield, D.A.; Di Domenico, F.; Perluigi, M.; Barone, E. Loss of biliverdin reductase-A favors Tau hyper-phosphorylation in Alzheimer's disease. *Neurobiol Dis* 2019, 125, 176-189, doi:10.1016/j.nbd.2019.02.003.
77. O'Brien, L.; Hosick, P.A.; John, K.; Stec, D.E.; Hinds, T.D., Jr. Biliverdin reductase isozymes in metabolism. *Trends Endocrinol Metab* 2015, 26, 212-220, doi:10.1016/j.tem.2015.02.001.
78. Tudor, C.; Lerner-Marmarosh, N.; Engelborghs, Y.; Gibbs, P.E.; Maines, M.D. Biliverdin reductase is a transporter of haem into the nucleus and is essential for regulation of HO-1 gene expression by haematin. *Biochem J* 2008, 413, 405-416, doi:10.1042/BJ20080018.
79. Durgin, B.G.; Hahn, S.A.; Schmidt, H.M.; Miller, M.P.; Hafeez, N.; Mathar, I.; Freitag, D.; Sandner, P.; Straub, A.C. Loss of smooth muscle CYB5R3 amplifies angiotensin II-induced hypertension by increasing sGC heme oxidation. *JCI Insight* 2019, 4, doi:10.1172/jci.insight.129183.
80. Haunhorst, P.; Hanschmann, E.M.; Brautigam, L.; Stehling, O.; Hoffmann, B.; Muhlenhoff, U.; Lill, R.; Berndt, C.; Lillig, C.H. Crucial function of vertebrate glutaredoxin 3 (PICOT) in iron homeostasis and hemoglobin maturation. *Mol Biol Cell* 2013, 24, 1895-1903, doi:10.1091/mbc.E12-09-0648.
81. Aslund, F.; Ehn, B.; Miranda-Vizueté, A.; Pueyo, C.; Holmgren, A. Two additional glutaredoxins exist in *Escherichia coli*: glutaredoxin 3 is a hydrogen donor for ribonucleotide reductase in a thioredoxin/glutaredoxin 1 double mutant. *Proc Natl Acad Sci U S A* 1994, 91, 9813-9817, doi:10.1073/pnas.91.21.9813.

82. Lillig, C.H.; Berndt, C.; Holmgren, A. Glutaredoxin systems. *Biochim Biophys Acta* 2008, 1780, 1304-1317, doi:10.1016/j.bbagen.2008.06.003.
83. Qi, W.; Li, J.; Chain, C.Y.; Pasquevich, G.A.; Pasquevich, A.F.; Cowan, J.A. Glutathione complexed Fe-S centers. *J Am Chem Soc* 2012, 134, 10745-10748, doi:10.1021/ja302186j.
84. Netz, D.J.; Stith, C.M.; Stumpfig, M.; Kopf, G.; Vogel, D.; Genau, H.M.; Stodola, J.L.; Lill, R.; Burgers, P.M.; Pierik, A.J. Eukaryotic DNA polymerases require an iron-sulfur cluster for the formation of active complexes. *Nat Chem Biol* 2011, 8, 125-132, doi:10.1038/nchembio.721.
85. Rudolf, J.; Makrantonis, V.; Ingledew, W.J.; Stark, M.J.; White, M.F. The DNA repair helicases XPD and FancJ have essential iron-sulfur domains. *Mol Cell* 2006, 23, 801-808, doi:10.1016/j.molcel.2006.07.019.
86. Kispal, G.; Sipos, K.; Lange, H.; Fekete, Z.; Bedekovics, T.; Janaky, T.; Bassler, J.; Aguilar Netz, D.J.; Balk, J.; Rotte, C., et al. Biogenesis of cytosolic ribosomes requires the essential iron-sulphur protein Rli1p and mitochondria. *EMBO J* 2005, 24, 589-598, doi:10.1038/sj.emboj.7600541.
87. Maio, N.; Rouault, T.A. Iron-sulfur cluster biogenesis in mammalian cells: New insights into the molecular mechanisms of cluster delivery. *Biochim Biophys Acta* 2015, 1853, 1493-1512, doi:10.1016/j.bbamcr.2014.09.009.
88. Wachnowsky, C.; Fidai, I.; Cowan, J.A. Cytosolic iron-sulfur cluster transfer-a proposed kinetic pathway for reconstitution of glutaredoxin 3. *FEBS Lett* 2016, 590, 4531-4540, doi:10.1002/1873-3468.12491.
89. Muhlenhoff, U.; Molik, S.; Godoy, J.R.; Uzarska, M.A.; Richter, N.; Seubert, A.; Zhang, Y.; Stubbe, J.; Pierrel, F.; Herrero, E., et al. Cytosolic monothiol glutaredoxins function in intracellular iron sensing and trafficking via their bound iron-sulfur cluster. *Cell Metab* 2010, 12, 373-385, doi:10.1016/j.cmet.2010.08.001.
90. Iwema, T.; Picciocchi, A.; Traore, D.A.; Ferrer, J.L.; Chauvat, F.; Jacquamet, L. Structural basis for delivery of the intact [Fe<sub>2</sub>S<sub>2</sub>] cluster by monothiol glutaredoxin. *Biochemistry* 2009, 48, 6041-6043, doi:10.1021/bi900440m.
91. Sen, S.; Rao, B.; Wachnowsky, C.; Cowan, J.A. Cluster exchange reactivity of [2Fe-2S] cluster-bridged complexes of BOLA3 with monothiol glutaredoxins. *Metallomics* 2018, 10, 1282-1290, doi:10.1039/c8mt00128f.
92. Xia, H.; Li, B.; Zhang, Z.; Wang, Q.; Qiao, T.; Li, K. Human glutaredoxin 3 can bind and effectively transfer [4Fe-4S] cluster to apo-iron regulatory protein 1. *Biochem Biophys Res Commun* 2015, 465, 620-624, doi:10.1016/j.bbrc.2015.08.073.
93. Li, H.; Mapolelo, D.T.; Dingra, N.N.; Naik, S.G.; Lees, N.S.; Hoffman, B.M.; Riggs-Gelasco, P.J.; Huynh, B.H.; Johnson, M.K.; Outten, C.E. The yeast iron regulatory proteins

- Grx3/4 and Fra2 form heterodimeric complexes containing a [2Fe-2S] cluster with cysteinyl and histidyl ligation. *Biochemistry* 2009, 48, 9569-9581, doi:10.1021/bi901182w.
94. Zhang, L.Q.; Cui, H.; Yu, Y.B.; Shi, H.Q.; Zhou, Y.; Liu, M.J. MicroRNA-141-3p inhibits retinal neovascularization and retinal ganglion cell apoptosis in glaucoma mice through the inactivation of Docking protein 5-dependent mitogen-activated protein kinase signaling pathway. *J Cell Physiol* 2019, 234, 8873-8887, doi:10.1002/jcp.27549.
95. Wu, B.; Teng, H.; Zhang, L.; Li, H.; Li, J.; Wang, L.; Li, H. Interaction of Hydrogen Sulfide with Oxygen Sensing under Hypoxia. *Oxid Med Cell Longev* 2015, 2015, 758678, doi:10.1155/2015/758678.
96. Greco, V.; Spalloni, A.; Corasolla Carregari, V.; Pieroni, L.; Persichilli, S.; Mercuri, N.B.; Urbani, A.; Longone, P. Proteomics and Toxicity Analysis of Spinal-Cord Primary Cultures upon Hydrogen Sulfide Treatment. *Antioxidants (Basel)* 2018, 7, doi:10.3390/antiox7070087.
97. Liu, X.; Pan, L.; Zhuo, Y.; Gong, Q.; Rose, P.; Zhu, Y. Hypoxia-inducible factor-1alpha is involved in the pro-angiogenic effect of hydrogen sulfide under hypoxic stress. *Biol Pharm Bull* 2010, 33, 1550-1554, doi:10.1248/bpb.33.1550.
98. Kai, S.; Tanaka, T.; Daijo, H.; Harada, H.; Kishimoto, S.; Suzuki, K.; Takabuchi, S.; Takenaga, K.; Fukuda, K.; Hirota, K. Hydrogen sulfide inhibits hypoxia- but not anoxia-induced hypoxia-inducible factor 1 activation in a von hippel-lindau- and mitochondria-dependent manner. *Antioxid Redox Signal* 2012, 16, 203-216, doi:10.1089/ars.2011.3882.
99. Xu, J.; Li, J.; Yu, Z.; Rao, H.; Wang, S.; Lan, H. HMGB1 promotes HLF-1 proliferation and ECM production through activating HIF1-alpha-regulated aerobic glycolysis. *Pulm Pharmacol Ther* 2017, 45, 136-141, doi:10.1016/j.pupt.2017.05.015.
100. Kim, J.W.; Tchernyshyov, I.; Semenza, G.L.; Dang, C.V. HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. *Cell Metab* 2006, 3, 177-185, doi:10.1016/j.cmet.2006.02.002.
101. Rak, M.; Rustin, P. Supernumerary subunits NDUFA3, NDUFA5 and NDUFA12 are required for the formation of the extramembrane arm of human mitochondrial complex I. *FEBS Lett* 2014, 588, 1832-1838, doi:10.1016/j.febslet.2014.03.046.
102. Peralta, S.; Torraco, A.; Wenz, T.; Garcia, S.; Diaz, F.; Moraes, C.T. Partial complex I deficiency due to the CNS conditional ablation of Ndufa5 results in a mild chronic encephalopathy but no increase in oxidative damage. *Hum Mol Genet* 2014, 23, 1399-1412, doi:10.1093/hmg/ddt526.
103. Nie, H.; Yu, X.; He, H.; Zhou, L.; Li, Q.; Song, C.; Wang, D.; Ren, T.; Chen, Z.; Huang, H., et al. Hepatocyte miR-33a mediates mitochondrial dysfunction and hepatosteatosis by suppressing NDUFA5. *J Cell Mol Med* 2018, 22, 6285-6293, doi:10.1111/jcmm.13918.

- 
104. Liu, Y.; Fiskum, G.; Schubert, D. Generation of reactive oxygen species by the mitochondrial electron transport chain. *J Neurochem* 2002, 80, 780-787, doi:10.1046/j.0022-3042.2002.00744.x.
105. Kudin, A.P.; Bimpong-Buta, N.Y.; Vielhaber, S.; Elger, C.E.; Kunz, W.S. Characterization of superoxide-producing sites in isolated brain mitochondria. *J Biol Chem* 2004, 279, 4127-4135, doi:10.1074/jbc.M310341200.
106. Fiones, I.H.; Fernandez-Vizarra, E.; Lykouri, M.; Brakedal, B.; Skeie, G.O.; Miletic, H.; Lilleng, P.K.; Alves, G.; Tysnes, O.B.; Haugarvoll, K., et al. Neuronal complex I deficiency occurs throughout the Parkinson's disease brain, but is not associated with neurodegeneration or mitochondrial DNA damage. *Acta Neuropathol* 2018, 135, 409-425, doi:10.1007/s00401-017-1794-7.
107. Burwell, L.S.; Brookes, P.S. Mitochondria as a target for the cardioprotective effects of nitric oxide in ischemia-reperfusion injury. *Antioxid Redox Signal* 2008, 10, 579-599, doi:10.1089/ars.2007.1845.
108. Bordt, E.A.; Clerc, P.; Roelofs, B.A.; Saladino, A.J.; Tretter, L.; Adam-Vizi, V.; Cherok, E.; Khalil, A.; Yadava, N.; Ge, S.X., et al. The Putative Drp1 Inhibitor mdivi-1 Is a Reversible Mitochondrial Complex I Inhibitor that Modulates Reactive Oxygen Species. *Dev Cell* 2017, 40, 583-594 e586, doi:10.1016/j.devcel.2017.02.020.
109. Papandreou, I.; Cairns, R.A.; Fontana, L.; Lim, A.L.; Denko, N.C. HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. *Cell Metab* 2006, 3, 187-197, doi:10.1016/j.cmet.2006.01.012.
110. Gasior, M.; Rogawski, M.A.; Hartman, A.L. Neuroprotective and disease-modifying effects of the ketogenic diet. *Behav Pharmacol* 2006, 17, 431-439, doi:10.1097/00008877-200609000-00009.
111. Van der Auwera, I.; Wera, S.; Van Leuven, F.; Henderson, S.T. A ketogenic diet reduces amyloid beta 40 and 42 in a mouse model of Alzheimer's disease. *Nutr Metab (Lond)* 2005, 2, 28, doi:10.1186/1743-7075-2-28.
112. Patel, N.V.; Gordon, M.N.; Connor, K.E.; Good, R.A.; Engelman, R.W.; Mason, J.; Morgan, D.G.; Morgan, T.E.; Finch, C.E. Caloric restriction attenuates A $\beta$ -deposition in Alzheimer transgenic models. *Neurobiol Aging* 2005, 26, 995-1000, doi:10.1016/j.neurobiolaging.2004.09.014.
113. Lim, G.P.; Calon, F.; Morihara, T.; Yang, F.; Teter, B.; Ubeda, O.; Salem, N., Jr.; Frautschy, S.A.; Cole, G.M. A diet enriched with the omega-3 fatty acid docosahexaenoic acid reduces amyloid burden in an aged Alzheimer mouse model. *J Neurosci* 2005, 25, 3032-3040, doi:10.1523/JNEUROSCI.4225-04.2005.

- 
114. Holmer, H.K.; Keyghobadi, M.; Moore, C.; Menashe, R.A.; Meshul, C.K. Dietary restriction affects striatal glutamate in the MPTP-induced mouse model of nigrostriatal degeneration. *Synapse* 2005, 57, 100-112, doi:10.1002/syn.20163.
115. Tieu, K.; Perier, C.; Caspersen, C.; Teismann, P.; Wu, D.C.; Yan, S.D.; Naini, A.; Vila, M.; Jackson-Lewis, V.; Ramasamy, R., et al. D-beta-hydroxybutyrate rescues mitochondrial respiration and mitigates features of Parkinson disease. *J Clin Invest* 2003, 112, 892-901, doi:10.1172/JCI18797.
116. Prins, M.L.; Fujima, L.S.; Hovda, D.A. Age-dependent reduction of cortical contusion volume by ketones after traumatic brain injury. *J Neurosci Res* 2005, 82, 413-420, doi:10.1002/jnr.20633.
117. Cahill, G.F., Jr.; Veech, R.L. Ketoacids? Good medicine? *Trans Am Clin Climatol Assoc* 2003, 114, 149-161; discussion 162-143.
118. Kashiwaya, Y.; Sato, K.; Tsuchiya, N.; Thomas, S.; Fell, D.A.; Veech, R.L.; Passonneau, J.V. Control of glucose utilization in working perfused rat heart. *J Biol Chem* 1994, 269, 25502-25514.
119. Sato, K.; Kashiwaya, Y.; Keon, C.A.; Tsuchiya, N.; King, M.T.; Radda, G.K.; Chance, B.; Clarke, K.; Veech, R.L. Insulin, ketone bodies, and mitochondrial energy transduction. *FASEB J* 1995, 9, 651-658, doi:10.1096/fasebj.9.8.7768357.
120. Wolf, S.; Arend, O.; Sponsel, W.E.; Schulte, K.; Cantor, L.B.; Reim, M. Retinal hemodynamics using scanning laser ophthalmoscopy and hemorheology in chronic open-angle glaucoma. *Ophthalmology* 1993, 100, 1561-1566, doi:10.1016/s0161-6420(93)31444-2.
121. Harju, M.; Vesti, E. Blood flow of the optic nerve head and peripapillary retina in exfoliation syndrome with unilateral glaucoma or ocular hypertension. *Graefes Arch Clin Exp Ophthalmol* 2001, 239, 271-277, doi:10.1007/s004170100269.
122. Yin, Z.Q.; Vaegan; Millar, T.J.; Beaumont, P.; Sarks, S. Widespread choroidal insufficiency in primary open-angle glaucoma. *J Glaucoma* 1997, 6, 23-32.
123. Flammer, J.; Mozaffarieh, M. Autoregulation, a balancing act between supply and demand. *Can J Ophthalmol* 2008, 43, 317-321, doi:10.3129/i08-056.
124. Galassi, F.; Giambene, B.; Varriale, R. Systemic vascular dysregulation and retrobulbar hemodynamics in normal-tension glaucoma. *Invest Ophthalmol Vis Sci* 2011, 52, 4467-4471, doi:10.1167/iovs.10-6710.
125. Zhao, W.; Wang, R. H<sub>2</sub>S-induced vasorelaxation and underlying cellular and molecular mechanisms. *Am J Physiol Heart Circ Physiol* 2002, 283, H474-480, doi:10.1152/ajpheart.00013.2002.

- 
126. Siragusa, M.; Fleming, I. The eNOS signalosome and its link to endothelial dysfunction. *Pflugers Arch* 2016, 468, 1125-1137, doi:10.1007/s00424-016-1839-0.
127. Sessa, W.C. eNOS at a glance. *J Cell Sci* 2004, 117, 2427-2429, doi:10.1242/jcs.01165.
128. Tasken, K.; Aandahl, E.M. Localized effects of cAMP mediated by distinct routes of protein kinase A. *Physiol Rev* 2004, 84, 137-167, doi:10.1152/physrev.00021.2003.
129. Aslam, M.; Hartel, F.V.; Arshad, M.; Gunduz, D.; Abdallah, Y.; Sauer, H.; Piper, H.M.; Noll, T. cAMP/PKA antagonizes thrombin-induced inactivation of endothelial myosin light chain phosphatase: role of CPI-17. *Cardiovasc Res* 2010, 87, 375-384, doi:10.1093/cvr/cvq065.
130. Collins, C.; Osborne, L.D.; Guilluy, C.; Chen, Z.; O'Brien, E.T., 3rd; Reader, J.S.; Burrige, K.; Superfine, R.; Tzima, E. Haemodynamic and extracellular matrix cues regulate the mechanical phenotype and stiffness of aortic endothelial cells. *Nat Commun* 2014, 5, 3984, doi:10.1038/ncomms4984.
131. Goldfinger, L.E.; Tzima, E.; Stockton, R.; Kiosses, W.B.; Kinbara, K.; Tkachenko, E.; Gutierrez, E.; Groisman, A.; Nguyen, P.; Chien, S., et al. Localized alpha4 integrin phosphorylation directs shear stress-induced endothelial cell alignment. *Circ Res* 2008, 103, 177-185, doi:10.1161/CIRCRESAHA.108.176354.
132. Bakre, M.M.; Zhu, Y.; Yin, H.; Burton, D.W.; Terkeltaub, R.; Deftos, L.J.; Varner, J.A. Parathyroid hormone-related peptide is a naturally occurring, protein kinase A-dependent angiogenesis inhibitor. *Nat Med* 2002, 8, 995-1003, doi:10.1038/nm753.
133. Jin, H.; Garmy-Susini, B.; Avraamides, C.J.; Stoletov, K.; Klemke, R.L.; Varner, J.A. A PKA-Csk-pp60Src signaling pathway regulates the switch between endothelial cell invasion and cell-cell adhesion during vascular sprouting. *Blood* 2010, 116, 5773-5783, doi:10.1182/blood-2010-07-296210.
134. Plitzko, B.; Havemeyer, A.; Bork, B.; Bittner, F.; Mendel, R.; Clement, B. Defining the Role of the NADH-Cytochrome-b5 Reductase 3 in the Mitochondrial Amidoxime Reducing Component Enzyme System. *Drug Metab Dispos* 2016, 44, 1617-1621, doi:10.1124/dmd.116.071845.
135. Wood, K.C.; Durgin, B.G.; Schmidt, H.M.; Hahn, S.A.; Baust, J.J.; Bachman, T.; Vitturi, D.A.; Ghosh, S.; Ofori-Acquah, S.F.; Mora, A.L., et al. Smooth muscle cytochrome b5 reductase 3 deficiency accelerates pulmonary hypertension development in sickle cell mice. *Blood Adv* 2019, 3, 4104-4116, doi:10.1182/bloodadvances.2019000621.
136. Rahaman, M.M.; Nguyen, A.T.; Miller, M.P.; Hahn, S.A.; Sparacino-Watkins, C.; Jobbagy, S.; Carew, N.T.; Cantu-Medellin, N.; Wood, K.C.; Baty, C.J., et al. Cytochrome b5 Reductase 3 Modulates Soluble Guanylate Cyclase Redox State and cGMP Signaling. *Circ Res* 2017, 121, 137-148, doi:10.1161/CIRCRESAHA.117.310705.

- 
137. Fukuto, J.M.; Carrington, S.J.; Tantillo, D.J.; Harrison, J.G.; Ignarro, L.J.; Freeman, B.A.; Chen, A.; Wink, D.A. Small molecule signaling agents: the integrated chemistry and biochemistry of nitrogen oxides, oxides of carbon, dioxygen, hydrogen sulfide, and their derived species. *Chem Res Toxicol* 2012, 25, 769-793, doi:10.1021/tx2005234.
138. Samaan, S.; Tranchevent, L.C.; Dardenne, E.; Polay Espinoza, M.; Zonta, E.; Germann, S.; Gratadou, L.; Dutertre, M.; Auboeuf, D. The Ddx5 and Ddx17 RNA helicases are cornerstones in the complex regulatory array of steroid hormone-signaling pathways. *Nucleic Acids Res* 2014, 42, 2197-2207, doi:10.1093/nar/gkt1216.
139. Dewundara, S.S.; Wiggs, J.L.; Sullivan, D.A.; Pasquale, L.R. Is Estrogen a Therapeutic Target for Glaucoma? *Semin Ophthalmol* 2016, 31, 140-146, doi:10.3109/08820538.2015.1114845.
140. Patel, P.; Harris, A.; Toris, C.; Tobe, L.; Lang, M.; Belamkar, A.; Ng, A.; Verticchio Vercellin, A.C.; Mathew, S.; Siesky, B. Effects of Sex Hormones on Ocular Blood Flow and Intraocular Pressure in Primary Open-angle Glaucoma: A Review. *J Glaucoma* 2018, 27, 1037-1041, doi:10.1097/IJG.0000000000001106.
141. Wickham, L.A.; Gao, J.; Toda, I.; Rocha, E.M.; Ono, M.; Sullivan, D.A. Identification of androgen, estrogen and progesterone receptor mRNAs in the eye. *Acta Ophthalmol Scand* 2000, 78, 146-153, doi:10.1034/j.1600-0420.2000.078002146.x.
142. Kobayashi, K.; Kobayashi, H.; Ueda, M.; Honda, Y. Estrogen receptor expression in bovine and rat retinas. *Invest Ophthalmol Vis Sci* 1998, 39, 2105-2110.
143. Chakrabarti, M.; Haque, A.; Banik, N.L.; Nagarkatti, P.; Nagarkatti, M.; Ray, S.K. Estrogen receptor agonists for attenuation of neuroinflammation and neurodegeneration. *Brain Res Bull* 2014, 109, 22-31, doi:10.1016/j.brainresbull.2014.09.004.
144. Jiang, M.; Ma, X.; Zhao, Q.; Li, Y.; Xing, Y.; Deng, Q.; Shen, Y. The neuroprotective effects of novel estrogen receptor GPER1 in mouse retinal ganglion cell degeneration. *Exp Eye Res* 2019, 189, 107826, doi:10.1016/j.exer.2019.107826.
145. Sarzi, E.; Seveno, M.; Angebault, C.; Milea, D.; Ronnback, C.; Quiles, M.; Adrian, M.; Grenier, J.; Caignard, A.; Lacroux, A., et al. Increased steroidogenesis promotes early-onset and severe vision loss in females with OPA1 dominant optic atrophy. *Hum Mol Genet* 2016, 25, 2539-2551, doi:10.1093/hmg/ddw117.
146. Pisano, A.; Preziuso, C.; Iommarini, L.; Perli, E.; Grazioli, P.; Campese, A.F.; Maresca, A.; Montopoli, M.; Masuelli, L.; Sadun, A.A., et al. Targeting estrogen receptor beta as preventive therapeutic strategy for Leber's hereditary optic neuropathy. *Hum Mol Genet* 2015, 24, 6921-6931, doi:10.1093/hmg/ddv396.
147. Chen, X.; Liu, Y.; Zhang, Y.; Kam, W.R.; Pasquale, L.R.; Sullivan, D.A. Impact of aromatase absence on murine intraocular pressure and retinal ganglion cells. *Sci Rep* 2018, 8, 3280, doi:10.1038/s41598-018-21475-x.

- 
148. Feola, A.J.; Fu, J.; Allen, R.; Yang, V.; Campbell, I.C.; Ottensmeyer, A.; Ethier, C.R.; Pardue, M. Menopause exacerbates visual dysfunction in experimental glaucoma. *Exp Eye Res* 2019, 186, 107706, doi:10.1016/j.exer.2019.107706.
149. Shah, A.; Schiffmacher, A.T.; Taneyhill, L.A. Annexin A6 controls neuronal membrane dynamics throughout chick cranial sensory gangliogenesis. *Dev Biol* 2017, 425, 85-99, doi:10.1016/j.ydbio.2017.03.011.
150. Klee, C.B. Ca<sup>2+</sup>-dependent phospholipid- (and membrane-) binding proteins. *Biochemistry* 1988, 27, 6645-6653, doi:10.1021/bi00418a001.
151. Kapustin, A.N.; Shanahan, C.M. Emerging roles for vascular smooth muscle cell exosomes in calcification and coagulation. *J Physiol* 2016, 594, 2905-2914, doi:10.1113/JP271340.
152. Hubner, C.A.; Schroeder, B.C.; Ehmke, H. Regulation of vascular tone and arterial blood pressure: role of chloride transport in vascular smooth muscle. *Pflugers Arch* 2015, 467, 605-614, doi:10.1007/s00424-014-1684-y.
153. Ye, S.; Chang, Y.; Kim, C.W.; Kwon, M.J.; Choi, Y.; Ahn, J.; Kim, J.M.; Kim, H.S.; Shin, H.; Ryu, S. Intraocular pressure and coronary artery calcification in asymptomatic men and women. *Br J Ophthalmol* 2015, 99, 932-936, doi:10.1136/bjophthalmol-2014-305925.
154. Borden, L.A. GABA transporter heterogeneity: pharmacology and cellular localization. *Neurochem Int* 1996, 29, 335-356, doi:10.1016/0197-0186(95)00158-1.
155. Crooks, J.; Kolb, H. Localization of GABA, glycine, glutamate and tyrosine hydroxylase in the human retina. *J Comp Neurol* 1992, 315, 287-302, doi:10.1002/cne.903150305.
156. Hilton, E.J.; Hosking, S.L.; Betts, T. The effect of antiepileptic drugs on visual performance. *Seizure* 2004, 13, 113-128, doi:10.1016/s1059-1311(03)00082-7.
157. Sakaew, W.; Tachow, A.; Thoungseabyoun, W.; Khrongyut, S.; Rawangwong, A.; Polsan, Y.; Masahiko, W.; Kondo, H.; Hipkaeo, W. Expression and localization of VIAAT in distal uriniferous tubular epithelium of mouse. *Ann Anat* 2019, 222, 21-27, doi:10.1016/j.aanat.2018.11.002.
158. Kononenko, N.L.; Puchkov, D.; Classen, G.A.; Walter, A.M.; Pechstein, A.; Sawade, L.; Kaempf, N.; Trimbuch, T.; Lorenz, D.; Rosenmund, C., et al. Clathrin/AP-2 mediate synaptic vesicle reformation from endosome-like vacuoles but are not essential for membrane retrieval at central synapses. *Neuron* 2014, 82, 981-988, doi:10.1016/j.neuron.2014.05.007.
159. Kononenko, N.L.; Classen, G.A.; Kuijpers, M.; Puchkov, D.; Maritzen, T.; Tempes, A.; Malik, A.R.; Skalecka, A.; Bera, S.; Jaworski, J., et al. Retrograde transport of TrkB-containing autophagosomes via the adaptor AP-2 mediates neuronal complexity and prevents neurodegeneration. *Nat Commun* 2017, 8, 14819, doi:10.1038/ncomms14819.

- 
160. Helbig, I.; Lopez-Hernandez, T.; Shor, O.; Galer, P.; Ganesan, S.; Pendziwiat, M.; Rademacher, A.; Ellis, C.A.; Humpfer, N.; Schwarz, N., et al. A Recurrent Missense Variant in AP2M1 Impairs Clathrin-Mediated Endocytosis and Causes Developmental and Epileptic Encephalopathy. *Am J Hum Genet* 2019, 104, 1060-1072, doi:10.1016/j.ajhg.2019.04.001.
161. Kittler, J.T.; Chen, G.; Honing, S.; Bogdanov, Y.; McAinsh, K.; Arancibia-Carcamo, I.L.; Jovanovic, J.N.; Pangalos, M.N.; Haucke, V.; Yan, Z., et al. Phospho-dependent binding of the clathrin AP2 adaptor complex to GABAA receptors regulates the efficacy of inhibitory synaptic transmission. *Proc Natl Acad Sci U S A* 2005, 102, 14871-14876, doi:10.1073/pnas.0506653102.
162. Froemke, R.C. Plasticity of cortical excitatory-inhibitory balance. *Annu Rev Neurosci* 2015, 38, 195-219, doi:10.1146/annurev-neuro-071714-034002.
163. Smith, K.R.; Muir, J.; Rao, Y.; Browarski, M.; Gruenig, M.C.; Sheehan, D.F.; Haucke, V.; Kittler, J.T. Stabilization of GABA(A) receptors at endocytic zones is mediated by an AP2 binding motif within the GABA(A) receptor beta3 subunit. *J Neurosci* 2012, 32, 2485-2498, doi:10.1523/JNEUROSCI.1622-11.2011.
164. Neal, M.J.; Shah, M.A. Development of tolerance to the effects of vigabatrin (gamma-vinyl-GABA) on GABA release from rat cerebral cortex, spinal cord and retina. *Br J Pharmacol* 1990, 100, 324-328, doi:10.1111/j.1476-5381.1990.tb15803.x.
165. Hosking, S.L.; Hilton, E.J. Neurotoxic effects of GABA-transaminase inhibitors in the treatment of epilepsy: ocular perfusion and visual performance. *Ophthalmic Physiol Opt* 2002, 22, 440-447, doi:10.1046/j.1475-1313.2002.00063.x.
166. Hilton, E.J.; Hosking, S.L.; Betts, T. Epilepsy patients treated with antiepileptic drug therapy exhibit compromised ocular perfusion characteristics. *Epilepsia* 2002, 43, 1346-1350, doi:10.1046/j.1528-1157.2002.44901.x.
167. Hammond, E.J.; Wilder, B.J. Gamma-vinyl GABA. *Gen Pharmacol* 1985, 16, 441-447, doi:10.1016/0306-3623(85)90002-3.
168. Wild, J.M.; Martinez, C.; Reinshagen, G.; Harding, G.F. Characteristics of a unique visual field defect attributed to vigabatrin. *Epilepsia* 1999, 40, 1784-1794, doi:10.1111/j.1528-1157.1999.tb01599.x.
169. Fujiwara, M.; Muramatsu, I. Gamma-aminobutyric acid receptor on vascular smooth muscle of dog cerebral arteries. *Br J Pharmacol* 1975, 55, 561-562, doi:10.1111/j.1476-5381.1975.tb07434.x.
170. Edvinsson, L.; Krause, D.N. Pharmacological characterization of GABA receptors mediating vasodilation of vertebral arteries in vitro. *Brain Res* 1979, 173, 89-97, doi:10.1016/0006-8993(79)91098-9.

- 
171. Hinds, K.; Monaghan, K.P.; Frolund, B.; McGeown, J.G.; Curtis, T.M. GABAergic control of arteriolar diameter in the rat retina. *Invest Ophthalmol Vis Sci* 2013, 54, 6798-6805, doi:10.1167/iovs.13-12362.
172. Deng, X.W.; Dubiel, W.; Wei, N.; Hofmann, K.; Mundt, K.; Colicelli, J.; Kato, J.; Naumann, M.; Segal, D.; Seeger, M., et al. Unified nomenclature for the COP9 signalosome and its subunits: an essential regulator of development. *Trends Genet* 2000, 16, 202-203, doi:10.1016/s0168-9525(00)01982-x.
173. Boussiotis, V.A.; Freeman, G.J.; Taylor, P.A.; Berezovskaya, A.; Grass, I.; Blazar, B.R.; Nadler, L.M. p27kip1 functions as an anergy factor inhibiting interleukin 2 transcription and clonal expansion of alloreactive human and mouse helper T lymphocytes. *Nat Med* 2000, 6, 290-297, doi:10.1038/73144.
174. Doronkin, S.; Djagaeva, I.; Beckendorf, S.K. The COP9 signalosome promotes degradation of Cyclin E during early *Drosophila* oogenesis. *Dev Cell* 2003, 4, 699-710, doi:10.1016/s1534-5807(03)00121-7.
175. Yan, J.; Walz, K.; Nakamura, H.; Carattini-Rivera, S.; Zhao, Q.; Vogel, H.; Wei, N.; Justice, M.J.; Bradley, A.; Lupski, J.R. COP9 signalosome subunit 3 is essential for maintenance of cell proliferation in the mouse embryonic epiblast. *Mol Cell Biol* 2003, 23, 6798-6808, doi:10.1128/mcb.23.19.6798-6808.2003.
176. Hetfeld, B.K.; Peth, A.; Sun, X.M.; Henklein, P.; Cohen, G.M.; Dubiel, W. The COP9 signalosome-mediated deneddylation is stimulated by caspases during apoptosis. *Apoptosis* 2008, 13, 187-195, doi:10.1007/s10495-007-0164-7.
177. da Silva Correia, J.; Miranda, Y.; Leonard, N.; Ulevitch, R.J. The subunit CSN6 of the COP9 signalosome is cleaved during apoptosis. *J Biol Chem* 2007, 282, 12557-12565, doi:10.1074/jbc.M609587200.
178. de Laat, W.L.; Jaspers, N.G.; Hoeijmakers, J.H. Molecular mechanism of nucleotide excision repair. *Genes Dev* 1999, 13, 768-785, doi:10.1101/gad.13.7.768.
179. Scharer, O.D. Nucleotide excision repair in eukaryotes. *Cold Spring Harb Perspect Biol* 2013, 5, a012609, doi:10.1101/cshperspect.a012609.
180. Zhang, M.; Du, W.; Acklin, S.M.; Jin, S.; Xia, F. SIRT2 protects peripheral neurons from cisplatin-induced injury by enhancing nucleotide excision repair. *J Clin Invest* 2020, 10.1172/JCI123159, doi:10.1172/JCI123159.
181. Barros, L.F.; San Martin, A.; Ruminot, I.; Sandoval, P.Y.; Fernandez-Moncada, I.; Baeza-Lehnert, F.; Arce-Molina, R.; Contreras-Baeza, Y.; Cortes-Molina, F.; Galaz, A., et al. Near-critical GLUT1 and Neurodegeneration. *J Neurosci Res* 2017, 95, 2267-2274, doi:10.1002/jnr.23998.

- 
182. Flammer, J.; Orgul, S.; Costa, V.P.; Orzalesi, N.; Krieglstein, G.K.; Serra, L.M.; Renard, J.P.; Stefansson, E. The impact of ocular blood flow in glaucoma. *Progress in retinal and eye research* 2002, 21, 359-393, doi:10.1016/s1350-9462(02)00008-3.
183. Osborne, N.N.; Melena, J.; Chidlow, G.; Wood, J.P. A hypothesis to explain ganglion cell death caused by vascular insults at the optic nerve head: possible implication for the treatment of glaucoma. *Br J Ophthalmol* 2001, 85, 1252-1259, doi:10.1136/bjo.85.10.1252.
184. Golubnitschaja-Labudova, O.; Liu, R.; Decker, C.; Zhu, P.; Haefliger, I.O.; Flammer, J. Altered gene expression in lymphocytes of patients with normal-tension glaucoma. *Curr Eye Res* 2000, 21, 867-876.
185. Flammer, J. [Glaucomatous optic neuropathy: a reperfusion injury]. *Klin Monbl Augenheilkd* 2001, 218, 290-291, doi:10.1055/s-2001-15883.
186. Perumal, N.; Funke, S.; Pfeiffer, N.; Grus, F.H. Proteomics analysis of human tears from aqueous-deficient and evaporative dry eye patients. *Scientific reports* 2016, 6, 29629, doi:10.1038/srep29629.
187. Perumal, N.; Funke, S.; Pfeiffer, N.; Grus, F.H. Characterization of lacrimal proline-rich protein 4 (PRR4) in human tear proteome. *Proteomics* 2014, 14, 1698-1709, doi:10.1002/pmic.201300039.

## **6 Appendix**

### **6.1 Contributions to the manuscripts**

#### Publication I:

Hanhan Liu developed the study design, organized and performed the experimental experiments and wrote the manuscript. Fabian Anders participated in the study design, assisted with the mass spectrometric measurements, proofread the manuscript and contributed important intellectual content. Carolina Mann and Aiwei Liu participated in sample recruitment, reviewed the manuscript. Solon Thanos, Norbert Pfeiffer and Franz H. Grus critically reviewed the manuscript and provided important intellectual input. Verena Prokosch performed the study coordination as well as the study design and participated in review and approval of the manuscript.

#### Publication II:

Hanhan Liu developed the study design and experimental set-up, organized and performed the experimental experiments and wrote the manuscript. Fabian Anders performed the MS-based affinity capture experiments as well as data analysis. Karl Mercieca aided in the study coordination, reviewed the manuscript and provided important intellectual knowledge. Verena Prokosch performed the study coordination and reviewed as well as approved the manuscript.

#### Publication III:

Hanhan Liu participated to write the manuscript, developed the study design, performed the animal experiments and assisted in the mass spectrometric measurements as well as data analysis. Natarajan Perumal and Caroline Manicam assisted in the development of the study design, performed the MS-based affinity capture experiments as well as data analysis and proofread the manuscript. Karl Mercieca reviewed the manuscript and provided important intellectual knowledge. Verena Prokosch participated in the study coordination as well as study design and critically reviewed the manuscript.

---

## 6.2 List of abbreviations

<b>RGC</b>	retinal ganglion cells	<b>OCT</b>	Optical Coherence Tomography
<b>IOP</b>	intraocular pressure	<b>EVO</b>	episcleral vein occlusion
<b>ROS</b>	reactive oxygen species	<b>RNFL</b>	retinal nerve fiber layer
<b>NO</b>	nitric oxide	<b>MS</b>	mass spectrometry
<b>H<sub>2</sub>S</b>	hydrogen sulfide	<b>FDR</b>	false discovery rate
<b>CO</b>	carbon monoxide	<b>DHA</b>	docosahexaenoic acid
<b>CBS</b>	cystathionine-β-synthase	<b>NAD<sup>+</sup></b>	nicotinamide adenine dinucleotide
<b>CSE</b>	cystathionine-γ-lyase	<b>PDH</b>	pyruvate dehydrogenase
<b>3MST</b>	3-mercaptopyruvate sulfurtransferase	<b>TCA</b>	tricarboxylic acid cycle
<b>NMDA</b>	N-methyl-D-aspartic acid	<b>ATP</b>	adenosine triphosphate
<b>AD</b>	Alzheimer's disease	<b>PKA</b>	protein kinase A
<b>PD</b>	Parkinson's disease	<b>sGC</b>	soluble guanylyl cyclase
<b>SNCA</b>	synuclein α	<b>SMC</b>	smooth muscle cell
<b>SNCB</b>	synuclein β	<b>AP-2</b>	adaptor protein complex 2
<b>SNCG</b>	synuclein γ	<b>NER</b>	nucleotide excision repair (NER)
<b>CNS</b>	central nervous system	<b>UPS</b>	ubiquitin proteasome system
<b>IPA</b>	ingenuity pathway analysis		

### 6.3 List of figures

<b>Figure 1.</b> Workflow overview.....	22
<b>Figure 2.</b> Changes in retinal metabolism and mitochondrial function.....	27
<b>Figure 3.</b> Changes in retinal vascular function.....	30
<b>Figure 4.</b> Overview of altered signalling pathways due to I/R injury and H <sub>2</sub> S treatment.....	35

### 6.5 Declaration of academic honesty

*I hereby declare that I wrote the dissertation submitted without any unauthorized external assistance and used only sources acknowledged in the work. All textual passages which are appropriated verbatim or paraphrased from published and unpublished texts as well as all information obtained from oral sources are duly indicated and listed in accordance with bibliographical rules. In carrying out this research, I complied with the rules of standard scientific practice as formulated in the statutes of Johannes Gutenberg-University Mainz to insure standard scientific practice. This is my first attempt to promotion.*

Mainz, den 24.09.2020

A handwritten signature in black ink, appearing to read 'Liu Hanhan', written over a horizontal line.

(Hanhan Liu)