Exchange of nitrogen dioxide (NO₂) between plants and the atmosphere under laboratory and field conditions

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"Il semble que la perfection soit atteinte non quand il n'y a plus rien à ajouter, mais quand il n'y a plus rien à retrancher."

"Perfektion ist nicht dann erreicht, wenn man nichts mehr hinzufügen, sondern wenn man nichts mehr weglassen kann."

> "Is seems that perfection is attained not when there is nothing more to add, but when there is nothing more to remove."

> > ANTOINE DE SAINT-EXUPERY

Contents

Danksagung	V
Zusammenfassung	VII
Summary	XI
List of tables	XIII
List of figures	XV
List of abbreviations	XX
1 Introduction	1
1.1 Nitrogen in the atmosphere	1
1.2 Chemistry of NO, NO ₂ and O ₃ in the troposphere	3
1.3 NO and NO ₂ biosphere-atmosphere exchange in ecosystem forest	5
1.3.1 Processes in soil	7
1.3.2 Interaction with plants	8
1.4 Objectives of thesis	14
2 Material and Methods	15
2.1 Basic considerations	16
2.1.1 Mass balance of the NO-NO ₂ -O ₃ triad of a dynamic plant chamber	16
2.1.2 Molar mass flux densities, deposition velocities and compensation point concentrations	19

II | CONTENTS

2.1.3 Constraints of precision	23
2.1.4 Constraints of design	30
2.2 Trace gas analyzers	31
2.3 Calibrations, limits of detection, standard errors and precision of trace gas concentration measurements	34
2.4 Dynamic chamber system	35
2.4.1 Design and construction	35
2.4.2 Implementation of concentration and flux density measurements	40
2.5 Experiments	41
2.5.1 Plant material	41
2.5.2 Field site description and set-up	42
2.5.3 Laboratory set-up	43

Specific chamł	cation and implementation of plant dynamic per system	45
3.1 Met	thods	46
Quality	assurance and error analysis	
3.1.1	Corrections for concentration changes in long tubing	46
3.1.2	Temporal response of analyzers	46
3.1.3	Temperature dependence of analyzers	46
3.1.4	Dynamic chamber: internal mixing, exchange rate of chamber volume, wall absorption and transmissivity	47
3.1.5	Significance of concentration differences	48
3.1.6	Bi-variate weighted linear least-squares fitting regression analysis	48
3.1.7	Standard errors of exchange flux densities, deposition velocities and compensation point concentrations	50
3.1.8	Significance of the compensation point concentrations	55
3.2 Resi	ılts	56
3.2.1	Analyzers and system performance	56
3.2.2	NO ₂ blending for fumigation experiment	59
3.2.3	Characterization of the dynamic plant chamber	59
	3.2.3.1 Radiation and NO ₂ photolysis rate	59
	3.2.3.2 Sorption effects and chamber volume exchange time	61

3.2.4.1 NO ₂ exchange flux density: Laboratory results	62
3.2.4.2 NO-NO ₂ -O ₃ exchange flux densities: Field results	66
3.3 Discussion	72
3.3.1 Overview of previous NO ₂ exchange flux measurements using dynamic plant chambers	72
3.3.2 Precision, data quality and photochemical reactions	74
3.3.2.1 Precision and data quality	74
3.3.2.2 Significance of concentration differences	77
3.3.2.3 Photo-chemical reactions in the dynamic plant chamber: impact on net exchange flux densities, deposition velocities and compensation point concentrations	78
3.3.3 Bi-variate weighted linear regression	83

4 Applica	ntion of plant dynamic chamber system to	
field n	neasurements	85
4.1 Met	thods	85
4.1.1	Photosynthesis rate, transpiration rate, stomatal conductance	85
4.1.2	Classification of data	86
4.1.3	Monitoring of plant-physiological processes due to chambers	87
4.1.4	Set-up at the ECHO project	88
4.2 Results		89
<u>EGER proj</u>	<u>ect</u>	
4.2.1	Microclimatic conditions	89
4.2.2	Plant physiology	90
4.2.3	Diurnal variations of gas exchange	92
4.2.4	Overview of plant chamber measurements	96
4.2.5	NO ₂ exchange flux density	99
4.2.6	O ₃ exchange flux density	111
ECHO proj	<u>ect</u>	
4.2.7	NO ₂ exchange flux density	122

4.3 Discussion	129
4.3.1 Effects on enclosed plants	129
4.3.2 NO_2 exchange to leaves	130
4.3.3 Deposition velocities of NO_2 and O_3	133
4.3.4 NO ₂ compensation point concentration	136
5 Conclusions and Perspectives	139
6 References	147
Curriculum Vitae	159

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Zusammenfassung

Stickstoff ist für Mensch, Tier und Pflanze ein essentieller Nährstoff, der ein wichtiges Bauelement von Proteinen und Nukleinsäuren ist. Obwohl der Großteil der Erdatmosphäre aus molekularem Stickstoff (N2) besteht (78 %), können nur wenige Mikroorganismen diesen direkt nutzen. Um für höhere Pflanzen oder Tiere verwertbar zu sein, muss der molekulare Stickstoff in eine reaktivere oxidierte Form überführt werden. Dies geschieht innerhalb des Stickstoffkreislaufs unter anderem durch freilebende Mikroorganismen, in Symbiose lebende Knöllchenbakterien oder durch elektrische Entladungen bei Gewittern. Dem Menschen ermöglicht das Haber-Bosch-Verfahren seit Anfang des 20. Jahrhunderts die Synthese von reaktivem Stickstoff. Damit konnte die Ernährungssicherung der Weltbevölkerung deutlich verbessert werden. Auf der anderen Seite hat der verstärkte Stickstoffeintrag die Versauerung und Eutrophierung von Ökosystemen und den Verlust an Biodiversität zu Folge. Für den Menschen ergeben sich nachteilige Auswirkungen auf die Gesundheit durch Feinstaubbildung und Sommersmog-Episoden. Reaktiver Stickstoff spielt zudem eine wichtige Rolle in der Atmosphärenchemie und deren globalen Kreisläufen von Schadund Nährstoffen.

Stickstoffmonoxid (NO) und Stickstoffdioxid (NO₂) gehören zu den reaktiven Spurengasen und werden unter der Bezeichnung NO_x zusammengefasst. Als wichtige Komponenten atmosphärischer oxidativer Prozesse beeinflussen sie aber auch die Lebenszeiten weniger reaktiver Treibhausgase. NO und NO₂ entstehen u.a. bei Verbrennungsvorgängen durch die Oxidation von atmosphärischem Stickstoff sowie durch biologische Vorgänge in Böden. NO wird in der Atmosphäre relativ schnell zum biologisch wirksameren NO₂ oxidiert. NO₂ wird in der Atmosphäre langsam weiter zu Nitrat (NO₃⁻) und zu Salpetersäure (HNO₃) aufoxidiert, lagert sich an Aerosole an und wird schließlich in der partikelgebundenen Form durch nasse und trockene Deposition aus der Atmosphäre ausgetragen. In der Atmosphärenchemie spielt NO_x zudem eine wichtige katalytische Rolle bei Bildung und Abbau von troposphärischem Ozon (O₃). NO, NO₂ und O₃ befinden sich in der Atmosphäre in einem photostationären Gleichgewicht, weshalb man von der NO-NO₂-O₃ Triade spricht. In Bereichen mit erhöhtem NO-Gehalt können beispielsweise Reaktionen mit anderen Luftschadstoffen die Bildung von NO₂ bewirken, wodurch das Gleichgewicht zur Ozonbildung hin verschoben wird.

Für Pflanzen stellt die NO_3^- -Aufnahme über die Wurzel die Hauptquelle des essentiellen Nährstoffs Stickstoff dar. Im Boden wird atmosphärischer Stickstoff mittels Bakterien über Stickstofffixierung oder Ammoniakbildung und Nitrifikation zu $NO_3^$ oxidiert. Zusätzlich nehmen Pflanzen atmosphärisches NO_2 direkt über ihre Spaltöffnungen auf. NO_2 wird im Apoplasten durch Disproportionierung als NO_3^- und Nitrit (NO_2^-) der Pflanze zur Verfügung gestellt. Mittels der Enzyme Nitrat- und Nitritreduktase erfolgt daraufhin eine weitere Umwandlung in Ammonium (NH_4^+). Beeinflusst wird der NO_2 -Gaswechsel vom Partialdruckgradienten in den Lufträumen der Blätter, dem Öffnungszustand der Spaltöffnungen und den im Blatt vorhandenen Widerständen. Die Regulierung der Spaltöffnungen geschieht unter anderem über klimatische Umweltbedingungen wie Lichtintensität, Temperatur und dem Wasserdampfsättigungsdefizit.

Die vorliegende Dissertation möchte dazu beitragen das Verständnis für die Rolle der Vegetation im NO₂-Zyklus der Atmosphäre zu verbessern und die Frage nach einem NO₂-Kompensationspunkt ($m_{comp,NO2}$) zu klären. Dazu wurde der NO₂-Austausch zwischen der Atmosphäre und Fichten (*Picea abies*) auf Blattebene mittels eines dynamischen Kammersystems unter Freiland- und Laborbedingungen untersucht. Die Messungen erfolgten im Rahmen des EGER-Projekts (Juni-Juli 2008). Zusätzlich wurden zur Verfügung gestellte NO₂-Messdaten ausgewertet, die während des ECHO-Projekts (Juli 2003) an Eichen (*Quercus robur*) aufgenommen wurden. Das verwendete Messsystem ermöglicht die gleichzeitige Bestimmung der Austauschraten von NO, NO₂, O₃, CO₂ und H₂O. Da die Flussberechnungen von NO, NO₂, und O₃ auf sehr kleinen Konzentrationsdifferenzen (Δm_i) beruhen, die zwischen Ein- und Ausgang der Messungen erforderlich. Um dies zu erreichen, wurde ein hoch spezifisches NO/NO₂ Messinstrument verwendet und das gesamte Messsystem dahingehend optimiert, dass eine hohe Messgenauigkeit dauerhaft gewährleistet werden konnte. Die Datenanalyse ergab, dass ein signifikanter $m_{comp,NO2}$ nur bestimmt werden kann, wenn die statistische Signifikanz von Δm_i gegeben ist. In Folge dessen wurde die Signifikanz von Δm_i als ein Qualitätskriterium für die Daten verwendet. Bei der Bestimmung der NO-, NO₂- und O₃-Austauschraten müssen die photochemischen Reaktionen der NO-NO₂-O₃ Triade innerhalb der Messkammer berücksichtigt werden, ansonsten werden Depositionsgeschwindigkeiten ($v_{dep,NO2}$) und $m_{comp,NO2}$ überschätzt. Für Fichten konnte unter Laborbedingungen kein signifikanter $m_{comp,NO2}$ bestimmt werden, unter Feldbedingungen lag $m_{comp,NO2}$ zwischen 0.17 und 0.65 ppb und $v_{dep,NO2}$ zwischen 0.07 und 0.42 mm s⁻¹. Die Analyse der Felddaten, gemessen an Eichen, ergab ebenfalls keinen NO₂-Kompensationspunkt, $v_{dep,NO2}$ lag zwischen 0.6 und 2.71 mm s⁻¹. Damit verdichten sich die Hinweise, dass Wälder hauptsächlich als Senken für NO₂ anzusehen sind und mögliche NO₂-Emissionen äußerst gering ausfallen. Nur bei hohen NO-Emissionen aus Böden, die in Reaktion mit Ozon mehr NO₂ liefern als von Pflanzen aufgenommen wird, könnten Wälder Quellen für NO₂ darstellen.

Summary

Nitrogen is an essential nutrient. It is for human, animal and plants a constituent element of proteins and nucleic acids. Although the majority of the Earth's atmosphere consists of elemental nitrogen (N₂, 78 %) only a few microorganisms can use it directly. To be useful for higher plants and animals elemental nitrogen must be converted to a reactive oxidized form. This conversion happens within the nitrogen cycle by free-living microorganisms, symbiotic living *Rhizobium* bacteria or by lightning. Humans are able to synthesize reactive nitrogen through the Haber-Bosch process since the beginning of the 20th century. As a result food security of the world population could be improved noticeably. On the other side the increased nitrogen input results in acidification and eutrophication of ecosystems and in loss of biodiversity. Negative health effects arose for humans such as fine particulate matter and summer smog. Furthermore, reactive nitrogen plays a decisive role at atmospheric chemistry and global cycles of pollutants and nutritive substances.

Nitrogen monoxide (NO) and nitrogen dioxide (NO₂) belong to the reactive trace gases and are grouped under the generic term NO_x . They are important components of atmospheric oxidative processes and influence the lifetime of various less reactive greenhouse gases. NO and NO₂ are generated amongst others at combustion process by oxidation of atmospheric nitrogen as well as by biological processes within soil. In atmosphere NO is converted very quickly into NO₂. NO₂ is than oxidized to nitrate (NO₃⁻) and to nitric acid (HNO₃), which bounds to aerosol particles. The bounded nitrate is finally washed out from atmosphere by dry and wet deposition. Catalytic reactions of NO_x are an important part of atmospheric chemistry forming or decomposing tropospheric ozone (O₃). In atmosphere NO, NO₂ and O₃ are in photostationary equilibrium, therefore it is referred as NO-NO₂-O₃ triad. At regions with elevated NO concentrations reactions with air pollutions can form NO₂, altering equilibrium of ozone formation.

The essential nutrient nitrogen is taken up by plants mainly by dissolved NO_3^- entering the roots. Atmospheric nitrogen is oxidized to NO_3^- within soil via bacteria by

nitrogen fixation or ammonium formation and nitrification. Additionally atmospheric NO_2 uptake occurs directly by stomata. Inside the apoplast NO_2 is disproportionated to nitrate and nitrite (NO_2^-) , which can enter the plant metabolic processes. The enzymes nitrate and nitrite reductase convert nitrate and nitrite to ammonium (NH_4^+) . NO_2 gas exchange is controlled by pressure gradients inside the leaves, the stomatal aperture and leaf resistances. Plant stomatal regulation is affected by climate factors like light intensity, temperature and water vapor pressure deficit.

This thesis wants to contribute to the comprehension of the effects of vegetation in the atmospheric NO₂ cycle and to discuss the NO₂ compensation point concentration $(m_{comp,NO2})$. Therefore, NO₂ exchange between the atmosphere and spruce (*Picea abies*) on leaf level was detected by a dynamic plant chamber system under laboratory and field conditions. Measurements took place during the EGER project (June-July 2008). Additionally NO₂ data collected during the ECHO project (July 2003) on oak (*Quercus robur*) were analyzed. The used measuring system allowed simultaneously determination of NO, NO₂, O₃, CO₂ and H₂O exchange rates. Calculations of NO, NO₂ and O₃ fluxes based on generally small differences (Δm_i) measured between inlet and outlet of the chamber. Consequently a high accuracy and specificity of the analyzer is necessary. To achieve these requirements a highly specific NO/NO₂ analyzer was used and the whole measurement system was optimized to an enduring measurement precision.

Data analysis resulted in a significant $m_{comp,NO2}$ only if statistical significance of Δm_i was detected. Consequently, significance of Δm_i was used as a data quality criterion. Photo-chemical reactions of the NO-NO₂-O₃ triad in the dynamic plant chamber's volume must be considered for the determination of NO, NO₂, O₃ exchange rates, otherwise deposition velocity ($v_{dep,NO2}$) and $m_{comp,NO2}$ will be overestimated. No significant $m_{comp,NO2}$ for spruce could be determined under laboratory conditions, but under field conditions $m_{comp,NO2}$ could be identified between 0.17 and 0.65 ppb and $v_{dep,NO2}$ between 0.07 and 0.42 mm s⁻¹. Analyzing field data of oak, no NO₂ compensation point concentration could be determined, $v_{dep,NO2}$ ranged between 0.6 and 2.71 mm s⁻¹. There is increasing indication that forests are mainly a sink for NO₂ and potential NO₂ emissions are low. Only when assuming high NO soil emissions, more NO₂ can be formed by reaction with O₃ than plants are able to take up. Under these circumstance forests can be a source for NO₂.

List of Tables

Table 1:	Overview of global sources of nitrogen oxides	1
Table 2:	Typical NO _x mixing ratios	2
Table 3:	Reported NO ₂ compensation points obtained from the literature	13
Table 4:	Interferences of chemiluminescent NO-NO ₂ -NO _x analyzers used different NO ₂ converters	32
Table 5:	Measured parameters and instrument specifications	33
Table 6:	Manufacturer details for parts of the dynamic chamber system	36
Table 7:	Derivatives $\partial y/\partial x_i$ of $y = F_{ex,NO2}$, $F_{ex,NO}$, $F_{ex,O3}$ for application of the generalized Gaussian error propagation to calculate the standard errors of $s_{Fex,NO2}$, $s_{Fex,NO}$ and $s_{Fex,O3}$	52
Table 8:	Derivatives $\partial y/\partial x_i$ of $y = v_{dep,NO2}$, $v_{dep,NO}$, $v_{dep,O3}$ for application of the generalized Gaussian error propagation to calculate the standard errors of s_{v,dep_NO2} , s_{v,dep_NO} and s_{v,dep_O3}	53
Table 9:	Derivatives $\partial y/\partial x_i$ of $y = m_{comp,NO2}$, $m_{comp,NO}$ and $m_{comp,O3}$ for application of the generalized Gaussian error propagation to calculate the standard errors of $s_{m,comp_NO2}$, $s_{m,comp_NO}$ and $s_{m,comp_O3}$	54
Table 10:	Results of the temperature dependence tests of the used analyzers	56
Table 11:	Parameters of sorption effects to the inner chamber walls	61
Table 12:	Parameters for NO ₂ laboratory measurements of simple and bi- variate weighted linear least-squares fitting regression analysis	65
Table 13:	Percentage of data m_i above $LOD(m_i)$ and significant differences $\Delta m_i = (m_{a,NO2} - m_{s,NO2})$ for field measurements	67
Table 14:	Parameters of bi-variate weighted linear least-squares fitting regression analysis for field measurements	71
Table 15:	Overview of studies that have performed dynamic chamber NO ₂ flux measurements on different plant species	73
Table 16:	Ambient conditions during time of field measurements (EGER)	89

XIV | LIST OF TABLES

Table 17:	Results of the nutrient content analysis of the needles	92
Table 18:	Overview of chamber measurements	97
Table 19:	Percentage of significant differences (Δm_i) of sample chamber 1 and 2	98
Table 20:	Conditions of the classes which were used for the classification of the measured data	98
Table 21:	Parameters of bi-variate weighted linear least-squares fitting regression analysis for data classes 1 -7 of sample chamber 1	108
Table 22:	Parameters of bi-variate weighted linear least-squares fitting regression analysis for data classes 1 -7 of sample chamber 2	109
Table 23:	Conditions of the classes which were used for the classification of the measured data within the ECHO project	122
Table 24:	Parameters of bi-variate weighted linear least-squares fitting regression analysis for data classes 1 -7 (ECHO data)	123
Table 25:	Measured and predicted leaf conductance to NO ₂ deposition	132
Table 26:	Averages of NO ₂ and O ₃ deposition velocities $(v_{dep,i})$ per ground area (LAI) and $v_{dep,NO2}$ corrected $(v_{dep,NO2})$ for NO ₂	
	compensation point concentration	135

List of Figures

Figure 1:	Scheme of oxidation of nitrogen oxides in the troposphere	4
Figure 2:	Biosphere-atmosphere interaction of the NO-NO ₂ -O ₃ -triad and volatile organic compounds (VOC) in a forest ecosystem	6
Figure 3:	Schematic representation of soil nitrogen cycle	8
Figure 4:	Illustrating the different pathways of trace gas uptake to a leaf	9
Figure 5:	Biochemical processes involved in foliar uptake and assimilation of NO_x and NH_3	11
Figure 6:	Schematic representation of the determination of bi-directional NO_2 exchange flux density, NO_2 deposition velocity and NO_2 compensation point concentration from measurements of NO_2 concentrations at the plant chamber's inlet and outlet under laboratory conditions.	25
Figure 7:	The dynamic plant chamber at well defined (laboratory) conditions: minimum detectable NO_2 compensation point concentration as function of NO_2 deposition velocity and the goodness of the ambient vs. sample NO_2 concentration measurements. Results are from data simulation.	28
Figure 8:	The dynamic plant chamber at well defined (laboratory) conditions: precision of NO_2 concentration measurements and precision of derived NO_2 exchange flux densities as function of the NO_2 concentration measured at the outlet of the dynamic chamber. Results are from data simulation.	29
Figure 9:	Photograph and schematic drawing of a dynamic chamber	38
Figure 10:	Schematic set-up of the system with three dynamic chambers	39
Figure 11	Distribution of forest species in Germany	41
Figure 12	Precision of the applied NO/NO ₂ analyzer during laboratory and field experiments and the precision of the blended NO_2 concentration used for fumigation of the young spruce trees in the laboratory.	57
Figure 13:	Response test for step changes between two different NO ₂ concentrations	58

XVI | LIST OF FIGURES

Figure 14:	Temporal course of blended NO ₂ concentrations used for fumigation of young spruce trees during the laboratory experiments	60
Figure 15:	Simultaneous measurements of radiation in and outside a chamber.	60
Figure 16:	Results of the response time test with helium.	62
Figure 17:	Laboratory NO ₂ fumigation of Norway Spruce under controlled conditions. NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber for application of 2σ -LOD-definition	64
Figure 18:	Switching scheme and time series of trace gas mixing ratios over two full measurement cycles during EGER field experiment	68
Figure 19:	Field measurements: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	69
Figure 20:	Field measurements: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	70
Figure 21:	Field measurements: NO concentration measured at the outlet of the dynamic plant chamber vs. NO concentration measured at the inlet of the dynamic plant chamber	71
Figure 22:	Percentage of gas-phase flux densities at the exchange flux densities for NO, NO_2 and O_3 .	80
Figure 23:	Photosynthetic light response curves of control and enclosed needles	91
Figure 24:	Temperature dependence of photosynthesis rate, in-situ measure- ments of CO_2 gas exchange on needle level in response to temperature	93
Figure 25:	Exchange flux densities of CO ₂ , H ₂ O, NO, NO ₂ , O ₃ with diurnal courses of PAR, leaf temperature and leaf conductance over the period from Jul 07 to Jul 08	94

Figure 26:	Exchange flux densities of CO ₂ , H ₂ O, NO, NO ₂ , O ₃ with diurnal courses of PAR, leaf temperature and leaf conductance over the period from Jun 29 to Jun 30	95
Figure 27:	NO ₂ measurements (EGER), data class 1 and 2 of sample chamber 1: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	100
Figure 28:	NO ₂ measurements (EGER), data class 3 and 4 of sample chamber 1: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	101
Figure 29:	NO ₂ measurements (EGER), data class 5 and 6 of sample chamber 1: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	102
Figure 30:	NO_2 measurements (EGER), data class 7 of sample chamber 1: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	103
Figure 31:	NO_2 measurements (EGER), data class 1 and 2 of sample chamber 2: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	104
Figure 32:	NO ₂ measurements (EGER), data class 3 and 4 of sample chamber 2: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	105
Figure 33:	NO_2 measurements (EGER), data class 5 and 6 of sample chamber 2: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	106
Figure 34:	NO ₂ measurements (EGER), data class 7 of sample chamber 2: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	107

XVIII | LIST OF FIGURES

Figure 35:	Variation of NO ₂ exchange flux density of sample chamber 1 for different leaf conductance classes	110
Figure 36:	Variation of NO ₂ exchange flux density of sample chamber 2 for different leaf conductance classes	111
Figure 37:	O_3 measurements (EGER), data class 1 and 2 of sample chamber 1: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	113
Figure 38:	O_3 measurements (EGER), data class 3 and 4 of sample chamber 1: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	114
Figure 39:	O_3 measurements (EGER), data class 5 and 6 of sample chamber 1: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	115
Figure 40:	O_3 measurements (EGER), data class 7 chamber 1: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	116
Figure 41:	O_3 measurements (EGER), data class 1 and 2 of sample chamber 2: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	117
Figure 42:	O_3 measurements (EGER), data class 3 and 4 of sample chamber 2: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	118
Figure 43:	O_3 measurements (EGER), data class 5 and 6 of sample chamber 2: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	119
		-

Figure 44:	O_3 measurements (EGER), data class 7 of sample chamber 2: O_3 concentration measured at the outlet of the dynamic plant chamber vs. O_3 concentration measured at the inlet of the dynamic plant chamber and O_3 exchange flux density vs. O_3 concentration measured at the outlet of the dynamic plant chamber	120
Figure 45:	Variation of O ₃ exchange flux density of sample chamber 1 for different leaf conductance classes	121
Figure 46:	Variation of O ₃ exchange flux density of sample chamber 2 for different leaf conductance classes	121
Figure 47:	NO_2 measurements (ECHO), data class 1 and 2: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	124
Figure 48:	NO_2 measurements (ECHO), data class 3 and 4: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	125
Figure 49:	NO_2 measurements (ECHO), data class 5 and 6: NO_2 concentration measured at the outlet of the dynamic plant chamber vs. NO_2 concentration measured at the inlet of the dynamic plant chamber and NO_2 exchange flux density vs. NO_2 concentration measured at the outlet of the dynamic plant chamber	126
Figure 50:	NO ₂ measurements (ECHO), data class 7: NO ₂ concentration measured at the outlet of the dynamic plant chamber vs. NO ₂ concentration measured at the inlet of the dynamic plant chamber and NO ₂ exchange flux density vs. NO ₂ concentration measured at the outlet of the dynamic plant chamber	127
Figure 51:	Variation of NO ₂ exchange flux density of measurements within ECHO for different leaf conductance classes	128

List of Abbreviations

General Abbreviations

BLC	Blue Light Converter
ECHO	Project: Emission and CHemical transformation of biogenic volatile Organic compounds
EGER	Project: ExchanGE processes in mountainous Regions
FEP	Fluorinated Ethylene Propylene
GOGAT	Glutamine OxoGlutarate AminoTransferase
GPT	Gas Phase Titration unit
GS	Glutamine Synthetase
IOP	Intensive Observation Period
LAI	Leaf Area Index
LOD	Limit Of Detection
NiR	Nitrite Reductase
NR	Nitrate Reductase
PAR	Photosynthetically Active Radiation
PFA	PerFluaroAlkoxy
ppm	parts per million (by volume)
ppb	parts per billion (by volume)
ppt	parts per trillion (by volume)
ppth	parts per thousand (by volume)
PTFE	PolyTetraFluoroEthylene
PVC	PolyVinyl Chloride

Roman symbols

A _{leaf}	leaf area	m ²
$F_{ex,i}$	exchange flux density of gas <i>i</i>	nmol $m^{-2} s^{-1}$
hv	photon's energy	J
$j(NO_2)$	photolysis rate of NO ₂	s ⁻¹
k	rate constant for chemical reactions	cm ³ molecule ⁻¹ s ⁻¹
m_i	slope of regression analysis of gas <i>i</i>	nmol m ⁻³
$m_{a,i}$	molar concentration in ambient air of gas <i>i</i>	nmol m ⁻³ , ppb
$m_{s,i}$	molar concentration within plant chamber of gas <i>i</i>	nmol m ⁻³ , ppb
<i>m_{comp,i}</i>	compensation point concentration of gas <i>i</i>	nmol m ⁻³ or ppb
M_i	molar mass of gas <i>i</i>	nmol s ⁻¹
n _i	intercept of regression analysis of gas <i>i</i>	nmol m ⁻³
Ν	number of samples	-
р	air pressure	hPa
Q	purging rate	$m^{3} s^{-1}$
R	universal gas constant	8.31441 J mol ⁻¹ K ⁻¹
R^2	regression coefficient	-
$S_{m_a,i}$	standard error	
$S_{m_s,i}$	standard error	
t	time	S
Т	temperature	°C or K
V	chamber volume	m ³
V _{dep,i}	deposition velocity of gas <i>i</i>	m s ⁻¹

XXII | LIST OF ABBREVIATIONS

Greek symbols

λ	wavelength of photon	nm
σ	standard deviation	
τ	characteristic time scale	S

Introduction

1.1 Nitrogen in the atmosphere

The earth's atmosphere consists to almost 78 % of non-reactive nitrogen (N_2) and 21 % of oxygen (O_2). The remaining 1 % of the atmospheric gases is characterized by a high diversity of low-concentrated so-called trace gases. Although trace gases compose only a small proportion of the atmosphere they contribute significantly to atmospheric chemical processes, the Earth's radiative budget and biogeochemical cycles.

Natural sources of atmospheric nitrogen compounds are nitrogen fixation by lightning and cosmic radiation as well as biogenic emissions of nitric oxide (NO) from natural and cultivated soils (SEINFELD and PANDIS 2006). Additionally anthropogenic sources of reduced nitrogen are fossil fuel combustion in industry and traffic, land-use and biomass burning. Table 1 presented an overview of the global nitrogen oxides (NO_x) sources.

sources	$\mathbf{NO}_{\mathbf{x}}$ (Tg-N yr ⁻¹)		
natural sources			
soil under natural vegetation	3.3		
lightning	5		
atmospheric chemistry	<0.5		
natural total	8.8		
anthropogenic sources			
fossil fuel combustion and industrial processes	33		
aircraft	0.7		
agriculture	2.3		
biomass and biofuel burning	7.1		
anthropogenic total	43.1		
total, all sources	51.9		

Table 1: Overview of global sources of nitrogen oxides (NO_x) (Tg-N yr⁻¹). Values are from the IPCC Fourth Assessment Report, according to DENMAN et al., 2007.

2 | INTRODUCTION

Over the past 100 years two anthropogenic activities have greatly increased the reactive nitrogen availability, food and energy production (GALLOWAY et al. 2004). And since the discovery to synthesize NH₃ from molecular N₂, known as the Haber-Bosch process, the human impact on the global nitrogen cycle raised drastically (ERISMAN et al. 2008). By now the emission of reactive nitrogen from human activities (food and energy production) increased by over a factor of 10 compared to the late 19th century (GALLOWAY et al. 2004).

Typical ambient nonurban NO₂ concentrations are 0.05 to 1 ppb (LERDAU et al. 2000). In regions of little industrial activity annual means of NO₂ mixing ratios are up to 5 ppb and in urban or industrialized regions about 20 ppb can be achieved. During smog events the NO₂ concentration may exceed 1 ppm (STULEN et al. 1998). An overview of NO_x found in the atmosphere of urban and rural sites is presented in Table 2. It is striking that at remote sites the concentrations are a few tenth of ppb up to 1000 ppb at urban environments. The human input is clearly visible in the sharply decreasing of the NO_x mixing ratios moving from urban to rural sites.

area	NO _x , ppb
urban - suburban	10 - 1000
rural	0.2 - 10
remote tropical forests	0.02 - 0.08
remote marine	0.02 - 0.04

Table 2: Typical NO_x mixing ratios (SEINFELD and PANDIS 2006).

This emission of reactive nitrogen influences biogeochemical processes in the atmosphere, in terrestrial ecosystems and in freshwater and marine aquatic ecosystems. The ecosystem productivity can be enhanced through fertilization or decreased through nutrient imbalance. The ecosystem biodiversity can be decreased through acidification and eutrophication. A higher reactive nitrogen concentration in the atmosphere can increase the incidence of human illness due to O_3 and particulate matter inhalation and the greenhouse potential of the atmosphere increases through N_2O production (GALLOWAY et al. 2004).

1.2 Chemistry of NO, NO₂ and O₃ in the troposphere

In the troposphere nitric oxide (NO), nitrogen dioxide (NO₂) and ozone (O₃) are one of the most important trace species of atmospheric chemistry (CRUTZEN 1979; LOGAN et al. 1983; WARNECK 1988; SEINFELD and PANDIS 2006). These gases are strongly related to each other and known as the NO-NO₂-O₃ triad.

Reactive nitrogen oxides are mainly emitted into troposphere as NO by natural and anthropogenic sources. In the atmosphere NO reacts with O_3 to form NO_2 (R1). Under daylight conditions NO_2 is photolyzed yielding NO (R2) again at wavelengths below 424 nm (*hv*). The formed oxygen atom reacts further to reproducing O_3 (R3):

$$NO + O_3 \rightarrow NO_2 + O_2 \tag{R1}$$

$$NO_2 + hv \rightarrow NO + O$$
 (R2)

$$O + O_2 + M \to O_3 + M \tag{R3}$$

where O is the oxygen radical and M symbolized a molecule which absorbs the released energy, like O₂, N₂ or H₂O.

Figure 1 illustrates the processing of nitrogen oxides in the troposphere. The reactions (R1) to (R3) are the main reactions of NO and NO₂ in the troposphere. All other reactions involving nitrogen oxides and O₃ are connected to this equilibrium. Further reactions of nitrogen oxides generally forms nitrate (NO₃⁻). This oxidation path is closely connected to the photochemistry of hydroxyl radicals (OH) and hydroperoxyl radicals (HO₂) with hydrocarbons, carbon monoxide (CO) (CRUTZEN 1979). Common intermediate products are nitrous acid (HNO₂), nitrate radicals (NO₃) and peroxyacetyl nitrate (PAN). There are two forms of nitrate present in the atmosphere, gaseous nitric acid (HNO₃) and nitrate bound to aerosol particles. Dry and wet deposition of these products are an effective sink of reactive nitrogen oxides.

4 | INTRODUCTION



Figure 1: Scheme of oxidation of nitrogen oxides in the troposphere. Photochemical processes are indicated by dashed arrows (after WARNECK 1988, MEIXNER 1994).

Nitrogen oxides are on special interest due to their regulation of the O_3 cycle and impact on the hydroxyl radical (OH) budget. In the absence of O_3 one of the dominant reaction pathways is the oxidation of NO by HO_2 :

$$HO_2 + NO \rightarrow NO_2 + OH$$
 (R4)

The OH radical is the main oxidant in the atmosphere and especially in the troposphere. It is extremely reactive and able to oxidize most of the chemical compounds found in the troposphere (SEINFELD and PANDIS 2006). Though NO is oxidized to NO_2 without the participation of O_3 the back reaction of NO_2 under daylight conditions produces O_3 . This is especially important in urban and industrial areas where high emissions of both NO and CO are present (SEINFELD and PANDIS 2006). Under low NO mixing ratios, the HO₂ radical reacts with O_3 which leads to an overall consumption of O_3 :

INTRODUCTION | 5

$$HO_2 + O_3 \rightarrow OH + 2 O_2 \tag{R5}$$

During night-time the NO-NO₂-O₃ triad is influenced by the formation of nitrate radicals (NO₃). These radicals are formed by the reaction of O₃ with NO₂ (R6). NO₃ comproportionates with NO back to NO₂ (R7):

$$O_3 + NO_2 \rightarrow NO_3 + O_2 \tag{R6}$$

$$NO + NO_3 \rightarrow 2 NO_2$$
 (R7)

Nitrate radicals react with organic molecules in the same way as OH radicals do. They remove a hydrogen atom from alkanes to form an organic alkyl radical (R), which then reacts with O_2 in the air to form peroxy radicals (RO₂). Formed RO₂ radicals are also able to oxidize NO. The availability of RO₂ and HO₂ depends on the oxidation path of CO, CH₄ and other hydrocarbons. Depending on the oxidation pathways RO₂ radicals can be formed or consumed. The used oxidation path depends on the nitrogen oxide mixing ratio in the air. Moreover, the mixing ratio of nitrogen oxides decides whether O₃ will be produced or consumed (CRUTZEN 1987). This is one of the main roles of nitrogen oxides in atmospheric chemistry. The lower limit of O₃ production is a nitrogen oxide mixing ratio of about 0.03 ppb. That means at higher values O₃ will be produced.

1.3 NO and NO₂ biosphere-atmosphere exchange in ecosystem forest

The exchange of reactive nitrogen between atmosphere and the ecosystem forest depends on turbulence, uptake by vegetation, deposition to soil, emission from soil and gas phase chemistry. The surface-atmosphere exchange of most gases is coupled to biological production and consumption processes. Figure 2 displays the NO-NO₂-O₃ triad in a forest ecosystem. NO is emitted from soil into atmosphere. If the ground is covered by vegetation a reduced wind velocity will extend the residence time of the gas inside the vegetation stand. Under these conditions parts of the emitted NO are able to react with O₃ to form NO₂ (see (R1)). Shading by plants reduces the photolysis of NO₂ (see (R2)) in this area. Within the vegetation stand surface exchange processes take

6 | INTRODUCTION

place and photochemical reactions continue in the atmosphere. Thus, the understanding of the separate production and destruction processes and their link with exchange processes is necessary for an understanding of the ecosystem forest.



Figure 2: Biosphere-atmosphere interaction (surface exchange and (photo-)chemistry processes) of the NO-NO₂-O₃-triad and volatile organic compounds (VOC) in a forest ecosystem (after COE et al 1993).

Nitrogen is essential to the nutrition of plants and animals. It is a constituent in all proteins and in the nucleic acids of all organisms. But the atmospheric nitrogen is not available for most biological organisms because the gaseous nitrogen molecules have very strong bonds, making the gas chemically stable (SEINFELD and PANDIS 2006). Accordingly, the availability of nitrogen is often a limiting factor for the biomass production of an ecosystem. To be useful for higher plants and animals, atmospheric nitrogen has to be converted to a reduced state. Some species of bacteria possess the enzyme nitrogenase which can convert atmospheric nitrogen into ammonium (NH4⁺) which can be metabolized by plants. These nitrogen-fixing organisms belong to the procaryotes (e.g. bacteria, cyanobabteria, actinomycetes) and may live freely in soil (e.g. *Azotobacter chroococcum, Clostridium pasteurianum*) or are symbionts (e.g. *Rhizobium, Bradyrhizobium*). Green plants can take up nitrogen directly from soil as nitrate (NO₃⁻) or as ammonium ions like mineral elements. The soil nitrate is derived

from natural mineral deposits, artificial fertilizers, animal waste or organic decay as the product of bacterial nitrification. Nitrates absorbed in this fashion are converted to nitrites (NO_2^{-}) by the enzyme nitrate reductase (NR), and then converted to ammonia by another enzyme called nitrite reductase (NiR).

1.3.1 Processes in soil

Within the soil NO can be produced and consumed. These processes are incorporated into the metabolism of microorganisms. Hereby two main biological processes are responsible, nitrification and denitrification (WILLIAMS et al. 1992). Figure 3 displays a schematic representation of the soil nitrogen cycle and the uptake of nitrogen compounds by plants.

Nitrification is a mainly aerobic process in which ammonium (NH_4^+) is oxidized to nitrite (NO_2^-) and nitrate (NO_3^-) . It is a two-step process in which two different groups of microorganisms are involved, *Nitroso*-bacteria (ammonia oxidizers) and *Nitro*-bacteria (nitrite oxidizers). In the first step *Nitroso*-bacteria oxidize NH_4^+ via hydroxylamine (NH_2OH) to NO_2^- and in the second step *Nitro*-bacteria oxidize NO_2^- to NO_3^- . Within the oxidation process formation of gaseous NO and N_2O as intermediate compounds is observed (ROBERTSON and GROFFMAN 2007). But the exact pathway of this formation is still not clarified (LUDWIG et al. 2001). NO can also be produced via NO_2^- reduction by *Nitroso*-bacteria (LUDWIG et al. 2001; ROBERTSON and GROFFMAN 2007). This occurs when O_2 is limited and the bacteria use NO_2^- as an electron acceptor (BOLLMANN and CONRAD 1998).

Denitrification occurs under anaerobic conditions and is the reduction of NO_3^- to N_2 and the intermediate compounds NO_2^- , NO and N_2O . These microorganisms use $NO_3^$ rather than O_2 as a terminal electron acceptor (ROBERTSON and GROFFMAN 2007). The denitrifiers are aerobic microorganisms, which can switch to anaerobic denitrification when O_2 is limited. Moreover O_2 is the more efficient electron acceptor, hence most denitrifies only carry out denitrification when O_2 is unavailable. This happens especially after rainfalls, when the soil pores become filled up with water and the O_2 diffusion through the soil is slow.



Figure 3: Schematic representation of soil nitrogen cycle. Nitrogen from the atmosphere is converted by bacteria into nitrogen compounds which are useable for plants.

1.3.2 Interaction with plants

Plants get nitrogen from the soil in form of nitrate (NO₃⁻) or ammonium (NH₄⁺) by absorption of their roots. If NO₃⁻ is absorbed, it is inside the cells first reduced to nitrite (NO₂⁻) by the enzyme nitrate reductase (NR) and then converted to NH₄⁺ by the enzyme nitrite reductase (NiR) for incorporation into amino acids, nucleic acids and chlorophyll (see below). Plants of the *Fabaceae* family (legumes) have a symbiotic relationship with rhizobia. These soil bacteria were located inside root nodules and convert atmospheric nitrogen into a form of nitrogen, which is usable to the host plant. For the nitrogen fixation the enzyme nitrogenase is used. After absorption of NH₄⁺ by the plant, ammonium is incorporated into amino acids by the enzyme glutamine synthetase (see below).

Another source of nitrogen for plants is the uptake of atmospheric nitrogen by leaves. In general, plants are considered as a sink of atmospheric nitrogen oxides. Since the studies by HILL (1971), the deposition of atmospheric NO and NO₂ is demonstrated for different plants. The uptake by plants depends on the plant species. HANSON and LINDBERG (1991) provide a comprehensive overview of the deposition of NO₂ to leaf and canopy surfaces, respectively. The authors show that the NO₂ leaf conductances under daytime conditions differ by two magnitudes. Continuative studies have identified some factors, which make a contribution to the different uptake rates. JENSEN and PILEGAARD (1993) found out that the nitrate supply in the soil have an effect on the NO₂ uptake rate. Also the salinity of the soil plays a role (FUHRER and ERISMANN 1980). Moreover, uptake rates differ also inside one species. Apparently the plant developmental stage is an important factor as indicated by studies of GRENNFELT et al. (1983) who investigated different ages of Scots pine needles. They identified NO₂ uptake variability of about 100 %.

The uptake of NO₂ proceeds mainly by diffusion through the stoma (SKÄRBY et al. 1981; SAXE 1986; HANSON et al. 1989; THOENE et al. 1991). A smaller fraction of NO₂ is absorbed by the cuticula of the leaves or the surface water film if present (KISSER-PRIESACK et al. 1987, 1990; BURKHARD and EIDEN 1994) (Figure 4).



Figure 4: Illustrating the different pathways of trace gas uptake to a leaf.

10 | INTRODUCTION

The deposition on a dry or wet cuticle is at least one or two orders of magnitude lower than the stomatal uptake (WELLBURN 1990). The NO₂ molecule can undergo irreversible as well as reversible reactions with phenolic components of the cuticle (LENDZIAN and KERSTIENS 1988; KISSER-PRIESACK et al. 1990) Hence, NO₂ can also be re-emitted into atmosphere. However, these processes seem to be negligible in comparison to the total NO₂ flux.

The gas uptake through the stomata is driven by the concentration gradient between the gas phase inside and outside the leaf. Several studies have demonstrated a linear increase of NO₂ uptake with rising atmospheric NO₂ concentration (JOHANSSON 1987; THOENE at al. 1991, 1996).

A plant can control the entry of gases into the leaf by varying the stomatal aperture. Number, distribution, size, shape and activity of the stoma are species specific. Additional they vary with adaptation to the place of growth and one individual to another as well. The stomatal movement is regulated mainly by two controlling cycles, carbon dioxide and water. The degree of opening is adjusted continuously to changes in the environment, such as light intensity, temperature, air humidity and carbon dioxide concentration.

In the substomatal cavity NO₂ dissolves rapidly in the aqueous phase of the apoplastic space where it disproportionates to NO₂⁻ and NO₃⁻ (LEE and SCHWARTZ 1981; RAMGE et al. 1993). Due to the better solubility in aqueous solutions NO₂ will be dissolved easier than NO. However, the observed uptake rate of NO₂ cannot be explained by the liquid solubility only. The disproportionation reaction appears too slow to explain the measured leaf fluxes of NO₂ (PARK and LEE 1988; RAMGE et al. 1993). Hence, the reduction of NO₂ by apoplastic antioxidants, particularly ascorbate, has been proposed (RAMGE et al. 1993). The theoretical calculations of RAMGE et al. (1993) demonstrated sufficient rates to explain observed NO₂ leaf fluxes if the reactions between water and NO₂ and between NO₂ has been experimentally demonstrated by TEKLEMARIAM and SPARKS (2006). They observed a significant correlation between leaf ascorbate concentrations and the leaf fluxes of NO₂. However, apoplastic ascorbate concentrations differ between species but also between individuals of the same species depending on environmental factors (POLLE et al. 1995; LUWE 1996).
Apoplastic NO₂ and NO₃⁻ can be incorporated into the general nitrogen metabolism of the leaves. Taking all these steps into account Figure 5 gives an overview about the different assimilation pathways for atmospheric NO, NO₂ and NH₃. After taken up into the apoplast the formed NO₂⁻ and NO₃⁻ are transported into the cytoplasm of mesophyll cells (AMMANN et al. 1995) where NO₃⁻ is reduced to NO₂⁻ by nitrate reductase (NR) (THOENE et al. 1991, TISCHNER 2000). NO₂⁻ is then transported into the chloroplast, where it is reduced to NH₄⁺ by nitrite reductase (NiR). Moreover, NH₄⁺ can be formed by assimilation of gaseous ammonia (NH₃). The NH₄⁺ is then incorporated into the amino acid glutamine by the enzyme glutamine synthetase (GS) inside the cytoplasm. Alternatively, NH₄⁺ can be incorporated by chloroplastic glutamate synthetase, a step also known as GOGAT cycle (LEA and MIFLIN 1974). Different studies revealed that GS is located in the cytoplasm (GS₁) and in the chloroplast (GS₂) of plant cells (MAECK 1995; SAKAKIBARA et al. 1996).



Figure 5: Biochemical processes involved in foliar uptake and assimilation of NO_x and NH_3 . Dashed lines indicate the possible role of cytosolic glutamine synthetase (GS₁) or chloroplastic glutamate synthetase (GS₂) in the assimilation of NH_4^+ derived from gaseous NH_3 (after LEA et al. 1994; STULEN et al. 1998)

12 | INTRODUCTION

The ability of plants to incorporate atmospheric NO₂ into free amino acids was demonstrated by numerous studies using ¹⁵N as a tracer (NUSSBAUM et al. 1993; WEBER et al. 1995; YONEYAMA et al. 2003). The studies have demonstrated that the primary assimilation of inorganic nitrogen into amino acids is largely through the GS/GOGAT pathway (YONEYAMA et al. 2003).

The question weather atmospheric nitrogen uptake by leaves affects the nitrogen uptake by roots is still under discussion. According to SCHULZE (1989) additional leaf uptake of atmospheric nitrogen causes a nutrient imbalance due to nitrogen to cation discrepancies, which result in decline symptoms like needle yellowing and loss. RENNENBERG and GEBLER (1999) reported on a down-regulation of the nitrogen uptake by roots to an extent that equals nitrogen uptake by leaves. We understand that the foliar uptake is significant enough (10 - 25 %) to influence plant metabolism but the complex regulation between root uptake, nitrogen availability and foliar uptake is a complex interaction of metabolic processes and physiological regulations and need further research (VALLANO and SPARKS 2008).

Moreover, plants can also act as a source of gaseous nitrogen compounds. The emission of ammonia (NH₃) by plants has been reported (KESSELMEIER et al. 1993; FANGMEIER et al. 1994; SCHJOERRING et al. 1998, 2000). Contrasting the emission of NO and NO₂ by plants is not accepted finally. NO emission is demonstrated only by a few studies (KLEPPER 1997; DEAN and HARPER 1986; WILDT et al. 1997). However, WILDT et al. (1997) estimated that plants emitted only 1 - 5 % NO compared to the global emission rate of NO from soils. Because NO has a lower water solubility than NO₂, the NO uptake by plants is lower than for NO₂ (MEIXNER 1994).

Similarly the potential emission of NO₂ by plants is still under discussion. A release may be expected when atmospheric NO₂ mixing ratios are below a certain compensation point concentration. LERDAU et al. (2000) reported that depending on the leaf area indices of the relevant sites only 25 to max. 80 % of the NO_x mainly derived from NO emission is escaping the forest (see JACOB and WOFSY 1990; YIENGER and LEVY 1995; WANG et al. 1998). However, such results do not agree with leaf-level measurements reporting about NO₂ emission from plants (besides plant uptake of NO₂). Corresponding compensation point concentrations of NO₂ between 0.3 and 3 ppb have been reported (RONDÓN et al. 1993; THOENE et al. 1996; WEBER and RENNENBERG 1996a; SPARKS et al. 2001; GEBLER et al. 2000, 2002; HEREID and MONSON 2001), suggesting plants to act as a sink for atmospheric NO₂ when ambient NO₂ concentrations are exceeding, or as a source of NO₂ when ambient NO₂ concentrations are below the NO₂ compensation point concentration. Table 3 gives an overview of reported NO₂ compensation points. According to LERDAU et al. (2000), these results and discussions contradict the reports of JACOB and WOFSY (1990), who demonstrated that even at ambient NO₂ concentrations of 0.2 to 0.4 ppb a strong uptake of NO₂ by plants (primary rainforest) is still required to align measured NO₂ concentrations in the canopy with the measured NO soil emission rates. LERDAU et al. (2000) emphasized the urgent need to find an explanation for this discrepancy, particularly in remote regions far away from anthropogenic NO_x sources (e.g. primary rain and boreal forests under low NO_x regimes). Thus it is required to investigate the contribution of the NO₂ uptake by plants and to ensure NO₂ compensation point concentrations and to ensure NO₂ compensation point concentrations at (sub-) ppb levels.

NO ₂ compensation point, ppb	plant species	author
0.03 - 17*	Scots pine	Raivonen et al. (2009)
1 - 3	Scots pine (Pinus sylvestris)	Johansson (1987)
1.8 - 1.9	Beech (Fagus sylvatica)	Geßler et al. (2000)
1.7	Norway spruce (Picea abies)	Geßler et al. (2002)
1.64 ± 0.3	Norway spruce (Picea abies)	Thoene et al. (1996)
0.53 - 1.6	tropical trees	Sparks et al. (2001)
1.0 - 1.2	Wheat (<i>Triticum aestivum</i>) Corn (<i>Zea mays</i>) Sunflower (<i>Helianthus annus</i>) Catharanthus (<i>Madagascar periwinkle</i>)	Teklemariam & Sparks (2006)
1.15	Wheat (Triticum aestivum)	Weber and Rennenberg (1996)
0.02 - 1.1	European tree species (Fagus sylvatica, Quercus robur, Quercus ilex, Betula pendula, Pinus sylvestris)	Chaparro-Suarez et al. (2011)
0.9	Corn (Zea mays)	Hereid & Monson (2001)
0.1 - 0.7	Scots pine (Pinus sylvestris) Norway spruce (Picea abies)	Rondón et al. (1993)
<0.1 - 0.6	Norway spruce (Picea abies)	Rondón & Granat (1994)
<0.1 - 0.3	Scots pine (Pinus sylvestris)	Rondón & Granat (1994)

Table 3: Reported NO₂ (or NO_y) compensation point concentrations obtained from literature.

1.4 Objectives of thesis

In this study measurements with a dynamic chamber system were performed to determine plant surface-atmosphere exchange fluxes of NO₂ (NO and O₃) under typical field conditions (uncontrolled) for a relatively remote, managed Norway spruce forest site as well as under controlled conditions including (laboratory) fumigation experiments. Because NO₂ compensation point concentrations were reported at (sub-)ppb levels, our laboratory NO₂ fumigation experiments were performed with 3- to 4-yr old Norway Spruce trees at 0.3 - 3.4 ppb. Also under field conditions, such low ambient NO₂ concentrations can be expected.

Moreover, exchange fluxes derived from dynamic chamber measurements are based on generally (very) small differences of NO₂ (NO, O₃) concentrations between inlet and outlet of the chamber. Consequently, detection limits of corresponding analyzers, statistical significance of the concentration differences, as well as the statistical goodness of measurements definitely have a substantial impact on the identification and quantification of statistically significant deposition velocities and compensation point concentrations, and have been considered correspondingly. Furthermore, as the exchange of NO₂ is a complex interaction of transport, chemistry and plant physiology, in our field experiments we determined fluxes of NO, NO₂, O₃, CO₂ and H₂O.

This thesis presents basic considerations of dynamic plant chamber system measurements and the constraints of precision and design for chamber measurements as well as results of laboratory and field measurements. In chapter 3 specification and implementation of the performed dynamic plant chamber system are presented. The performance of data analysis is demonstrated on values from laboratory experiments and selected results from field measurements. The application of the chamber system to field measurements and the results for the trace gas exchange between plants and the atmosphere under field conditions are described in chapter 4.

Material and Methods

The commonly used technique for leaf-level exchange measurements of NO₂ is the dynamic chamber technique (a technique also used for many non-reactive (e.g. CO₂, H₂O, COS) and reactive trace gases (e.g. NO, O₃, VOCs, DMS, CS₂, HONO, HNO₃, CH₂O, HCOOH, CH₃COOH)). Here, an entire plant (or parts of a plant) is enclosed in a (transparent) chamber which is purged by (preferably ambient) air. Two measurements of NO_2 concentration are usually performed, namely (1) at the entrance of the chamber (= ambient NO_2 concentration) and (2) within the chamber. If the chamber is well mixed, the latter measurement can be replaced by that of the outlet NO₂ concentration. Alternatively, a set of two chambers, one enclosing the plant the other being empty, can be used. To relate these two concentration measurements to the exchange (i.e. the unior bi-directional flux) of NO₂ between the (chamber) atmosphere and the enclosed plant (or parts of plant), the full mass balance of the dynamic chamber must be considered, i.e. NO₂ fluxes entering and leaving the chamber, as well as all other fluxes due to NO₂ sinks and sources within the chamber's volume. Under typical field conditions (i.e. ambient air enters the dynamic chamber), not only NO₂ is purged through the chamber, but also ambient NO and O₃. Fast reaction between NO and O₃ forms a "chemical" source of NO₂, while (under daylight conditions) photolysis of NO₂ ($\lambda = 420$ nm) is a "chemical" sink. Depending on actual ambient NO₂, NO and O₃ concentrations as well on UV irradiation intensity, corresponding "gas phase fluxes" may reach the magnitude of the NO₂ flux from/to the enclosed plant(s) (MEIXNER et al. 1997; PAPE et al. 2009). Consequently, simultaneous measurements of NO₂, NO and O₃ concentrations at the outlet of the chamber are required. However, since there is substantial uptake of O₃ by the plants (to a much lesser extent also of NO), NO₂, NO and O₃ concentrations at the inlet of the chamber have to be measured, too. As a positive "by-product" of these additional concentration measurements, deposition velocities of O₃ (and NO) may be inferred considering the dynamic chamber's mass balances of O₃ and NO.

2.1 Basic considerations

A small branch of a tree (leaf area A_{leaf}), which is enclosed in a transparent plant chamber of volume V was considered. The air within the plant chamber is well mixed by action of one (or more) fan(s). Ambient air (containing NO₂, NO and O₃) is entering the plant chamber at the inlet, flushing the chamber with the purging rate Q (m³ s⁻¹) and leaving the chamber at the outlet. Within plant chamber trace gases of the NO-NO₂.O₃ triad may be (a) emitted and/or taken up from/by leaves, (b) deposited to the inner walls of the plant chamber and (c) destroyed and/or generated by (fast) photo-chemical reactions.

2.1.1 Mass balance of the NO-NO₂-O₃ triad of a dynamic plant chamber

Considering the molar mass flux of the trace gas i ($i = NO_2$, NO, O₃), i.e. the derivative of molar mass M_i with respect to time ($\partial M_i / \partial t$ in nmol s⁻¹), the individual flux components of the dynamic plant chamber system are defined as follows:

 $\partial M_{in,i}/\partial t$:= molar mass flux of trace gas *i* entering the plant chamber

 $\partial M_{out,i}/\partial t :=$ molar mass flux of trace gas *i* leaving the plant chamber

- $\partial M_{wall,i}/\partial t :=$ molar mass flux of trace gas *i* to the inner wall of the plant chamber (due to ad-/absorption of trace gas *i*)
- $\partial M_{em,i}/\partial t :=$ molar mass flux of trace gas *i* caused by (biogenic) emission from the leaves
- $\partial M_{dep,i}/\partial t :=$ molar mass flux of trace gas *i* caused by uptake to the leaves (e.g. cuticular, stomatal and/or mesophyllic uptake)
- $\partial M_{prod,i}/\partial t$:= molar mass flux of trace gas *i* into the plant chamber's volume caused by gas phase production, i.e. from photochemical decay or fast chemical reaction of other trace gas(es)
- $\partial M_{dest,i}/\partial t :=$ molar mass flux of trace gas *i* out of the plant chamber's volume caused by gas-phase destruction, i.e. by photochemical decay of trace gas *i* or by fast chemical reaction with other trace gas(es).

Under steady-state conditions (i.e. concentrations of trace gas i are constant (have reached equilibrium)) and considering the convention, that mass fluxes into (out) of the plant chamber's volume are counted positive (negative), the molar mass flux balance of the trace gas i is given by

$$+\frac{\partial M_{in,i}}{\partial t} - \frac{\partial M_{out,i}}{\partial t} - \frac{\partial M_{wall,i}}{\partial t} + \frac{\partial M_{em,i}}{\partial t} - \frac{\partial M_{dep,i}}{\partial t} + \frac{\partial M_{prod,i}}{\partial t} - \frac{\partial M_{dest,i}}{\partial t} = 0$$
(1)

While the first three and the last two left-hand terms of Eq. (1) may be known and/or are determined by laboratory or *in-situ* measurements, $\partial M_{em,i}/\partial t$ and $\partial M_{dep,i}/\partial t$ are the unknown fluxes of trace gas *i*. We combine these two fluxes to the bi-directional "exchange flux" $\partial M_{ex,i}/\partial t$

$$\frac{\partial M_{ex,i}}{\partial t} = + \frac{\partial M_{em,i}}{\partial t} - \frac{\partial M_{dep,i}}{\partial t} \qquad i = NO_2, NO, O_3$$
(2)

Considering the purging rate Q (m³ s⁻¹) and the molar concentration $m_{a,i}$ (nmol m⁻³) of trace gas *i* in ambient air, the ingoing flux is

$$\frac{\partial M_{in,i}}{\partial t} = Q \cdot m_{a,i} \qquad \qquad i = NO_2, NO, O_3$$
(3)

The molar concentration at the outlet of the plant chamber is equivalent to the molar concentration within the plant chamber ($m_{s,i}$ in nmol m⁻³), provided the plant chamber's volume is well mixed by one (or more) appropriate fan(s) (see MEIXNER et al. 1997; PAPE et al. 2009). Then, the flux leaving the chamber is defined by

$$\frac{\partial M_{out,i}}{\partial t} = Q \cdot m_{s,i} \qquad \qquad i = NO_2, NO, O_3$$
(4)

The flux to the inner walls can be easily determined by corresponding laboratory experiments (e.g. LUDWIG 1994; MEIXNER et al. 1997). If the material of the plant chamber is consisting of chemically inert material, the flux $\partial M_{wall,i}/\partial t$ can usually be

neglected. In case of the NO-NO₂-O₃ triad, the relevant photochemical reactions controlling the gas-phase production and destruction of the respective trace gas are

$$NO + O_3 = NO_2 + O_2, \qquad k_{R1} := k = 1.4 \cdot 10^{-12} \cdot e^{(-1310/T)}$$
 (see R1)

$$NO_2 + h\nu = NO + O$$
, $k_{R2} \coloneqq j(NO_2)$, $\lambda \le 420 nm$ (see R2)

Applying simple reaction kinetics, the corresponding fluxes $\partial M_{prod,i}/\partial t$ and $\partial M_{dest,i}/\partial t$ are given by

$$\frac{\partial M_{prod,NO2}}{\partial t} = \frac{\partial M_{dest,NO}}{\partial t} = \frac{\partial M_{dest,O3}}{\partial t} = V \cdot k \cdot m_{s,NO} \cdot m_{s,O3}$$
(5)

and

$$\frac{\partial M_{dest,NO2}}{\partial t} = \frac{\partial M_{prod,NO}}{\partial t} = \frac{\partial M_{prod,O3}}{\partial t} = V \cdot j(NO_2) \cdot m_{s,NO2}$$
(6)

where V is the plant chamber's volume (m³), k is the (temperature-dependent) reaction coefficient of the NO + O₃ reaction (m³ nmol⁻¹ s⁻¹) (ATKINSON et al. 2004) and $j(NO_2)$ (s⁻¹) is the photolysis rate of reaction (R2), which can be measured *in-situ* (or parameterized from data of global radiation; see TREBS et al. 2009).

Considering Eqs. (1) - (6), the molar mass flux balances of the trace gas triad $NO-NO_2-O_3$ (under steady state conditions) can be formulated as follows:

$$\frac{\partial M_{ex,NO2}}{\partial t} = Q \cdot m_{s,NO2} - Q \cdot m_{a,NO2} - V \cdot k \cdot m_{s,NO} \cdot m_{s,O3} + V \cdot j(NO_2) \cdot m_{s,NO2}$$
(7.1)

$$\frac{\partial M_{ex,NO}}{\partial t} = Q \cdot m_{s,NO} - Q \cdot m_{a,NO} + V \cdot k \cdot m_{s,NO} \cdot m_{s,O3} - V \cdot j(NO_2) \cdot m_{s,NO2}$$
(7.2)

$$\frac{\partial M_{ex,O3}}{\partial t} = Q \cdot m_{s,O3} - Q \cdot m_{a,O3} + V \cdot k \cdot m_{s,NO} \cdot m_{s,O3} - V \cdot j(NO_2) \cdot m_{s,NO2}$$
(7.3)

Equations (7.1) - (7.3) explicitly define the molar mass fluxes (in nmol s⁻¹) of the NO₂, NO and O₃ surface exchange between the plant chamber's atmosphere and the enclosed leaves in terms of measured and/or *a priori* known quantities only.

2.1.2 Molar mass flux densities, deposition velocities and compensation point concentrations

Equations (7.1) - (7.3) are formulated in terms of molar mass fluxes (in nmol s⁻¹). However, considering the exchange of reactive trace gases between the plant chamber's atmosphere and the enclosed leaves, the exchange flux density ($F_{ex,i}$) of the molar mass (in nmol m⁻² s⁻¹) is commonly used rather than the molar mass flux itself. In case of plant chamber studies, the appropriate reference surface (reference area) is the surface area (A_{leaf} , in m²) of the leaves. Therefore, the exchange flux density $F_{ex,i}$ is defined as $F_{ex,i} := (\partial M_i / \partial t) / A_{leaf}$, and the corresponding balance equations will read as follows:

$$F_{ex,NO2} = -\frac{Q}{A_{leaf}} \left(m_{a,NO2} - m_{s,NO2} + \frac{V}{Q} k m_{s,NO} m_{s,O3} - \frac{V}{Q} j (NO_2) m_{s,NO2} \right)$$
(8.1)

$$F_{ex,NO} = -\frac{Q}{A_{leaf}} \left(m_{a,NO} - m_{s,NO} - \frac{V}{Q} k m_{s,NO} m_{s,O3} + \frac{V}{Q} j (NO_2) m_{s,NO2} \right)$$
(8.2)

$$F_{ex,O3} = -\frac{Q}{A_{leaf}} \left(m_{a,O3} - m_{s,O3} - \frac{V}{Q} k m_{s,NO} m_{s,O3} + \frac{V}{Q} j (NO_2) m_{s,NO2} \right)$$
(8.3)

In case of defined laboratory experiments, where plants may be fumigated with only one of the three trace gases (i.e., gas-phase production and/or destruction of the trace gas can be ruled out), Eqs. (8.1) - (8.3) will reduce to the well-known form of

$$F^*_{ex,i} = -\frac{Q}{A_{leaf}} (m_{a,i} - m_{s,i}) \qquad i = NO_2, NO, O_3$$
(8.4)

In case of bi-directional exchange (see Eq. (2)), the exchange between the plant chamber's atmosphere and the leaves can be directed to or away from the leaves. This exchange process can be subject to the so-called "compensation point concentration" ($m_{comp,i}$, in nmol m⁻³). According to CONRAD (1994), $m_{comp,i}$ is "that concentration at which the consumption rate reaches the same value as the production rate, so that the result of both processes is zero flux". The exchange flux density $F_{ex,i}$ is commonly parameterized (e.g. HICKS et al. 1987) by the so-called "deposition velocity" $v_{dep,i}$ (in m s⁻¹ or mm s⁻¹) of trace gas *i* and its compensation point concentration, $m_{comp,i}$:

$$F_{ex,NO2} = -v_{dep,NO2} \left(m_{s,NO2} - m_{comp,NO2} \right)$$
(9.1)

$$F_{ex,NO} = -v_{dep,NO} \left(m_{s,NO} - m_{comp,NO} \right)$$
(9.2)

$$F_{ex,O3} = -v_{dep,O3} \quad \left(m_{s,O3} - m_{comp,O3}\right) \tag{9.3}$$

Note, that (by convention) $F_{ex,i}$ is directed "downward" to the leaves, if $m_{s,i} > m_{comp,i}$, $F_{ex,i}$ is zero, if $m_{s,i} = m_{comp,i}$ and $F_{ex,i}$ is directed "upward" from the leaves, if $m_{s,i} < m_{comp,i}$.

Given, that the quantities Q, A_{leaf} , k and $j(NO_2)$ are *a priori* known and/or simultaneously measured with $m_{s,i}$ and $m_{a,i}$. Then, the desired quantities, $v_{dep,i}$ and $m_{comp,i}$, are commonly determined from the linear relationship between $F_{ex,i}$ and $m_{s,i}$, where $v_{dep,i}$ is the slope and $m_{comp,i}$ is the intersect of $F_{ex,i}$ with the $m_{s,i}$ -axis (see RONDÓN and GRANAT 1994; THOENE et al. 1996; WEBER and RENNENBERG 1996a; SPARKS et al. 2001; HEREID and MONSON 2001; GEBLER et al. 2002).

However, due to the fact, that $F_{ex,i}$ (see Eqs. (8.1) - (8.3)) contains the term $Q/A_{leaf} \cdot (m_{a,i} - m_{s,i})$, the calculation of any form of linear regression between F_{ex} and $m_{s,i}$ is mathematically not correct, because the dependent variable $F_{ex,i}$ contains the independent variable $(m_{s,i})$.

This problem can be resolved by returning to the originally measured quantities, $m_{a,i}$ and $m_{s,i}$. If we combine Eqs. (8.1) - (8.3) and Eqs. (9.1) - (9.3) and resolve these equations for $m_{s,NO2}$, $m_{s,NO}$ and $m_{s,O3}$, we yield three linear relationships between the measured variables $m_{s,NO2}$ and $m_{a,NO2}$, $m_{s,NO}$ and $m_{a,NO}$ and $m_{a,O3}$:

$$m_{s,NO2} = n_1 + m_1 \cdot m_{a,NO2} \tag{10.1}$$

$$m_{s,NO} = n_2 + m_2 \cdot m_{a,NO} \tag{10.2}$$

$$m_{s,03} = n_3 + m_3 \cdot m_{a,03} \tag{10.3}$$

using the definitions:

$$n_{1} \coloneqq \frac{\overline{A}_{leaf} v_{dep,NO2} m_{comp,NO2} + V \overline{k} \overline{m}_{s,NO} \overline{m}_{s,O3}}{\overline{Q} + \overline{A}_{leaf} v_{dep,NO2} + V \overline{j} (NO_{2})} ; \quad m_{1} \coloneqq \frac{\overline{Q}}{\overline{Q} + \overline{A}_{leaf} v_{dep,NO2} + V \overline{j} (NO_{2})}$$
(11.1)

$$n_{2} := \frac{\overline{A}_{leaf} v_{dep,O3} m_{comp,O3} + V \,\overline{j} (NO_{2}) \overline{m}_{s,NO2}}{\overline{Q} + \overline{A}_{leaf} v_{dep,O3} + V \,\overline{k} \,\overline{m}_{s,O3}} ; \quad m_{2} := \frac{\overline{Q}}{\overline{Q} + \overline{A}_{leaf} v_{dep,NO} + V \,\overline{k} \,\overline{m}_{s,O3}}$$
(11.2)

$$n_{3} := \frac{\overline{A}_{leaf} v_{dep,O3} m_{comp,O3} + V \,\overline{j} (NO_{2}) \overline{m}_{s,NO2}}{\overline{Q} + \overline{A}_{leaf} v_{dep,O3} + V \,\overline{k} \,\overline{m}_{s,NO}} ; \quad m_{3} := \frac{\overline{Q}}{\overline{Q} + \overline{A}_{leaf} v_{dep,O3} + V \,\overline{k} \,\overline{m}_{s,NO}}$$
(11.3)

The quantities n_i and m_i may be evaluated (graphically) as the intercept and the slope of the plot of measured $m_{s,i}$ versus measured $m_{a,i}$. Application of different forms of linear regression analysis delivers n_i and m_i and bi-variate weighted linear least-squares fitting (which considers uncertainties of both, $m_{s,i}$ and $m_{a,i}$) provides also their standard errors $s_{n,i}$ and $s_{m,i}$ (see Sect. 3.1.6).

The linear relationships between $F_{ex,i}$ and $m_{s,i}$ are still maintained. This can be shown by resolving Eqs. (10.1) - (10.3) for $m_{a,i}$ and making use of Eqs. (8.1) - (8.3):

$$F_{ex,NO2} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{n_1}{m_1} - \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,NO} \overline{m}_{s,O3} \right) + \frac{\overline{Q}}{\overline{A}_{leaf}} \left(1 - \frac{1}{m_1} + \frac{V}{\overline{Q}} \,\overline{j} (NO_2) \right) \cdot m_{s,NO2} \tag{12.1}$$

$$F_{ex,NO} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{n_2}{m_2} - \frac{V}{\overline{Q}} \,\overline{j} (NO_2) \,\overline{m}_{s,NO2} \right) + \frac{\overline{Q}}{\overline{A}_{leaf}} \left(1 - \frac{1}{m_2} + \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,O3} \right) \cdot m_{s,NO} \tag{12.2}$$

$$F_{ex,O3} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{n_3}{m_3} - \frac{V}{\overline{Q}} \,\overline{j} (NO_2) \,\overline{m}_{s,NO2} \right) + \frac{\overline{Q}}{\overline{A}_{leaf}} \left(1 - \frac{1}{m_3} + \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,NO} \right) \cdot m_{s,O3} \tag{12.3}$$

Finally, the desired deposition velocities $(v_{dep,i})$ of the NO-NO₂-O₃ triad result from Eqs. (11.1) - (11.3), resolving for $v_{dep,i}$,

$$v_{dep,NO2} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{1}{m_1} - 1 - \frac{V}{\overline{Q}} \, \overline{j}(NO_2) \right)$$
(13.1)

$$v_{dep,NO} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{1}{m_2} - 1 - \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,O3} \right)$$
(13.2)

$$v_{dep,O3} = \frac{\overline{Q}}{\overline{A}_{leaf}} \left(\frac{1}{m_3} - 1 - \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,NO} \right) \tag{13.3}$$

and the desired compensation point concentrations $(m_{comp,i})$ of the NO-NO₂-O₃ triad result from combining Eqs. (11.1) - (11.3) and Eqs. (13.1) - (13.3):

$$m_{comp,NO2} = \frac{n_1 - m_1 \frac{V}{\overline{Q}} \bar{k} \,\overline{m}_{s,NO} \,\overline{m}_{s,O3}}{1 - m_1 - m_1 \frac{V}{\overline{Q}} \,\bar{j}(NO_2)}$$
(14.1)

$$m_{comp,NO} = \frac{n_2 - m_2 \frac{V}{\overline{Q}} \overline{j}(NO_2) \overline{m}_{s,NO2}}{1 - m_2 - m_2 \frac{V}{\overline{Q}} \overline{k} \overline{m}_{s,O3}}$$
(14.2)

$$m_{comp,O3} = \frac{n_3 - m_3 \frac{V}{\overline{Q}} \,\overline{j} (NO_2) \,\overline{m}_{s,NO2}}{1 - m_3 - m_3 \frac{V}{\overline{Q}} \,\overline{k} \,\overline{m}_{s,NO}}$$
(14.3)

The quantities n_1 , n_2 , n_3 and m_1 , m_2 , m_3 cannot be determined (graphically or numerically) from single pairs of $m_{a,i}$ and $m_{s,i}$, but from a (statistically sufficient) set of measured $m_{a,i}$ and $m_{s,i}$ (i.e. data sets classified for defined conditions of irradiation, temperature, humidity, concentrations, respectively). Therefore, n_1 , n_2 , n_3 and m_1 , m_2 , m_3 represent mean values for these data sets. Consequently, the quantities Q, A_{leaf} , $j(NO_2)$, k, $m_{s,NO2}$, $m_{s,NO}$ and $m_{s,O3}$ in Eqs. (12.1) - (12.3), (13.1) - (13.3) and (14.1) - (14.3) must be averaged over the same (time) period (the same data set) of $m_{a,i}$ and $m_{s,i}$ measurements from which the quantities n_i and m_i have been derived.

 $F_{ex,i}$, $v_{dep,i}$ and $m_{comp,i}$ of the NO-NO₂-O₃ triad were calculated from trace gas concentrations which were normalized for temperature and barometric pressure (0 °C, 1013.25 hPa).

2.1.3 Constraints of precision

Exchange flux densities $F_{ex,i}$ are determined from molar concentrations of the NO-NO₂-O₃ triad, ambient ones $(m_{a,i})$ as well as those in the plant chamber $(m_{s,i})$ (see Eqs. (8.1) - (8.3)). They are all measured with one set of analyzers only. The calculation procedure of exchange flux densities, deposition velocities as well as compensation point concentrations is based on linear regression analysis of $m_{a,i}$ and $m_{s,i}$, which are (a) both error-prone and (b) not very different of each other, i.e. their difference is usually (very) small. The uncertainties of these differences depend mainly on the precision of the analyzers; the uncertainties might be large and consequently those of the derived quantities $F_{ex,i}$, $v_{dep,i}$ and $m_{comp,i}$.

For the sake of simplicity we assume well defined laboratory conditions. Then, the trace gas exchange flux densities $F_{ex,i}$ are described by Eq. (8.4), which is equivalent to (a) only pre-scribed concentrations of trace gas $i (= m_{a,i})$ will enter the dynamic plant chamber, (b) the enclosed leaves are only exposed to corresponding $m_{s,i}$, (c) purging rate Q and leaf area A_{leaf} are known and constant and (d) sample concentrations of the other trace gases $(m_{s,j\neq i})$, photolysis rate $j(NO_2)$ as well as wall-sorptions of trace gas i are negligible. After evaluation of the linear relationship between $m_{a,i}$ and $m_{s,i}$, corresponding exchange flux densities $F^*_{ex,i}$, deposition velocities $v^*_{dep,i}$ and compensation point concentrations $m^*_{comp,i}$ are given by

$$F^*_{ex,NO2} = \frac{\overline{Q}}{\overline{A}_{leaf}} m_1 \left(n_1 + (m_1 - 1) \cdot m_{s,NO2} \right)$$
(15.1.1)

$$F^*_{ex,NO} = \frac{\overline{Q}}{\overline{A}_{leaf}} m_2 \left(n_2 + (m_2 - 1) \cdot m_{s,NO} \right)$$
(15.1.2)

$$F^*_{ex,O3} = \frac{Q}{\overline{A}_{leaf}} m_3 \left(n_3 + (m_3 - 1) \cdot m_{s,O3} \right)$$
(15.1.3)

$$v^*_{dep,NO2} = \frac{\overline{Q}}{\overline{A}_{leaf}} \frac{1 - m_1}{m_1}$$
 (15.2.1)

$$v^*_{dep,NO} = \frac{\overline{Q}}{\overline{A}_{leaf}} \frac{1 - m_2}{m_2}$$
(15.2.2)

$$v^*_{dep,O3} = \frac{\overline{Q}}{\overline{A}_{leaf}} \frac{1 - m_3}{m_3}$$
 (15.2.3)

$$m^*_{comp,NO2} = \frac{n_1}{1 - m_1} \tag{15.3.1}$$

$$m^*_{comp,NO} = \frac{n_2}{1 - m_2}$$
(15.3.2)

$$m^*_{comp,O3} = \frac{n_3}{1 - m_3} \tag{15.3.3}$$

Confining to NO₂, a schematic representation (using simulated data) of how the quantities defined by Eqs. (15.1.1), (15.2.1) and (15.3.1) are determined from genuine measurements of $m_{a,NO2}$ and $m_{s,NO2}$ is given in Figure 6a. Since the "1:1"-line is equivalent to $m_{a,NO2} = m_{s,NO2}$ (i.e. $F_{ex,NO2} = 0$, see Eq. (8.4)), the intersect of the linear regression line and the "1:1"-line is the NO₂ compensation point concentration, $m_{comp,NO2}$. Here, the dilemma of the experimental proof of a (highly) significant $m_{comp,NO2}$ becomes obvious. The lower $m_{comp,NO2}$ will be, the more the intersect shifts down the "1:1"-line, closer and closer to the limit of detection of the NO₂ concentration measurements $(LOD(m_{a,NO2}), LOD(m_{s,NO2}); 3\sigma$ -definition). This dilemma becomes even more obvious, if we consider the schematic representation of Eq. (8.4) in Figure 6b, where $LOD(F_{ex,NO2})$ has been calculated from corresponding $s_{m s,NO2}$ and $s_{m a,NO2}$ by Gaussian error propagation. Here, $m_{comp,NO2}$ ($F_{ex,NO2} = 0$) is the intersect of the $m_{s,NO2}$ -axis with the best-fit line of $F_{ex,NO2}$ vs. $m_{s,NO2}$ (which is mathematically not correct, see above). For high NO₂ compensation point concentrations (as in Figure 6), $m_{comp,NO2}$ can still be evaluated by interpolation from significant data pairs (i.e. data pairs, where $> LOD(m_{NO2}), \geq +LOD(F_{ex,NO2})$ or $\leq -LOD(F_{ex,NO2})$, respectively). If $m_{comp,NO2}$ falls below $LOD(m_{s,NO2})$ and F_0 is consequently below $+LOD(F_{ex,NO2})$, $m_{comp,NO2}$ could only be determined by extrapolation from significant data pairs.



Figure 6: Schematic representation of the determination of bi-directional NO₂ exchange flux density ($F_{ex,NO2}$), NO₂ deposition velocity ($v_{dep,NO2}$) and NO₂ compensation point concentration ($m_{comp,NO2}$) from measurements of NO₂ concentrations at the plant chamber's inlet ($m_{a,NO2}$) and outlet ($m_{s,NO2}$) under laboratory conditions ($m_{a,NO} = m_{a,O3} = j(NO_2) \approx 0$). (**a**) by linear regression of $m_{s,NO2}$ with $m_{a,NO2}$. (**b**) by plotting $F_{ex,NO2}$ vs. $m_{s,NO2}$. Dashed lines represent the limits of detection (3σ -definition) for NO₂ concentration measurements ((a) and (b) panel) and the determination of the NO₂ exchange flux density ((b) panel), which are both defined by the sensitivity of the applied NO₂ analyzer (note: $LOD(m_{a,NO2}) = LOD(m_{s,NO2})$). Data points and error bars of NO₂ concentrations have been simulated to match $R^2(m_{a,NO2}, m_{s,NO2}) = 0.9925$, error bars of NO₂ exchange flux have been calculated by Gaussian error propagation (c.f. Eq. (8.4)). Filled circles identify data points > LODs, hollow circles those \leq LODs.

According to Eqs. (15.1.1), (15.2.1) and (15.3.1), the errors of $F_{ex,NO2}$, v_{depNO2} and $m_{comp,NO2}$ are entirely due to the errors of n_1 and m_1 , which are in turn entirely due to the goodness of the linear relationship between $m_{a,NO2}$ and $m_{s,NO2}$ as well as to the errors of $m_{a,NO2}$ and $m_{s,NO2}$ and $m_{s,NO2}$ ($s_{m_a,NO2}$ and $s_{m_s,NO2}$ see Sect. 3.1.7). This leads to the simple conclusion, that the determination of $F_{ex,NO2}$, v_{depNO2} and $m_{comp,NO2}$ is as more precise, as higher the regression coefficient $R^2(m_{s,NO2}, m_{a,NO2})$ and as lower the standard errors $s_{m_s,NO2}$ and $s_{m_a,NO2}$ are.

Only one NO₂ analyzer is used for the measurements of both concentrations, $m_{a,NO2}$ and $m_{s,NO2}$. As shown below (Sect. 2.3), the standard error $s_{m_a,NO2}$ ($s_{m_s,NO2}$) was found to be a weak exponential function of $m_{a,NO2}$ ($m_{s,NO2}$), starting with a fixed value $s_{m,LOD(NO2)}$ at $m_{a,NO2} = m_{s,NO2} = 0$. To demonstrate, how the goodness ($R^2(m_{s,NO2}, m_{a,NO2})$) of the linear relationship between $m_{a,NO2}$ and $m_{s,NO2}$ and how the magnitude of $s_{m_a,NO2}$ and $s_{m_s,NO2}$ impact the NO₂ exchange measurements, we consider (a) the determination of the minimum possible, but still highly significant NO₂ compensation point concentration ($m_{comp,NO2}$) and (b) the precision of the NO₂ exchange flux density ($F_{ex,NO2}$).

For that we simulated data sets of $m_{a,NO2}$ and $m_{s,NO2}$ within the range $LOD(m_{s,NO2}) \le m_{s,NO2} \le 615$ nmol m⁻³ (15 ppb) for prescribed NO₂ deposition velocities $(0.1 \le v_{dep,NO2} \le 0.8 \text{ mm s}^{-1}, \text{ per leaf area})$ and for pre-scribed $R^2(m_{s,NO2}, m_{a,NO2})$ between 0.999 and 0.6. The latter was achieved by random number application to the $m_{a,NO2}$ data. Standard errors $s_{m_s,NO2}$ and $s_{m_a,NO2}$ were calculated from $m_{a,NO2}$ and $m_{s,NO2}$ (see Eq. (16.1), Sect. 2.3), while the standard error of $F_{ex,NO2}$ (s_{F_ex,NO2}) was calculated from $s_{m_s,NO2}$, $s_{m_a,NO2}$ and $r(m_{s,NO2}, m_{a,NO2}) = [R^2(m_{s,NO2}, m_{a,NO2})]^{1/2}$ by application of the general form of Gaussian error propagation (see Sect. 3.1.7).

Application of bi-variate linear regression analysis to this simulated data set delivers the quantities n_1 and m_1 as well their standard errors $s_{n,1}$ and $s_{m,1}$ (which depend on $s_{m_s,NO2}$, $s_{m_a,NO2}$ and $R^2(m_{s,NO2}, m_{a,NO2})$). Application of the general form of Gaussian error propagation (see Sect. 3.1.7) to Eq. (15.3.1) delivers the standard error of the NO₂ compensation point concentration ($s_{m_comp,NO2}$). The "detectable existence" of $m_{comp,NO2}$ (i.e. testing the hypothesis $m_{comp,NO2} \neq 0$) has been statistically secured by application of the t-test to the values of $m_{comp,NO2}$, $s_{m_comp,NO2}$ and N (number of ($m_{s,NO2}, m_{a,NO2}$) data pairs). In Figure 7, the minimum detectable NO₂ compensation point concentration, i.e. the lowest, but still highly significant $m_{comp,NO2}$ (P ≥ 0.999) is shown for a pre-scribed

range of NO₂ deposition velocities as function of the regression coefficient $R^{2}(m_{s,NO2}, m_{a,NO2})$ and for three different values of $LOD(m_{s,NO2})$, namely 0.4, 4.5 and 44.6 nmol m⁻³ (0.01, 0.1, 1.0 ppb). These three values represent a certain "history" of NO/NO₂ chemiluminescence analyzers: $LOD(m_{s,NO2}) = 44.6 \text{ nmol m}^{-3}$ (1 ppb) reprethe state-of-art of commercial NO_2 analyzers of 1985-1995, sents $LOD(m_{s,NO2}) = 4.5 \text{ nmol m}^{-3}$ (0.1 ppb) the best performance between 1995-2005's, while $LOD(m_{s,NO2}) = 0.4 \text{ nmol m}^{-3}$ (0.01 ppb) is characteristic for the most advanced NO/NO₂ analyzers which have been recently applied over the remote Southern Atlantic Ocean (HOSAYNALI BEYGI et al. 2011). For typical ranges of laboratory measurements, i.e. $0.9 \le R^2 \le 0.99$, minimum detectable NO₂ compensation point concentrations range $17.5 - 99.4 \text{ nmol m}^{-3}$ (0.39 - 2.23 ppb), between if NO_2 analyzers with $LOD(m_{s NO2}) = 44.6 \text{ nmol m}^{-3}$ (1.0 ppb) have been used. Best performance of presentday NO₂ analyzers allow minimum detectable $m_{comp,NO2}$ between 3.6 and 21.3 nmol m⁻³ (0.08 - 0.48 ppb). Very low minimum detectable $m_{comp,NO2}$ (0.8 - 4.0 nmol m⁻³ or 0.02 - 0.09 ppb) may be reached if the most advanced state of NO₂ analyzers is considered. It should be noted that, due to the potential goodness of the measurements, the minimum detectable $m_{comp,NO2}$ could be lower than the actual $LOD(m_{s,NO2})$, but statistically still highly significant.

The impact of $s_{m_s,NO2}$, $s_{m_a,NO2}$ and $R^2(m_{s,NO2}, m_{a,NO2})$ on the precision of the NO₂ exchange flux density (= $s_{F_ex,NO2}$)/ $F_{ex,NO2}$) is demonstrated in Figure 8. For the sake of clarity, another data set has been simulated (random number application), namely for pre-scribed NO₂ deposition velocities ($0.3 \le v_{dep,NO2} \le 0.6 \text{ mm s}^{-1}$, per leaf area), a pre-scribed NO₂ compensation point concentration ($m_{comp,NO2} = 67 \text{ nmol m}^{-3}$ (1.5 ppb)) and for $0.99 \le R^2 \le 0.9$. Also shown in Figure 8 is the precision of $m_{s,NO2}$ (= $s_{m_s,NO2}/m_{s,NO2}$; right axis) for the "history" of $LOD(m_{s,NO2})$ values, namely $LOD(m_{s,NO2}) = 44.6$, 4.5 and 0.4 nmol m⁻³ (1.0, 0.1, 0.01 ppb). Before 1995 ($LOD(m_{NO2}) = 1$ ppb), a precision of $m_{s,NO2}$ better than 10 % could hardly be achieved in the lower ppb-range. Best performing present-day NO₂ chemiluminescence analyzers ($LOD(m_{NO2}) = 0.1$ ppb) exceed the 10 % level of $m_{s,NO2}$ precision not before $m_{s,NO2}$ falls below 14.8 nmol m⁻³ (0.33 ppb), while another step of magnitude can be reached with most advanced NO₂ analyzers ($s_{m_s,NO2}/m_{s,NO2} > 10$ % not before $m_{s,NO2} < 1.5 \text{ nmol m}^{-3}$ (0.03 ppb)). The "history" of NO₂ analyzers is also mirrored in the precision of $F_{ex,NO2}$

 $F_{ex,NO2}$ (= s_{*F*_ex,NO2})/ $F_{ex,NO2}$) reaches infinity at $m_{s,NO2} = m_{comp,NO2}$, since there the NO₂ exchange flux density equals zero. Otherwise, the precision of $F_{ex,NO2}$ rapidly falls (very) well below the 10 % level. This is a consequence of the fact, that $m_{a,NO2}$ and $m_{s,NO2}$ are the decisive quantities for the determination of $F_{ex,NO2}$. Since $m_{a,NO2}$ and $m_{s,NO2}$ are highly correlated, the standard error of $F_{ex,NO2}$ is proportional to $[s^2_{m_a,NO2} + s^2_{m_s,NO2}]^{1/2} - 2 s_{m_a,NO2} s_{m_s,NO2} [R^2(m_{s,NO2}, m_{a,NO2})]^{1/2}$, rather than proportional to $[s^2_{m_a,NO} + s^2_{m_s,NO2}]^{1/2}$ alone (see Sect. 3.1.7). In other words, the error of $F_{ex,NO2}$ benefits from the compensation of the errors of $m_{a,NO2}$ and $m_{s,NO2}$.



Figure 7: The dynamic plant chamber at well defined (laboratory) conditions: minimum detectable NO₂ compensation point concentrations ($m_{comp,NO2}$ at P \ge 0.999, i.e. "highly significant") as function of NO₂ deposition velocity ($v_{dep,NO2}$; per leaf area) and the goodness (R^2) of the ambient vs. sample NO₂ concentration measurements (standard errors of NO₂ concentration measurements considered). Results are from data simulation (random number application) $v_{dep,NO2}$ (0.999 $\leq R^2 \leq 0.6$ matching pre-scribed $R^2(m_{a,NQ2}, m_{s,NO2})$ and prescribed and $v_{dep,NO2} = 0.1, 0.2, \dots, 0.8 \text{ mm s}^{-1}$). The greenish range represents simulated data of a NO₂ analyzer with $LOD(m_{NO2}) = 0.4$ nmol m⁻³ (0.01 ppb), bluish the range for $LOD(m_{NO2}) = 4.5 \text{ nmol m}^{-3}(0.1 \text{ ppb})$, the reddish range for $LOD(m_{NO2}) = 44.6 \text{ nmol m}^{-3}(1.0 \text{ ppb})$.

Finally, it should be emphasized, that the estimates of this sub-section are made on the basis of Eqs. (15.1.1), (15.2.1) and (15.3.1) for (best) defined laboratory conditions. Under field conditions, however, the equations for the determination of $F_{ex,NO2}$, v_{depNO2} and $m_{comp,NO2}$ will contain also average quantities of $m_{s,NO}$, $m_{s,O3}$, $j(NO_2)$ and k (see Eqs. (12.1), (13.1), (14.1)). It is obvious, that their variability (standard errors) will enlarge standard errors of n_1 and m_1 and diminish $R^2(m_{s,NO2}, m_{a,NO2})$. Consequently, corresponding minimum detectable NO₂ compensation point concentrations will certainly be higher and precisions of $F_{ex,NO2}$ will be lower than those given in Figure 7 and Figure 8.



Figure 8: The dynamic plant chamber at well defined (laboratory) conditions: precision of NO₂ concentration measurements (= $s_{m,s_NO2}/m_{s,NO2}$; right axis) and precision of derived NO₂ exchange flux densities (= $s_{Fex_NO2}/F_{ex,NO2}$, left axis) as function of the NO₂ concentration measurement at the outlet of the dynamic chamber (precision $m_{s,NO2}$, right axis). Results are from data simulation (random number application), which considers standard errors of NO₂ concentration measurements, and which matches pre-scribed $R^2(m_{a,NO2},m_{s,NO2})$ and pre-scribed $m_{comp,NO2} = 67$ nmol m⁻³ (1.5 ppb). Dark purple, purple and pink lines (= precision of $m_{s,NO2}$) represent data for a NO₂ analyzer characterized by $LOD(m_{s,NO2}) = 44.6$ nmol m⁻³ (1.0 ppb), $LOD(m_{s,NO2}) = 4.5$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (0.01 ppb), respectively. Ranges of the precision of derived NO₂ exchange flux densities are identified by reddish, bluish and greenish areas for $LOD(m_{s,NO2}) = 44.6$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 44.6$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (0.01 ppb), $LOD(m_{s,NO2}) = 4.5$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 4.6$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (1.0 ppb), $LOD(m_{s,NO2}) = 4.5$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (1.0 ppb), $LOD(m_{s,NO2}) = 4.5$ nmol m⁻³ (0.1 ppb) and $LOD(m_{s,NO2}) = 0.4$ nmol m⁻³ (0.01 ppb). The width of the colored areas stands for all considered combinations of R^2 and $v_{dep,NO2}$ (0.99 $\leq R^2 \leq 0.9$ and $0.3 \leq v_{dep,NO2} \leq 0.6$ mm s⁻¹). The respective upper boundary of each colored area represents the combination $v_{dep,NO2} = 0.3$ mm s⁻¹ and $R^2 = 0.9$, while the lower boundary represents $v_{dep,NO2} = 0.6$ mm s⁻¹ and $R^2 = 0.99$.

2.1.4 Constraints of design

Aside the strong demand for precise and highly sensitive measurements of NO_2 concentration, there are more requirements to the dynamic leaf chamber system and the measurements of the surface exchange fluxes of NO_2 (NO, O_3):

- The environment in the chamber should as closely as possible represent the surrounding (ambient) environment.
- (2) Enclosing the plant (part of plants) by the chamber should not affect the plant itself, neither through mechanical stress nor due to changed environmental conditions. Changes in concentrations of relevant trace gases should be small in order to prevent affecting plant metabolism and stomata regulation.
- (3) Primary plant-physiological processes, such as CO₂ surface exchange fluxes (assimilation) and H₂O surface exchange fluxes (transpiration) should be closely followed, measured and finally related to the NO₂ (NO, O₃) surface exchange.
- (4) Losses of NO₂ (NO, O₃) on chamber materials must be negligible (if not: must be quantified).
- (5) The chamber system should be applicable for laboratory and field measurements without substantial modifications.
- (6) Simultaneous measurements of surface exchange fluxes of NO₂, O₃, NO, CO₂ and H₂O should be feasible.
- (7) Differences of NO₂ (NO, O₃) concentrations between inlet and outlet of the dynamic chamber, which are expected to be (very) small, must be resolved with statistical significance.

Furthermore, fumigation experiments to study the NO₂ surface exchange in the laboratory (NO₂ exchange under controlled conditions) demand the generation of very low (ppb- and sub-ppb levels) and temporally stable NO₂ concentrations in order to identify statistically significant NO₂ compensation point concentrations. These low NO₂ concentrations have to be reproducible and verifiable.

2.2 Trace gas analyzers

NO and NO₂ concentrations were measured by a gas-phase chemiluminescence NO analyzer (Model 42C, Thermo Electron Corporation, USA). In a low pressure reaction chamber, the NO of the air sample reacts with ozone (provided by the analyzer) forming electronically excited NO₂ molecules. Decaying to the ground state, the excited NO₂ molecule emits a photon (chemiluminescence) and the total light intensity in the reaction chamber, detected by a photomultiplier, is proportional to the NO concentration. NO_2 in the air sample is also measured by the NO analyzer after conversion of NO_2 to NO. In most commercial NO/NO₂ analyzers a molybdenum converter is applied (heated to 300 - 400 °C), where NO₂ is catalytically reduced to NO at the converter's surface. However, previous studies demonstrated that molybdenum converters are non-specific for NO₂ because other oxidized nitrogen compounds of ambient air, like gaseous nitrous acid (HONO), nitric acid (HNO₃), the nitrate radical (NO₃), dinitrogen pentoxide (N_2O_5) , peroxyacetyl nitrate (PAN) and other organic nitrates were found to be also converted to NO, which leads to systematic and considerable overestimation of the measured NO₂ values (WINER et al. 1974; MATTHEWS et al. 1977; GROSJEAN and HARRISON 1985; GEHRIG and BAUMANN 1993; STEINBACHER et al. 2007). During some studies hydrated, crystalline ferrous sulfate (FeSO₄) for the surface reduction of NO₂ to NO were used. However, FeSO₄ converter also overestimates the mixing ratio of NO and NO₂ (RIDLEY et al. 1988). Significant interferences of *n*-propyl nitrate, nitrous acid (HNO₂) and PAN were reported (KELLY et al. 1980; COX et al. 1983; FEHSENFELD et al. 1987). As a consequence FEHSENFELD et al. (1987) did not recommend FeSO₄ converter for measuring NO₂. Another frequently used analyzer to measure NO₂ is the Luminox detector (LMA-3, Scintrex/Unisearch Inc.). Its measurement principle is based on the chemiluminescent reaction of NO₂ with luminol in aqueous solution (MAEDA et al. 1980; WENDEL et al. 1983; SCHIFF et al. 1986). The luminol technique is noted for interferences by ambient O₃ and PAN, and exhibits non-linear response at low NO₂ concentrations. The interferences due to O_3 and PAN are significant especially at low NO₂ concentrations (KELLY et al. 1990). Table 4 shows an overview about commonly used NO₂ converters and their reported interferences. No interferences or any artifacts were reported for photolytic converters, where NO₂ is photolyzed by ultraviolet light < 420 nm (FEHSENFELD et al. 1990) or were negligible, respectively (RYERSON et al. 2000). Consequently, we used a highly NO₂ specific blue light converter which

photodissociates NO_2 into NO at a wavelength of approximately 395 nm (manufactured by Droplet Measurement Technologies Inc., Colorado, USA). To obtain a better accuracy and precision of the NO_2 (and NO) measurements at sub-ppb concentrations, the NO/NO_2 analyzer has always been operated with pure oxygen (instead with the oxygen of ambient air) for the internal generation of ozone, necessary for the reaction with NO in the low pressure reaction chamber.

NO ₂ converter	conversion principle	compound	Response % of concn	author
luminol	NO ₂ reacts with luminol solution	PAN	25 %	Drummond et al., 1989
		O ₃	0.0033 ppb NO ₂ (per ppb O ₃)	Kelly et al., 1990
molybdenum (Mo)	heated ~ 400 °C surface oxidation	PAN ethyl nitrate ethyl nitrite HNO ₃	92 % 103 % 92 % not quantified	Winer et al., 1974
		HNO ₃ PAN methyl nitrate n-propyl nitrate n-butyl nitrate		Grosjean & Harrison, 1985
		hydrocarbons	negative interferences	Kurtenbach et al., 2001
ferrous sulfate (FeSO ₄)	surface oxidation	PAN	20 %	Kelly et al., 1980
		HONO	100 %	Cox et al., 1983
		n-propyl nitrate PAN	32 % 35 - 45 %	Fehsenfeld et al., 1987
photolytic	ultraviolet light (320 - 500 nm)	none		Fehsenfeld et al., 1990
photolytic	ultraviolet light (> 350 nm)	$\begin{array}{c} \text{HONO} \\ \text{BrONO}_2 \\ \text{NO}_3 \\ \text{N}_2\text{O}_5 \\ \text{HO}_2\text{NO}_2 \end{array}$	37 % 5 % 10 % 3 % 12 %	Ryerson et al., 2000

Table 4: Interferences of chemiluminescent NO-NO₂-NO_x analyzers used different NO₂ converters.

Measurements of CO_2 and H_2O concentrations were performed by infrared dual channel gas analyzer for difference measurements between the outlet of an empty reference chamber and the sample gas (LI-7000, LiCor, Lincoln, NE, USA). An additional gas analyzer (LI-6262, LiCor, Lincoln, NE, USA) monitored the absolute CO_2 and H_2O concentrations to deliver a base signal for the LI-7000 operating in differential mode. O_3 concentration was detected using an UV-absorption analyzer (Model 49C, Thermo Electron Corporation, USA). All measured parameters are listed in Table 5.

Table 5: Measured parameters and instrument specifications. Limit of detection $(LOD(m_i), 3\sigma$ -definition) for the gas concentrations were determined under field and laboratory conditions.

parameter	symbol	unit	$LOD(m_i)$		instrument (model)
			lab	field	
nitric oxide	NO	ppb	0.23 ppb	0.10 ppb	ThermoElectron, 42C
nitrogen dioxide	NO_2	ppb	1.01 ppb	0.31 ppb	ThermoElectron, 42C
ozone	O_3	ppb	0.8 ppb	0.98 ppb	ThermoElectron, 49C
carbon dioxide	CO_2	ppm	1.2 ppm	1.5 ppm	LiCor, LI-6262 / LI-7000
water vapor	H_2O	ppth	0.3 ppth	0.2 ppth	LiCor, LI-6262 / LI-7000
air temperature	Т	°C			thermocouple
relative humidity	rH	%			Rotronic, MP100A
photosynthetic active radiation	PAR	μ mol m ⁻² s ⁻¹			LiCor, LI-190SA
photolysis rate	$j(NO_2)$	s ⁻¹			filter radiometer
air pressure	Р	hPa			Ammonit

2.3 Calibrations, limits of detection, standard errors and precision of trace gas concentration measurements

For the calibration of the NO/NO₂ analyzer (field conditions), a NO standard $(5.09 \pm 0.1 \text{ ppm}, \text{Air Liquide, Germany})$ was applied. The standard was diluted by synthetic air, which has been additionally cleaned with activated charcoal and Purafil® (Purafil, Inc., USA) to remove any potential NO and NO₂ contaminations. For the dilution of the NO standard a gas phase titration unit was applied (GPT, 146C Dynamic Gas Calibrator, Thermo Electron Corporation, USA). In the GPT, NO₂ calibration gas is produced by titration (see Reaction (R1)) of the diluted NO standard with O₃ (generated by a UV lamp in the GPT). The BLC's efficiency was determined by the ratio of measured NO₂ and the known value of NO₂ obtained by titration of NO. The O₃ analyzer was calibrated by the GPT-generated O₃, where the exact O₃ concentration is known from the gas phase titration of the NO standard. For the calibration of the CO₂/H₂O analyzers three gaseous CO₂ standards were used (355.4 ppm, 401.1 ppm, 453.8 ppm, Air Liquid, Germany); the H₂O signal has been calibrated by a dew point generator (LI-610, LiCor, Lincoln, NE, USA). To maintain high quality concentration measurements even under long-term field conditions, it was necessary to control and to service the system frequently. In the field, calibrations were performed once a week to ensure stability of the analyzers (quantifying potential drifts), while in the laboratory calibrations were performed just before the start of the experiment.

The determination of the limit of detection (LOD) is particularly important for the exchange measurements of NO and NO₂, as (very) low concentrations have been encountered under both, laboratory and field conditions. According to MACDOUGALL and CRUMMETT (1980) the "limit of detection" is the lowest concentration level that can be determined to be statistically different from a measurement of "zero" concentration. Here we define $LOD(m_{NO2})$, $LOD(m_{NO})$ and $LOD(m_{O3})$ as three times that standard deviation ($s_{m_{NO2,0}}$, $s_{m_{NO,0}}$, $s_{m_{O3,0}}$), which has been obtained through a statistically significant number (laboratory: 360, field: 160 - 360) of zero-air measurements. In Table 5 the $LOD(m_i)$ of the instruments are summarized. The conversion efficiency of the BLC for NO₂ was around 25 % during laboratory measurements and 32 - 36.5 % under field conditions.

Besides the determination and rigorous control of the LOD's, the quantification of the analyzers' reproducibility (precision) is as more necessary, as exchange fluxes of the NO-NO₂-O₃ triad are evaluated from very small differences of concentrations measured at the inlet and the outlet of the dynamic plant chamber. We define the precision of the analyzers as the ratio of the standard errors $s_{m,i}$ and the corresponding concentrations m_i ($i = NO, NO_2, O_3$). The standard errors of NO and NO₂ measurements have been found to be a (weak) function of the NO and NO₂ concentrations themselves:

$$s_{m,NO2} = s_{m NO2,0} \cdot \exp(b_{NO2} \cdot m_{NO2}) \tag{16.1}$$

$$s_{m,NO} = s_{m_NO,0} \cdot \exp(b_{NO} \cdot m_{NO})$$
(16.2)

where $s_{m_NO2,0}$ and $s_{m_NO,0}$ are the standard errors at $m_{NO2} = 0$ and $m_{NO} = 0$, b_{NO2} and b_{NO} (in nmol⁻¹ m³) have been derived from calibration exercises.

2.4 Dynamic chamber system

2.4.1 Design and construction

The open (flow through), dynamic chamber system was a further development of the systems operated in previous studies (SCHÄFER et al. 1992; KESSELMEIER et al. 1996; KUHN et al. 2002). The system was designed for measurements of trace gas exchange in the field with minimal effects on the gases. The system has been demonstrated to be easily handled under field conditions. The design of the chambers is illustrated in Figure 9 and details of the used materials and parts are listed in **Table 6**. The chambers had an inner diameter of 40 cm. The height of the chambers could be varied by extending the frame and could be adjusted to the plant specimen. Our initial height was 45 cm and we used extensions of 15 cm at field measurements. The chamber frame and the lid were made of PVC and acrylic glass.

	part	manufacturer	specifications
(1) + (2)	chamber frame and lid	MPI workshop, Germany	PVC, acrylic glass
(3)	inner chamber wall	Saint Gobain, Germany	FEP (fluorinated ethylene propylene) film, thickness 0.05 mm, chemically inert, transparent for visible and UV light
(4)	clamps	Holex, Germany	parallel clamp, typ 25
(5)	silicon straps	Dichtungstechnik Bensheim GmbH, Germany	transparent MVQ-silicone cord, diameter 5 mm
(6)	inlet fan	Micronel, Switzerland	axial fan, model D344T012GK-2
(7)	air mass flow sensor	Honeywell International Inc., USA	model AWM 700
(8)	propeller	APC Propellers, USA	Sport Prop, 10x7, Teflon® coating by MPI workshop
(9)	mixing fan	Micronel, Switzerland	ultra slim fan, model F62MM012GK-9, Teflon® coating by MPI workshop
(10)	tubing	diverse	1/4" PFA tubing
(11)	in-line filter case	Entegris Inc., USA	Galtek® Integral Ferrule in-line filters
	particulate membrane filter	Pall Corporation, USA	Zefluor TM membrane disc filters, model P5PJ047, pore size 2 μ m, diameter 47 mm
	solenoid valves	Entegris Inc., USA	Galtek® diaphragm valves, 3-way, 1/4" orifice
	sample pump	Vakuubrand, Germany	diaphragm pump, model MZ4C, chemical resistant
	heating tape	EHT Haustechnik AEG, Germany	typ HT SLH 15/L300, self limiting, max. holding temperature 60°C, heat output 15 W/m

Table 6: Manufacturer details for parts of the dynamic chamber system.

The inner walls consisted of a thin transparent Teflon film (FEP). Previous investigations of the spectral transmissivity of the FEP film have shown that photosynthetically active radiation (PAR) nearly completely transmits this film: in the spectral range of PAR (400 - 700 nm) transmissivity is about 95 %. In the range of $\lambda \le 400$ nm, the transmissivity of the FEP film is about 90 % (SCHÄFER et al. 1992; PAPE et al. 2009). A consequence of the horizontal installation of the chamber during field measurement is that transmissivities of the acrylic glass parts of the chamber play only a very minor role. Furthermore, the Teflon film was reported to show no interferences with trace gases tested such as organic acids (SCHÄFER et al. 1992; KESSELMEIER et al. 1997), monoterpenes and isoprene (KESSELMEIER et al. 1996, 1997; KUHN et al. 2000) and reduced sulfur compounds (KESSELMEIER et al. 1993).

The FEP film was fixed with elastic silicone straps around the outer side of the frame. The inner side of the lid was covered by the Teflon film as well. The lid was

fixed to the chamber with four clamps. Several holes in the lid allowed the installation of tubes, mixing fans and the intake system of purging air. The purging air flow through the chamber was established in the field by a blowing axial inlet fan which was controlled by an air mass flow sensor installed outside the chamber frame. At laboratory we used pressurized air for flushing the chamber. For a continuous turbulent mixing of the air inside the chamber a Teflon propeller driven by a magnetically coupled motor attached outside as well as two Teflon coated mixing fans were used. This design ensured that the air pumped through the chamber only came into contact with parts made of Teflon (PFA or PTFE). For the measurements several chambers were combined (Figure 10). As in former studies on the NO_2 exchange with different plants, an extra empty ("reference") chamber was also applied. The empty chamber was used to detect basic contamination in the system, adsorption/desorption, as well as to investigate gas-phase chemical reactions within the chamber volume and at the wall surface. A central V25 microprocessor unit (PASCAL based code) controlled the power supply for the mass flow sensors, purging and mixing fans, and signal recording by a PC card. Each chamber could be controlled independently. Furthermore, the V25 operated a number of environmental sensors for air and needle temperature, photosynthetically active radiation (PAR) and relative humidity, and recorded their signals.



Figure 9: Photograph and schematic drawing of a dynamic chamber consisting of: (1) PVC (grey parts) frame, (2) acrylic glass (blue parts)n lid, (3) FEP film (red parts in the scheme), (4) clamp to attach lid to frame, (5) silicon straps, (6) inlet fan, (7) air mass flow sensor, (8) Teflon propeller, (9) mixing fan, (10) sample tube for chamber air, (11) filter, (12) closure, (13) plant material.



Figure 10: Schematic set-up of the system with three dynamic chambers. Open lines are PFA sampling tubes, black lines are cables for data acquisition and control.

2.4.2 Implementation of concentration and flux density measurements

Exchange flux densities of the NO-NO₂-O₃ triad as well as of CO₂ and H₂O are determined from the difference of molar concentrations measured at the inlet and outlet of the dynamic chambers. Ideally, a total of 10 analyzers per dynamic chamber would guarantee simultaneous concentration measurements at all these positions. However, full simultaneity is usually prohibited not only for cost arguments; operation of two trace gas analyzers with an agreement (in their absolute accuracy) much less of the expected difference between inlet and outlet concentration is currently not feasible. Therefore, only one set of analyzers was used operating in a mode of continuous switching between the inlet and outlet position(s) of the (different) dynamic chamber(s). For gas piping the tubes from the different positions at the chambers were combined to one insulated and heated (above ambient temperature) bundle to prevent water vapor condensation. To ensure similar conditions for all lines, all tubes were set to the same length (in this field study 37 m). The sampling air flow was maintained by Teflon membrane pumps with an air flow of 8 - 10 L min⁻¹. To avoid contamination of tubes and analyzers a PTFE particulate filter (pore size 2 µm) was installed in front of the intake line. Switching between the different intake lines was maintained by several 3-way PFA solenoid valves. The necessary quantity of valves depends on the number of dynamic chambers in operation. The sample line connected the valve block to the analyzers. Even when an individual intake line was not switched to the analyzers, the air flow through it was kept constant. A second V25 unit was used to control the solenoid valves and the cycle times and recorded the data of the trace gas analyzers. Measurement cycle times and switching (during field experiments) is shown in Figure 18a. The shown cycling time of 4 minutes is a result of optimization between fast switching and the analyzers' and system's capabilities: the most important issues in this respect are the analyzers' (moving) averaging times of 30 s and the temporal response of the analyzers to switching concentrations.

Air temperature and needle surface temperatures inside the chambers were continuously recorded by Teflon covered thermocouples (0.005", ChromegaTM-Constantan, Omega, UK). PAR was detected outside the chamber with a LiCor quantum sensor (model LI-190SA, LiCor, Lincoln, NE, USA). Relative humidity was measured with a combined temperature and relative humidity probe (Model MP100A, Rotronic, Switzerland).

2.5 Experiments

The results of this study are based on datasets obtained during the second intensive observation period (IOP II) of the project EGER (ExchanGE processes in mountainous Regions) and laboratory measurements.

2.5.1 Plant material

Measurements of NO-NO₂-O₃ trace gas exchange fluxes were done at Norway spruce (*Picea abies* L.) also commonly known as the European spruce. Spruce is a coniferous evergreen tree of the genus *Picea* in the family *Pinaceae*. The Norway spruce grows throughout Europe from Norway in the northwest to Poland eastward, and also in the mountains of central Europe, southwest to the western end of the Alps, and southeast in the Carpathians and Balkans to the extreme north of Greece. It prefers a damp and cool climate, therefore it is a mountains tree in the south part of the distribution area. The primary habitat requirement is the water supply and a sufficient ventilation of the soil.

The Norway spruce is one of the most economically important coniferous species in Europe. The distribution of forest species in Germany is displayed in Figure 11. Spruce is the widespread species with 28.2 % of the total population followed by pine and beech. 58.1 % of the German forests consist of conifer due to economical reasons.





Laboratory experiments were performed with 3- to 4-yr old Norway spruce trees (*Picea abies* L.) grown in pots in a commercial soil mixture. All specimens originated from the EGER field site and were dug out half a year before the measurements started. For the laboratory studies the above-ground parts of the whole tree were enclosed in the chamber. A typical young tree had a leaf area (A_{laef}) of 0.16 m² in total (projected leaf area). For the field experiments branches of adult Norway spruces were investigated. The front part of an intact branch with older needles and new shoots, still attached to the tree, was enclosed to around 40 cm length in the chamber. Two plant chambers on different trees were used for the field studies. At the end of the studies the enclosed leaf area was measured to be 0.36 m² (tree 1, projected leaf area) and 0.37 m² (tree 2, projected leaf area) with a dry weight of 66 g (tree 1) and 78 g (tree 2). All exchange measurements started one day after enclosure in order to allow an acclimatization of the branch or plant.

At the end of the experiments leaves of the enclosed branches were harvested for determination of leaf area and dry weight. Leaves were scanned by a calibrated scanner system (DeskSCAN II, Hewlett-Packard, USA; area determining software SIZE, Müller, Germany). Dry leaf weight was obtained after drying for 2 days at 70 °C in an oven (Heraeus, Germany). The needles of spruce have stomata on the entire needle surface, therefore the area of the whole surface was used. For needle surface area calculation the single surface area was multiplied by factor 2.74 according to RIEDERER et al. (1988). Leaf area during the field measurements varied with the leaf flushing, therefore we interpolated the leaf area retroactively.

2.5.2 Field site description and set-up

The field experiment was conducted within the project EGER. The project was focused on the role of process interactions among the different scales of soil, in-canopy and atmospheric exchange processes of mass, energy, and non-reactive as well as reactive trace substances. It took place in summer 2008 (01 June - 15 July) in northeast Bavaria, Germany (Fichtelgebirge), a mountainous area, covered mainly with forest, agricultural area and including meadows and lakes. The research site "Weidenbrunnen" (50°08'31" N, 11°52'01" E; 774 m a.s.l.) was part of a spruce forest ecosystem, which resulted from intensive reforestation in the last century. The plant cover was dominated by Norway spruce (*Picea abies*). The main understory types were moss, grass (*Deschampsia flexuosa* and *Calamagrostis villosa*), blueberries (*Vaccinium myrtillus*) and young spruce. The stand-age was 56 years (according to ALSHEIMER 1997) and the mean canopy height was 23 m (SERAFIMOVICH et al. 2008). The tree density of the stand was 1007/ha (ALSHEIMER 1997), with a leaf area index (LAI) of 5.2 (THOMAS and FOKEN 2007). The Fichtelgebirge is located in the transition zone from maritime to continental climates with maritime impact (FOKEN 2003). The annual average temperatures are 5.0 °C (1971-2000; FOKEN 2003) with extreme values of -20 °C during wintertime and 30° C during summer. In the summer period, Atlantic air masses account for the temperate climate, whereas during winter continental influence due to easterly winds can result in short but extreme cold periods. The annual precipitation is 1162 mm (1971-2000; FOKEN 2003). The main wind direction is a west or south west wind (GERSTBERGER et al. 2004). This field site is maintained for more than 10 years by the University of Bayreuth and a lot of studies have been conducted there.

2.5.3 Laboratory set-up

For laboratory experiments the plant chambers were installed inside a thermostatted cabinet (Heraeus, Germany), which was kept under controlled temperature and humidity conditions (day: 25 °C, 60 %; night: 20 °C, 50 %) with a light/dark regime of 12/12 hours. In addition to the cabinet irradiation (Osram Powerstar HQI-BT 400 W/D) we used a set of light emitting diodes with a spectral bandwidth of 400 - 700 nm. The total measured PAR in the middle of the chamber was about 450 μ mol photons m⁻² s⁻¹. The plant chambers were continuously flushed with purified air, obtained by passing compressed air through a gas purification system consisting of several columns in series, filled with silica gel (2 - 5 mm, Merck, Germany), molecular sieve (0.3 nm perlform, Merck, Germany), charcoal (0.3 mm LS-Labor Service, Germany) and glass wool (Merk, Germany). The purified air was then led through a glass tank filled with demineralized water to humidify the air. Different NO₂ concentrations (between 0.3 and 4 ppb) were generated by mixing NO₂ from a pressurized standard cylinder $(m_{std,NO2} = 41151 \pm 2049 \text{ nmol m}^{-3} (1.004 \pm 0.050 \text{ ppm}) \text{ NO}_2 \text{ in } N_2; \text{ Air Liquide,}$ Germany) into the purified air stream. Mixing was performed by adjustment of two mass flow controllers (MKS Instruments, USA), one to keep the flow of NO₂ standard

gas ($Q_{std,NO2}$), the other the flow of the purified air stream (Q_{dil}) constant. The blended NO₂ concentration ($m_{blend,NO2}$) and its standard error ($s_{m_blend,NO2}$) are given by

$$m_{blend,NO2} = \frac{\left(m_{std,NO2} \ Q_{std,NO2} + m_{dil,NO2} \ Q_{dil}\right)}{\left(Q_{std,NO2} + Q_{dil}\right)}$$
(17.1)

$$s_{m_blend,NO2} = \pm \frac{(m_{blend,NO2})^2}{m_{std,NO2} Q_{std,NO2}} \sqrt{\left(\frac{s_{Q_std,NO2} Q_{dil}}{Q_{std,NO2}}\right)^2 + (s_{Q,dil})^2}$$
(17.2)

where $s_{m_blend,NO2}$ results of Gaussian error propagation applied to Eq. (17.1); concentrations (and standard errors) of $m_{std,NO2}$, $m_{blend,NO2}$ and $m_{dil,NO2}$ are in nmol m⁻³, flow rates (and standard errors) of in $Q_{std,NO2}$ and Q_{dil} are in m³ s⁻¹. For calculation of $s_{m_blend,NO2}$ it is assumed, that $m_{std,NO2}$ is constant (during the time of the laboratory experiment) and m_{dil} is zero.

The NO₂ mixture was directed into the dynamic plant chambers (without using the blowing axial inlet fan as for our field studies). For the laboratory measurements one plant chamber and one empty chamber with a volume (V) of 57 L were used. Each chamber was flushed at a constant flow (Q) of 14 L min⁻¹, controlled by mass flow controllers (MKS Instruments, USA), resulting in an exchange of the entire chamber's volume every 4 minutes. For two minutes each, air samples were directed to the analyzers from three different intake lines (purging NO₂ mixture (upstream of the chambers), outlet of empty and plant chambers). All analyzers were placed inside a cabinet (GKPv 6522, Liebherr, Germany) thermostatted at 25 °C to minimize variations of the analyzers

Specification and implementation of dynamic plant chamber system

In this chapter a dynamic chamber system based on previous measurements of volatile organic compounds, formaldehyde, formic and acetic acid and sulfur compounds (e.g. KESSELMEIER et al. 1993, 1996, 1998; KUHN et al. 2000) is presented. The dynamic chamber system allows exchange measurements of NO_2 (O_3 and NO) under field conditions (uncontrolled) as well as studies under controlled conditions including (laboratory) fumigation experiments.

Because NO₂ compensation point concentrations were reported at (sub-)ppb levels, our laboratory NO₂ fumigation experiments were performed with 3- to 4-yr old Norway Spruce trees at 0.3 - 3.4 ppb. Also under field conditions, such low ambient NO₂ concentrations can be expected. Moreover, exchange fluxes derived from dynamic chamber measurements are based on generally (very) small differences of NO₂ (NO, O₃) concentrations between inlet and outlet of the chamber. Consequently, detection limits of corresponding analyzers, statistical significance of the concentration differences, as well as the statistical goodness of measurements definitely have a substantial impact on the identification and quantification of statistically significant deposition velocities and compensation point concentrations, and have been considered correspondingly. Furthermore, as the exchange of NO₂ is a complex interaction of transport, chemistry and plant physiology, fluxes of NO, NO₂, O₃, CO₂ and H₂O were determined in the field experiments.

3.1 Methods

Quality assurance and error analysis

3.1.1 Corrections for concentration changes in long tubing

Long intake lines (mostly necessary for field experiments) may impact the trace gas concentrations (BEIER and SCHNEEWIND 1991). Trace gases may ad- or absorb on the inner walls of the tubing and/or react with each other according reactions (R1) and (R2). Therefore, we used opaque tubing to completely prevent photolysis of NO₂. Hence, reaction (R1) (NO + O_3) was the most important reaction to consider. For a known residence time, temperature and pressure in the tubes, the mixing ratios of NO, NO₂ and O₃ can be corrected according to BEIER and SCHNEEWIND (1991). To proceed, the residence time of the individual trace gas in the tubing as well as the characteristic chemical reaction time (τ_i ; $i = NO, O_3$) must be known. The latter is calculated by $\tau_{NO} = (k N_{O3})^{-1}$ $\tau_{\rm O3} = (k N_{\rm NO})^{-1}$ molecules cm⁻³, respectively (N_{O3}) and N_{NO} in and $k_{\text{R1}} = k = 1.4 \times 10^{-12} \exp(-1310/\text{T})$ in cm³ molecules⁻¹ s⁻¹; see ATKINSON et al. 2004).

3.1.2 Temporal response of analyzers

Response tests were carried out to check the response of analyzers to changes of concentrations when switching between intake lines with low concentration of the respective trace gas (NO, NO₂, O₃) to another intake line with high trace gas concentration (after stabilization) and back to the intake line of low concentration.

3.1.3 Temperature dependence of analyzers

The signals of analyzers are sensitive to the surrounding temperature. These effects are of special importance for field studies where it is more difficult to keep temperatures constant. Thus a series of tests were performed to determine the temperature dependence of all trace gas analyzers. The tests were done inside the conditioning cabinet (Heraeus, Germany) under different temperature conditions (temperature range: 18 - 46 °C). For each analyzer a calibration was carried out at each temperature level. We considered the correction of the analyzers' signals necessary if the observed drift
with temperature exceeded the maximum signal noise measured with zero air. We did not perform a correction when the drift was below 1 % for the entire temperature range or the analyzer's noise was greater than the temperature drift.

3.1.4 Dynamic chamber: internal mixing, exchange rate of chamber volume, wall absorption and transmissivity

The effective turbulent mixing as well as the fast exchange of the plant chamber's volume is essential for the determination of exchange flux densities of reactive as well as non-reactive trace gases (see MEIXNER 1994; MEIXNER et al. 1997). Particularly, the derivation of accurate NO2 and O3 leaf conductances from NO2 and O3 deposition velocities obtained by dynamic chamber measurements critically depends from the effectiveness of internal mixing and the chamber volume's exchange rate (see PAPE et al. 2009). Fast internal mixing of the chamber's volume has been assured by operation of three fans (see Figure 9) inside the chamber. A similar procedure was chosen by PAPE et al. (2009), who quantified complete mixing of the chamber volume in less than 2 s. The exchange rate of the chamber's volume is primarily determined by the volume V and the purging rate Q. However, due to delay effects of the sampling lines and due to the limited response times of the analyzers after switching between the different intakes, it is not possible to directly observe the trace gas' mixing in the plant chamber. Therefore, the time needed for temporal equilibrium of trace gas concentrations in an empty plant chamber was determined by measurements of a fast-response helium detector (Pico leak detector, MKS Instrument Inc., USA). A helium pulse was released into the purging stream of the chamber and the needed time for equilibration was determined.

Sorption effects (ad-, ab-, desorption) to and from the inner wall materials of the dynamic chamber should not modify the concentrations of (reactive) trace gases. Using the laboratory set-up, we investigated potential sorption effects to the inner walls of an empty chamber by fumigating it consecutively with different NO, NO₂ and O₃ concentrations. There were no desorption effects observed. Wall absorption was quantified in form of "blank" deposition velocities, where $v_{dep_wall,i} = Q (m_{a,i} - m_{s,i}) / (A_{wall} m_{s,i})$ $(i = NO_2, NO, O_3)$. In the field, the transmissivity of the FEP film (the dynamic chamber's wall) for PAR and the NO₂ photolysis rate $j(NO_2)$ has been monitored by continuous and simultaneous measurements of corresponding radiation fluxes inside and outside the chamber. PAR was measured with a LiCor quantum sensor (model LI-190SA, LiCor, Lincoln, NE, USA) and $j(NO_2)$ was determined as an omni-directional actinic UV radiation flux using a $j(NO_2)$ -sensor (filter radiometer, Meteorologie Consult GmbH, Königstein, Germany).

3.1.5 Significance of concentration differences

Particularly in the laboratory, the exchange flux density is directly proportional to $\Delta m_i = (m_{a,i} - m_{s,i})$, the difference of trace gas concentrations at the inlet and the outlet of the dynamic chamber (see Eq. (8.4)). Even under field conditions, the major component of the exchange flux density $F_{ex,i}$ is $Q/A_{leaf}\Delta m_i$. Keeping in mind, that (a) the sign of Δm_i determines direction of the exchange flux density and (b) the errors of $m_{a,i}$ and $m_{s,i}$ are decisively controlling the error of Δm_i (and consequently that of $F_{ex,i}$), it is more than obvious to control the significance of Δm_i . The corresponding statistical test requires the number of individual measurements, the averages and standard errors of $m_{s,i}$ and $m_{a,i}$. These were provided and calculated from the individual concentration measurements during one measurement cycle (laboratory: 30 min, field: 4 min). Prior to this, we identified outliers in the data sets by application of the Nalimov-test, a variant of Grubbs' test. Then, the significance of differentiation between the two averages of $m_{s,i}$ and $m_{a,i}$ was statistically secured by application of the t-test. Δm with statistical significance below 99 % ($\alpha < 0.99$) were correspondingly flagged and not included in subsequent calculations.

3.1.6 Bi-variate weighted linear least-squares fitting regression analysis

Since the concentrations $m_{a,i}$ and $m_{s,i}$ are measured with identical analyzers (see above), corresponding standard errors $s_{ms,i}$ and $s_{ma,i}$ are of the same order of magnitude. Therefore, bi-variate weighted linear least-squares fitting (which considers uncertainties of both, $m_{s,i}$ and $m_{a,i}$) is preferred to any standard forms of linear regression analysis (which consider, at best, uncertainties in the *y*-values, but no uncertainties in the *x*-values). The preferred algorithm delivers corresponding values of intersect (n_i) and slope (m_i) and other statistical quantities, like the standard errors of n_i and m_i $(s_{n,i}, s_{m,i})$, as well as correlation and regression coefficients, $r(m_{s,i}, m_{a,i})$ and $R^2(m_{s,i}, m_{a,i})$. YORK et al. (2004) presented the original set of equations for bi-variate weighted linear least-squares fitting regression analysis, where the slope m_i has to be solved iteratively (see below). For the iterative calculation a Microsoft Excel[®] spreadsheet was used, which has been provided by CANTRELL (2008) as a supplement of his paper (http://www.atmos-chem-phys.net/8/5477/2008/acp-8-5477-2008-supplement.zip).

Field data of concentrations in particular, have usually not all the same uncertainty. All kinds of linear least square fitting methods (considering errors in *y* and *x*) account for the fact, that data with the least uncertainty should have the greatest influence on the intercept *n* and the slope *m* of the fitted line. This is achieved by weighting each of the data points ($m_{a,i}$, $m_{s,i}$) with a factor ω_i , which is usually set to the inverse of the square of standard errors (standard deviations) of *x* and *y*-values (here: $s_{ma,i}^{-2}$ and $s_{ms,i}^{-2}$).

YORK et al. (2004) have provided a very detailed description of the bi-variate weighted linear least-squares fitting method. Here, only those equations are presented which are necessary to calculate the intersect *n* and the slope *m* of the best straight line (and related standard errors, s_n and s_m). For the sake of comparability with YORK et al. (2004), $m_{a,i} := X_i$ and $m_{s,i} := Y_i$, $s_{ma,i}^{-2} := \omega X_i$ and $s_{ms,i}^{-2} = \omega Y_i$ were set The method of YORK et al. (2004) to calculate the intercept *n* (*s_n*) and the slope *m* (*s_m*) comprises the following set of four equations:

$$n = \overline{Y} - m \, \overline{X}$$
; $i = 1, 2, ..., N$ (18.1)

$$m = \frac{\sum W_i \beta_i (Y_i - \overline{Y})}{\sum W_i (X_i - \overline{X})}$$
(18.2)

$$s_n^2 = \frac{1}{\sum W_i} + \bar{x}^2 s_m^2$$
(18.3)

$$s_m^2 = \frac{1}{\sum W_i (x_i - \bar{x})_i^2}$$
(18.4)

where,

$$x_{i} = \overline{X} + \beta_{i}; \quad y_{i} = \overline{Y} + \beta_{i};$$

$$\overline{X} = \frac{\sum W_{i}X_{i}}{W_{i}}; \quad \overline{Y} = \frac{\sum W_{i}Y_{i}}{W_{i}}; \quad \overline{x} = \frac{\sum W_{i}x_{i}}{W_{i}}; \quad \overline{y} = \frac{\sum W_{i}y_{i}}{W_{i}}$$

$$W_{i} = \frac{\omega(X_{i})\omega(Y_{i})}{\omega(X_{i}) + m^{2}\omega(Y_{i})}; \qquad \omega(X_{i}) = s_{X,i}^{-2}; \qquad \omega(Y_{i}) = s_{Y,i}^{-2}$$

$$\beta_{i} = W_{i} \left(\frac{X_{i} - \overline{X}}{\omega(Y_{i})} + \frac{m(Y_{i} - \overline{Y})}{\omega(X_{i})}\right); \qquad (18.5)$$

The original set of equations presented by YORK et al. (2004) contain additional terms in the equations for W_i and β_i for consideration of potential correlations between $s_{X,i}$ and $s_{Y,i}$, which are set to zero here (i.e. $s_{ma,i}$ and $s_{ms,i}$ are assumed to be uncorrelated). Since the equation for the slope *m* (Eq. 18.2) contains the variables W_i and β_i , which are in turn functions of *m* (see Eq. (18.5)), Eq. (18.2) has to be solved iteratively.

3.1.7 Standard errors of exchange flux densities, deposition velocities and compensation point concentrations

Standard errors of exchange flux densities $F_{ex,i}$, deposition velocities $v_{dep,i}$ and compensation point concentrations $m_{comp,i}$ of the NO-NO₂-O₃ triad may be derived by applying standard Gaussian error propagation. For that the standard errors of all variables on the right hand side of Eqs. (8.1) - (8.3), (13.1) - (13.3) and (14.1) - (14.3) must be known, and all variables of each individual equation should be independent of each other. However, the latter is not the case for (at least) $m_{s,i}$ and $m_{a,i}$ (see Eqs. (8.1) - (8.3)). Therefore, application of the generalized form of the Gaussian error propagation is preferred, which considers the mutual dependence of each pair variables (TAYLOR 1982; PHILLIPS et al. 2002). The general formulation of the standard error s_y of a quantity $y = f(x_1, x_2, x_3, ..., x_n)$ reads as follows:

$$s_{y}^{2} = \sum_{i=1}^{n} \left(\frac{\partial y}{\partial x_{i}} \cdot s_{x,i} \right)^{2} + 2 \cdot \sum_{i=1}^{n-1} \sum_{j=i+1}^{n} \frac{\partial y}{\partial x_{i}} \cdot \frac{\partial y}{\partial x_{j}} \cdot s_{x,i} \cdot s_{x,j} \cdot r(x_{i};x_{j})$$
(19)

where $r(x_i; x_j)$ are the correlation coefficients between each pairs of all x_i and x_j .

The individual variables x_i for the quantities $y = F_{exNO2}$, F_{exO3} , $v_{dep,NO2}$, $v_{dep,NO2}$, $v_{dep,NO2}$, $m_{comp,NO2}$, $m_{comp,NO2}$, $m_{comp,NO2}$, $m_{comp,NO2}$, $m_{comp,NO2}$, $m_{comp,O3}$ are defined by Eqs. (8.1) - (8.3), (13.1) - (13.3) and (14.1) - (14.3).

During field experiments, all $m_{a,i}$ and $m_{s,i}$ of the NO-NO₂-O₃ triad have been measured in cycles of 4 minutes. During this time period, it has been shown, that the error of the purging rate Q is negligible. The volume V of the chambers is *a priori* known, its error is considered to be zero. Standard errors of $m_{a,i}$ and $m_{s,i}$ are known for each data pair of measurements. Averages and standard errors of A_{leaf} , $j(NO_2)$, k and conjugated concentrations $m_{s,j}$ (j \neq i) have to be calculated individually from each data set which is used for the determination of $F_{ex,i}$, $v_{dep,i}$ and $m_{comp,i}$.

Therefore, according to Eq. (8.1), the mass exchange flux density F_{exNO2} is a function of 7 error-prone variables, namely $x_1 = m_{a,NO2}$, $x_2 = m_{s,NO2}$, $x_3 = j(NO_2)$, $x_4 = k$, $x_5 = m_{s,NO}$, $x_6 = m_{s,O3}$ and $x_7 = A_{leaf}$. Analogously to $F_{ex,NO2}$, the 7 variables for $F_{ex,NO}$ $(F_{ex,O3})$ in Eq. (8.2) (Eq. 8.3) are $x_1 = m_{a,NO}(m_{a,O3}), x_2 = m_{s,NO}(m_{s,O3}), x_3 = j(NO_2), x_4 = k$, $x_5 = m_{s,NO2}$, $x_6 = m_{s,O3}$ ($m_{s,NO}$) and $x_7 = A_{leaf}$. Considering Eq. (13.1), the deposition velocity $v_{dep,NO2}$ is a function of 3 error-prone variables, $x_1 = m_1$, $x_2 = j(NO_2)$ and $x_3 = A_{leaf}$, while the deposition velocity $v_{dep,NO}$ ($v_{dep,O3}$) depends on 4 error-prone variables, namely $x_1 = m_2$ (m_3), $x_2 = k$, $x_3 = m_{s,O3}$ ($m_{s,NO}$) and $x_4 = A_{leaf}$. The compensation point concentrations $m_{comp,NO2}$ ($m_{comp,NO}$, $m_{comp,O3}$) are each functions of 6 error-prone variables (see Eqs. (14.1) - (14.3)). These are $x_1 = n_1$ (n_2, n_3) , $x_2 = m_1$ (m_2, m_3) , $x_3 = i(NO_2)$, $x_4 = k$, $x_5 = m_{s,NO} (m_{s,NO2}, m_{s,NO2})$ and $x_6 = m_{s,O3} (m_{s,O3}, m_{s,NO})$. Bi-variate weighted linear leastsquares fitting regression analysis of measured $m_{s,i}$ versus $m_{a,i}$ (which considers both, $s_{ma,i}$ and $s_{ms,i}$) delivers the quantities n_1 , n_2 , n_3 and m_1 , m_2 , m_3 as well as their standard errors s_{n1} , s_{n2} , s_{n3} and s_{m1} , s_{m2} , s_{m3} . To calculate the standard errors $s_{Fex,NO2}$, s_{Fex $s_{Fex,O3}$, $s_{v,dep NO2}$, $s_{v,dep NO}$, $s_{v,dep O3}$, $s_{m,comp NO2}$, $s_{m,comp NO}$ and $s_{m,comp O3}$ by application of the general Gaussian error propagation (Eq. (19)), one have to calculate all the derivatives of $y_i = F_{ex,i}$, $y_i = v_{dep,i}$ and $y_i = m_{comp,i}$, $(i = NO_2, NO, O_3)$ with respect to the corresponding variables $x_1, x_2, ..., x_n$ mentioned above. The derivatives of $\partial y/\partial x_i$ are given in Table 7, Table 8 and Table 9.

Table 7: Derivatives $\partial y/\partial x_i$ of $y = F_{ex,NO2}$, $F_{ex,NO}$, $F_{ex,O3}$ with respect to the variables x_i in Eqs. (8.1) – (8.3) for application of the generalized Gaussian error propagation to calculate the standard errors of $s_{Fex,NO2}$, $s_{Fex,NO2}$ and $s_{Fex,O3}$ according to Eq. (19).

variable	dependent variable y							
X	F _{ex,NO2}	$F_{ex,NO}$	$F_{ex,O3}$					
m _{a,NO2}	$-rac{Q}{A_{leaf}}$							
m _{s,NO2}	$+\frac{Q}{A_{leaf}}\left(1+\frac{V}{Q}j(NO_2)\right)$	$-\frac{V}{A_{leaf}}j(NO_2)$	$-rac{V}{A_{leaf}}j(NO_2)$					
<i>m</i> _{a,NO}		$-rac{Q}{A_{leaf}}$						
m _{s,NO}	$-\frac{V}{A_{leaf}}k m_{s,O3}$	$+\frac{Q}{A_{leaf}}\left(1+\frac{V}{Q}k\ m_{s,O3}\right)$	$+ \frac{V}{A_{leaf}} k m_{s,O3}$					
<i>m</i> _{<i>a</i>,<i>O</i>3}			$-rac{Q}{A_{leaf}}$					
<i>m</i> _{<i>s</i>,<i>O</i>3}	$-\frac{V}{A_{leaf}}k m_{s,NO}$	$+ \frac{V}{A_{leaf}} k m_{s,NO}$	$+\frac{Q}{A_{leaf}}\left(1+\frac{V}{Q}k\ m_{s,NO}\right)$					
j(NO ₂)	$+ \frac{V}{A_{leaf}} m_{s,NO2}$	$-\frac{V}{A_{leaf}}m_{s,NO2}$	$-\frac{V}{A_{leaf}}m_{s,NO2}$					
k	$-\frac{V}{A_{leaf}}m_{s,NO} m_{s,O3}$	$+\frac{V}{A_{leaf}} m_{s,NO} m_{s,O3}$	$+\frac{V}{A_{leaf}} m_{s,NO} m_{s,O3}$					
A_{leaf}	$-rac{F_{ex,NO2}}{A_{leaf}}$	$-rac{F_{ex,NO}}{A_{leaf}}$	$-rac{F_{ex,O3}}{A_{leaf}}$					

Table 8: Derivatives $\partial y/\partial x_i$ of $y = v_{dep,NO2}$, $v_{dep,NO}$, $v_{dep,O3}$ with respect to the variables x_i in Eqs. (13.1) – (13.3) for application of the generalized Gaussian error propagation to calculate the standard errors of $s_{v,dep,NO2}$, $s_{v,dep,NO2}$ and $s_{v,dep,O3}$ according to Eq. (19).

variable			
X	V _{dep,NO2}	V _{dep,NO}	V _{dep,O3}
$\overline{m}_{s,NO}$			$-rac{V}{\overline{A}_{leaf}}\overline{k}$
$\overline{m}_{s,O3}$		$-rac{V}{\overline{\mathcal{A}}_{leaf}}ar{k}$	
$\overline{j}(NO_2)$	$-rac{V}{\overline{A}_{leaf}}$		
\overline{k}		$-rac{V}{\overline{A}_{leaf}}\overline{m}_{s,O3}$	$-rac{V}{\overline{A}_{leaf}}\overline{m}_{s,NO}$
\overline{A}_{leaf}	$-rac{v_{dep,NO2}}{\overline{A}_{leaf}}$	$-rac{v_{dep,NO}}{\overline{A}_{leaf}}$	$-\frac{v_{dep,O3}}{\overline{A}_{leaf}}$
m_1	$-rac{\overline{Q}}{\overline{A}_{leaf}} m_1^2$		
<i>m</i> ₂		$-rac{\overline{Q}}{\overline{A}_{leaf}} m_2^2$	
<i>m</i> ₃			$-\frac{\overline{Q}}{\overline{A}_{leaf}} m_3^2$

variable		dependent variable y	
X	<i>m_{comp,NO2}</i>	<i>m</i> _{comp,NO}	<i>mcomp,03</i>
$\overline{m}_{s,NO2}$		$-\frac{m_2V}{\overline{Q}}\bar{j}\big(NO_2\big)\cdotD_2^{-1}$	$-\frac{m_3V}{\overline{Q}}\overline{j}\big(NO_2\big)\cdotD_3^{-1}$
$\overline{m}_{s,NO}$	$-\frac{V}{\overline{Q}}m_1\overline{k}\overline{m}_{s,O3}\cdot D_1^{-1}$		$\left(n_3 - m_3 \frac{V}{\overline{Q}} \overline{j} (NO_2) \overline{m}_{s,NO2}\right) \frac{m_3 V \overline{k}}{\overline{Q}} \cdot D_3^{-2}$
$\overline{m}_{s,O3}$	$-rac{V}{\overline{\overline{Q}}}m_1\overline{k}\overline{m}_{s,NO}\cdot D_1^{-1}$	$\left(n_2 - m_2 \frac{V}{\overline{Q}} \overline{j} (NO_2) \overline{m}_{s,NO2}\right) \frac{m_2 V \overline{k}}{\overline{Q}} \cdot D_2^{-2}$	
$\bar{j}(NO_2)$	$\left(n_1 - m_1 \frac{V}{\overline{Q}} \overline{k} \overline{m}_{s,NO} \overline{m}_{s,O3}\right) \frac{m_1 V}{\overline{Q}} \cdot D_1^{-2}$	$-\frac{m_2 V}{\overline{Q}}\overline{m}_{s,NO2}\cdot D_2^{-1}$	$-\frac{m_3 V}{\overline{Q}}\overline{m}_{s,NO2}\cdot D_3^{-1}$
k	$-\frac{V}{\overline{Q}}m_1\overline{m}_{s,NO}\overline{m}_{s,O3}\cdot D_1^{-1}$	$\left(n_2\overline{m}_{s,O3} - m_2 \frac{V}{\overline{Q}} \overline{j}(NO_2)\overline{m}_{s,NO2} \overline{m}_{s,O3}\right) \frac{m_2 V}{\overline{Q}} \cdot D_2^{-2}$	$\left(n_{3}\overline{m}_{s,NO}-m_{3}\frac{V}{\overline{Q}}\overline{j}(NO_{2})\overline{m}_{s,NO2}\overline{m}_{s,NO}\right)\frac{m_{3}V}{\overline{Q}}\cdot D_{3}^{-2}$
n_1	$\left(1-m_1-m_1\frac{V}{\overline{Q}}\overline{j}(NO_2)\right)^{-1}:=D_1^{-1}$		
<i>n</i> ₂		$\left(1-m_2-m_2\frac{V}{\overline{Q}}\overline{k}\overline{m}_{s,O3}\right)^{-1}:=D_2^{-1}$	
<i>n</i> ₃			$\left(1 - m_3 - m_3 \frac{V}{\overline{Q}} \overline{k} \overline{m}_{s,NO}\right)^{-1} := D_3^{-1}$
m_1	$\left[n_{1}\left(1+\frac{V}{\overline{Q}}\overline{j}(NO_{2})\right)-\frac{V}{\overline{Q}}\overline{k}\overline{m}_{s,NO}m_{s,O3}\right]\cdot D_{1}^{-2}$		
<i>m</i> ₂		$\left[n_2 \left(1 + \frac{V}{\overline{Q}} \overline{k} \overline{m}_{s,O3} \right) - \frac{V}{\overline{Q}} \overline{j} (NO_2) \overline{m}_{s,NO2} \right] \cdot D_2^{-2}$	
<i>m</i> ₃			$\left\lfloor n_3 \left(1 + \frac{V}{\overline{Q}} \overline{k} \overline{m}_{s,NO} \right) - \frac{V}{\overline{Q}} \overline{j} (NO_2) \overline{m}_{s,NO2} \right\rfloor \cdot D_3^{-2}$

Table 9: Derivatives $\partial y/\partial x_i$ of $y = m_{comp,NO2}$, $m_{comp,NO2}$ and $m_{comp,O3}$ with respect to the variables x_i in Eqs. (14.1) – (14.3) for application of the generalized Gaussian error propagation to calculate the standard errors of $s_{m,comp_NO2}$, $s_{m,comp_NO2}$, $s_{m,comp_NO2}$ and $s_{m,comp_O3}$ according to Eq. (19).

3.1.8 Significance of the compensation point concentrations

The bi-variate weighted linear least-squares regression analysis of $m_{a,i}$ and $m_{s,i}$ delivers the intercept n_i , the slope m_i and their standard errors $s_{n,i}$ and $s_{m,i}$. According to Eqs. (14.1) - (14.3), each of the compensation point concentrations $m_{comp,i}$ of the NO-NO₂-O₃ triad can be considered as a random variable, represented by the average of $m_{comp,i}$ and the standard error $s_{m,comp,i}$. The decision whether or not a compensation point concentration exists is equivalent to the test of the hypothesis whether or not the average of $m_{comp,i}$ is highly significantly ($\alpha = 0.999$), significantly ($\alpha = 0.99$) or likely ($\alpha = 0.95$) different from $m_{comp,i}^* = 0$.

For that, it is assumed that each of the test quantities T_i

$$T_{i} = \left(\overline{m}_{comp,i} - m^{*}_{comp,i}\right) \cdot \frac{\sqrt{N}}{s_{m,comp,i}} \qquad i = NO_{2}, NO, O_{3}$$

$$(20)$$

matches the *t*-distribution with N-1 degrees of freedom. Depending on α , the hypothesis $m_{comp,i} = m^*_{comp,i}$ must be rejected, if

$$\left|\overline{m}_{comp,i} - m^*_{comp,i}\right| \ge \frac{S_{m,comp,i}}{\sqrt{N}} \cdot t_{\alpha;N-1}; \qquad \left(i.e. \ \frac{t_{\alpha;N-1}}{T_i} \le 1\right)$$
(21)

where $t_{\alpha;N-1}$ are the values of the *t*-distribution (*N*-1) for $\alpha = 0.999$, 0.99, 0.95, respectively.

3.2 Results

3.2.1 Analyzers and system performance

The results for the test of temperature dependence of all analyzers (see Sect. 3.1.3) are listed in Table 10. Between 18 and 46 °C the efficiency of the BLC drifted at from 37.0 % to 47.4 % over the whole temperature range. This means that for an initial concentration of 10 ppb NO₂ a drift of 2.2 ppb over the whole temperature range would be observed, which is equivalent to 3.6 nmol m⁻³/K (0.08 ppb/K). For NO the signal drift was 2.8 nmol m⁻³/K (0.07 ppb/K). The data of the CO₂ and O₃ analyzers did not need to be corrected because the signal drift was below 1 % for the entire temperature range, in contrast to the NO and NO₂ values. For the mathematical correction the slope of the regression line of the temperature tests (trace gas concentration versus temperature) was used.

Table 10: Results of the temperature dependence tests of the used analyzers. The temperatures are internal temperatures of the analyzers. The drift specifies the signal change during the whole temperature range. The signal noise is the maximum noise (3σ) detected with zero air during the test.

analyzer	trace gas	temperature range	drift	signal noise (3σ)
LI-7000	CO_2	22 – 44 °C	+ 0.97 ppm	0.25 ppm
LI-6262	CO_2	22 – 44 °C	- 3.5 ppm	0.23 ppm
TEI 49C	O_3	21 – 46 °C	+ 0.4 ppb	0.7 ppb
TEI 42C	NO	18 – 46 °C	- 1.9 ppb	0.2 ppb
TEI 42C/BLC	NO_2	18 – 46 °C	- 10.4 %	0.5 ppb

On the basis of the results of calibration procedures it was found, that the standard error of the O₃ concentration measurements could be considered as constant (±13.3 nmol m⁻³ or ±0.32 ppb) for the observed range of O₃ concentrations (719 - 2866 nmol m⁻³ or 19 - 77 ppb). The standard errors of NO₂ and NO concentration measurements are described by Eqs. (16.1) and (16.2); the parameters $s_{m_{\rm NO2,0}}$ and $s_{m_{\rm NO,0}}$ are given in Table 5 (3 σ -definition: $LOD(m_i) = 3 s_{m,i,0}$), $b_{\rm NO2} = 3.42 \times 10^{-4}$ nmol⁻¹ m³ (1.40×10⁻² ppb⁻¹) and $b_{\rm NO} = 7.88 \times 10^{-4}$ nmol⁻¹ m³ (3.23×10⁻² ppb⁻¹).

In Figure 12, the precision $(s_{m,i}/m_i)$ of the concentration measurements is exemplified for NO₂ during laboratory (red curve) and field experiments (green curve). The precision of m_{NO2} was only approx. 35 % during laboratory experiments at $LOD(m_{NO2}) = 1.04$ ppb (46.4 nmol m⁻³). After considerable improvement of the NO/NO₂ analyzer precision at 1 ppb improved to nearly 10 % in the field (however, precision was still 35 % at $LOD(m_{NO2}) = 0.31$ ppb (13.8 nmol m⁻³)). For further comparison, we consider that concentration m_i , where corresponding precision curves fall short of the 10 %-precision lines. These concentrations were 161.9 nmol m⁻³ (3.63 ppb; laboratory conditions), 45.9 nmol m⁻³ (1.03 ppb; field conditions), and they would be 14.7 nmol m⁻³ (0.33 ppb) and 1.3 nmol m⁻³ (0.03 ppb), if analyzers could be applied with $LOD(m_{NO2}) = 0.1$ and 0.01 ppb, respectively. For the NO and O₃ analyzers applied under field conditions, corresponding NO and O₃ concentrations (< 10 % precision) were 15.2 nmol m⁻³ (0.34 ppb; $LOD(m_{NO}) = 0.10$ ppb) and 144.5 nmol m⁻³ (3.24 ppb; $LOD(m_{O3}) = 0.98$ ppb), respectively.



Figure 12: Precision ($s_{m,NO2}/m_{NO2}$) of the applied NO/NO₂ analyzer during laboratory (red curve) and field experiments (green curve). For comparison, curves for precisions of hypothetical analyzers with $0.01 \le LOD(m_{NO2}) \le 2$ ppb are also shown (numbers on black and grey curves). The blue curve is the precision of the blended NO₂ concentration used for fumigation of the young spruce trees in the laboratory.

The performance of the dynamic chamber system depends critically on the temporal delay of concentrations (measured by only one set of analyzers) which are caused by switching between different intake lines of considerable length and by chemical reactions inside corresponding tubing (see Sect. 3.1.1). The tubing residence time for the 36.5 m long tubes of the field experiment was ≤ 4.1 s under ambient temperature and pressure conditions, calculated from sample flow (1.42 - 1.67 m³ s⁻¹ or 8.5 - 10 L min⁻¹), the length of the tubes and the tubes' inner diameter (0.00435 m). Since a considerable high flow through the intake filters and the long, thin tubes caused a distinct pressure drop (approx. 490 hPa), the actual residence time was consequently shorter (1.9 s). The characteristic chemical time scale (τ_{chem} ; e-fold time) for the NO + O₃ reaction (see (R1)) was within $20 < \tau_{chem} < 120$ s during the entire field experiment. Since τ_{chem} was always much longer than the tubing's residence time, any effects of the NO + O₃ reaction on measured concentrations could be neglected (as well as for the NO₂+hvreaction (R2), since opaque tubes have been used). However, the flow rate between the valve block (see Figure 10) and the analyzers is about 1/10 of the tubing purge flow; therefore, the "response time" of the entire system for a sudden change of concentrations was tested. Results are shown in Figure 13 for NO₂ (step change from 41 to 861 nmol m⁻³). Immediately after switching some typical pressure effects (valves) could be observed, but a temporally stable concentration was reached after 90 s. For the return switch a quite similar effect were observed and "response times" of NO, O₃, CO₂ and H₂O were comparable (data not shown). Based on these tests, the first 90 s of each concentration measurement were skipped from further data processing.



Figure 13: Response test for step changes between two different NO₂ concentrations (m_{NO2}). The red dashed line marks the switching point.

3.2.2 NO₂ blending for fumigation experiment

For laboratory NO₂ fumigation experiments very low (ppb- and sub-ppb levels) and temporally stable NO₂ concentrations have to be made available. That is essentially necessary to significantly identify any NO₂ compensation point whose concentrations are expected at these low concentration levels. Blended NO₂ concentrations ($m_{blend,NO2}$) of 13.4, 26.8, 44.6, 80.3 and 151.7 nmol m⁻³ (0.3, 0.6, 1.0, 1.8, 3.4 ppb) were provided by diluting an NO₂ standard into purified air (see Sect. 2.5.3). A typical course of these concentrations are shown in Figure 14, where the vertical dashed lines indicate times where blending was changed to obtain the next NO₂ concentration. A stable course of the new NO₂ concentration level was reached after max. 60 min. Fluctuation of the blended NO₂ concentration was between 8.0 and 16.1 nmol m^{-3} (0.18 - 0.36 ppb). These fluctuations do not depend on the analyzers' temperature (see Sect. 3.2.1). During laboratory measurements, the temperature variation of the instrument was only ± 0.5 °C, which would be equivalent to a change of $m_{blend,NO2} = 44.6$ nmol m⁻³ (1 ppb) of less than 1 %. The measured fluctuations could be also due to the precision of $m_{blend,NO2}$ which depends on the precision of the applied mass flow controllers. According to the manufacturer, the precision of the mass flow controllers $s \pm 0.8$ % of full scale. Using this information, the precision of $m_{blend,NO2}$ has been calculated through Eqs. (17.1) and (17.2) and is also shown in Figure 12. Uncertainty of the mass flow controllers may have added < 20 % to the observed variation of measured the blended NO₂ concentration.

3.2.3 Characterization of the dynamic plant chamber

3.2.3.1 Radiation and NO₂ photolysis rate

Transmissivity of PAR through the chamber walls (FEP film) is one of the fundamental requirements that the plant will be not affected by the chamber itself. Moreover, the calculation of the exchange flux density $F_{ex,i}$ (see Eqs. (8.1) - (8.3)) has to consider the NO₂ + hv reaction. For this, the photolysis rate $j(NO_2)$ inside the chamber volume has to be known. Therefore the transmissivity was controlled by simultaneous measurements inside and outside the chamber. While PAR was 10 % lower inside the chamber than outside, $j(NO_2)$ was 30 % lower inside the chamber (Figure 15).

RESULTS



Therefore, 70 % of ambient $j(NO_2)$ was used for the calculations of $F_{ex,i}$, $v_{dep,i}$, $m_{comp,i}$ and their standard errors.

Figure 14: Temporal course of blended NO₂ concentrations (12.3, 24.6, 41.0, 73.8 and 139.4 nmol m^{-3} (0.3, 0.6, 1.0, 1.8, 3.4 ppb)) used for fumigation of young spruce trees during the laboratory experiments. NO₂ concentrations were provided by diluting a NO₂ standard into purified air. Red dashed lines indicate times where blending was changed to obtain the next NO₂ concentration.



Figure 15: Simultaneous measurements of radiation in and outside a chamber. (a) Photosynthetically active radiation *PAR* (slope = 0.94, $R^2 = 0.98$, n = 456), (b) photolysis rate j_{NO2} (slope = 0.66, $R^2 = 0.99$, n = 1440). The black line indicates the 1:1 line and the red line represents the linear fit on the data points.

3.2.3.2 Sorption effects and chamber volume exchange time

An empty dynamic chamber has been exposed to various concentrations of NO₂, NO and O₃ concentrations and "blank flux densities" have determined according to Eq. (8.4). "Blank flux densities" for NO, NO₂ and O₃ are listed in Table 11. They were always negative (i.e. no desorption from the chamber's inner surfaces) and revealed very low values. Expressed in corresponding "wall deposition velocities" -2.12×10^{-3} (NO), -2.92×10^{-3} (NO₂) and -1.94×10^{-3} mm s⁻¹ (O₃) were found. These values were two orders of magnitude lower than $v_{dep,i}$ observed under laboratory as well as under field conditions. Comparing incoming and outgoing concentrations of the NO-NO₂-O₃ triad, a maximum of 2 % of the trace gases might have been absorbed by the inner surfaces of the plant chamber. Therefore, with regard to the mass balance of the dynamic plant chamber, neglecting of any mass fluxes to the walls of the chamber ($\partial M_{wall,i}/\partial t$ see Sect. 2.1.1) is certainly justified.

Table 11: Parameters of sorption effects to the inner chamber walls determined by laboratory experiments. q_{10} and q_{90} denote the 10 % and 90 % quantiles of the entire blank flux density $F_{wall,i}$ data, concentration ranges represent applied fumigation concentrations during the experiment, Δc_{mean} denotes the mean concentration difference of incoming and outgoing chamber air in % (range of differences in %).

$F_{wall,i}$, pn	nol $m^{-2} s^{-1}$		concentrations			
mean (±σ)	q ₁₀ q ₉₀	$v_{dep_wall,i}$, m s ⁻¹	range, ppb	Δc_{mean}		
-4.47 (±3.52)	-7.951.13	-2.12×10^{-6}	10 - 62	0.8 % (0.3 - 1.6)		
-4.43 (±3.11)	-9.111.51	-2.92×10^{-6}	6 - 47	1.8 % (0.4 - 3.4)		
-4.88 (±2.47)	-7.052.05	-1.94x10 ⁻⁶	7 - 45	1.6 % (0.5 - 3.7)		
	<i>F_{wall,i}</i> , pn mean (±σ) -4.47 (±3.52) -4.43 (±3.11) -4.88 (±2.47)	$F_{wall,i}$, pmol m ⁻² s ⁻¹ mean (± σ) $q_{10}q_{90}$ -4.47 (±3.52)-7.951.13-4.43 (±3.11)-9.111.51-4.88 (±2.47)-7.052.05	$F_{wall,i}$, pmol m ⁻² s ⁻¹ mean (± σ) $q_{10}q_{90}$ $v_{dep_wall,i}$, m s ⁻¹ -4.47 (±3.52)-7.951.13-2.12x10 ⁻⁶ -4.43 (±3.11)-9.111.51-2.92x10 ⁻⁶ -4.88 (±2.47)-7.052.05-1.94x10 ⁻⁶	$F_{wall,i}$, pmol m ⁻² s ⁻¹ concemean (± σ) q_{10} q_{90} $v_{dep_wall,i}$, m s ⁻¹ range, ppb-4.47 (±3.52)-7.951.13-2.12x10 ⁻⁶ 10 - 62-4.43 (±3.11)-9.111.51-2.92x10 ⁻⁶ 6 - 47-4.88 (±2.47)-7.052.05-1.94x10 ⁻⁶ 7 - 45		

The chamber volume exchange time was determined from an experiment, where a short pulse of (chemically inert) helium (He) has been added to the purging flow of the dynamic chamber (see Sect. 3.1.4). Results are shown in Figure 16. For the time of complete exchange (i.e., a constant level of He is observed), we used the time interval to reach 98 % of the final He concentration (t_{98}). Due to the limited temporal resolution of the He detector (5 s), t_{98} might have been between 80 and 85 s. This result was nearly identical to the time calculated from chamber volume (V = 79 L) and purging rate (Q = 60 L min⁻¹), which equals 79 s.



Figure 16: Results of the response time test with helium. The chamber ($V = 0.079 \text{ m}^3$) was operated with purging air flow rate $Q = 60 \text{ L min}^{-1}$. The red lines represent start and end of the helium addition, the black dashed line marks the end of equilibration. For the approximation of a complete exchange we used the time interval for 98 % approximation (t_{98}).

3.2.4 Demonstration of exchange flux density measurements

3.2.4.1 NO₂ exchange flux density: Laboratory results

Here, we confine ourselves to the results of "daytime" experiments, i.e. fumigation of the 3- to 4-yr old Norway Spruce trees with $13 < m_{a,NO2} < 152$ nmol m⁻³ (0.3 - 3.4 ppb), controlled temperature (25 °C), relative humidity (60 %) and PAR (450 µmol photons m⁻² s⁻¹, for 12 h) conditions. During experiment no significant difference of m_{O3} or m_{NO} between reference and plant chamber could be detected, also the amount of $j(NO_2)$ inside the chamber was negligible with respect to any measurable effects due to reaction (R2). As shown in Sect. 3.2.1, the performance of the NO₂ analyzer was definitely sub-optimal ($LOD(m_{NO2}) = 1.04$ ppb; 3 σ -definition). Therefore,

we based our evaluations of $F_{ex,NO2}$, $v_{dep,NO2}$ and $m_{comp,NO2}$ on a 2σ NO₂ detection limit (28.5 nmol m⁻³ or 0.6 ppb) for the observed concentrations ($m_{a,NO2}$, $m_{s,NO2}$). A total of 51 pairs of $m_{a,NO2}$ and $m_{s,NO2}$ have been obtained during the fumigation experiments. 17 data pairs passed the $LOD(m_{NO2})$ criterion, where another three of them had to be rejected due to the significance criterion for $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Fourteen data pairs of $m_{a,NO2}$ and $m_{s,NO2}$ have been subjected to a bi-variate weighted regression analysis 3.1.6), which resulted in $R^2 = 0.9706$, $n_1 = 1.7 \pm 2.63$ nmol m⁻³, Sect. (see $m_1 = 0.71 \pm 0.035$, $v_{dep,NO2} = 0.22 \pm 0.013$ mm s⁻¹ and $m_{comp,NO2} = 5.9 \pm 9.13$ nmol m⁻³. The significance probability of $m_{comp,NO2} \neq 0$ is 96.87 % ("likely"). NO₂ exchange flux densities ($F_{ex,NO2}$) and their standard errors have been calculated according to Eq. (11) and are shown in Figure 17. Figure 17a displays results of $F_{ex,NO2}$ where the 2σ -LOD(m_{NO2})-definition, Figure 17b where the 1σ -LOD(m_{NO2})-definition has been applied. Furthermore, in both panels $F_{ex,NO2}$ data were separated for the significance of (significant: blue circles, non-significant: reddish diamonds); Δm_{NO2} the $(F_{ex,NO2}; m_{s,NO2})$ -regression lines have been calculated according to Eq. (8.1.1) for all $F_{ex,NO2}$ data (pink line) and for those $F_{ex,NO2}$ data, where Δm_{NO2} is significant (blue line). Corresponding NO₂ compensation point concentrations $m_{comp,NO2}$ were calculated according Eq.(8.3.1) and are represented by red filled circles (significant Δm_{NO2}) and pink hollow circles (all data). Details of statistical evaluation are listed in Table 12. The most striking result is, that (regardless of which linear least-square fitting algorithm and which $LOD(m_{NO2})$ -definition is applied) the values of $m_{comp,NO2}$ are always highly significant, if all $F_{ex,NO2}$ data were used. Applying the simple linear least-square fitting algorithm (without considering $s_{m_a,NO2}$ nor $s_{m_s,NO2}$) $m_{comp,NO2}$ remains highly significant, even if only those $F_{ex,NO2}$ data are considered where Δm_{NO2} is significant. However, applying linear least-square fitting algorithms which consider either $s_{m s,NO2}$ or $s_{m a,NO2}$ and $s_{m s,NO2}$, the existence of $m_{comp,NO2}$ becomes "unlikely" ("likely"). With the exception of applying the 2σ NO₂ detection limit to all $F_{ex,NO2}$ data, the impact of different statistical treatments on the evaluation of NO₂ deposition velocities is small $(0.19 \le v_{dep,NO2} \le 0.22 \text{ mm s}^{-1}).$



Figure 17: Laboratory NO₂ fumigation of 3 - 4yr old Norway Spruce trees (*Picea abies* L.) under controlled conditions (25 °C, 60 %, 450 µmol photons m⁻² s⁻¹): NO₂ exchange flux density (*F*_{ex,NO2}) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber (*m*_{s,NO2}) for application of 2σ -*LOD*(*m*_{s,NO2})-definition (**(a)** panel) and 1σ -*LOD*(*m*_{s,NO2})-definition (**(b)** panel). *F*_{ex,NO2} data were calculated according Eq. (8.4), their standard errors according to Eq. (19). Blue circles identify *F*_{ex,NO2} where *m*_{s,NO2} > *LOD*(*m*_{s,NO2}), white circles stand for *F*_{ex,NO2} where *m*_{s,NO2} ≤ *LOD*(*m*_{s,NO2}) and reddish diamonds for those *F*_{ex,NO2} data, which have to be rejected for non-significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line (considering blue circle data) and pink line (considering blue circle and reddish diamond data) are calculated according to Eq. (15.1.1). NO₂ compensation point concentration *m*_{comp,NO2} is calculated according to Eq. (15.3.1) and is represented by red filled circles (considering blue circle and plue circle and reddish diamond data). More details of statistical evaluation are listed in Table 12.

Table 12: Parameters for NO ₂ laboratory measurements of simple (no errors considered), simple (standard error of m _{s,NO2} considered) and bi-variate
weighted (standard error of $m_{s,NO2}$ and $m_{a,NO2}$ considered) linear least-squares fitting regression analysis. Data were separated for all data of Δm_{NO2}
$(m_{a,NO2} - m_{s,NO2})$ and the significance of Δm_{NO2} . 2 σ , 1 σ and no NO ₂ detection limit was applied to the data.

			all	$(m_{a,NO2}-m_{s,NO2})$) data	only significant $(m_{a,NO2} - m_{s,NO2})$ data					
			linear lea	st-squares fittin	g algorithm	linear lea	linear least-squares fitting algorithm				
<i>LOD</i> (<i>m</i> _{NO2}) definition	statistical quantity	unit	simple, no errors considered	simple, only <i>s_{m_s,NO2}</i> considered	bi-variate & weighted, s _{m,a,NO2} & s _{m,s,NO2} considered	simple, no errors considered	simple, only s _{m_s,NO2} considered	bi-variate & weighted, s _{m,a,NO2} & s _{m,s,NO2} considered			
	N	[1]	17	17	17	14	14	14			
$LOD(m_{NO2})$	$R^2(m_{a,NO2},m_{s,NO2})$	[1]	0.9692	0.9716	0.9610	0.9794	0.9778	0.9706			
$2 imes \sigma_{m_NO2,0}$	$m_{comp,NO2}$	nmol m ⁻³	16.5 ± 1.81	14.2 ± 12.15	17.3 ± 7.29	6.8 ± 2.22	2.2 ± 16.76	5.9 ± 9.13			
definition	$m_{comp,NO2} \neq 0?$	%	99.99 (HS)	99.99 (HS)	99.99 (HS)	99.99 (HS)	37.1 (UL)	96.6 (L)			
	V _{dep,NO2}	mm s ⁻¹	0.27 ± 0.007	0.24 ± 0.016	0.26 ± 0.014	0.21 ± 0.006	0.20 ± 0.015	0.22 ± 0.013			
	N	[1]	45	45	45	33	33	33			
$LOD(m_{NO2})$	$R^2(m_{a,NO2},m_{s,NO2})$	[1]	0.9695	0.9754	0.9605	0.9847	0.9851	0.9782			
$1 \times \sigma_{m_NO2,0}$	m _{comp,NO2}	nmol m ⁻³	6.8 ± 0.52	7.3 ± 5.95	8.1 ± 3.46	-1.8 ± 0.63	-0.7 ± 7.82	0.6 ± 3.67			
definition	$m_{comp,NO2} \neq 0?$	%	99.99 (HS)	99.99 (HS)	99.99 (HS)	99.99 (HS)	39.5 (UL)	61.8 (UL)			
	$v_{dep,NO2}$	mm s ⁻¹	0.21 ± 0.004	0.22 ± 0.012	0.22 ± 0.010	0.19 ± 0.003	0.20 ± 0.012	0.20 ± 0.009			
	Ν	[1]	51	51	51	36	36	36			
$IOD(m_{MOD})$	$R^2(m_{a,NO2},m_{s,NO2})$	[1]	0.9682	0.9728	0.9575	0.9819	0.9815	0.9719			
not	$m_{comp,NO2}$	nmol m ⁻³	7.1 ± 0.44	6.8 ± 4.72	7.6 ± 3.07	-1.6 ± 0.60	-0.4 ± 6.22	0.5 ± 3.67			
considered	$m_{comp,NO2} \neq 0?$	%	99.99 (HS)	99.99 (HS)	99.99 (HS)	99.99 (HS)	27.6 (UL)	60.4 (UL)			
	Vdep,NO2	mm s ⁻¹	0.22 ± 0.004	0.22 ± 0.012	0.22 ± 0.010	0.19 ± 0.003	0.20 ± 0.011	0.20 ± 0.010			

RESULTS

3.2.4.2 NO-NO₂-O₃ exchange flux densities: Field results

In Figure 18, typical time series of trace gas mixing ratios are shown, measured at two different spruce branches during the EGER field campaign. The observed mixing ratio changes were due to switching between the different intakes. After switching, concentrations showed the delay effects mentioned above (see Sect. 3.2.1). Due to this, the first 90 s after valve switching were skipped from subsequent data processing (these first 90 s interval indicated as grey shaded vertical bars in Figure 18). Values for CO_2 and H_2O were measured as the difference between empty chamber and each switched intake. The temporal variation of CO_2 and H_2O concentrations of the plant chambers versus ambient air or empty chamber represented the physiological activity of the plants, since the CO_2 exchange flux density represents the photosynthetic CO_2 assimilation and the H_2O flux density the transpiration of the enclosed plant parts.

During the field experiment nearly 3000 pairs of $m_{a,i}$ and $m_{s,i}$ have been obtained. Applying the $LOD(m_i)$ (3 σ -definition) and the significance criterion for $\Delta m_i = (m_{a,i} - m_{s,i})$, around 60 % of the NO₂ data pairs remained. In Table 13 the details of the data pairs selection for both trees are listed for NO, NO₂ and O₃. Classification according to measurements during day and night demonstrated, that during night less data pairs were distinguishable from each other, especially those of NO. Between the spruce branches in both sampling chambers no differences were noticeable.

After classification of all individual concentration data into different classes of leaf conductance (approx. identical to different classes of radiation conditions), bi-variate weighted regression analysis between classified pairs of $m_{a,i}$ and $m_{s,i}$ was performed (see Sect. 3.1.6). The data pairs were additionally screened for singular concentration peaks of NO, NO₂ and O₃, which mainly occurred due to advection of automobile exhaust gases from a busy country road (2000 cars/h) in a distance of about 1-2 km from the site. The problem of advection at this field site is well known and has been documented through profile measurements of in- and above canopy concentrations, as well as through eddy covariance flux measurements of NO-NO₂-O₃ performed simultaneously to our dynamic chamber measurements (PLAKE et al. 2009). For the analysis of dynamic chamber derived O₃ flux densities, we assumed $m_{comp,O3} = 0$ ($n_3 = 0$), since emissions of O₃ from plants are not known so far.

For the present study, we restrict our results to one spruce branch (chamber 1) and one class with high PAR radiation (mean PAR = 355 μ mol photons m⁻² s⁻¹). The analysis for NO₂ resulted in $R^2(m_{a,NO2}, m_{s,NO2}) = 0.9480$, $n_1 = 6.5 \pm 1.59$ nmol m⁻³, $m_1 = 0.79 \pm 0.016$, $v_{dep,NO2} = 0.18 \pm 0.034$ mm s⁻¹ and $m_{comp,NO2} = -9.5 \pm 14.75$ nmol m⁻³. The probability of $m_{comp,NO2} \neq 0$ is 99.99 % ("highly significant"); however, a negative NO₂ compensation point concentration is physically meaningless. For O₃ the $R^2(m_{a,O3}, m_{s,O3}) = 0.9847, \qquad m_3 = 0.80 \pm 0.005$ analysis resulted in and $v_{dep.O3} = 0.32 \pm 0.018$ mm s⁻¹. In Figure 19a (Figure 20a), results of bi-variate weighted regression analysis between $m_{a,NO2}$ and $m_{s,NO2}$ ($m_{a,O3}$ and $m_{s,O3}$) are shown, while in Figure 19b (Figure 20b) those of $F_{ex,NO2}$ ($F_{ex,O3}$) versus $m_{s,NO2}$ ($m_{s,O3}$). In Figure 19a and b, data can be individually identified for their significance of Δm_{NO2} by corresponding color coding. For O₃, there is no corresponding color coding, since all Δm_{O3} were significant (see Table 13). Linear relationships between $F_{ex,NO2}$ and $m_{s,NO2}$ were calculated by Eq. 12.1 for data pairs owing significant Δm_{NO2} and for all data pairs. In Table 14 all results of statistical analysis of $F_{ex,NO2}$ and $F_{ex,O3}$ data are listed. Results of bi-variate weighted regression analysis for NO are shown in Figure 21. A large part of m_{NO} was lower than $LOD(m_{NO})$ (grey diamonds) or corresponding data pairs were non-significant with respect to $\Delta m_{NO} = (m_{a,NO} - m_{s,NO})$ (reddish diamonds). The regression coefficient $R^2(m_{a,NO}, m_{s,NO})$ was only 0.5355. Therefore, consecutive analyses are biased: probabilities of significant $m_{comp,NO}$ and $v_{dep,NO}$ becomes unlikely (51.7 and 22.4 %, respectively). Hence, there were no further evaluations for $F_{ex,NO}$, $v_{dep,NO}$ and $m_{comp,NO}$.

	$m_i > \text{LOD}$	tree 1 + significant Δ (number of tota	m_i % of total al)	$m_i > \text{LOD}$	tree 2 $m_i > \text{LOD} + \text{significant } \Delta m_i \% \text{ of total}$ (number of total)			
	all (2988)	day (1885)	night (1103)	all (2993)	day (1887)	night (1106)		
NO	24	33	7	24	33	8		
NO ₂	57	62	48	67	69	63		
O_3	96	98	93	98	99	97		

Table 13: Percentage of data m_i above $LOD(m_i)$ (3 σ -definition) and significant differences $\Delta m_i = (m_{a,NO2} - m_{s,NO2})$ of tree 1 and 2 for field measurements.



Figure 18: Switching scheme and time series of trace gas mixing ratios over two full measurement cycles during EGER field experiment. Data were corrected for calibration factors, temperature dependency and offset of analyzers. **(a)** Control scheme indicating periods of skipped data (first 90 s) for data processing (grey bars), sampling/analysis of ambient air (yellow bars), sampling/analysis of plant chamber 1 (green bars), sampling/analysis of reference chamber (red bars) and sampling/analysis of plant chamber 2 (blue bars). **(b-c)** Time series of CO_2 and H_2O mixing ratios measured as difference between reference chamber and respectively switched intake. **(d-f)** Time series of O_3 , NO_2 and NO mixing ratios. **(g)** Photosynthetic active radiation (PAR).



Figure 19: Field measurements: **(a)** NO₂ concentration measured at the outlet of the dynamic plant chamber $(m_{s,NO2})$ vs. NO₂ concentration measured at the inlet of the dynamic plant chamber $(m_{a,NO2})$. Light blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$ and reddish diamonds for those data pairs, which have to be rejected for non-significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$ and reddish diamonds for those data pairs, which have to be rejected for non-significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line (considering blue circle data) is calculated according to bi-variate weighted linear least-squares fitting regression analysis (see Sect. 3.1.6). **(b)** NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). $F_{ex,NO2}$ data were calculated according Eq. (8.4), their standard errors according to Eq.(19). Reddish diamonds stand for those $F_{ex,NO2}$ data, which have to be rejected for non-significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line (considering blue circle data) and pink line (considering blue circle and reddish diamond data) are calculated according to Eq. (15.1.1).



Figure 20: Field measurements: (a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis (see Sect. 3.1.6). (b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$. $F_{ex,O3}$ data were calculated according Eq. (8.4), their standard errors according to Eq. (19). Dark red line is calculated according to Eq. (15.1.1).



Figure 21: Field measurements: NO concentration measured at the outlet of the dynamic plant chamber $(m_{s,NO})$ vs. NO concentration measured at the inlet of the dynamic plant chamber $(m_{a,NO})$. Light green circles identify data pairs for significance of $\Delta m_{NO=} = (m_{a,NO} - m_{s,NO})$, reddish diamonds stand for those data pairs, which have to be rejected for non-significance of Δm_{O3} and grey diamonds for data pairs where $m_{NO} \leq LOD(m_{NO})$. Green line (considering green circle data) is calculated according to bi-variate weighted linear least-squares fitting regression analysis (see Sect. 3.1.6).

Table 14: Parameters of bi-variate weighted linear least-squares fitting regression analysis for
field measurements. NO ₂ data were separated for all data of Δm_{NO2} ($m_{a,NO2} - m_{s,NO2}$) and the
significance of Δm_{NO2} . Data of O ₃ were almost significant for $\Delta m_{O3} (m_{a,O3} - m_{s,O3})$. 3 σ detection
limit was applied to the data.

		all	only significant	only significant
		$(m_{a,NO2}-m_{s,NO2})$ data	$(m_{a,NO2}-m_{s,NO2})$ data	$(m_{a,O3}-m_{s,O3})$ data
statistical quantity	unit	NO ₂	NO ₂	O ₃
N	[1]	154	123	155
$R^2(m_{a,i};m_{s,i})$	[1]	0.9404	0.9480	0.9847
<i>m</i> _{comp,i}	nmol m ⁻³	-18.2 ± 17.57	-9.5 ± 14.75	0^*
$m_{comp,i} \neq 0?$	%	99.99 (HS)	99.99 (HS)	-
V _{dep,i}	mm s ⁻¹	0.14 ± 0.031	0.18 ± 0.034	0.32 ± 0.018

* assumption for O₃: $m_{comp,O3} = 0$.

DISCUSSION

3.3 Discussion

3.3.1 Overview of previous NO₂ exchange flux measurements using dynamic plant chambers

Table 15 shows a list of past dynamic chamber studies that have focused on NO₂ exchange between different plant species and the atmosphere. Most of these measurements were made with NO₂ converters which were not specific for NO₂ detection. Some authors used heated molybdenum converters (THOENE et al. 1991, 1996; TEKLEMARIAM and SPARKS 2006; RAIVONEN et al. 2009), heated ferrous sulphate converters (RONDÓN et al. 1993, RONDÓN and GRANAT 1994), or a detector based on chemiluminescence on liquid surfaces (HANSON et al. 1989; HEREID and MONSON 2001; SPARKS et al. 2001). All these converters overestimate NO₂ concentrations because of interferences with other (oxidized) nitrogen compounds (see Sect. 2.2). Only the application of photolytic converter guarantees the interference-free determination of particularly (very) low NO₂ concentrations.

During most of the field studies filtered air was used for purging the dynamic chambers. In most cases, this air was free of O_3 and NO_x , and known NO_2 concentrations were delivered to the dynamic chamber by diluting standard mixtures of NO_2 from a cylinder (GEBLER et al. 2000, 2002; SPARKS et al. 2001; HEREID and MONSON 2001). Some studies additionally controlled the CO_2 and water vapor concentrations of the purging air, the irradiance and temperature conditions inside the chamber (HEREID and MONSON 2001; SPARKS et al. 2001). Filtered and/or synthetic air (i.e. home-made H₂O and CO_2 concentrations, free of non target reactive trace gases) hardly represents ambient air. Therefore, a potential influence on the physiological behavior of the plant cannot entirely be excluded.

author	plant species	measured gases	location	wall material ¹	purging air ²	NO ₂ concentration in purging air, ppb	chamber volume, L	NO ₂ converter ³	analyzer	LOD⁴, ppb 3σ-definition	DIS
Hanson et al. 1989	deciduous, coniferous	NO ₂	lab	glass	pure $air^w + CO_2 + NO_2$	60 - 70	22.7	Luminol	LMA-3, Luminox	n.s.	CUSS
Thoene et al. 1991,1996	spruce	NO_2	lab	glass	zero air ^w + NO_2	1.6 - 125	3	Мо	Thermo Electron, 14B/E	NO ₂ : 1.0*	ION
Neubert et al. 1993	sunflower, tobacco	NO, NO ₂ , O ₃	lab	PTFE	zero air ^w + NO/NO ₂ /O ₃	< 100	160	PLC	Tecan, CLD 770 AL ppt	NO: 0.02; NO ₂ : 0.1*	
Rondón et al. 1993	pine, spruce	NO, NO ₂ , O ₃	field	FEP	ambient air, O_3 free + NO_2 ambient air + NO_2	0.25 - 120	10	FeSO ₄ Mo	Teco, 14D Tecan, CLD 770 AL ppt	NO: 0.3* NO: 0.06*	
Rondón & Granat 1994	pine, spruce	NO, NO ₂ , O ₃	lab	FEP	zero air ^w + CO ₂ + NO ₂ +O ₃	0.2 - 25	12.6	FeSO ₄ /PLC	Tecan, CLD 770 AL ppt	NO: 0.075; NO ₂ : 0.3	
Weber & Rennenberg 1996a,b	wheat	NO, NO ₂	lab	PMMA	zero air ^w + NO ₂	0 - 90	18-124	PLC	Tecan, CLD 770 AL ppt	NO: 0.075**	
Geßler et al. 2000, 2002	beech, spruce	NO ₂ , NH ₃	field, lab	BG	zero air ^w + NO ₂ /NH ₃	0.2 - 37	3	PLC	Tecan, CLD 770 AL ppt	NO ₂ : < 0.1*	
Sparks et al. 2001	tropical trees	NO ₂	field	n.s. (L)	pure $air^w + CO_2 + NO_2$	0.1 - 13	n.s.	Luminol	LMA-3, Luminox	NO ₂ : 0.005*	
Hereid & Monson 2001	corn	NO, NO ₂	field	n.s. (L)	pure air ^w + CO ₂ +NO/NO ₂	0.1->10	n.s.	Luminol	LMA-3, Luminox	NO ₂ : 0.005*	
Gut et al. 2002	tropical trees	NO, NO ₂ , O ₃	lab	FEP	ambient air	5 - 18	75	PLC	Eco-Physics, CLD 780 TR	NO: 0.052*	
Teklemariam & Sparks 2006	corn, sunflower, wheat	NO, NO ₂	lab	n.s. (L)	pure air ^w + CO ₂ +NO/NO ₂	1 - 5	n.s.	Мо	TEI 42	NO ₂ : 0.5*	YNA
Raivonen et al. 2009	Scots pine	NO, NO ₂	field	FEP, QG	ambient air	< 1	1	Мо	TEI 42S	n.s.	MIC
Chaparro-Suarez 2008	deciduous, coniferous	NO, NO ₂ , O ₃	lab	FEP	zero air ^w + NO_2	0 - 5	7.3	PLC	Eco-Physics, CLD 780 TR	NO: 0.06	۲LA
this study	spruce	NO, NO ₂ , O ₃	field lab	FEP	ambient air zero air ^w + NO ₂	0.4 - 21 0.3 - 4	75 60	BLC	TEI 42C	NO: 0.1; NO ₂ :0.31 NO: 0.2; NO ₂ :1.0	

Table 1	5: Overview	v of studies	that have	performed	dynamic cha	amber NO ₂ flu	x measurements	on different	plant species
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n.s. = not specified

 1 QG = quartz glass; BG = borosilicate glass; FEP, PFA, PTFE = Teflon materials; PMMA = polymethylmethacrylate (Plexiglas); L = dynamic leaf chamber of gas exchange system Model LI-6400, LiCor, Lincoln, Nebraska, USA

² w air humidified; pure air = air from a pure air generator; zero air = reactive trace gases removed with filters (NO_x, NH₃, H₂S, SO₂, O₃)

³ Mo = molybdenum converter; PLC = photolytic converter; $FeSO_4$ = ferrous sulphate converter; BLC = blue light converter

⁴ * LOD definition unknown; ** manufacturer's data

DISCUSSION

For field measurements of the NO-NO₂-O₃ triad under ambient conditions, fast gas phase reactions inside the chambers have to be considered. Therefore, NO, NO₂ and O₃ concentrations have to be measured simultaneously, even if only one of the trace gases is of interest (PAPE et al. 2009). All previous field studies described corrections of the calculated exchange flux densities not in detail. RONDÓN et al. (1993) specified some corrections for measured NO concentrations only, although O₃ and UV radiation were present in their dynamic chamber. In those cases where measurements of exchange flux densities were performed applying a simultaneously operated empty chamber (as "reference" chamber), corresponding flux densities were calculated from the concentration differences Δm_{NO2} between the outlet of the plant and empty chambers, respectively. This allowed a certain correction for chamber specific wall absorption and/or desorption processes (GEBLER et al. 2000, 2002; RAIVONEN et al. 2009). However, this procedure may not rule out adverse effects of fast gas-phase reactions on the evaluated flux densities, deposition velocities and compensation point concentrations (see below).

3.3.2 Precision, data quality and photochemical reactions

3.3.2.1 Precision and data quality

As shown in Sect. 3.2.1, the precision of NO₂ concentration measurements of our NO₂ analyzer improves from 35 % (at its limits of detection) rapidly to < 10 % at 162 nmol m⁻³ (3.63 ppb; laboratory) and 46 nmol m⁻³ (1.03 ppb; field). In Sect. 2.1.2 we presented the expected precision of the NO₂ exchange flux density for NO₂ concentrations up to 200 nmol m⁻³, for pre-scribed $m_{comp,NO2} = 67$ nmol m⁻³ (1.5 ppb), pre-scribed NO₂ deposition velocities (0.3 - 0.6 mm s⁻¹) and typical $R^2(m_{a,NO2}; m_{s,NO2})$ ranging from 0.99 to 0.9 (see Figure 8). Since $F_{ex,NO2}$ approaches zero at $m_{s,NO2} = m_{comp,NO2}$, the exchange flux density's precision ($\sigma_{Fex,NO2} / F_{ex,NO2}$) will become indefinite there. Consequently, the uncertainty of $F_{ex,NO2}$ will become as higher as closer $m_{s,NO2}$ approaches $m_{comp,NO2}$ (from either side). Analogously to the results shown in Figure 8, we determined which NO₂ concentration difference, $\pm |m_{s,NO2} - m_{comp,NO2}|$, will be necessary to keep the NO₂ exchange flux density's precision for our NO₂ analyzer under 10 %. For laboratory conditions ($LOD(m_{NO2}) = 45$ nmol m⁻³ or 1.01 ppb), this difference was ± 13.8 nmol m⁻³ or ± 0.31 ppb ($v_{dep,NO2} = 0.6$ mm s⁻¹; $R^2(m_{a,NO2}; m_{s,NO2}) = 0.99$) and

±91 nmol m⁻³ or ±2.05 ppb ($v_{dep,NO2} = 0.3 \text{ mm s}^{-1}$; $R^2(m_{a,NO2}; m_{s,NO2}) = 0.9$). During the EGER field experiment ($LOD(m_{NO2}) = 13.8 \text{ nmol m}^{-3}$ or 0.31 ppb) corresponding values were ±4.5 and ±8.5 nmol m⁻³ (0.1 and ±0.19 ppb), respectively. It is a serious consequence of these calculations, that, for a given detection limit, there is a well defined limit of $m_{comp,NO2}$ where the NO₂ compensation point concentration can be inferred from flux density data ($\sigma_{Fex,NO2} / F_{ex,NO2} \leq 10\%$) by interpolation of data measured on both sides of $m_{comp,NO2}$ lelow that limit, due to the obvious conflict of the requested $|m_{s,NO2} - m_{comp,NO2}|$ and $LOD(m_{NO2})$, $m_{comp,NO2}$ can only be inferred from flux density data at $m_{s,NO2} > m_{comp,NO2}$ by extrapolation, owing the risk of (much) higher uncertainties. These limits were for our NO₂ analyzer 33.5 and 133.8 nmol m⁻³ (0.75 and 3.0 ppb; laboratory) and 13.4 and 44.6 nmol m⁻³ (0.3 and 1.0 ppb; field) for the above mentioned combinations of $v_{dep,NO2}$ and $R^2(m_{a,NO2}; m_{s,NO2})$.

In previous studies the NO_2 sensitivity (a proxy for precision) of corresponding NO_x or NO₂ analyzers has been specified through their detection limit only (see Table 15). NEUBERT et al. (1993) and GEBLER et al. (2000), who used analyzers equipped with photolytic NO₂ converters mentioned a $LOD(m_{NO2})$ of 4.5 nmol m⁻³ (0.1 ppb); however, the corresponding definition of LOD (1 σ , 2 σ or 3 σ of σ_{NO20}) is not reported. Based on the manufacturer's data of the analyzers and on our experience, we assume that the reported values correspond to the 1σ -definition (P = 0.68). This assumption is in agreement with the values of RONDÓN and GRANAT (1994), who have used the same NO₂ analyzer model, namely with $LOD(m_{NO2}) = 8.9 \text{ nmol m}^{-3}$ (0.2 ppb; 2σ definition). Using the same LOD-definition (2σ) , RONDÓN and GRANAT (1994) reported a four times lower LOD for NO of 2.2 nmol m⁻³ (0.05 ppb). WEBER and RENNENBERG (1996a; 1996b) using also a photolytic NO₂ converter, have not reported any specifications about their instrument's sensitivity; therefore, we assumed that, based on the manufacturer's information about the applied NO/NO₂ analyzer, the LOD for NO was 33.5 nmol m⁻³ (0.075 ppb; 3σ-definition). According to RONDÓN and GRANAT (1994), and based on our experience the corresponding LOD for NO2 can be assumed to have not been better than 10 nmol m⁻³ (0.225 ppb; $3 \times LOD(m_{NO})$). Using the results of our simulation of the minimum detectable NO₂ compensation point concentration (see Sect. 2.1.3), we can state that NO₂ compensation point concentrations \geq 44.6 nmol m⁻³ $(\geq 1 \text{ ppb})$ can be detected with high significance, if NO₂ analyzers with $LOD(m_{NO2}) \approx 13.4 \text{ nmol m}^{-3}$ (0.3 ppb) were used (as WEBER and RENNENBERG 1996a and GEBLER et al. 2002) and $R^2(m_{a,NO2}, m_{s,NO2})$ was in a typical range (0.9 - 0.99) of laboratory measurements. Using NO₂ analyzers with $LOD(m_{NO2}) \approx 44.6$ nmol m⁻³ (\approx 1 ppb; e.g. analyzers with molybdenum converters) the significant detection of $m_{comp,NO2}$ > 44.6 nmol m⁻³ (1 ppb) would already be difficult, if the $v_{dep,NO2}$ is very small (< 0.3 mm s⁻¹). For example, THOENE et al. (1996) reported $m_{comp,NO2} = 73.1$ nmol m⁻³ (1.64 ppb) which has most likely be detected with high significance, because they $v_{dep,NO2} = 0.8 \text{ mm s}^{-1}$. On reported the other hand. the detection of $m_{comp,NO2} = 13.4 - 31.2 \text{ nmol m}^{-3}$ (0.3 - 0.7 ppb; RONDÓN et al. 1993) at $v_{dep,NO2} = 0.8 \text{ mm s}^{-1}$ seems now, from a statistical point of view, to be unlikely.

The data quality of exchange flux densities requires the control of quantifiable parameters of the measurement technique. To these belong the results of regular calibrations of the applied analyzers, their detection limits and those parameters which quantify the dependence of the analyzers' signals from other external factors like the ambient temperature. Our studies showed that the temperature dependency of the applied chemiluminescence NO/NO₂ analyzer can not be neglected (0.08 ppb/K). Hence, constant ambient temperature is definitely necessary to operate the analyzers at the requested level of precision. For our laboratory experiments we solved this problem with a commercial thermostat housing the analyzers. During field experiments this may be not always feasible. There, we used an air conditioning system for the entire instruments' shelter (container). Since the still remaining fluctuations of temperature were large enough to affect the precision of the NO/NO₂ analyzer, we corrected the analyzer's signals (see Sect. 3.2.1) It should be stated, that all mentioned previous studies on NO₂ exchange flux densities have even not mentioned this problem.

Laboratory measurements at very low concentrations demand low and stable blended NO₂ concentrations for fumigation of the plants. During our experiments we observed substantial fluctuations of the blended NO₂ concentration which entered the dynamic plant chamber. These fluctuations were due to the blending procedure (and the limited sensitivity of the NO/NO₂ analyzer). As shown in Figure 12 (blue line), the noise of NO₂ concentrations caused by the blending procedure itself will substantially affect the precision of the NO₂ concentration measurements (and consequently those of NO₂ flux density), particularly if the detection limit of future NO₂ analyzers will be improved to be better than 10 nmol m⁻³ (0.25 ppb). Then, the improved precision of the NO_2 concentration measurements will fall short of the noise of the blended NO_2 concentration at the inlet of the dynamic chamber (see Figure 12) and the improvement of the blending procedure (e.g. by application of more precise flow controllers) will become necessary.

3.3.2.2 Significance of concentration differences

The error of NO₂ exchange flux density measurements by the dynamic chamber method mainly depends on the error of trace gas concentration differences, Δm_i between the inlet and the outlet of the dynamic plant chamber. In contrast to laboratory conditions, NO₂ concentrations in the field were relative high and rarely conflicted $LOD(m_{NO2})$. However, during field measurements about 30 to 40 % of daytime Δm_{NO2} data were found to be not significantly different from each other (Table 13) and had to be rejected from further analysis. This rather high percentage of rejected data was mostly due to the temporal variation of ambient NO₂ concentration ($m_{a,NO2}$) during the 4 min measurement interval, rather than due to the precision or to $LOD(m_{NO2})$. Ambient NO₂ mixing ratio can rapidly change due to the spatially and temporally varying sources within area surrounding the site of measurements (nearby country roads). In our laboratory studies the percentage of non-significant Δm_{NO2} "daytime" data was 37 % for $m_{a,NO2} < 44.6$ nmol m⁻³ (1 ppb) and vanished for $m_{a,NO2} \ge 71.4$ nmol m⁻³ (1.6 ppb).

In some of the previous studies means or data sets were compared for significant differences by analysis of variance (e.g. WEBER and RENNENBERG 1996a, 1996b; HEREID and MONSON 2001; SPARKS et al. 2001). However, actual numbers on significant Δm_{NO2} were not reported. We like to emphasize, that (1) our approach to apply a significance test on the measured concentrations directly is rather novel, and (2) the control of the significance of Δm_{NO2} is one of the fundamental quality control criteria for highly significant NO₂ exchange flux densities, NO₂ deposition velocities and above all the detection of highly significant NO₂ compensation point concentrations. When using data without significance control of Δm_{NO2} , NO₂ compensation point concentrations will be overestimated (see below) and therefore be (highly) significant but not true.

DISCUSSION

3.3.2.3 Photo-chemical reactions in the dynamic plant chamber: impact on net exchange flux densities, deposition velocities and compensation point concentrations

In the mentioned previous studies, the impact of photo-chemical reactions was mostly not considered, neither for the calculation of $v_{dep,NO2}$ nor for that of $m_{comp,NO2}$. Partly, not all components of the NO-NO₂-O₃ triad were always measured. Furthermore, most field studies have not used ambient air as purging air. Instead, ambient air was filtered to remove reactive trace gases, particularly O₃ and NO_x. Afterwards, the desired NO₂ concentration was blended (e.g. GEBLER et al. 2000). Using filtered air, free of NO and O₃, allows to neglect reaction (R1), but photolysis of NO₂ (R2) will still occur, as soon as appreciable amounts of $j(NO_2)$ are present in the plant chamber. Consideration of photo-chemical reactions, like the NO₂ loss by reaction (R2) and the formation of NO₂ by reaction (R1) were mentioned by NEUBERT et al. (1993), the production and destruction of NO by RONDÓN et al. (1993).

With the framework of equations developed in Sects. 2.1.2 and 2.1.3, we provide a straightforward tool to examine the impact of photo-chemical reactions on the determination of exchange flux densities, deposition velocities and compensation point concentrations. While actual $F_{ex,i}$, $v_{dep,i}$ and $m_{comp,i}$ are described by Eqs. (12.1) - (12.3), (13.1) - (13.3) and (14.1) - (14.3), the quantities $F^*_{ex,i}$, $v^*_{dep,i}$ and $m^*_{comp,i}$ are given by Eqs. (15.1.1) - (15.1.3), (15.2.1) - (15.2.3) and (15.3.1) - (15.3.3). The latter are those quantities, which would have been observed if no photo-chemical reactions had taken place (e.g. for NO₂ during our laboratory experiments, see Sect. 3.2.1). According to Eqs. (8.4), (15.1.1), (15.2.1) and (15.3.1), the exchange flux densities $F^*_{ex,i}$ are identical to the so-called "chamber flux densities", $F_{cham,I} = -Q/A_{leaf}(m_{a,i} - m_{s,i})$.

In previous experiments, where photo-chemical reactions have not been considered, the actual exchange flux densities $F_{ex,i}$ have just been substituted by $F_{cham,i}$ alone. During some of the more recent experiments photo-chemical reactions have been either (partially) excluded by corresponding set-ups or were "considered" by application of an empty chamber ("reference chamber") (RONDÓN et al. 1993; GEBLER et al. 2000, 2001; HEREID and MONSON 2001; Sparks et al. 2001; RAIVONEN et al. 2009). However, photochemical reactions within the latter chamber will be definitely different from those in the dynamic plant chamber, simply for the fact, that neither $j(NO_2)$, nor $m_{s,NO2}$, $m_{s,NO}$ or

DISCUSSION

 $m_{s,O3}$ are identical in both chambers. In order to examine potential under/overestimation of simple "chamber flux densities" $F_{cham,i}$, by neglecting NO-NO₂-O₃ gas-phase production and destruction fluxes, we combine the mentioned equations to obtain:

$$F_{ex,NO2} = F_{cham,NO2} - \frac{V}{\overline{A}_{leaf}} \left(\overline{k} \, \overline{m}_{s,NO} \overline{m}_{s,O3} - \overline{j} \left(NO_2 \right) \overline{m}_{s,NO2} \right)$$
(22.1)

$$F_{ex,NO} = F_{cham,NO} - \frac{V}{\overline{A}_{leaf}} \left(\overline{j} \left(NO_2 \right) \overline{m}_{s,NO2} - \overline{k} \ \overline{m}_{s,NO} \overline{m}_{s,O3} \right)$$
(22.2)

$$F_{ex,O3} = F_{cham,O3} - \frac{V}{\overline{A}_{leaf}} \left(\overline{j} (NO_2) \overline{m}_{s,NO2} - \overline{k} \, \overline{m}_{s,NO} \overline{m}_{s,O3} \right)$$
(22.3)

Whether actual exchange flux densities $F_{ex,i}$ are higher, equal or lower than corresponding $F_{cham,i}$ depends whether the difference of the corresponding gas-phase destruction and production fluxes (second term, right hand side of Eqs. (22.1) - (22.3)) is positive, negative and different from zero.

If we differentiate our calculated exchange flux densities $F_{ex,i}$ of the field experiment into the chamber flux densities $F_{cham,i}$ and the gas-phase flux densities $F_{gas,i}$, which comprised the gas-phase production and destruction of NO-NO₂-O₃, we can identify the fraction of $F_{gas,i}$ of each $F_{ex,i}$. For the selected leaf conductance class (see Sect. 3.2.4.2), the percentage of $F_{gas,i}$ is displayed in Figure 22 for NO, NO₂ and O₃. The fraction of $F_{gas,O3}$ at the exchange flux density of O₃ is very small (±1 %); therefore, it can be neglected. For the NO₂ exchange flux density the fraction of $F_{gas,NO2}$ becomes much more important. The median contribution of $F_{gas,NO2}$ to $F_{ex,NO2}$ was just +8 %, but in particular cases it could be +22 % or -12 %, respectively. Quite clear becomes the impact of the gas-phase reactions for the NO exchange flux density. Here, $F_{gas,NO}$ amounted +42 % (median value), but ranging from +85 % to -170 %. That means, that under certain conditions $F_{ex,NO}$ can change its sign, if $F_{gas,NO}$ will not be considered: the estimated NO emission will convert to a NO deposition (or vice versa).



Figure 22: Percentage of gas-phase flux densities $F_{gas,i}$ at the exchange flux densities $F_{ex,i}$ for NO (green diamond), NO₂ (blue diamond) and O₃ (orange diamond). Results are from the field experiment, restricted to one selected data class (see Sect. 3.2.4.2). The apexes of the diamonds represented the upper (75 %) and the lower (25 %) quantile and the black dash in the middle of the diamonds the median. $F_{gas,NO}$ and $F_{gas,NO2}$ were applied to the left y-axis and $F_{gas,O3}$ to the right y-axis.

Similar relations can be developed for deposition velocities $v_{dep,i}$ by combining Eqs. (13.1) - (13.3) with Eqs. (15.2.1) - (15.2.3):

$$v_{dep,NO2} = v_{dep,NO2}^{cham} - \frac{V}{\overline{A}_{leaf}} \overline{j} (NO_2)$$
(23.1)

$$v_{dep,NO} = v_{dep,NO}^{cham} - \frac{V}{\overline{A}_{leaf}} \overline{k} \,\overline{m}_{s,O3}$$
(23.2)

$$v_{dep,O3} = v_{dep,O3}^{cham} - \frac{V}{\overline{A}_{leaf}} \overline{k} \,\overline{m}_{s,NO}$$

$$(23.3)$$

where the quantities with the superscript "*cham*" are those which be derived from using "chamber flux densities" $F_{cham,i}$ instead of actual exchange flux densities $F_{ex,i}$. The actual deposition velocities $v_{dep,i}$ are in any case lower than $v^{cham}_{dep,i}$ with the exception $m_{s,O3} = 0$, $m_{s,NO} = 0$ and $j(NO_2) = 0$ (i.e. during nighttime). To examine how much the

gas-phase reactions will affect $v_{dep,i}$, we split our calculated deposition velocity $v_{dep,i}$ for the field data into $v^{cham}_{dep,i}$ and the complementary part caused by gas-phase reactions. The contribution of photolysis (see Eq. 23.1) to $v_{dep,NO2}$ was 80 %, that of reaction (R1) on $v_{dep,O3}$ only 3 %. Corresponding estimates on $v_{dep,NO}$ were not performed, since NO deposition velocities were not significant during the EGER field experiment. For their experimental conditions, NEUBERT et al. (1993) identified an error of about 20 % for their v_{dep,NO2} determination, if they would neglect photolysis of NO₂. However, our results should be compared to those of previous studies with caution: in most of the previous studies it is not clear whether the photolysis of NO₂ was correctly taken into account. Nevertheless, we tried to estimate the potential impact of NO₂ photolysis on these, previously reported $v_{dep,NO2}$. For that, the quantities A_{leaf} , V, $j(NO_2)$ and $v_{dep,NO2}$ have to be a priori known or they must be derived from other (accompanying) data. Most of the authors have not reported any data of A_{leaf} . So, we estimated the unknown A_{leaf} on the basis of available information about chamber design and our experience concerning the ratio between length of branch and needle area. Moreover, most authors did not specify the used chamber wall material nor its transmissivity for the wavelength range of $i(NO_2)$. Therefore, we estimated the transmissivity on basis of available material information. THOENE et al. (1991, 1996) and GEBLER et al. (2002) used borosilicate glass (Schott Glaswerke, Mainz, Germany). Combining the manufacturer's specification (http://www. schott.com/tubing) and our experience with different wall materials (including glass) we estimated the $i(NO_2)$ transmissivity of borosilicate glass to 60 %. For FEP-Teflon film, used by RONDÓN et al. (1993), we estimated 70 % transmissivity (related to our Teflon film). If NO₂ photolysis would not have been considered at all, THOENE et al. (1991,1996) and RONDÓN et al. (1993) would have potentially overestimated their $v_{dep,NO2}$ values by 17 - 81 % and GEBLER et al. (2002) by up to 100 % (according to Eq. (23.1), depending on prevailing radiation conditions). However, since these authors have applied an empty ("reference") chamber (see Sect. 3.3.1), the impact on NO₂ photolysis on their reported $v_{dep,NO2}$ values might be smaller if the underlying assumption is correct that the effect of NO₂ photolysis is identical in the plant and in the empty chamber. The results of field measurements by SPARKS et al. (2001) and HEREID and MONSON (2001) most likely have not been affected by NO₂ photolysis because they used a leaf chamber system with red light-emitting diodes which produce no appreciable radiation in the wavelength range of $i(NO_2)$.

82 | DYNAMIC PLANT CHAMBER SYSTEM

DISCUSSION

The corresponding relations for the compensation point concentrations $m_{comp,i}$ are obtained by combining Eqs. (14.1) - (14.3) with Eqs. (15.3.1) - (15.3.3):

$$m_{comp,NO2} = m_{comp,NO2}^{cham} \cdot \frac{1 - m_1 \left[1 + \frac{V}{n_1 \overline{Q}} \ \overline{k} \ \overline{m}_{s,NO} \overline{m}_{s,O3} (1 - m_1) \right]}{1 - m_1 \left(1 + \frac{V}{\overline{Q}} \ \overline{j} (NO_2) \right)}$$
(24.1)

$$m_{comp,NO} = m_{comp,NO}^{cham} \cdot \frac{1 - m_2 \left[1 + \frac{V}{n_2 \overline{Q}} \, \overline{j} (NO_2) \overline{m}_{s,NO2} \left(1 - m_2 \right) \right]}{1 - m_2 \left(1 + \frac{V}{\overline{Q}} \, \overline{k} \, \overline{m}_{s,O3} \right)}$$
(24.2)

$$m_{comp,O3} = m_{comp,O3}^{cham} \cdot \frac{1 - m_3 \left[1 + \frac{V}{n_3 \overline{Q}} \ \overline{j} (NO_2) \overline{m}_{s,NO2} (1 - m_3) \right]}{1 - m_3 \left(1 + \frac{V}{\overline{Q}} \ \overline{k} \ \overline{m}_{s,NO} \right)}$$
(24.3)

Here, the value of the fraction (right hand side of Eqs. (24.1) - (24.3)) determines whether the actual compensation point concentrations $m_{comp,i}$ are higher, equal, or lower than $m^{cham}_{comp,i}$.

For our experimental conditions, $m_{comp,NO2}$ would be overestimated by 10 %, if the gas-phase reactions would not have been considered (i.e. assuming $m_{comp,NO2} = m^{cham}_{comp,NO2}$). For the compensation point concentration of O₃ the overestimation would be only 1 %. The $m_{comp,NO2}$ values reported in previous studies (THOENE et al. 1991, 1996; RONDÓN et al. 1993, GEBLER et al. 2002) would be overestimated between 3 and 17 %, if the photolysis of NO₂ was not considered.

When the value of the fractions on the right hand side of Eqs. (24.1) - (24.3) are examined for being greater, equal, or lower than unity, the following relations are obtained:

$$m_{comp,NO2} > (=,<) m_{comp,NO2}^{cham}$$
, if $m_{comp,NO2}^{cham} > (=,<) \frac{k \overline{m}_{s,NO} \overline{m}_{s,O3}}{\overline{j}(NO_2)}$ (25.1)

$$m_{comp,NO} > (=,<) m_{comp,NO}^{cham}$$
, if $m_{comp,NO}^{cham} > (=,<) \frac{\overline{j}(NO_2)\overline{m}_{s,NO2}}{\overline{k} \overline{m}_{s,O3}}$ (25.2)
$$m_{comp,O3} > (=,<) m_{comp,O3}^{cham}$$
, if $m_{comp,O3}^{cham} > (=,<) \frac{\overline{j}(NO_2)\overline{m}_{s,NO2}}{\overline{k} \overline{m}_{s,NO}}$ (25.3)

The relevance of these relations consists in their potential for the easy check, whether or not the correct evaluation of compensation point concentrations has to consider photochemical reactions. Having evaluated measured concentrations $m_{a,i}$ and $m_{s,i}$ by bi-variate weighted linear regression (which delivers n_i and m_i), the quantities $m^{cham}_{comp,i}$ are determined. Using the simultaneously measured averages of k, $j(NO_2)$, $m_{s,NO2}$, $m_{s,NO}$ and $m_{s,O3}$, the right hand fractions of relations (25.1) - (25.3) can be calculated, which provide the necessary quantities to test whether or not $m^{cham}_{comp,i}$ have to be corrected for photo-chemical reactions in the dynamic plant chamber (by Eqs. (24.1) - (24.3)).

3.3.3 Bi-variate weighted linear regression

The determination of deposition velocities $v_{dep,i}$, as well as compensation point concentrations $m_{comp,i}$ is based on linear regression of the measured concentration of trace gas *i* in ambient air and within the dynamic plant chamber. Therefore, it is indispensable that errors of both variables were considered in the determination of $v_{dep,i}$ and $m_{comp,i}$. For our laboratory results (see Sect. 3.2.4.1) we have shown the effect of applying simple linear regression (no errors considered at all), linear regression (y-errors considered) and bi-variate weighted linear regression (y- and x-errors considered) on the significance of derived $v_{dep,NO2}$ and $m_{comp,NO2}$ data (see Table 12). Generally speaking, applying a simple linear least-square fitting algorithm, the probability of $m_{comp,i} \neq 0$ can be highly significant, while applying the bi-variate weighted linear least-square fitting algorithm the probability for the existence of $m_{comp,i}$ could easily become "likely" or even "unlikely". In the fewest cases previous authors have applied the bi-variate algorithm (e.g. GEBLER et al. 2000, 2002). Finally, it should be stated that in all previous studies values of $v_{dep,NO2}$ and $m_{comp,NO2}$ have been derived from linear relationships between $F_{ex,NO2}$ and $m_{s,NO2}$ which is mathematically not correct, since the dependent variable $F_{ex,NO2}$ contains the independent variable $m_{s,NO2}$ (see Sect. 2.1.2).

Application of dynamic plant chamber system to field measurements

In this chapter the application of the dynamic plant chamber system to field measurements is presented. The chamber system was used during the second intensive observation period of the EGER project from 01 June to 15 July 2008. It took place in northeast Bavaria, Germany (see Sect. 2.5.2). Additionally, data measured during the project ECHO (Emission and CHemical Transformation of Biogenic Volatile Organic Compounds) within the German Atmospheric Research Program (AFO 2000) were provided to me for analysis. For this study the exchange rates of NO, NO₂, O₃, CO₂ and H₂O on oak (*Quercus robur*) under environmental conditions were measured.

4.1 Methods

4.1.1 Photosynthesis rate, transpiration rate, stomatal conductance

Considering the exchange of the non-reactive trace gases CO₂ and H₂O between the plant chamber's atmosphere and the enclosed leaves, the exchange flux densities of the net rate of photosynthesis (photosynthesis rate minus the simultaneously proceeding photorespiration) $F_{ex,CO2}$ (in µmol m⁻² s⁻¹) and the transpiration rate $F_{ex,H2O}$ (in mmol m⁻² s⁻¹) were calculated after VON CAEMMERER and FARQUHAR (1981):

$$F_{ex,n} = -\frac{Q}{A_{leaf}} (m_{r,n} - m_{s,n}) \qquad n = CO_2, H_2O$$
(26)

The calculation is based on the difference between the molar concentration $m_{r,n}$ and $m_{s,n}$ (µmol m⁻³ or mmol m⁻³) of trace gas *n* within the empty reference chamber and the plant

METHODS

chamber, respectively, the enclosed leaf area (A_{leaf}) and the chamber purging rate (Q). Equation (26) is comparable with Eq. (8.4), which describes the exchange flux density $F_{ex,i}$ of the reactive trace gas i ($i = NO_2$, NO, O₃) if gas-phase production and/or destruction of the reactive trace gas can be ruled out.

The stomatal conductance for H₂O (g_{H2O} in m s⁻¹) was calculated from the transpiration rate (in kg m⁻² s⁻¹) and the humidity gradient, which is the difference between the absolute humidity inside the leaf (ah_{leaf} in kg m⁻³) and the absolute humidity of ambient air (ah_a in kg m⁻³). The humidity inside the leaf was calculated as saturated vapor concentration at leaf temperature:

$$g_{H2O} = \frac{F_{H2O}}{ah_{leaf} - ah_a}$$
(27)

The predicted leaf conductance of the gas $i (g_{i,p})$ ($i = NO_2, O_3$) was estimated from stomatal conductance to water vapor (g_{H2O}) by scaling it

$$g_{i,p} = g_{H2O} \cdot R_D \tag{28}$$

where R_D is the ratio of diffusivities (cm² s⁻¹) of NO₂ or O₃ and water vapor in air. According to MASSMAN (1998) values of R_D were amounted to 0.62 for NO₂ and 0.66 for O₃ at conditions near standard temperature and pressure (T = 0 °C, p = 101.325 kPa).

4.1.2 Classification of data

Consideration of a possible compensation point concentration and determination of the deposition velocity require a certain amount of measuring points. During data selection it is necessary to find comparable conditions for the plant. This avoids comparison of, for example, trace gas exchange rates measured during a physiologically active phase of the plant with wide opened stoma and exchange rates measured at a phase of closed stoma. A suitable parameter with which to select data points is the stomatal conductance for H₂O (g_{H2O}) because this parameter gives information about the condition of the plant and indirect information about air temperature, radiation and water vapor deficit (VPD). Furthermore, it is known that the NO₂ exchange is strongly regulated by stomatal conductance (THOENE et al. 1991; GEBLER et al. 2000; TEKLEMARIAM and SPARKS 2006; CHAPARRO-SUAREZ 2008). Hence, the data were classified into seven classes of g_{H2O} . The interval between the several classes is based on a logarithmical scale.

4.1.3 Monitoring of plant-physiological processes due to chambers

Working with chambers and enclosed plants (parts of plants) necessitates control of the plant living conditions. The design and operation of the chambers needs to guarantee an undisturbed metabolism. For example, if the purging air flow is too low, it would disturb the gas exchange of the plant because CO_2 and H_2O concentrations would be reduced or accumulate inside the chamber. Plants will react with some regulation mechanisms on the modified conditions. Adequate experiments are needed to show that the plant is unaffected. This must be done for each plant and site. Simultaneous measurements of CO_2 surface exchange fluxes (assimilation), H_2O surface exchange fluxes (transpiration) and determination of stomatal conductance can provide an indication of the plant condition. For long-term field measurements further control experiments including unenclosed plants (part of the plants) would be advantageous to ensure the same behavior of control and enclosed plants even after long enclosure periods. For example, measurements of the photosynthetic capacity in response to temperature, radiation, CO_2 mixing ratio and relative humidity or analysis of the nutrient composition of enclosed and control plants could be tested.

We proved the photosynthetic capacity of the enclosed needles in comparison to control needles. Measurements of *in-situ* CO₂ and H₂O needle gas exchange in response to temperature, radiation, CO₂ mixing ratio and relative humidity were made using a portable gas exchange system (WALZ GFS3000, Walz, Effeltrich/Germany). From these light-response curves the light compensation point (I_c) and the light-saturated point (I_s) could be identified. The I_c marked the irradiance level where CO₂ uptake and respiratory CO₂ release were in equilibrium. By definition the I_s was reached at 90 % of maximal yield of photosynthesis. Additionally we have analyzed the nutrient composition (calcium, potassium, magnesium, manganese, phosphorus, sulfur, carbon and nitrogen) of control and enclosed needles according to validated analytical methods by the *Bayreuth Center of Ecology and Environmental Research* (BayCEER).

4.1.4 Set-up at the ECHO project

For the measurements within the ECHO project a comparable dynamic chamber system was used. The measurement site was located in an urban area (Jülich, Germany) in a deciduous forest stand and took place from 12 to 19 July 2003. For a detailed description of the field site and the measurement system see DINDORF et al. (2006).

During the measurements, a branch of the oak tree (*Quercus robur*) was enclosed in a dynamic plant chamber with a height of 60 cm and a diameter of 40 cm, equivalent to ~75 L. The total enclosed leaf area was 0.26 m^2 and the total dry-leaf weight 24.0 g. The sample chamber and an empty reference chamber of identical volume were continuously flushed with 35 L min⁻¹ of ambient air, which results in a residence time of 129 s. The chambers were installed close to each other at a height of 18 m. Air was sampled alternately from the reference and the plant chamber outlets for 90 s.

NO and NO₂ concentrations were measured with a chemiluminescence NO analyzer (Eco Physics CLD 780 TR). The $LOD(m_{NO})$ of the instrument amounts to 30 ppt for a 3 s integration time. O₃ concentrations were measured with an UV-absorption analyzer (Model 49C, Thermo Environment). CO₂ and H₂O concentrations were measured with an infrared gas analyzer (LI-6262, LiCor, Lincoln, NE, USA).

4.2 Results

EGER project

4.2.1 Microclimatic conditions

The ambient concentrations of NO, NO₂, O₃, CO₂ and H₂O and the relative humidity and air temperature were recorded in addition to the chamber measurements. Global radiation was detected on the top of the tower. A summary of the ambient measurements is given in Table 16. NO concentrations were mostly close to the detection limit. Sporadic NO peaks occurred due to advection of automobile exhaust gases from a busy country road (2000 cars/h) at a distance of about 1 - 2 km from the site. The NO₂ concentration varied between 0.4 and 21.5 ppb including concentration peaks from the road. A diurnal distribution was seen with higher NO₂ concentrations at night. O₃ mixing ratios ranged between 3 and 78 ppb. In the morning hours a gradual decline of O₃ concentration was observed. Weather conditions during measurements were characteristic of the region. Air temperature ranged from 4 to 28 °C with a mean temperature of 14 °C.

	average	range
NO, ppb	0.19 ± 0.17	0.07 - 2.89
NO ₂ , ppb	2.46 ± 1.42	0.42 - 21.49
O ₃ , ppb	47.12 ± 11.67	19.0 - 77.1
CO ₂ , ppm	380 ± 8	293 - 409
H ₂ O, ppth	13 ± 2.6	7 - 25
relative humidity, %	68.3 ± 17.4	32.3 - 99.9
temperature, °C	14.4 ± 4.5	3.8 - 27.7
global radiation, $W m^{-2}$	232 ± 276	0 - 1005

Table 16: Ambient conditions during time of field measurements. Given are mean values and minimum and maximum data over the measuring period (1 June to 15 July 2008).

4.2.2 Plant physiology

The photosynthetic capacity control measurements of enclosed and control needles as performed to detect effects caused by the chambers on the enclosed plant parts are presented in Figure 23 for one example tree. We distinguished between control and enclosed needles and between young and old needles. The control needles were outside the chambers during the whole field campaign. The photosynthesis rate was measured under different light conditions at ambient CO₂ concentrations (370 - 390 ppm). It is obvious that enclosed and control needles have the same photosynthesis rates. Young and old needles behaved similarly. I_c was in a range of 40 to 70 µmol photons m⁻² s⁻¹ and I_s between 500 and 1100 µmol photons m⁻² s⁻¹.

The results of the nutrient composition analysis (see Table 17) of the needles exhibited no obvious differences between control and enclosed needles for total carbon and total nitrate concentration as well for magnesium, manganese, phosphate and sulfur. Only for potassium and calcium were major differences recognizable between young control and enclosed needles. As the low concentration of potassium in the young needles does not go below the limit of potassium (PFLÜGER and MENGEL 1972; SIEGHARDT 1988; LARCHER 2003), these differences were not a sign of a harmful effect of the chamber. Furthermore, young plants have higher potassium concentrations than older plants because potassium is needed during leaf development. Potassium ions are responsible for the maintenance of the status of plasma swelling. Potassium deficit can be identified by fading of leaves and later leaf die off (LARCHER 2003). These symptoms were not observed.



Figure 23: Photosynthetic light response curves at ambient CO_2 concentration (370 - 390 ppm) of control and enclosed needles. (a) young control needles, (b) young enclosed needles, (c) older control needles, (d) older enclosed needles.

		Tre	ee 1		Tree 2					
	yo	oung	old		young		(old		
	control	enclosed	control	control enclosed		control enclosed		enclosed		
$mg g^{-1} dw$										
Ca	2.05	2.11	3.07	2.83	2.99	2.20	6.85	6.70		
K	6.91	8.66	4.65	5.06	4.58	5.87	2.61	2.53		
Mg	0.67	0.78	0.54	0.64	0.63	0.58	0.66	0.69		
Mn	0.17	0.17	0.28	0.24	0.12	0.09	0.27	0.26		
Р	1.40	1.61	1.15	1.15	1.21	1.34	0.88	0.89		
S	0.67	0.74	0.83	0.83	0.62	0.64	0.76	0.74		
%										
Ν	1.13	1.35	1.40	1.49	1.09	1.10	1.27	1.26		
C	47.48	47.63	48.75	48.87	47.94	48.72	49.51	49.53		

Table 17: Results of the nutrient content analysis of the needles.

4.2.3 Diurnal variations of gas exchange

The diurnal variation of exchange flux densities of CO₂, H₂O, NO, NO₂ and O₃ for Tree 1 for the period from 7 to 8 July are presented in Figure 25. At these two days maximal PAR reached 575 µmol photons m⁻² s⁻¹ and leaf temperature (T_{leaf}) was always below 18 °C. The O₃ exchange flux density ($F_{ex,O3}$) ranged between -0.02 and -0.61 nmol m⁻² s⁻¹. On both days a gradual increase of $F_{ex,O3}$ was observed, started rising at sunrise at 06:00 and reaching maximum values at 14:00. Then the $F_{ex,O3}$ declined to minimum values, as did the leaf conductance, before it began to increase again the next morning. Compared to O₃ the diurnal course of NO₂ exchange flux density ($F_{ex,NO2}$) was not so pronounced. Moreover, for NO₂ more $F_{ex,NO2}$ data had to be rejected for nonsignificance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$ than for O₃. The range of $F_{ex,NO2}$ was between -0.04 and 0.01 nmol m⁻² s⁻¹. The diurnal NO distribution was not identifiable. Most of the $F_{ex,NO}$ measurements had to be rejected as $\Delta m_{NO} = (m_{a,NO} - m_{s,NO})$ was not significant or the concentrations were under the $LOD(m_{NO})$. The positive values of $F_{ex,NO}$ occurred by corrections for the chemical gas phase reactions and not by higher NO concentrations inside the sample chamber compared to concentrations in ambient air.

Exchange flux densities of NO and NO_2 between 06:00 and 12:00 have to be interpreted with caution because in this time interval advection of automobile exhaust

gases from a country road cannot be excluded. As mentioned above this problem of advection is well known. It has been identified in simultaneously-performed profile measurements of in- and above canopy concentrations and eddy covariance flux measurements of NO-NO₂-O₃ (PLAKE et al. 2009).

Figure 26 presents the diurnal variations of the period from 29 to 30 June. During these two days PAR reached maximal values of 1800 μ mol photons m⁻² s⁻¹ with an average of 400 μ mol photons m⁻² s⁻¹ under daylight conditions. Compared to the two days presented before, these days were much sunnier and therefore T_{leaf} reached values above 20 °C around noon. The distribution of the leaf conductance (Figure 26c) is strongly connected to the leaf temperature. If T_{leaf} exceeds values of around 23 °C stomata began to close. This temperature effect has been documented through independent measurements of temperature dependence of photosynthesis on leaf level (see Figure 24, data provided by E. Falge, personal communication, 2009). To that effect the diurnal course of $F_{ex,O3}$ only reached maximum values of -0.51 nmol m⁻² s⁻¹ and began to decrease earlier. During such days diurnal distributions of $F_{ex,NO2}$ could hardly be seen, because when stomata closed around noon only very small NO₂ exchange fluxes could be detected. In most cases $F_{ex,NO2}$ data had to be rejected for non-significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. There was an additional problem with the measurements of NO for these two days: Most of the measured NO concentrations were below the detection limit of the NO analyzer or the concentration differences Δm_{NO} were not significant, therefore we can not calculate any exchange fluxes.



Figure 24: Temperature dependence of photosynthesis rate ($F_{ex,CO2}$). *In-situ* measurements of CO₂ gas exchange on needle level in response to temperature, using a portable gas exchange system (WALZ GFS3000, Walz, Effeltrich/Germany). Here photosynthesis rates were represented with positive sign. Data provided by E. Falge, (personal communication, 2009).



Figure 25: Exchange flux densities (F_{ex}) of CO₂ (panel (**a**), green line), H₂O (panel (**c**), blue line), NO, NO₂, O₃ (panel (**d-f**)) with diurnal courses of PAR (panel (**a**), orange line), leaf temperature (T_{leaf}) (panel (**b**)) and leaf conductance (g_{H2O}) (panel (**c**) black line) over the period from Jul 07 to Jul 08. The flux data are 16 min averages. Grey circles identify $F_{ex,i}$ data, which have to be rejected for non-significance of $\Delta m_i = (m_{a,i} - m_{s,i})$. Grey bars indicate time interval of possible advection of automobile exhaust gases from a country road.



Figure 26: Exchange flux densities (F_{ex}) of CO₂ (panel (**a**), green line), H₂O (panel (**c**), blue line), NO, NO₂, O₃ (panel (**d-f**)) with diurnal courses of PAR (panel (**a**), orange line), leaf temperature (T_{leaf}) (panel (**b**)) and leaf conductance (g_{H2O}) (panel (**c**) black line) over the period from Jun 29 to Jun 30. The flux data are 16 min averages. Grey circles identify $F_{ex,i}$ data, which have to be rejected for non-significance of $\Delta m_i = (m_{a,i} - m_{s,i})$. Grey bars indicate time interval of possible advection of automobile exhaust gases from a country road.

4.2.4 Overview of plant chamber measurements

The field measurements took place over a period of six weeks. Table 18 presents the results of both dynamic sample chambers (trace gas concentration measurements of NO, NO₂, O₃, photosynthesis, transpiration, exchange flux densities of NO, NO₂, O₃, leaf conductance, light and temperature conditions). The NO concentration ($m_{s,NO}$) inside the two sample chambers was on average 0.16 ppb at day and 0.1 ppb at night which approached limit of detection of the analyzer ($LOD(m_{NO}) = 0.1$ ppb = 4.46 nmol m⁻³). The NO₂ concentration $m_{s,NO2}$ were in contrast always above the limit of detection ($LOD(m_{NO2}) = 0.31$ ppb = 13.8 nmol m⁻³). At day and night the mean NO₂ values were around 2 ppb. Some high concentration peaks were observable, especially for NO₂, up to a maximum of 17 ppb. This temporary concentration rise resulted from the traffic road near by the site, frequently during the rush-hour traffic in the morning between 06:00 and 12:00. The O₃ concentrations $m_{s,O3}$ reached averages of 40 ppb. Both branches demonstrated similar photosynthesis, respiration and transpiration activities. Leaf conductances of H₂O (g_{H2O}) were also comparable. Consequently there was no evidence for different behavior of the two trees or enclosed parts of the plants.

The data selection of statistically-significant differences of trace gas concentrations at the inlet and the outlet of the dynamic chamber (see Sect. 3.1.5) resulted in different numbers for NO, NO₂ and O₃, which were used to calculate the exchange flux densities. Table 19 presents the percentage of significant concentration differences $\Delta m_i = (m_{a,i} - m_{s,i})$ measured at the inlet and outlet of the dynamic sample chamber 1 and 2. It becomes apparent that for O₃ most of the concentration differences were significant. Most statistically insignificant concentration differences were found for NO, especially at night. Overall, only one fourth of the NO data pairs passed the significance criterion. For NO₂ measurements the percentage of significant data pairs was 60 - 70 %.

The bi-variate weighted regression analysis for NO resulted in very small $R^2(m_{a,NO}, m_{s,NO})$ between 0.0173 and 0.9031. Moreover, the probabilities of significant NO compensation point concentrations $m_{comp,NO}$ and NO deposition velocities $v_{dep,NO}$ are generally unlikely. Continuative evaluations for $m_{comp,NO}$ and $v_{dep,NO}$ were not practical.

	sample cl	hamber 1	sample chamber 2			
	day ^a	night	day ^a	night		
m _{s,NO} , ppb	0.16 ± 0.12	0.10 ± 0.04	0.16 ± 0.13	$0.09\pm\!\!0.04$		
	(0.10* - 1.53)	(0.10* - 0.35)	(0.10* - 1.75)	(0.10* - 0.35)		
$F_{\text{ex,NO}}$, nmol m ⁻² s ⁻¹	-0.006 ± 0.015	0.009 ± 0.005	-0.005 ± 0.007	0.010 ± 0.004		
	(-0.110 - 0.044)	(0.002 - 0.019)	(-0.026 - 0.090)	(0.002 - 0.023)		
$m_{s,NO2}$, ppb	2.19 ±1.35	2.28 ± 1.31	2.13 ±1.27	2.30 ±0.91		
	(0.73 - 17.19)	(0.76 - 12.28)	(0.77 - 11.91)	(0.66 - 7.63)		
$F_{ex,NO2}$, nmol m ⁻² s ⁻¹	-0.011 ±0.015	-0.014 ± 0.025	-0.019 ± 0.020	-0.013 ±0.022		
	(-0.079 - 0.058)	(-0.414 - 0.085)	(-0.341 - 0.045)	(-0.205 - 0.155)		
$v_{dep,NO2}$, mm s ⁻¹	0.19 ± 0.11		0.24 ± 0.11			
	(0.07 - 0.35)		(0.14 - 0.42)			
<i>m</i> _{s,O3} , ppb	40.80 ± 11.88	37.41 ±8.23	40.16 ± 11.88	40.42 ± 10.80		
	(17.76 - 72.41)	(21.31 - 63.41)	(15.58 - 72.95)	(19.41 - 70.27)		
$F_{ex,O3}$, nmol m ⁻² s ⁻¹	-0.367 ± 0.174	-0.019 ± 0.316	-0.386 ±0.156	-0.180 ± 0.123		
	(-1.153 - 0.086)	(-0.889 - 0.293)	(-1.167 - 0.152)	(-1.141 - 0.255)		
$v_{dep,O3}$, mm s ⁻¹	0.22 ± 0.11	0.20 ± 0.09				
	(0.07 - 0.38)		(0.06 - 0.32)			
$F_{ex,CO2}$, µmol m ⁻² s ⁻¹	-0.57 ± 0.47	$0.09\pm\!\!0.07$	-0.59 ± 0.45	0.13 ±0.07		
	(-2.66 - 0.20)	(-0.05 - 0.34)	(-2.01 - 0.24)	(-0.77 - 0.52)		
$F_{ex,H20}$, mmol m ⁻² s ⁻¹	$0.07\pm\!\!0.06$	0.01 ± 0.01	$0.09\pm\!\!0.06$	0.01 ± 0.01		
	(0 - 0.39)	(0 - 0.03)	(0 - 0.28)	(0 - 0.03)		
\boldsymbol{g}_{H2O} , cm s ⁻¹	0.03 ± 0.04	0.01 ± 0.03	0.05 ± 0.06	0.01 ± 0.014		
	(0 - 0.54)	(0 - 0.07)	(0 - 0.83)	(0 - 0.17)		
$g_{NO2,p}$, cm s ⁻¹	0.020 ± 0.022	0.007 ± 0.006	0.031 ± 0.040	0.006 ± 0.005		
	(0 - 0.34)	(0 - 0.042)	(0 - 0.513)	(0 - 0.107)		
<i>T</i> _{leaf} , °C	17.9 ± 4.7	11.3 ±2.8	18.3 ±4.9	13.3 ±3.3		
	(6.5 - 38.7)	(6.3 – 16.7)	(6.3 - 33.1)	(6.3 - 22.4)		
rH_{out} , %	66.7 ± 17.5	85.4 ± 11.1	$66.0\pm\!\!17.8$	$79.0 \pm \! 14.2$		
	(32.3 - 99.9)	(62.5 - 99.9)	(32.6 - 99.9)	(40.3 - 99.9)		
PAR , μ mol m ⁻² s ⁻¹	231 ±273	-	255 ± 280	-		
	(0 - 1875)	-	(0 - 1848)	-		

 Table 18: Overview of chamber measurements. Given are mean data from 4 minute average values of day and night measurements.

^a daytime values were used when global radiation $> 5 \text{ W m}^{-2}$

* limit of detection (LOD)

	sa signifi (1	mple chamb icant Δ <i>m_i</i> % number of tot	er 1 of total tal)	sample chamber 2 significant Δm_i % of total (number of total)			
	all (2988)	day (1885)	night (1103)	all (2993)	day (1887)	night (1106)	
NO	24	33	7	24	33	8	
NO_2	57	62	48	67	69	63	
O ₃	96	98	93	98	99	97	

Table 19: Percentage of significant differences (Δm_i) of sample chamber 1 and 2. Only concentrations above LOD were considered.

Table 20: Definition of the classes, which were used for the classification of measured data. All displayed data are mean values. Leaf conductance (g_{H2O}) were calculated on basis of projected leaf area and total leaf surface area.

-	class		1	2	3	4	5	6	7
	g _{H2O} projected	cm s ⁻¹	0.01	0.025	0.06	0.08	0.1	0.13	0.16
_	A _{leaf}		0.025	0.06	0.08	0.1	0.13	0.16	1.0
	g н20 total	cm s ⁻¹	0.004	0.01	0.02	0.03	0.04	0.05	0.06
	A_{leaf}		0.01	0.02	0.03	0.04	0.05	0.06	0.4
-	PAR	μ mol m ⁻² s ⁻¹	130 ±261	200 ± 334	253 ±311	$279\pm\!\!300$	297 ±312	355 ± 335	319 ± 365
	T _{air}	°C	18.8 ±4.9	16.8 ±4.9	16.5 ±4.2	15.7 ±3.7	14.3 ±3.8	13.9 ±3.6	12.0 ± 3.4
chamber 1	r.H.	%	54 ±17	64 ± 18	64 ±16	67 ± 14	69 ± 14	70 ± 13	80 ± 14
	F _{ex,CO2}	μ mol m ⁻² s ⁻¹	-0.15 ±0.12	-0.37 ±0.22	-0.62 ±0.26	-0.74 ±0.31	-0.86 ±0.37	-1.02 ±0.42	-1.05 ±0.46
	F _{ex,H20}	mmol m ⁻² s ⁻¹	0.03 ±0.02	0.05 ±0.04	$\begin{array}{c} 0.08 \\ \pm 0.05 \end{array}$	0.09 ±0.05	0.10 ±0.06	0.11 ±0.07	0.09 ±0.08
-	PAR	$\mu mol m^{-2} s^{-1}$	51 ±158	157 ±251	279 ±353	336 ±387	278 ±290	320 ± 307	322 ±329
	T _{air}	°C	16.9 ±4.7	17.4 ±5.1	17.4 ±4.7	16.8 ±4.2	15.8 ±3.9	14.6 ±3.7	12.6 ±3.5
chamber 2	r.H.	%	63 ±19	61 ±19	59 ±17	61 ±16	66 ± 14	69 ± 14	77 ± 16
	F _{ex,CO2}	μ mol m ⁻² s ⁻¹	-0.03 ±0.11	-0.25 ±0.22	-0.53 ±0.26	-0.67 ±0.31	-0.77 ±0.31	-0.88 ±0.36	-0.98 ±0.42
	F _{ex,H20}	mmol m ⁻² s ⁻¹	$\begin{array}{c} 0.02 \\ \pm 0.02 \end{array}$	0.06 ±0.05	0.10 ±0.06	0.11 ±0.07	0.11 ±0.06	0.11 ±0.06	0.09 ±0.06

Before calculating exchange flux densities, compensation point concentrations and deposition velocities, all data were not only controlled for significant concentration differences but also classified (see Sect. 4.1.2). Table 20 displays the ambient and plant conditions of the single classes for each chamber. Depending on the placement of chamber installation the ambient conditions might differ between the two chambers, especially for PAR. Using leaf conductance of 0.01 cm s⁻¹ (0.004 cm s⁻¹ per total A_{leaf}) as lower limit excluded situations of condensation inside the chamber from further considerations.

4.2.5 NO₂ exchange flux density

The exchange flux densities of NO₂ ($F_{ex,NO2}$) were calculated according to Eq. (8.4) and their standard errors according to Eq. (19). NO₂ deposition velocities ($v_{dep,NO2}$) and NO₂ compensation point concentrations ($m_{comp,NO2}$) were determined using the bi-variate regression analysis (see Sect. 2.1.2). The results of the single classes 1 -7 are displayed in Figure 27 to Figure 30 for data from sample chamber 1 and in Figure 31 to Figure 34 for data from sample chamber 2 (refer to the schematic representation in Figure 6). Panels 1a -7a represent the results of bi-variate weighted linear regression analysis between NO₂ concentrations at the plant chamber's inlet $(m_{a,NO2})$ and outlet $(m_{s,NO2})$, while at panels 1b - 7b show these data for $F_{ex,NO2}$ versus $m_{s,NO2}$. The blue line represents $v_{dep,NO2}$ and the red filled circle marks $m_{comp,NO2}$. The blue circles identify $F_{ex,NO2}$ data which were significant for $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. The bi-variate regression analysis (see Sect. 3.1.6) of the data from sample chamber 1 resulted in $R^{2}(m_{aNO2}, m_{s,NO2})$ between 0.8709 and 0.9401 and $v_{dep,NO2}$ between 0.07 and 0.35 mm s⁻¹. The $v_{dep,NO2}$ increased with the number of the classes, that is, as the class number and g_{H2O} gets higher the deposition velocity increases. The determined $m_{comp,NO2}$ ranged between 2.4 ± 9.63 and 29.0 ± 16.30 nmol m⁻³ (0.05 - 0.65 ppb). The significance probabilities of $m_{comp,NO2} \neq 0$ vary from 96.90 % ("likely") to 99.99 % ("highly significant"). The results of sample chamber 2 were mostly comparable. $R^2(m_{a,O3}, m_{s,O3})$ reached values between 0.8106 and 0.9702. $v_{dep,NO2}$ reached values mainly in the same range but the determination of $m_{comp,NO2}$ became complicated at this point. For classes 2, 3 and 6, $m_{comp,NO2}$ results in negative values with a probability of 99.99 % ("highly significant"). However a negative NO₂ compensation point concentration is physically meaningless.





(1a), (2a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (1b), (2b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.





(3a), (4a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (3b), (4b) NO2 exchange flux density (Fex,NO2) vs. NO2 concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.

102 | FIELD MEASUREMENTS





(5a), (6a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (5b), (6b) NO₂ exchange flux density (Fex.NO2) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.



Figure 30: NO₂ measurements, data class 7 of sample chamber 1

(7a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (7b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.





4.0

2.0

qdd 3.0



 $\mathbf{m}_{a,NO2}$, ppb

3

2

0

200

150

100

(1a)

(1a), (2a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (1b), (2b) NO2 exchange flux density (Fex,NO2) vs. NO2 concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.





(3a), (4a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber $(m_{a,NO2})$. Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (3b), (4b) NO2 exchange flux density (Fex.NO2) vs. NO2 concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.





(5a), (6a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (5b), (6b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.



Figure 34: NO₂ measurements, data class 7 of sample chamber 2

(7a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the inlet of the dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (7b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$). NO₂ compensation point concentration $m_{comp,NO2}$ is represented by red filled circle.

108 | FIELD MEASUREMENTS

Details of statistical evaluation for NO_2 and O_3 data of sample camber 1 are listed in Table 21 and for data of sample chamber 2 in Table 22.

Table 21: Parameters of bi-variate weighted linear least-squares fitting regression analysis (see Sect. 3.1.6) for data classes 1 -7 of sample chamber 1. Only significant data of $\Delta m_i = (m_{a,i} - m_{s,i})$ were applied.

statistical quantity	unit		NO ₂	O ₃		NO ₂	O ₃
N	[1]		91	278		55	226
$R^2(m_{a,i};m_{s,i})$	[1]		0.8939	0.9963		0.9248	0.9872
n	nmol m ⁻³	1	6.5 ± 1.75		S	8.6 ± 2.09	
т	[1]	ass	0.91 ± 0.025	0.95 ± 0.002	ass	0.79 ± 0.021	$0.82\pm\!\!0.003$
m _{comp,i}	nmol m ⁻³	<u>ت</u>	14.0 ± 33.38		C	23.1 ±14.43	
$m_{comp,i} \neq 0$?	%		99.98 (HS)			99.99 (HS)	
$V_{dep,i}$	mm s ⁻¹		0.07 ± 0.059	0.07 ± 0.004		0.25 ±0.045	0.27 ± 0.0008
N	[1]		102	377		35	185
$R^2(m_{a,i};m_{s,i})$	[1]		0.8886	0.9932	class 6	0.9263	0.9910
n	nmol m ⁻³	7	6.6 ± 1.89			11.4 ± 2.81	
m	[1]	lass	0.89 ± 0.021	0.91 ± 0.003		0.74 ± 0.034	0.79 ± 0.004
m _{comp,i}	nmol m ⁻³	ు	22.7 ± 30.49			29.0 ± 16.30	
$m_{comp,i} \neq 0$?	%		99.99 (HS)			99.99 (HS)	
$V_{dep,i}$	mm s ⁻¹		0.09 ± 0.062	0.12 ± 0.004		0.30 ± 0.079	0.32 ±0.017
N	[1]		47	211		75	306
$R^2(m_{a,i};m_{s,i})$	[1]		0.8709	0.9926		0.8861	0.9811
n	nmol m ⁻³	3	7.1 ±3.19		٢	6.3 ± 1.99	
т	[1]	lass	$0.86\pm\!\!0.041$	0.87 ± 0.004	lass	0.74 ± 0.025	
$m_{comp,i}$	nmol m ⁻³	5	13.9 ±36.65		5	2.4 ±9.63	
$m_{comp,i} \neq 0?$	%		98.73 (S)			96.90 (L)	
$V_{dep,i}$	mm s ⁻¹		0.13 ±0.071	0.18 ±0.009		0.35 ± 0.034	0.38 ±0.014
N	[1]		52	210			
$R^2(m_{a,i};m_{s,i})$	[1]		0.9401	0.9932			
n	nmol m ⁻³	4	4.0 ± 2.36				
т	[1]	ass	0.87 ± 0.026	0.85 ± 0.003			
<i>m</i> _{comp,i}	nmol m ⁻³	cl	-24.3 ±35.61				
$m_{comp,i} \neq 0?$	%		99.99 (HS)				
$\mathcal{V}_{dep,i}$	mm s ⁻¹		0.11 ± 0.076	0.22 ± 0.009			

* assumption for O₃: $m_{comp,O3} = 0$ (n₃ = 0).

Table 22: Parameters of bi-variate weighted linear least-squares fitting regression analysis (see Sect. 3.1.6) for data classes 1 -7 of sample chamber 2. Only significant data of $\Delta m_{i=}(m_{a,i}-m_{s,i})$ were applied.

statistical quantity	unit		NO ₂	O ₃		NO ₂	O ₃
N	[1]		43	152		74	274
$R^2(m_{a,i};m_{s,i})$	[1]		0.9702	0.9938		0.8393	0.9876
n	nmol m ⁻³	1	-4.7 ±3.24		S	7.5 ± 2.56	
т	[1]	ass	1.00 ± 0.031	0.95 ± 0.004	lass	0.78 ± 0.030	0.82 ± 0.004
m _{comp,i}	nmol m ⁻³	َنَ ت	6860 ± 12428		́с)	14.5 ± 13.20	
$m_{comp,i} \neq 0$?	%		28.08 (UL)			99.99 (HS)	
$V_{dep,i}$	mm s ⁻¹		-0.002 ± 0.035	0.06 ± 0.004		0.29 ± 0.061	0.26 ± 0.011
N	[1]		102	443		43	195
$R^2(m_{a,i};m_{s,i})$	[1]		0.9075	0.9850		0.8912	0.9885
n	nmol m ⁻³	7	3.7 ± 1.40		9	1.4 ±2.76	
т	[1]	lass	0.89 ± 0.014	0.91 ± 0.003	lass	0.85 ± 0.033	0.80 ± 0.005
m _{comp,i}	nmol m ⁻³	َنَ ت	-16.7 ±13.94		5	-34.0 ± 22.60	
$m_{comp,i} \neq 0$?	%		99.99 (HS)			99.99 (HS)	
$v_{dep,i}$	mm s ⁻¹		0.14 ± 0.037	0.11 ±0.004		0.19 ± 0.050	0.28 ± 0.012
Ν	[1]		87	283		140	455
$R^2(m_{a,i};m_{s,i})$	[1]		0.8783	0.9913		0.8106	0.9538
n	nmol m ⁻³	e	4.1 ±1.99		۲	8.1 ±1.73	
т	[1]	lass	0.88 ± 0.024	0.87 ± 0.004	lass	0.71 ± 0.021	0.77 ± 0.005
m _{comp,i}	nmol m ⁻³	5	-13.6 ±19.45		5	7.4 ± 6.40	
$m_{comp,i} \neq 0$?	%		99.99 (HS)			99.99 (HS)	
$v_{dep,i}$	mm s^{-1}		0.14 ±0.015	0.17 ± 0.007		0.42 ± 0.067	0.32 ± 0.008
Ν	[1]		59	208			
$R^2(m_{a,i};m_{s,i})$	[1]		0.8545	0.9928			
n	nmol m ⁻³	4	8.5 ±2.61				
т	[1]	ass	0.82 ± 0.027	0.84 ± 0.004			
m _{comp,i}	nmol m ⁻³	င	16.5 ±15.25				
$m_{comp,i} \neq 0?$	%		99.99 (HS)				
$V_{dep,i}$	mm s ⁻¹		0.25 ± 0.047	0.22 ± 0.009			

* assumption for O₃: $m_{comp,O3} = 0$ (n₃ = 0).

RESULTS

On closer consideration of the variation of $F_{ex,NO2}$ referring to the leaf conductance classes it became apparent that $F_{ex,NO2}$ increased with raising classes of g_{H2O} . In Figure 35 the median of $F_{ex,NO2}$ for each class is presented as well as the interquartile ranges and the minimum and maximum of $F_{ex,NO2}$ for data measured at sample chamber 1. The data of sample chamber 2 are displayed in Figure 36. Both enclosed branches showed the same trend. At very low leaf conductance (g_{H2O}) the median of $F_{ex,NO2}$ reached values of -0.002 nmol m⁻² s⁻¹ (sample chamber 1) and -0.015 nmol m⁻² s⁻¹ (sample chamber 2) respectively. With increasing leaf conductance $F_{ex,NO2}$ rose up almost linear to 0.020 and 0.024 nmol m⁻² s⁻¹ respectively. The connection between NO₂ exchange rates and stomatal conductance can be identified.



Figure 35: Variation of NO₂ exchange flux density ($F_{ex,NO2}$) of sample chamber 1 for different leaf conductance classes (increasing values of leaf conductance from class 1 to 7). Blue lines denote the median, the boxes the interquartile ranges (0.25 – 0.75), the black lines the minimum and maximum.



Figure 36: Variation of NO₂ exchange flux density ($F_{ex,NO2}$) of sample chamber 2 for different leaf conductance classes (increasing values of leaf conductance from class 1 to 7). Blue lines denote the median, the boxes the interquartile ranges (0.25 – 0.75), the black lines the minimum and maximum.

4.2.6 O₃ exchange flux density

The analysis of the O₃ data was done in the same way as the analysis of the NO₂ data (see Sects. 2.1.2 and 2.1.3). The results for the evaluation of O₃ exchange flux densities ($F_{ex,O3}$) and O₃ deposition velocities ($v_{dep,O3}$) of the classes 1 -7 are presented in Figure 37 to Figure 40 for sample chamber 1 and in Figure 41 to Figure 44 for sample chamber 2. Panels 1a - 7a represent the results of bi-variate weighted linear regression analysis between O₃ concentrations at the plant chamber's inlet ($m_{a,O3}$) and outlet ($m_{s,O3}$), while in panels 1b - 7b those of $F_{ex,O3}$ versus $m_{s,O3}$ are shown. The orange line represents $v_{dep,O3}$. The red circles identify $F_{ex,O3}$ data which were significant for $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. For the analysis of O₃ data an O₃ compensation point concentration $m_{comp,O3} = 0$ ($n_3 = 0$) was assumed.

The bi-variate regression analysis for the data from sample chamber 1 resulted in $R^2(m_{a,O3},m_{s,O3})$ between 0.9811 and 0.9963 and $v_{dep,O3}$ ranged between 0.07 ±0.059 and 0.38 ±0.014 mm s⁻¹. Values of $v_{dep,O3}$ increased with raising class of leaf conductance g_{H2O} . The results of sample chambers 1 were comparable with sample chamber 2. Analysis of O₃ data from sample chamber 2 resulted in $R^2(m_{a,O3},m_{s,O3})$ between 0.9538 and 0.9938. Deposition velocity of O₃ for the seven classes ranged between 0.06 ±0.004 and 0.32 ±0.008 mm s⁻¹. In Table 21 and Table 22 the details of the statistical analysis of the O₃ data are listed.

In Figure 45 and Figure 46 the median, the interquartile ranges and the minimum and maximum of $F_{ex,O3}$ for each class of g_{H2O} are displayed. It is obviously that values of $F_{ex,O3}$ increased when g_{H2O} became greater. Therefore, O₃ deposition increased with raising leaf conductance. The increasing of $F_{ex,O3}$ were more pronounced than the observed increasing of $F_{ex,NO2}$. The median of $F_{ex,O3}$ ranged from -0.17 to -0.52 nmol m⁻² s⁻¹ for sample chamber 1 and from -0.16 to -0.49 nmol m⁻² s⁻¹ for sample chamber 2. The rate of increase between the single classes declined with raising class: at high leaf conductances a trend of saturation for the O₃ uptake can be expected. The O₃ exchange rates seem to depend on stomatal conductance like NO₂ to a certain degree. Compared to NO₂ deposition of O₃ were higher at low g_{H2O} , this may have been caused by O₃ deposition to leaf, petiole and bark surfaces of the enclosed branches (due to high reactivity of ozone).



Figure 37: O₃ measurements, data class 1 and 2 of sample chamber 1

(1a), (2a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (1b), (2b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.





(3a), (4a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (3b), (4b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.





(5a), (6a) O_3 concentration measured at the outlet of the dynamic plant chamber ($m_{s,O3}$) vs. O_3 concentration measured at the inlet of the dynamic plant chamber ($m_{a,O3}$). Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (5b), (6b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber ($m_{s,O3}$).

RESULTS



Figure 40: O₃ measurements, data class 7 of sample chamber 1

(7a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (7b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.





(1a), (2a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (1b), (2b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.



Figure 42: O₃ measurements, data class 3 and 4 of sample chamber 2

(3a), (4a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (3b), (4b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.


Figure 43: O₃ measurements, data class 5 and 6 of sample chamber 2

(5a), (6a) O_3 concentration measured at the outlet of the dynamic plant chamber ($m_{s,O3}$) vs. O_3 concentration measured at the inlet of the dynamic plant chamber ($m_{a,O3}$). Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (5b), (6b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber ($m_{s,O3}$).



Figure 44: O₃ measurements, data class 7 of sample chamber 2

(7a) O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$ vs. O_3 concentration measured at the inlet of the dynamic plant chamber $(m_{a,O3})$. Orange circles identify data pairs for significance of $\Delta m_{O3} = (m_{a,O3} - m_{s,O3})$. Orange line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (7b) O_3 exchange flux density ($F_{ex,O3}$) vs. O_3 concentration measured at the outlet of the dynamic plant chamber $(m_{s,O3})$.



Figure 45: Variation of O_3 exchange flux density ($F_{ex,O3}$) of sample chamber 1 for different leaf conductance classes (increasing values of leaf conductance from class 1 to 7). Red lines denote the median, the boxes the interquartile ranges (0.25 - 0.75), the black lines the minimum and maximum.



Figure 46: Variation of O_3 exchange flux density ($F_{ex,O3}$) of sample chamber 2 for different leaf conductance classes (increasing values of leaf conductance from class 1 to 7). Red lines denote the median, the boxes the interquartile ranges (0.25 - 0.75), the black lines the minimum and maximum.

ECHO project

The analysis of the data measured at the ECHO project was performed with the same calculations and criterions as the analysis of the data measured at the EGER project. Here, only exchange flux densities, deposition velocities and compensation point concentrations of NO₂ were determined. The data were also controlled for the significance criterion for $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$ and classified into the same seven classes of g_{H2O} (see Sect. 4.1.2). Conditions of the classes are displayed in Table 23.

Table 23: Conditions of the classes which were used for the classification of the measured data within the ECHO project. All displayed data are mean values.

class		1	2	3	4	5	6	7
g H20	cm s ⁻¹	0.01 - 0.025	0.025 - 0.06	0.06 - 0.08	0.08 - 0.1	0.1	0.13 - 0.16	0.16 - 1.0
PAR	μ mol m ⁻² s ⁻¹	48	110	236	449	397	553	398
T _{air}	°C	17.5 ±2.0	18.5 ±2.3	19.8 ±2.1	19.9 ± 2.8	19.0 ±3.4	18.2 ±3.0	17.6 ±2.4
F _{ex,CO2}	μ mol m ⁻² s ⁻¹	-0.06 ±0.78	-1.30 ±1.09	-2.62 ±1.18	-3.71 ±1.07	-3.81 ±1.45	-4.49 ±1.52	-4.80 ±1.48
F _{ex,H20}	mmol m ⁻² s ⁻¹	$\begin{array}{c} 0.03 \\ \pm 0.02 \end{array}$	0.09 ±0.05	0.18 ±0.08	0.28 ±0.15	0.30 ±0.14	0.37 ±0.20	0.42 ±0.18

4.2.7 NO₂ exchange flux densities

During the ECHO measurements 636 data pairs of $m_{a,NO2}$ and $m_{s,NO2}$ have been obtained under daytime conditions. After applying the significance criterion nearly 52 % of the NO₂ data pairs remained. An additional screening for singular concentration peaks was not done.

The results of the bi-variate weighted linear regression analysis for the classes are presented in Figure 47 to Figure 50. In panels 1a - 7a the bi-variate regression analysis between NO₂ concentrations at the plant chamber's inlet ($m_{a,NO2}$) and outlet ($m_{s,NO2}$) are shown, where $m_{a,NO2}$ is represented by the concentration measured at the outlet of the empty reference chamber. Panels 1b - 7b represent $F_{ex,NO2}$ versus $m_{s,NO2}$. The blue line represents $v_{dep,NO2}$.

In Table 24 the details of statistical evaluation are listed. The analysis resulted in $R^2(m_{aNO2}, m_{s,NO2})$ between 0.8970 and 0.9951 and $v_{dep,NO2}$ between 0.60 and 2.71 mm s⁻¹. In these results $v_{dep,NO2}$ also increased with raising leaf conductance g_{H2O} . The determined $m_{comp,NO2}$ always gave negative values. Even if the significance probability of $m_{comp,NO2} \neq 0$ is varied from 99.51 % ("significant") to 99.99 % ("highly significant") a negative NO₂ compensation point concentration is unrealistic and physically meaningless, but nevertheless indicates the absence of a compensation point.

statistical quantity	unit	NO ₂		NO ₂		
N	[1]		21		25	
$R^2(m_{a,i};m_{s,i})$	[1]		0.9574	lass 5	0.9951	
n	nmol m ⁻³	1	-6.9 ±2.54		-4.0 ±1.39	
m	[1]	lass	0.78 ± 0.007		0.54 ± 0.007	
$m_{comp,i}$	nmol m ⁻³	C	-198.0 ±42.22	0	-85.8 ±13.42	
$m_{comp,i} \neq 0$?	%		99.99 (HS)		99.99 (HS)	
$v_{dep,i}$	$mm s^{-1}$		0.60 ± 0.026		1.81 ±0.047	
N	[1]		38		15	
$R^2(m_{a,i};m_{s,i})$	[1]		0.9434	class 6	0.9857	
n	nmol m ⁻³	lass 2	-0.4 ±2.46		-2.9 ±2.23	
m	[1]		0.70 ± 0.012		0.48 ± 0.014	
$m_{comp,i}$	nmol m ⁻³	S	-107.3 ±27.47		-39.4 ±8.31	
$m_{comp,i} \neq 0?$	%		99.99 (HS)		99.99 (HS)	
$v_{dep,i}$	$mm s^{-1}$		0.88 ±0.049		2.18 ±0.126	
N	[1]		35		12	
$R^2(m_{a,i};m_{s,i})$	[1]		0.8970	class 7	0.9803	
n	nmol m ⁻³	3	-0.9 ±2.92		6.0 ±2.13	
m	[1]	lass	0.61 ± 0.012		0.43 ± 0.026	
$m_{comp,i}$	nmol m ⁻³	c	-50.9 ± 10.28		-5.3 ± 5.19	
$m_{comp,i} \neq 0?$	%		99.99 (HS)		99.51 (S)	
$v_{dep,i}$	mm s ⁻¹		1.37 ± 0.068		2.71 ±0.285	
N	[1]		33			
$R^2(m_{a,i};m_{s,i})$	[1]		0.9812			
n	nmol m ⁻³	4	-4.9 ±2.27			
m	[1]	ass	0.53 ± 0.009			
$m_{comp,i}$	nmol m ⁻³	င	-89.8 ±22.86			
$m_{comp,i} \neq 0?$	%		99.99 (HS)			
$V_{dep,i}$	$mm s^{-1}$		1.89 ±0.065			

Table 24: Parameters of bi-variate weighted linear least-squares fitting regression analysis for data classes 1 -7. Only significant data of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$ were applied.





(1a), (2a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the outlet of the reference dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (1b), (2b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$).



Figure 48: NO₂ ECHO field measurements, data class 3 and 4

(3a), (4a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the outlet of the reference dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (3b), (4b) NO₂ exchange flux density (F_{ex NO2}) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber $(m_{s,NO2})$.





(5a), (6a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the outlet of the reference dynamic plant chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (5b), (6b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$).



Figure 50: NO₂ ECHO field measurements, data class 7

(7a) NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant reference chamber ($m_{a,NO2}$). Blue circles identify data pairs for significance of $\Delta m_{NO2} = (m_{a,NO2} - m_{s,NO2})$. Blue line is calculated according to bi-variate weighted linear least-squares fitting regression analysis. (7b) NO₂ exchange flux density ($F_{ex,NO2}$) vs. NO₂ concentration measured at the outlet of the dynamic plant chamber ($m_{s,NO2}$).

The development of the NO₂ exchange flux densities $F_{ex,NO2}$ for the different classes of leaf conductance g_{H2O} are displayed in Figure 51. $F_{ex,NO2}$ increased with rising g_{H2O} and the median reached a maximum value of -0.49 nmol m⁻² s⁻¹ at class 5 $(g_{H2O} = 0.10 - 0.13)$. At class 6 and 7 $F_{ex,NO2}$ declines and become lower than the values of class 1.



Figure 51: Variation of NO₂ exchange flux density ($F_{ex,NO2}$) of measurements within ECHO for different leaf conductance classes (increasing values of leaf conductance from class 1 to 7). Blue lines denote the median, the boxes the interquartile ranges (0.25 – 0.75), the black lines the minimum and maximum.

4.3 Discussion

4.3.1 Effects on enclosed plants

Application of a chamber system with enclosed plants or parts of plants requires a control of plant condition to be certain observations and data are transferable and not created under unnatural conditions. To achieve the dynamic chamber requirement that plant physiological processes are not affected by the chamber the chamber walls were made of transparent and almost chemically inert material. The reduction of photosynthetic active radiation (PAR) by the chamber (~10 %) was not a crucial factor because the correlation between CO₂ exchange and PAR intensity indicated that the system is light saturated around 500 µmol photons m⁻² s⁻¹ (PAR). The PAR transmittance and the high purging air flow rate afford that the physiological processes of the enclosed plant parts are working under natural conditions.

It is important to make sure that the plant is not affected by the chamber, especially for long-term studies. Consequently, we controlled the status of the plants after field experiments. We could not identify visual differences between enclosed and not enclosed plant material. Moreover, no variations in physiological performance were detectable. The photosynthetic capacities of enclosed and control needles were similar. The minor differences were still within a normal spread compared to literature values. The average maximum values for CO₂ uptake of coniferous evergreen trees ranged from 7 to $12 \mu mol m^{-2} s^{-1}$ (LARCHER 2003). In literature light compensation points were specified as 30 to 40 μ mol photons m⁻² s⁻¹ and light saturation points are denoted as 800 to 1000 μ mol photons m⁻² s⁻¹ for sun shoots of coniferous trees under conditions of ambient CO₂ and optimal temperature. For shade shoots of coniferous trees light saturation points were declared between 150 and 200 µmol photons m⁻² s⁻¹ and light compensation points between 2 and 10 μ mol photons m⁻² s⁻¹ (LARCHER 2003). Our shoots are better classified rather as sun shoots, although because of their position at the bottom of the crown they conform also in some aspects to shade shoots. The value of our calculated light compensation point was between 40 to 70 $\mu mol \ photons \ m^{-2} \ s^{-1}$ and our calculated light saturation point was between 500 and 1100 µmol photons m⁻² s⁻¹. Our values were higher than literature values but the differences were small between our enclosed and control needles. Hence, these differences were not a sign of a harmful effect of the chamber.

Furthermore, nutrient compositions of needles did not differ. Only for potassium and calcium were differences noticeable. The higher concentration of potassium was found for the young enclosed needles but the concentration was in a normal range, which is specified in literature to be between 5 and 70 mg g⁻¹dw (FREY 1998). Potassium is needed during leaf development and it is responsible for the maintenance of the status of plasma swelling. A potassium deficit can be identified by tips of needles drying out and by premature shedding of needles (LARCHER 2003). Such symptoms were not observed. Usual calcium content in plants was 0.4 to 13 mg g⁻¹dw. Symptoms of deficiency would be drying of buds, young shoots dying off and chlorosis of the tips of fir trees followed by browning of needles (LARCHER 2003). It can be assumed that the difference between enclosed and controlled needles was not harmful.

Our data sets give good reasons to assume that the enclosed branch behaved normally. Contrasting, in many chamber studies plant conditions were monitored only by measuring CO₂ and H₂O exchange of the plant and calculating leaf conductance (e.g., THOENE et al. 1996; SPARKS et al. 2001; GEBLER et al. 2002). These measurements allowed the actual plant conditions to be inferred but in contrast to our work no comparison of plants inside and outside the chamber was performed.

4.3.2 NO₂ exchange to leaves

With increasing ambient NO_2 concentrations an increase of NO_2 exchange flux densities could be observed. This agrees with previous studies (RONDÓN et al. 1993; THOENE et al. 1991; WEBER and RENNENBERG 1996; GEBLER et al. 2002) and with the assumption that NO_2 exchange is driven by the NO_2 concentration difference between atmosphere and the gaseous phase of the leaf interior.

Emission fluxes of NO₂ have been measured by several studies. TEKLEMARIAM and SPARKS (2006) reported emissions from four species (wheat, corn, sunflower, Madagascar periwinkle) between 36.8 and 101.0 pmol m⁻² s⁻¹. SPARKS et al. (2001) observed NO₂ emissions up to 50 pmol m⁻² s⁻¹ from several tropical trees and HEREID and MONSON (2001) from field-grown corn. NO₂ emissions from spruce needles were reported by RONDÓN at al. (1993) and GEBLER et al. (2002). In the present study the leaf emission of NO₂ from spruces varied between 0.07 and 58 pmol m⁻² s⁻¹ measured at averaged ambient NO₂ concentration of 67.3 nmol m⁻³ (1.5 ppb). For oak no emission fluxes have been observed for ambient NO₂ concentrations as low as 0.5 ppb, which is in close accordance with CHAPARRO-SUAREZ et al. (2011) who found no emission from four deciduous trees and a pine species as well.

Our measured deposition fluxes of NO₂ for spruce were on average two or three times higher than the detected emission fluxes. The NO₂ deposition fluxes varied between -0.078 and -0.018 nmol m⁻² s⁻¹. NO₂ deposition fluxes reported by THOENE et al. (1996) were in a range between -1.88 and -0.03 nmol m⁻² s⁻¹ and SPARKS et al. (2001) detected uptake rates on average between -1.55 and -0.15 nmol m⁻² s⁻¹ for several tropical trees. Thus, NO₂ fluxes of these studies were clearly higher than our measured NO₂ fluxes. For both studies analyzers with NO₂ converters with known interferences due to other oxidized nitrogen compounds were used (see Sect. 2.2). The comparison to the results of these studies could indicate that our performed measuring system, using a blue light converter, is more specific for NO₂ measurements. Similar deposition fluxes up to -0.3 nmol m⁻² s⁻¹ (at 5 ppb) were reported by CHAPARRO-SUAREZ et al. (2011) using a very similar instrumentation. However, GEBLER et al. (2002) using a photolytic converter reported NO₂ deposition fluxes at a range between -0.12 and -0.02 nmol m⁻² s⁻¹, which are obviously like our measured ranges.

Exchange rates of NO₂ depend not only on atmospheric NO₂ concentration but also on stomatal conductance (NEUBERT et al. 1993; HEREID and MONSON 2001; SPARKS et al. 2001; CHAPARRO-SUAREZ et al. 2011). SPARKS et al. (2001) observed for tropical tree species a kind of saturation for NO₂ uptake at higher stomatal conductance. Such observations have not been found in the present study for spruce. However, for oak a decline of the NO₂ exchange flux density, though not a saturation, was recognizable at higher leaf conductance. The NO₂ uptake may be additionally limited by internal resistances (mesophyllic), as reported by several authors (THOENE et al. 1991, 1996; SPARKS et al. 2001). This circumstance is reflected in a smaller measured leaf conductance to NO₂ ($g_{NO2,m}$) than the predicted NO₂ leaf conductance ($g_{NO2,p}$), which is calculated from leaf conductance to water vapor (see Eq. (28)). The measured leaf conductance $g_{NO2,m}$ was calculated from quotient of $F_{ex,NO2}$ and the NO₂ concentration at the outlet of the sample chamber. For oak we found higher measured NO₂ leaf conductance than the predicted conductance. Similar results were found by GEBLER et al. (2002) for spruce. They assumed that the reaction of ascorbate with NO₂ is responsible for maintaining high fluxes of NO₂ into the leaf and preventing high internal resistances. HANSON et al. (1993) found for broadleaf and conifer species an underestimation of measured leaf conductance by the predicted conductance of between 15 and 30 % as well. Our results for spruce looked different. The measured and predicted NO₂ leaf conductances are listed in Table 25. At higher leaf conductance classes the measured NO₂ conductance was lower than the predicted, however, at the low leaf conductance classes the results were reversed. A common trend for spruce and oak is the decrease in the different. Only for spruce predicted NO₂ leaf conductances exceed measured leaf conductances with higher class.

Table 25: Measured and predicted leaf conductance to NO₂ deposition. Measured leaf conductance $(g_{NO2,m})$ was calculated from quotient of $F_{ex,NO2}$ and NO₂ concentration at the outlet of the sample chamber, predicted leaf conductance $(g_{NO2,p})$ was calculated from g_{H2O} according to Eq. (28).

	Spruce			Spruce			Oak		
	chamber 1				chamber	2			
class	$g_{NO2,m}$ mm s ⁻¹	$g_{NO2,p}$ mm s ⁻¹	difference %	$g_{NO2,m}$ mm s ⁻¹	$g_{NO2,p}$ mm s ⁻¹	difference %	$g_{NO2,m}$ mm s ⁻¹	$g_{NO2,p}$ mm s ⁻¹	difference %
1	0.054	0.038	+41.5	0.128	0.062	+108.5	0.746	0.109	+582.2
2	0.053	0.096	-45.3	0.150	0.096	+55.5	0.881	0.273	+222.2
3	0.102	0.156	-34.4	0.163	0.157	+3.8	1.053	0.429	+145.4
4	0.136	0.204	-33.3	0.178	0.204	-12.8	1.257	0.558	+25.4
5	0.143	0.256	-44.0	0.206	0.256	-19.3	1.335	0.694	+92.3
6	0.144	0.326	-55.9	0.250	0.323	-22.7	1.369	0.876	+56.3
7	0.269	0.630	-57.3	0.301	0.0835	-64.0	1.416	1.409	+0.5

It seems that at higher leaf conductance the internal resistance becomes more important for the limitation of the NO_2 uptake particularly for spruce. However, we should keep in mind that for dynamic chamber measurements at high leaf conductance, which is equivalent to a low resistance for gas exchange, the impact of the chamber resistances could become a significant factor because the values of the low plant resistances and the chamber resistances can be in the same order of magnitude. In this case both resistances add a similar rate to the total resistance and cannot be neglected anymore. The deposition of NO_2 inside the chamber is then not only limited by stomatal and mesophyllic resistance. Using predicted NO_2 leaf conductance calculated from known leaf conductance to water vapor to model NO_2 exchange fluxes would lead to under- or overestimations of the NO_2 fluxes.

It was particularly noticeable that at the EGER field site spruces showed a temperature dependence of stomatal conductance and consequently of gas exchange, because stomata close at high leaf temperatures to avoid drought. Hence, very low NO₂ (NO, O₃) exchange flux densities were detected during daytime if leaf temperature exceeded 23 °C. This plant behavior illustrates the need for following and measuring the plant-physiological processes like photosynthesis (CO₂ surface exchange flux) and transpiration (H₂O surface exchange flux) and the need to related them to the NO₂ (NO, O₃) surface exchange. Moreover, it is beneficial to know the plant's optimal conditions in order to interpret the NO₂ (NO, O₃) exchange measurements without ignorance of the plant's characteristics (e.g. temperature effects on stomatal closure at high temperatures would lead to underestimation of NO₂ (NO, O₃) exchange behavior if generalized to normal plant behavior).

4.3.3 Deposition velocities of NO₂ and O₃

The measurements on spruce branches were made for two different trees at the same time. Between both enclosed branches no differences in the deposition velocity of NO_2 and O_3 were observed.

Until now, reported $v_{dep,NO2}$ values were not differentiated into variable classes of leaf conductance. The measured $v_{dep,NO2}$ values between 0.07 and 0.42 mm s⁻¹ for spruce were of the same order of magnitude as values described by GEBLER et al. (2002) for field measurements under controlled conditions. GEBLER et al. (2002) described NO₂ deposition velocities for spruces of 0.09 mm s⁻¹, which is in the range of our minimal measured value. In contrast THOENE et al. (1991, 1996) described values of 0.4 - 0.9 mm s⁻¹ for laboratory measurements. However, RONDÓN et al. (1993) specified values of 1.8 to 2.1 mm s⁻¹, which were five times higher than our maximal measured

DISCUSSION

value. Interestingly, the determined $v_{dep,NO2}$ for oak under field conditions were clearly higher than for spruce but in the same range as reported by CHAPARRO-SUAREZ et al. (2011) for oak.

The differences in $v_{dep,NO2}$ between our measured values and previous studies may have been caused by unspecific NO₂ analyzers or by neglecting the photolysis of NO₂ for the determination of these previously reported $v_{dep,NO2}$. As mentioned in Sect. 3.3.2.3 we estimated how much the gas-phase reactions will affect $v_{dep,NO2}$ under the reported experimental conditions. For the reported values of RONDÓN et al. (1993) a highest expected error of 20 % were calculated according to Eq. (23.1), but the corrected $v_{dep,NO2}$ would be still above our measured values. The values of THOENE et al. (1991, 1996) would be in our range if the estimated error would be taken into account. Another reason for different $v_{dep,NO2}$ measured on leaf level can be different ages of the enclosed leafs or needles. GRENNFELT et al. (1983) reported of higher $v_{dep,NO2}$ for 1-year-old needles compared to current year needles. Unfortunately, studies about different performance of gas exchange depending on the needle ages are very seldom.

The classification into different leaf conductance classes resulted in increasing $v_{dep,NO2}$ values with rising leaf conductance. Consequently $v_{dep,NO2}$ is positively correlated to the leaf conductance. This finding agrees with a number of previous studies (NEUBERT et al. 1993; HEREID and MONSON 2001; SPARKS et al. 2001; GEBLER et al. 2002; CHAPARRO-SUAREZ et al. 2011).

The O₃ deposition velocities $v_{dep,O3}$ measured for spruce were between 0.06 and 0.38 mm s⁻¹ depending on the leaf conductance class. These values are in the same magnitude as the deposition velocities determined for NO₂. However, RONDÓN et al. (1993) reported about $v_{dep,O3}$ which were an order of magnitude higher than $v_{dep,NO2}$.

Most of the reported deposition velocities (NO₂ and O₃) are based on canopy deposition velocity measurements for both foliar and non-foliar sites using eddy correlation technique. Consequently the gas exchange of the soil, reactions with surfaces and reactions with radicals, for example VOCs emitted from plants, are taken into account. For a deciduous forest $v_{dep,O3}$ of 10 mm s⁻¹ in the summer and 3 mm s⁻¹ in the winter are reported from PADRO (1991). He also mentioned the values of $v_{dep,O3}$ over a vineyard, a cotton field and a senescent grass field the values are 5, 8 and 2 mm s⁻¹, respectively (PADRO 1996). For a spruce forest values of 7 mm s⁻¹ for $v_{dep,O3}$ were determined by

PILEGAARD et al. (1995), and the results of a study at a fruit orchard (several varieties of apple) resulted in $v_{dep O3}$ values of 3 to 5 mm s⁻¹ (WALTON et al. 1997). For NO₂ deposition velocities less values are reported at publications than for O₃. For wheat fields values of 0.35 mm s⁻¹ are reported by PILEGAARD et al. (1998) and for the fruit orchard 2 to 6 mm s⁻¹ by WALTON et al. (1997). Monthly means (Jan to Oct) of $v_{dep,NO2}$ for an oak forest were between 0.2 and 6.4 mm s⁻¹ (PUXBAUM and GREGORI 1998) and for a deciduous forest $v_{dep,NO2}$ values of 2 mm s⁻¹ were reported by HORII et al. (2004). To compare rudimentary deposition velocities determined from canopy or leaf-level measurements the values of the deposition velocity per projected needle area can be converted by multiplying the measured deposition velocity by the leaf area index (LAI). Additionally the existence of a NO₂ compensation point concentration can be considered by a correction of the NO₂ deposition velocity. According to RONDÓN et al. (1993) this correction can be calculated as $v_{depNO2}^{LAI,cor} = v_{depNO2}^{LAI} \cdot (1 - m_{comp,NO2}/m_{s,NO2})$, where $m_{comp,NO2}$ is the NO₂ compensation point concentration and $m_{s,NO2}$ is the mean NO₂ concentration during the period. Table 26 presents the NO₂ and O₃ deposition velocities, corrected and not corrected, to the canopy for the different classes of leaf conductance and species.

		Spruce			Oak		
		chamber 1					
class	$v_{dep,NO2}^{LAI}$	$v_{dep,NO2}^{LAI,cor}$	$\mathcal{V}_{dep,O3}^{LAI}$	$v_{dep,NO2}$ LA	$v_{dep,NO2}^{LAI,cor}$	$v_{dep,O3}^{LAI}$	$v_{dep,NO2}$ LAI
1	0.37	0.30	0.35	n.a.	no <i>m_{comp,NO2}</i>	0.29	2.84
2	0.46	0.34	0.62	0.71	no <i>m_{comp,NO2}</i>	0.58	4.16
3	0.68	0.56	0.94	0.74	no <i>m_{comp,NO2}</i>	0.88	6.43
4	0.56	0.73	1.14	1.30	1.04	1.12	8.90
5	1.28	0.91	1.40	1.50	1.21	1.36	8.48
6	1.58	0.99	1.67	1.00	no <i>m_{comp,NO2}</i>	1.46	10.24
7	1.82	1.75	2.00	2.20	1.96	1.65	12.73

Table 26: Averages of NO₂ and O₃ deposition velocities ($v_{dep,i}$ ^{LAI} in mm s⁻¹) per ground area (LAI) and $v_{dep,NO2}$ ^{LAI} corrected ($v_{dep,NO2}$ ^{LAI,cor} in mm s⁻¹) for NO₂ compensation point concentration when existent. LAI of Spruce forest (EGER) = 5.2, LAI of Oak forest (ECHO) = 4.7.

DISCUSSION

The converted NO₂ deposition velocity for the oak stand resulted in values between 2.8 and 17.7 mm s⁻¹ and an average of 7.7 mm s⁻¹. This averaged $v_{dep,NO2}^{LAI}$ is in a good agreement with the values reported by PUXBAUM and GREGORI (1998) who found monthly averaged NO₂ deposition velocities of 6.4 mm s⁻¹ at the month July for an Oak forest.

For the spruce stand $v_{dep,NO2}^{LAI}$ values were corrected for the NO₂ compensation point concentration when a compensation point was determined (see Sect. 4.2.5). The average value of $v_{dep,NO2}^{LAI,cor}$ was 0.98 mm s⁻¹, which is one order of magnitude lower than the reported averaged and corrected NO₂ deposition velocity per ground area for a spruce stand by RONDÓN et al. (1993). The determination of $v_{dep,O3}^{LAI}$ resulted in an average value of 1.10 mm s⁻¹, which come up to seventh part of the O₃ deposition velocity reported by PILEGAARD et al. (1995) for a Spruce forest.

It should be mentioned that, at least for spruce, the measurements were made at branches in the middle of the canopy. The radiation intensity and thus the stomatal conductance probably differs upwards to the top of canopy and downwards to the ground of forest. Accordingly, values of deposition velocities can differ over the whole tree stand. Therefore RONDÓN et al. (1993) considered their converted deposition velocities to be upper limits as they measured at the tree top.

4.3.4 Compensation point concentration

In literature a wide range of NO₂ compensation point concentrations ($m_{comp,NO2}$) were reported between 0.1 and 3 ppb (see Table 3). In this study the range of highly significant $m_{comp,NO2}$ determined for spruce needles under field conditions was between 7.4 ±6.40 and 29.0 ±16.30 nmol m⁻³ (0.17 - 0.65 ppb). The bi-variate weighted linear least-square fitting regression analysis of the laboratory measurements resulted in $m_{comp,NO2} = 5.9 \pm 9.13$ nmol m⁻³ (0.13 ppb) and the significance probability for the existence of $m_{comp,NO2}$ was 96.6 % ("likely") (see Sect. 3.2.4.1). The analysis of the data for oak resulted in negative NO₂ compensation point concentrations, which are unrealistic. Both results, measured under field and laboratory conditions, challenge the existence of a NO₂ compensation point concentration for spruce as well for oak. However, if a compensation point for NO₂ uptake exists the concentration will be less than 1 ppb.

These considerations are in close accordance with laboratory experiments as performed by CHAPARRO-SUAREZ et al. (2011) who also question the existence of a compensation point.

For coniferous trees NO₂ compensation point concentrations between 0.1 and 0.7 ppb were reported by RONDÓN et al. (1993) and RONDÓN and GRANAT (1994). These values are comparable to the values determined in this study. However, THOENE et al. (1996) determined $m_{comp,NO2}$ of 1.64 ppb for spruce and GEBLER et al. (2002) values of 1.7 ppb. Such large values (above 1 ppb) would imply an almost constant NO₂ emission from the forest at regions with small ambient NO₂ concentrations, which is not reported so far. These differences in the estimation of an exact compensation point concentration had led to some discussion (LERDAU et al. 2000). The discrepancy between the values determined in this study and those high values reported by previous studies (THOENE at al. 1996; GEBLER et al. 2002) may be explained by using different measurement techniques to detect NO₂ concentrations. As mentioned above (also see Sect. 2.2) most of the commonly used converters for the conversion of NO_2 to NO are not highly specific for NO₂. Converters, which consist of molybdenum, iron sulfate or predicted on liquid phase reaction (luminol), show interferences with other compounds as nitrous acid (HNO₂), nitric acid (HNO₃), peroxyacetyl nitrate (PAN) and other organic nitrates, therefore NO₂ concentrations will be overestimated. The highly NO₂ specific blue light converter performed in this study should minimize these source of error. Another reason for different estimations of compensation point concentrations can be the application of different measurement setups and data analysis. For the most results of other authors it is not clear if photochemistry of the NO-NO₂-O₃ triad was taken into account or if an experimental setup was used which excluded reactions of NO₂ photochemistry. The results of this study account for those potential sources of error. For the laboratory measurements a chamber design and an experimental setup were used which excluded photochemistry. During field measurements chemical reactions of the NO-NO₂-O₃ triad were part of the natural conditions, therefore the measured values were corrected. The impact of gas-phase reactions on compensation point concentrations is less than on deposition velocity. The overestimation of NO₂ compensation point concentration would be between 3 and 17 % for the values reported by THOENE et al. (1996) and GEBLER et al. (2002) (see Sect. 3.3.2.3). However, this would not suffice to explain the high values of NO₂ compensation point concentration.

DISCUSSION

Furthermore, the determination of deposition velocity and compensation point concentration by applying simple linear regression (no errors considered at all) or bi-variate weighted linear regression (y- and x-errors considered) could be another reason for the discrepancy. In most of the previous studies simple linear regression between exchange flux density $F_{ex,i}$ and the trace gas concentration at the outlet of the sample chamber $m_{s,i}$ were applied (RONDÓN et al. 1993; RONDÓN and GRANAT 1994; THOENE et al. 1996; SPARKS et al. 2001; HEREID and MONSON 2001) only GEBLER et al. (2000, 2002) applied a bi-variate algorithm.

Moreover, the observed difference between laboratory and field measurements and between the reported values from literature could have resulted from different plant materials used or different habitat conditions. Previous studies suggest that mesophyllic characteristics like leaf ascorbate concentration may influence NO₂ exchange rates (RAMGE et al. 1993; TEKLEMARIAM and SPARKS 2006). The apoplastic ascorbate concentration varies with species, environmental conditions (POLLE et al. 1995; SCHWANZ et al. 1996) and stage of development (LUWE 1996). The differences may be due to different ascorbate concentrations. Another reason could be a different colonization of the trees by chemolithoautotrophic nitrifying bacteria. It is known that these bacteria colonize the phyllosphere of trees. HEUSER and ZIMMER (2003) demonstrated autotrophic nitrite oxidizers on leaf surface of English oak (Quercus robur L.) and PAPEN et al. (2002) detected them at spruce needles. TEUBER (2003) was able to verify nitrifying bacteria living inside the apoplast of spruce needles. These organisms are able to metabolize NH_4^+ and NO_2^- which is formed when NO_2 dissolved in water. It is to be assumed that NO₂ uptake and compensation point concentration will be differing if plants are colonized by nitrifiers or not. From previous studies (PAPEN et al. 2002) it is known that NH₃ deposition fluxes significantly increased as consequence of metabolic activity of nitrifying bacteria. Possibly, this observation is also valid for NO₂.

Conclusions and Perspectives

In this study a dynamic chamber system for surface exchange flux measurements of reactive and non-reactive trace gases on plants under field and laboratory conditions was presented. I would like to conclude the findings as follows:

- One of the most important characteristics of the dynamic chamber system is the minimal disturbance of plant physiology and growth. Changes in concentrations of relevant trace gases should be small in order to be comparable to the outer environment. Furthermore, small changes prevent enclosure induced artifacts on plant metabolism and stomata regulation. Reliable investigations should not only focus on a few interesting trace gases but always include CO₂ and water vapor exchange because of plant physiological feedback regulations.
- 2. According to the "blank" measurements, the wall material of our plant chamber can be considered as chemically inert. I like to emphasize, that mass fluxes to the walls of the chamber can basically not be neglected and must be considered in the mass flux balance of the dynamic plant chamber, if there are any appreciable effects of ad- or desorption.
- 3. The performance of the dynamic chamber system must be controlled and, if necessary, suitable parameterized correction algorithms have to be applied to maintain/improve the precision of NO₂ concentration and exchange flux density measurements. The sensitivity of the NO/NO₂ analyzer to changes of ambient temperature is one of these parameters. Our analyzer drifted 0.07 ppb/K (NO) and 0.08 ppb/K (NO₂). The precision of the NO₂ exchange flux densities is almost entirely determined by the precision of the NO₂ concentration measurements, which in turn depends on the sensitivity (limit of detection) of the NO₂ analyzer. Considering best performance of our system, a flux density precision

140 | CONCLUSIONS AND PERSPECTIVES

of ≤ 10 % can be reached, as long as NO₂ concentrations in the plant chamber differ by 0.1 ppb from the expected NO₂ compensation point concentration.

- 4. Determination of NO₂ concentrations at sub-ppb level and of NO₂ exchange flux densities at the thousandths (hundredths) of nmol m⁻² s⁻¹ level definitely require (a) a NO₂ specific converter (photolytic converter) and (b) a highly sensitive NO/NO₂ analyzer (lower detection limit (3σ) of at least 13 nmol m⁻³ (0.3 ppb), better 4.5 nmol m⁻³ (0.1 ppb)).
- 5. The significance of concentration differences Δm_i (between trace gas concentrations measured at the inlet and the outlet of the dynamic chamber) is the fundamental quality criterion for the determination of high quality exchange flux densities and deposition velocities, but particularly for the detection of (highly) significant compensation point concentrations. Especially under field measurements, the percentage of non-significant Δm_i can be rather high due to the temporal variation of ambient concentrations during the measurement interval.
- 6. Laboratory measurements for the identification of NO₂ compensation point concentrations under controlled conditions require low, reproducible and verifiable NO₂ concentration for NO₂ fumigation experiments. The precision of corresponding NO₂ concentration measurements is not only limited by the noise of the NO/NO₂ analyzer, but also by the noise of the NO₂ blending procedure. Application of future NO/NO₂ analyzers (lower detection limit $(3\sigma) < 2.2 \text{ nmol m}^{-3} (< 0.05 \text{ ppb})$ will be useless, unless the uncertainty of the NO₂ blending for fumigation experiments will be improved significantly.
- 7. Photo-chemical reactions in the dynamic plant chamber's volume must be considered (or be excluded by corresponding set-ups). Otherwise, particularly the exchange of the NO-NO₂-O₃ triad with the plants could be seriously over- or underestimated. This is particularly important for the determination of the NO₂ deposition velocity. Under our experimental conditions in the field, the overestimation of the NO₂ deposition velocity had reached about 80 % if photolysis of NO₂ has been neglected. Excluding the chemical reaction of NO with O₃ by corresponding experimental design (e.g. using NO and O₃ free purging air), effects of NO₂ photolysis would still be present, as long as there is appreciable illumination of the plants. This can hardly be avoided because for plant

physiological studies photosynthetically active radiation is essential. The only way out would be to use a chamber wall material where the transmissivity for PAR is high and in the wavelength range of $j(NO_2)$ negligible. For laboratory studies, the application of light-emitting diodes which do not emit in the wavelength range of $j(NO_2)$ seems to be very promising.

- 8. Using an empty ("reference") chamber for considering (compensating) photochemical reactions would imply that NO₂-photolysis, as well as the concentrations of NO₂, NO and O₃ in the empty and in the plant chambers are identical; however, this is definitely not the case.
- 9. For mathematical correctness, deposition velocities and compensation point concentrations should be derived from linear relationships between the originally measured quantities, namely the NO, NO₂ and O₃ concentrations at the inlet and the outlet of the dynamic chamber. A straight-forward and thorough statistical treatment of measured data will result in high-quality and reliable data of exchange flux densities, deposition velocities and compensation point concentrations, if solid characterization and quantification of trace gas concentration errors as well as errors of all other quantities (necessary for calculation of the exchange flux densities) is achieved and general Gaussian error propagation as well as bi-variate weighted linear least-squares fitting regression analysis is applied.
- 10. It is recommended, that results from previous studies on NO₂ exchange flux densities, NO₂ deposition velocities and NO₂ compensation point concentrations which have been obtained by dynamic plant chambers should be handled with care owing to neglecting (at least) the effects of NO₂ photolysis in the plant chamber's volume and insufficient characterization of the specificity and precision of the NO₂ analyzers. A re-evaluation would be helpful.
- 11. The control of plant conditions and the plant nutrient composition after field measurements indicated that the enclosed branches were not harmed by the dynamic plant chambers and behaved normally still after six weeks of enclosure.

- 12. The plant's optimal conditions have to be known to avoid errors in analyzing of the NO₂ (NO, O₃) exchange measurements. Ignoring temperature effects on stomatal closure, plant gas exchange intensity would be misjudge if interpreting these fluxes as plant behavior without limitation. An underestimation would occur if gas exchange is only measured at high temperatures or an overestimation happens if measuring is performed only at low temperatures.
- 13. Exchange rates of NO₂ increased with raising NO₂ ambient concentrations and depend also on stomatal conductance. With raising leaf conductance for water vapor the NO₂ uptake increased linearly in case of spruce, whereas oak leaves exhibited a considerable decrease of NO₂ deposition at high leaf conductances at observed NO₂ ambient mixing ratios. Exchange rates of O₃ for spruce were non-linearly related to stomatal conductance and saturated at high leaf conductances.
- 14. The uptake of NO₂ is also limited by internal resistances. For spruce our results let us assume that the internal resistances add a small percentage to the total resistance of leaf gas exchange at low leaf conductance but increase at higher conductances. For oak the internal resistance seems to play a minor role. But for dynamic chamber measurements the chamber resistances may not be neglected if plant resistances and chamber resistances are in the same order of magnitude. NO₂ fluxes modeled by predicted NO₂ leaf conductances, which were calculated from known leaf conductances to water vapor, resulted in over- or underestimations of NO₂ fluxes.
- 15. Determination of deposition velocity and compensation point concentration requires a classification of the data, for example by leaf conductance. This is the only way to ensure comparable plant conditions and to assure reliable interpretation.
- 16. NO₂ deposition velocities ($v_{dep,NO2}$) are positively correlated to leaf conductance. For spruce $v_{dep,NO2}$ ranged between 0.07 and 0.42 mm s⁻¹ and for oak between 0.06 and 2.71 mm s⁻¹. NO₂ deposition velocities of spruce are within the lowest reported range of other reported data. NO₂ and O₃ deposition velocities determined for spruce were of the same magnitude.

- 17. Estimates of NO₂ deposition velocity per ground area (on a LAI basis) amounted to 7.7 mm s⁻¹ for oak and to 0.98 mm s⁻¹ for the spruce stand. Oak data were in a good agreement with reported values, whereas the estimates in case of spruce were lower than reported.
- 18. Highly significant NO₂ compensation point concentration ($m_{comp,NO2}$) determined for spruce needles under field conditions ranged between 7.4 ±6.40 and 29.0 ±16.30 nmol m⁻³ (0.17 - 0.65 ppb). Laboratory measurements resulted in $m_{comp,NO2} = 5.9 \pm 9.13$ nmol m⁻³ (0.13 ppb) with a significance probability for the existence of $m_{comp,NO2}$ of 96.6 % ("likely"). For oak no $m_{comp,NO2}$ was found. The results for spruce under field and laboratory conditions challenge the existence of a NO₂ compensation point concentration. There is increasing indication that forests are mainly a sink for NO₂ and potential NO₂ emissions are low. Only when assuming high NO soil emissions, more NO₂ can be formed by reaction with O₃ than plants are able to take up. Under these circumstances forests can be a source for NO₂.
- 19. The constant lower values of NO₂ gas exchange flux densities, NO₂ deposition velocities and NO₂ compensation point concentrations in comparison to most previous studies probably based up on usage of more specific NO₂ analyzer with a blue light converter.

This study demonstrated, that the determination of significant NO₂ exchange fluxes, deposition velocities and compensation point concentrations highly depend on the resolution of the NO₂ concentration differences measured at the inlet and outlet of the dynamic chamber. For further research the precision of the NO₂ concentration measurements could be enhanced by using NO₂ analyzers with an improved detection limit, maybe to be better than 10 nmol m⁻³ (0.25 ppb). Measurements under laboratory conditions at very low NO₂ concentrations only take advantage of an improved NO₂ analyzer if the blending procedure is improved too. Otherwise the fluctuations due to the blending procedure will affect the precision of the NO₂ concentration measurements.

The field experiment presented in this study illustrated the problems of instationarity of trace gas concentrations during non-simultaneous concentration measurements.

144 | CONCLUSIONS AND PERSPECTIVES

Stationarity can only be achieved if trace gas concentration is constant during measuring period without concentration peaks. To approach this problem another independent measuring system can be used to determine the ambient trace gas concentration constantly. However, according to the available technique the use of one single instrument is preferable to guarantee precise simultaneous concentration measurements at the inlet and outlet of the dynamic chamber. Operation of two trace gas analyzers with an absolute accuracy and precision much better than the expected difference between inlet and outlet concentration is currently not available. Another option would be a two channel analyzer, which could measure directly the concentration difference between the inlet and outlet of the dynamic chamber. A different option would be the reduction of the switching time between measurements of chamber inlet and outlet. However, as a consequence tubing lengths would have to be reduced, which is hard to handle for field measurements, especially when analyzers are affected by temperature fluctuations and therefore should be operated under constant temperature conditions.

A main conclusion of this study is that photo-chemical reactions in the dynamic plant chamber's volume must be considered. So all reactions inside the chamber have to be known for an accurate chemical correction. For detailed understanding it would be necessary to verify if the reactions of the NO-NO₂-O₃ triad cover the main production and destruction processes inside the chambers. Additional processes include reactions of the reactive trace gases with plant surfaces and water films on the surface and reactions with volatile organic compounds (VOC's) emitted by the plant itself. In this context another interesting point is the formation of nitrous acid (HONO) by disproportionation of NO₂ and the rapidly decomposing of HONO into NO. This reaction would provide a NO source inside the chamber. This possibility is indicated by changing signs of NO fluxes from negative to positive (equivalent to emission) just by chemical gas phase correction.

This study exhibits some interesting findings about the specific gas exchange behavior of spruce and oak. Issues like different leaf types (sun and shade leaves), plant and leaf age or the impact of stem and bark deserve closer attention. For further investigations another question will be if NO₂ exchange behavior would change if the nitrogen supply via soil changes. A higher nitrogen fertilization of soil could cause a lower nitrogen uptake by leaves and vise versa. On the other hand the bottom up way of analysis of the nitrogen metabolism raises the question of nitrogen uptake limitation. One reason maybe changes in plant metabolism affecting nitrate reductase activity or apoplastic ascorbate concentration. Further investigations of this issue is needed especially for forest forming species like spruce, beech and oak as well as plants at the subcanopy level.

More process related gas exchange measurements at leaf level are required to compare results with flux measurements above canopy performed as eddy covariance measurements. This could help to discover misunderstandings in the biosphere-atmosphere exchange model of the ecosystem forest. A further method linking leaf and canopy level can be the use of open-top chambers. This method is well known for exposing plant mesocosms to elevated CO_2 and could be used as well for NO_2 fumigation of a whole plant.

147 | References

References

- Alsheimer, M.: Charakterisierung räumlicher und zeitlicher Heterogenität der Transpiration unterschiedlicher montaner Fichtenbestände durch Xylemflussmessungen, Bayreuther Forum Ökologie **49**, 1-143, 1997.
- Amman, M., Stalder, M., Sutter, M., Brunold, C., Baltensperger, U., Jost, D.T., Turler, A. and Gaggeler, H.W.: Tracing uptake and assimilation of NO₂ in spruce needles with ¹³N, Journal of Experimental Botany **46**, 1685-1691, 1995.
- Atkinson, R., Baulch, D.L., Cox, R.A., Crowley, J.N., Hampson, R.F., Hynes, R.G., Jenkin, M.E., Rossi, M.J. and Troe, J.: Evaluated kinetic and photochemical data for atmospheric chemistry: Volume I – gas phase reactions of O_x, HO_x, NO_x and SO_x species, Atmospheric Chemistry and Physics 4, 1461-1738, 2004, http://www.atmoschem-phys.net/4/1461/2004/.
- Beier, N. and Schneewind, R.: Chemical reactions of gases in tubes of probing systems and their influence on measured concentrations, Ann. Geophysicae 9, 703-707, 1991.
- Bollmann, A. and Conrad, R.: Influence of O₂ availability on NO and N₂O release by nitrification and denitrification in soils, Global Change Biology **4**, 387-396, 1998.
- Burkhard, J. and Eiden, R.: Thin water films on coniferous needles, Atmospheric Environment **28**, 2001-2017, 1994.
- von Caemmerer, S. and Farquhar, G.D.: Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves, Planta **153**, 376-387, 1981.
- Cantrell, C.A.: Technical note: Review of methods for linear least-squares fitting of data and application to atmospheric chemistry problems, Atmospheric Chemistry and Physics **8**, 5477-5487, 2008.
- Chaparro-Suarez, I.G.: Untersuchungen zum Austausch von Stickstoffdioxid (NO₂) zwischen Pflanzen und Atmosphäre, Phd Thesis, Johannes Gutenberg University Mainz, Mainz, Germany 110 p, 2008.
- Chaparro-Suarez, I.G, Meixner, F.X., Kesselmeier, J.: Nitrogen dioxide (NO₂) uptake by vegetation controlled by atmospheric concentrations and plant stomatal aperture, Atmospheric Environment, 2011, doi: 10.1016/j.atmosenv.2011.07.021 (in press).
- Coe, H., Gallagher, M.W. and Choularton, T.W.: NO_x and O₃ exchange above a forest canopy in southern Scottland. In: Slanina, J., Angeletti, G. and Beilke, S. (Eds.), General assessment of biogenic emissions and deposition of nitrogen compounds,

sulphur compounds and oxidants in Europe, Air Pollution Research Report 47, Commission of the European Communities, Directorate-General for Science, Research and Development, Brussels, Belgium, pp. 189-200, 1993.

- Conrad, R.: Compensation concentration as critical variable for regulating the flux of trace gases between soil and atmosphere, Biogeochemistry **27**, 155-170, 1994.
- Cox, R.A., Jones, B.M.J., Penkett, S.A. and Sheppard, D.A.: Mechanism of photochemical and free radial oxidation of sulfur compounds in the gas phase, paper presented at the 5th International Conference of the Commission on Atmospheric Chemistry and Global Pollution, Oxford, England, Aug. 28 to Sept. 3, 1983
- Crutzen, P.J.: The role of NO and NO₂ in the chemistry of the troposphere and stratosphere, Annual Reviews Earth Planet, Science 7, 443-472, 1979.
- Crutzen, P.J.: Role of the tropics in atmospheric chemistry. In: The geophysiology of Amazonia, edited by R.E. Dickinson, 107-132, John Wiley & Sons, New York, 1987.
- Dean, J.V. and Harper, J.E.: Nitric oxide and nitrous oxide production by soybean and winged bean during in vivo nitrate reductase assay, Plant Physiology **82**, 718-732, 1986.
- Denman, K.L., G. Brasseur, G., A. Chidthaisong, P. Ciais, P.M. Cox, R.E. Dickinson, D. Hauglustaine, C. Heinze, E. Holland, D. Jacob, U. Lohmann, S Ramachandran, P.L. da Silva Dias, S.C. Wofsy and X. Zhang, 2007: Couplings Between Changes in the Climate System and Biogeochemistry. In: Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change [Solomon, S., D. Qin, M. Manning, Z. Chen, M. Marquis, K.B. Averyt, M.Tignor and H.L. Miller (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Dindorf T., Kuhn, U., Ganzeveld, L., Schebeske, G., Ciccioli, P., Holzke, C., Köble, R., Seufert, G. and Kesselmeier, J.: Significant light and temperature dependent monoterpene wmissions from European beech (*Fagus sylvatica* L.) and their potential impact on the European volatile organic compound budget, Journal of Geophysical Research **111**, D16305, 2006, doi:10.1029/2005JD006751.
- Drummond, J.W., Castledine, C., Green, J., Denno, R., Mackay, G.I. and Schiff, H.I.: New technologies for use in acid deposition networks. In Monitoring Methods for Toxics in the Atmosphere. ASTM Special Technical Publication No. 1052, Philadelphia, 1989.

Erisman, J.W., Sutton, M.A., Galloway, J., Klimont, Z. and Winiwarter, W.: How a century of ammonia synthesis changed the world, Nature Geoscience 1, 636-639, 2008.

Fangmeier A., Hadwiger-Fangmeier A., Van der Eerden L. and Jäger H.-J.: Effects of atmospheric ammonia on vegetation - A review, Environmental Pollution 86, 43-82, 1994.

- Fehsenfeld, F.C., Dickerson, R.R., Hübler, G., Luke, W.T., Nunnermacker, L.J., Williams, E.J., Roberts, J.M., Calvert, J.G., Curran, C.M., Delany, A.C., Eubank, C.S., Fahey, D.W., Fried, A., Gandrud, B.W., Langford, A.O., Murphy, P.C., Norton, R.B., Pickering, K.E. and Ridley, B.A.: A Ground-Based Intercomparison of NO, NO_x and NO_y Measurement Techniques, Journal of Geophysical Research **92**(D12), 14,710-14,722, 1987.
- Fehsenfeld, F.C., Drummond, J.W., Roychowdhury, U.K., Galvin, P.J., Williams, E.J., Buhr, M.P., Parrish, D.D., Hübler, G., Langford, A.O., Calvert, J.G., Ridley, B.A., Grahek, F., Heikes, B.G., Kok, G.L., Shetter, J.D., Walega, J.G., Elsworth, C.M., Norton, R.B., Fahey, D.W., Murphy, P.C., Hovermale, C., Mohnen, V.A., Demerjian, K.L., Mackay, G.I. and Schiff, H.I.: Intercomparison of NO₂ measurement techniques, Journal of Geophysical. Research **95**, 3579-3597, 1990.
- Foken, T.: Lufthygenisch-bioklimatische Kennzeichnung des oberen Egertales (Fichtelgebirge bis Karlovy Vary), Bayreuther Forum Ökologie **100**, 1-70, 2003.
- Frey W., Lösch R.: Lehrbuch der Geobotanik. Pflanze und Vegetation in Raum und Zeit. Spektrum Akademischer Verlag, München. 528 p., 2004.
- Fuhrer, J. and Erismann, K.H.: Uptake of NO₂ by plants grown at different salinity levels, Experientia **36**, 409-410, 1980.
- Galloway, J.N., Dentener, F.J., Capone, D.G., Boyer, E.W., Howarth, R.W., Seitzinger, S.P., Asner, G.P., Cleveland, C.C., Green, P.A., Holland, E.A., Karl, D.M., Michaels, A.F., Porter, J.H., Townsend, A.R. and Vorosmarty, C.J.: Nitrogen cycles: past, present and future, Biogeochemistry 70, 153-226, 2004.
- Gehrig, R. and Baumann, R.: Comparison of 4 different types of commercially available monitors for nitrogen oxides with test gas mixtures of NH₃, HNO₃, PAN and VOC and in ambient air, paper presented at EMEP Workshop on Measurements of Nitrogen-Containing Compounds, EMEP/CCC Report 1/93, Les Diablerets, Switzerland, 1993.
- Gerstberger, P., Foken, T. and Kalbitz, K.: Biogeochemistry of forested catchments in a changing environment: A German case study, Ecological studies, Springer, chapter The Lehstenbach and Steinkreuz catchments in NE Bavaria, Germany, 15-41, 2004.
- Geßler, A., Rienks, M. and Rennenberg, H.: NH₃ and NO₂ fluxes between beech trees and the atmosphere - correlation with climatic and physiological parameters, New Phytologist **147**, 539-560, 2000.
- Geßler, A., Rienks, M. and Rennenberg, H.: Stomatal uptake and cuticular adsorption contribute to dry deposition of NH₃ and NO₂ to needles of adult spruce (Picea abies) trees, New Phytologist **156**, 179-194, 2002.
- Grennfelt, P., Bengtson, C. and Skärby, L.: Dry deposition of nitrogen dioxide to scots pine needles. In: Precipitation, scavenging, dry deposition and resuspension, edited by R. Pruppacher, H.R.G. Semonin and W.G.N. Slinn, Elsevier, New York 1983.

- Grosjean, D. and Harrison, J.: Response of chemiluminescence NO_x analyzers and ultraviolet ozone analyzers to organic air pollutants, Environmental Science & Technology **19**, 862-865, 1985.
- Gut, A., Scheibe, M., Rottenberger, S., Rummel, U., Welling, M., Ammann, C., Kirkman, G.A., Kuhn, U., Meixner, F.X., Kesselmeier, J., Lehmann, B.E., Schmidt, W., Müller, E. and Piedade, M.T.F.: Exchange fluxes of NO₂ and O₃ at soil and leaf surfaces in an Amazonian rain forest, Journal of Geophysical Research 107(D20), 8060, doi: 10.1029/2001JD000654, 2002.
- Hanson, P.J., Rott, K., Taylor, G.E.Jr., Gunderson, C.A., Lindberg, S.E. and Ross-Todd, B.M.: NO₂ deposition to elements representative of a forest landscape, Atmospheric Environment 23, 1783-1794, 1989.
- Hanson, P.J. and Lindberg, S.E.: Dry deposition of reactive nitrogen compounds: A rewiev of leaf, canopy and non-foliar measurements, Atmospheric Environment **25A**, 8, 1615-1634, 1991.
- Hereid, D.P. and Monson, R.K.: Nitrogen oxide fluxes between corn (*Zea mays* L.) leaves and the atmosphere, Atmospheric Environment **35**, 975-983, 2001.
- Heuser, T. and Zimmer, W.: Genus- and Isolate-Specific Real-Time PCR Quantification of *Erwinia* on Leaf Surfaces of English Oaks (*Quercus robur* L.), Current Microbiology 47, 214-219, 2003.
- Hicks, B. B., Baldocchi, D.D., Meyers, T.P., Hosker, R.P., Matt, D.R.: A preliminary multiple resistance routine for deriving dry deposition velocities from measured quantities, Water, Air, & Soil Pollution **36**, 311, 1987.
- Hill, C.: Vegetation: a sink for atmospheric pollutants, Journal of the Air Pollution Control Association **21**, 341-346, 1971.
- Horii, C.V., Munger, j.W., Wofsy, C., Zahniser, M., Nelson, D. and McManus, J.B.: Fluxes of nitrogen oxides over a temperate deciduous forest, Journal of Geophysical Research **109**, D08305, 2004, doi:10.1029/2003JD004326.
- Hosaynali Beygi, Z., Fischer, H., Harder, H.D., Martinez, M., Sander, R., Williams, J., Brookes, D.M., Monks, P.S. and Lelieveld, J.: Oxidation photochemistry in the Southern Atlantic boundary layer: Unexpected deviations of photochemical steady state, Atmospheric Chemistry and Physics Discuss. 11, 7045-7093, 2011, doi:10.5194/acpd-11-7045-2011.
- Jacob, D.J. and Wofsy, S.C.: Budgets of Reactive Nitrogen, Hydrocarbons, and Ozone over the Amazon Forest during the Wet Season, Journal of Geophysical Research 95(D10), 16737-16754, 1990.
- Jensen, E.S. and Pilegaard, K.: Absorption of nitrogen dioxide by barley in open-top chambers, New Phytologist **123**, 359-364, 1993

- Johansson, C.: Pine forest: a negligible sink for atmospheric NO_x in rural Sweden, Tellus **39B**, 426-438, 1987.
- Jones, H.G.: Plants and microclimate, 2nd ed., Cambridge (Cambr. Univ. Pr.), 1992.
- Kelly, T.J., Stedman, D.H., Ritter, J.A. and Harvey, R.B.: Measurements of Oxides of Nitrogen and Nitric Acid in Clean Air, J. Geophys. Res., 85, C12, 7417-7425, 1980.
- Kelly, J., Spicer, C.W. and Ward, G.F.: An assessment of the luminol chemiluminescence technique for measurement of NO₂ in ambient air, Atmospheric Environment **24A**, 2397-2403, 1990.
- Kesselmeier J., Merk L., Bliefernicht M.,and Helas G.: Trace gas exchange between terrestrial plants and atmosphere: carbon dioxide, carbonyl sulfide and ammonia under the rule of compensation points. In: General Assessment of Biogenic Emissions and Deposition of Nitrogen Compounds, Sulphur Compounds and Oxidants in Europe, CEC Air Pollution Research Report 47 (edited by Slanina J., Angeletti G. and Beilke S.), pp. 71-80, 1993.
- Kesselmeier, J., Meixner, F.X., Hofmann, U., Ajavon, A., Leimbach, S. and Andreae, M.O.: Reduced sulfur compound exchange between the atmosphere and tropical tree species in southern Cameroon, Biogeochemistry 23, 23-45, 1993.
- Kesselmeier, J., Schäfer, L., Ciccioli, P., Branceleoni, E., Cecinato, A., Frattoni, M., Foster, P., Jacob, V., Denis, J., Fugit, J.L., Dutaur, L. and Torres, L.: Emission of monoterpenes and isoprene from a Mediterranean oak species Quercus ilex L. measured within the BEMA (Biogenic Emissions in the Mediterranean Area) project, Atmospheric Environment **30**, 1841-1850, 1996.
- Kesselmeier, J., Bode, K., Hofmann, U., Mueller, H., Schaefer, L., Wolf, A., Ciccioli, P., Brancaleoni, E., Cecinato, A., Frattoni, M., Foster, P., Ferrari, C., Jacob, V., Fugit, J.L., Dutaur, L., Simon, V. and Torres, L.: Emission of short chained organic acids, aldehydes and monoterpenes from Quercus ilex L. and Pinus pinea L. in relation to physiological activities, carbon budget and emission algorithms, Atmospheric Environment **31** (SI), 119-134, 1997.
- Kesselmeier, J., Bode, K., Gerlach, C. and Jork, E.M.: Exchange of atmospheric formic and acetic acids with trees and crop plants under controlled chamber and purified air conditions, Atmospheric Environment **32**, 1765-1775, 1998.

Kisser-Priesack, G.M., Scheunert, I. and Gnatz, G.: Uptake of ¹⁵NO₂ and ¹⁵NO by plant cuticles, Naturwissenschaften **74**, 550-551, 1987.

Kisser-Priesack, G.M., Bienek, D. and Ziegler, H.: NO₂ binding to defined phenolics in the plant cuticle, Naturwissenschaften **77**, 492-493, 1990.

Klepper; L.: Nitric oxide (NO) and nitrogen dioxide (NO₂) emissions from herbicide-treated soybean plants, Atmospheric Environment **13**, 537-542, 1979.

- Kuhn, U., Wolf, A., Gries, C., Nash, T.H. and Kesselmeier, J.: Field measurements on the exchange of carbonyl sulfide between lichens and the atmosphere, Atmospheric Environment **34**, 4867-4878, 2000.
- Kuhn, U., Rottenberger, S., Biesenthal, T., Wolf, A., Schebeske, G., Ciccioli, P., Brancaleoni, E., Frattoni, M., Tavares, T.M. and Kesselmeier, J.: Isoprene and monoterpene emissions of Amazonian tree species during the wet season: Direct and indirect investigations on controlling environmental functions, Journal of Geophysical Research 107(D20), 8071, doi: 10.1029/2001JD000978, 2002.
- Kurtenbach, R., Becker, K.H., Gomes, J.A.G., Kleffmann, J., Lörzer, J.C., Sittler, M., Wiesen, P., Ackermann, R., Geyer, A., Platt, U.: Investigations of emissions and heterogeneous formation of HONO in a road traffic tunnel, Atmospheric Environment 35, 3385-3394, 2001.
- Larcher, W.: Physiological Plant Ecology, 4th ed., Springer-Verlag, Berlin Heidelberg New York, 2003.
- Lea, P.J. and Miflin, B.J.: Alternative route for nitrogen assimilation in higher plants, Nature **251**, 614-616, 1974.
- Lea, P.J., Wolfenden, J., Wellburn, A.R.: Influence of air pollutants upon nitrogen metabolism, In: Alscher, R., Wellburn, A.R., eds. Plant Responses to the Gaseous Environment. London: Chapman & Hall, 279-299, 1994.

Lee, Y.N. and Schwartz, S.E.: Evaluation of the rate of uptake of nitrogen dioxide by atmospheric and surface liquid waters, Journal of Geophysical Research **86**, 11971-11973, 1981.

- Lendzian, K.J. and Kerstiens, G.: Interactions between plant cuticles and gaseous air pollutants, Aspects of Applied Biology **17**, 97-104, 1988.
- Lerdau, M.T., Munger, J.W. and Jacob, D.J.: The NO₂ Flux Conundrum, Science **289**(5488), 2291-2293, 2000.
- Logan, J.A.: Nitrogen oxides in the troposphere: Global and regional budgets, Journal of Geophysical Research Atmospheres **88**, 10785-10807, 1983.
- Ludwig, J.: Untersuchungen zum Austausch von NO und NO₂ zwischen Atmosphäre und Biosphäre, PhD Thesis, University of Bayreuth, Bayreuth, Germany, 251 pp, 1994.
- Ludwig, J., Meixner, F.X., Vogel, B. and Forstner, J.: Soil-air exchange of nitric oxide: An overview of processes, environmental factors and modeling studies, Biogeochemistry **52**, 225-257, 2001.
- Luwe, M.: Antioxidants in the apoplast and symplast of beech (*Fagus sylvatica* L.) leaves: seasonal variations and responses to changing ozone concentrations in air, Plant Cell and Environment **19**(3), 321-328, 1996.

- MacDougall, D. and Crummett, W.B.: Guidelines for Data Acquisition and Data Quality Evaluation in environmental chemistry, Analytical Chemistry **52**, 2242-2249, doi: 10.1021/ac5064a004, 1980.
- Massman, W.J.: A review on the molecular diffusivities of H₂O, CO₂, CH₄, CO, O₃, SO₂, NH₃, N₂O, NO and NO₂ in air, O₂ and N₂ near STP, Atmospheric Environment **32**, 1111-1127, 1998.
- Matthews, R.D., Sawyer R.F. and Schefer R.W. Interferences in Chemiluminescent Measurement of NO and NO₂ Emissions from Combustion Systems, Environmental Science & Technology **11**, 1092-1096, 1977.
- Maeck, G.: Organ-specific changes in the activity and subunit composition of glutamine-synthetase isoforms of barley (*Hordeum vulgare* L.) after growth on different levels of NH₄⁺, Planta **196**, 231-238, 1995.
- Maeda, Y.K., Aoki, K. and Munemori, M.: Chemiluminescence method for the determination of nitrogen dioxide, Analytical Chemistry **52**, 307-311, 1980.
- Meixner, F.X.: Surface Exchange of Odd Nitrogen Oxides, Nova Acta Leopoldina 70(288), 299-348, 1994.
- Meixner, F.X., Fickinger, Th., Marufu, L., Serca, D. Nathaus, F.J., Makina, E., Mukurumbira, L., Andreae, M.O.: Preliminary results on nitric oxide emission from a southern African savanna ecosystem, Nutrient Cycling in Agroecosystems 48, 123-138, 1997.
- Neubert, A., Kley, D. and Wildt, J.: Uptake of NO, NO₂ and O₃ by sunflower (Helianthus annuus L.) and Tobacco plants (Nicotiana tabacum L.): dependence on stomatal conductivity, Atmospheric Environment **27A**(14), 2137-2145, 1993.
- Nussbaum, S., Von Ballmoos, P., Gfeller, H., Schlunegger, U.P., Fuhrer, J., Rhodes, D. and Brunold, C.: Incorporation of atmospheric ¹⁵NO₂-nitrogen into free amino acids by
- Norway spruce Picea abies (L.) Karst, Oecologia 94, 408-414, 1993.
- Padro, J.: Seasonal contrast in modeled and observed dry deposition velocities of O₃, SO₂ and NO₂ over three surfaces, Atmospheric Environment **27A(**6), 807-814, 1993.
- Padro, J.: Summary of ozone dry deposition velocity measurements and model estimates over vineyard, cotton, grass and deciduous forest in summer, Atmospheric Environment **30**(13), 2363-2369, 1996.
- Pape, L., Ammann, C., Nyfeler-Brunner, A., Spirig, C., Hens, K. and Meixner, F.X.: An automated dynamic chamber system for surface exchange measurement of nonreactive and reactive trace gases of grassland ecosystems, Biogeosciences 6, 405-429, http://www.biogeosciences.net/6/405/2009/, 2009.

- Papen, H., Geßler, A., Zumbusch, E., Rennenberg, H.: Chemolithoautotrophic Nitrifiers in the Phyllosphere of a Spruce Ecosystem Receiving High Atmospheric Nitrogen Input, Current Microbiology 44, 56-60, 2002.
- Park, J.Y. and Lee, Y.N.: Solubility and decomposition kinetics of nitrous acid in aqueous solution, Journal of Physical Chemistry 92, 6294-6302, 1988.
- Pflüger, R. and Mengel, K.: Die photochemische Aktivität von Chlroplasten aus unterschiedlich mit Kalium ernährten Pflanzen, Plant and Soil **36**, 417-425; 1972.
- Phillips, N., Bond, B.J., McDowell, N.G., Ryan, M.G.: Canopy and hydraulic conductance in young, mature and old Douglas-fir trees, Tree Physiology 22, 205-211, 2002.
- Pilegaard, K., Jensen, N.O. and Hummelshøj, P.: Seasonal and diurnal variation in the deposition velocity of ozone over a spruce forest in Denmark, Water, Air and Soil Pollution 85, 2223-2228, 1995.
- Pilegaard, J., Hummelshøj, P., Jensen, N.O.: Fluxes of ozone and nitrogen dioxide measured by eddy correlation over a harvested wheat field, Atmospheric Environment **32**(7), 1167-1177, 1998.
- Plake, D.: Vertikale Konzentrationsprofile und Flüsse von reaktiven und nicht reaktiven Spurengasen im Fichtelgebirge, Diplomarbeit Thesis, Universität Münster, Münster, 144 pp, 2009.
- Polle, A., Wieser, G. and Havranek, W.M.: Quantification of ozone influx and apoplastic ascorbate content in needles of Norway spruce trees (*Picea abies* L., Karst) at high altitude, Plant, Cell and Environment **18**(6), 681-688, 1995.
- Puxbaum, H. and Gregori, M.: Seasonal and annual deposition rates od sulphur, nitrogen and chloride species to an oak forest in north-eastern Austria (Wolkersdorf, 240 m A.S.L.), Atmospheric Environment **32**(20), 3557-3568, 1998.
- Raivonen, M., Vesala, T., Pirjola, L., Altimir, N., Keronen, P., Kulmala, M. and Hari, P., Compensation point of NOx exchange: Net result of NOx consumption and production, Agricultural and Forest Meteorology 149, 1073-1081, 2009.
- Ramge, P., Badeck, F.W., Plöchl, M. and Kohlmaier, G.H.: Apoplastic antioxidants as decisive elimination factors within the uptake process of nitrogen dioxide into leaf tissues, New Phytologist 125, 771-785, 1993.
- Rennenberg, H. and Geßler, A.: Consequences of N deposition to forest ecosystems -Recent results and future research needs, Water, Air and Soil Pollution **116**, 47-64, 1999.
- Ridley, B.A., Carroll, M.A., Torres, A.L., Condon, E.P., Sachse, G.W., Hill, G.F. and Gregory, G.L. An intercomparison of results from ferrous sulphate and photolytic converter techniques for measurements of NO_x made during the NASA GTE/CITE 1 aircraft program, Journal of Geophysical Research **93**(D12), 15,803-15,811, 1988.
- Riederer, M., Kurbasik, K., Steinbrecher, R. and Voss, A.: Surface areas, lengths and volumes of *Picea abies* (L.) Karst. Needles: determination, biological variability and effect of environmental factors, Trees 2, 165-172, 1988.
- Robertson, G.P. and Groffman, P.M.: Nitrogen transformation, in: Soil microbilogy, ecology, and biochemistry, edited by: Paul, E.A., Elsevier, Heidelberg, 2007.
- Rondón, A. and Granat, L.: Studies on the dry deposition of NO_x to coniferous species at low NO₂ concentrations, Tellus **46B**, 339-352, 1994.
- Rondón, A., Johansson, C. and Granat, L.: Dry deposition of nitrogen dioxide and ozone to coniferous forest, Journal of Geophysical Research **98**, 5159-5172, 1993.
- Ryerson, T.B., Williams, E.J. and Fehsenfeld, F.C.: An efficient photolysis system for fast-response NO₂ measurements, Journal of Geophysical Research **105**(D21), 26,447-26,461, 2000.
- Sakakibara, H., Shimizu, H., Hase, T., Yamazaki, Y., Takao T., Shimonishi, Y., Sugiyama, T.: Molecular Identification and Characterization of Cytosolic Isoforms of Glutamine Synthetase in Maize Roots, Journal of Biological Chemistry 271(47), 29561-29568, 1996.
- Saxe, H.: Stomatal-dependent and stomatal-independent uptake of NO_x, New Phytologist **103**, 199-205, 1986.
- Schäfer, L., Kesselmeier, J. and Helas, G.: Formic and Acetic acid emission from conifers measured with a "cuvette" technic, in CeC Air Pollution Research 39: Field Measurements and Interpretation of Species Related to Photooxidants and Acid Deposition, edited by G. Angeletti, S. Beilke, and J. Slanina, 319-323, Eur. Comm., Brussels, 1992.
- Schiff, H.I., Mackay, G.I., Castledine, C., Harris, G.W. and Tran, Q.: Atmospheric measurements of nitrogen dioxide with a sensitive luminol instrument, Water, Air and Soil Pollution 30, 105-114, 1986.
- Schjoerring, J.K., Husted, S., Mack, G., Nielsen, K.H., Finnemann, J., Mattsson, M.: Physiological regulation of plant-atmosphere ammonia exchange, Plant and Soil **221**, 95-102, 2000.
- Schjoerring, J.K., Husted, S., Mattsson, M.: Physiological parameters controlling plantatmosphere ammonia exchanges, Atmospheric Environment 32, 491–498, 1998.
- Schulze, E.-D.: Air pollution and forest decline in a spruce (*Picea abies*) forest, Science **244** (4906), 776-783, 1989, doi:10.1126/science.244.4906.776.
- Schwanz, P., Picon, C., Vivien, P., Dreyer, E., Guehl, J.-M. and Polle, A.: Responses of antioxidative systems to drought stress in pendunculate oak and maritime pine as modulated by elevated CO₂, Plant Physiology **110**, 393-402, 1996.

- Seinfeld, J.H. and Pandis, S.N.: Atmospheric Chemistry and Physics: From Air Pollution to Climate Change, 2nd ed., John Wiley & Sons, Inc., Hoboken, New Jersey, 2006.
- Serafimovich, A, Siebicke, L, Staudt, K, et al.:, ExchanGE processes in mountainous Regions (EGER) - Documentation of the Intensive Observation Period (IOP2) June, 1st to July, 15th 2008, Arbeitsergebnisse. 37, Universität Bayreuth, Abteilung Mikrometeorologie ISSN 1614-8916, Bayreuth. 147pp, 2008.
- Sieghardt; H.: Schwermetall- und Nährelementgehalte von Pflanzen und Bodenproben schwermetallhaltiger Halden im Raum Bleiberg in Kärnten (Österreich). II. Holzpflanzen, Zeitschrift für Pflanzenernährung und Bodenkunde **151**, 21-26, 1988.
- Skärby, L., Bengtson, C., Boström, C.A., Grennfelt, P. and Troeng, E.: Uptake of NO_x in Scots pine, Silva Fennica **15**, 396-398, 1981.
- Sparks, J.P., Monson, R.K., Sparks, K.L. and Lerdau, M.: Leaf uptake of nitrogen dioxide (NO₂) in a tropical wet forest: implications for tropospheric chemistry, Oecologia 127, 214-221, 2001.
- Steinbacher, M., Zellweger, C., Schwarzenbacher, B., Bugmann, S., Buchmann, B., Ordóñez, C., Prevot, A.S.H. and Hueglin C.: Nitrogen oxide measurements at rural sites in Switzerland: Bias of conventional measurement techniques, Journal of Geophysical Research 112, D11307, doi:10.1029/2006JD007971, 2007.
- Stulen, I., Perez-Soba, M., De Kok, L.J., Van der Eerden, L.: Impact of gaseous nitrogen deposition on plant functioning, New Phytologist **139**, 61-70, 1998.
- Taylor, J.R.: An introduction to error analysis: The study of uncertainties in physical measurements, Oxford University Press, Mill Valley, CA, 270 p, 1982.
- Teklemariam, T.A. and Sparks, J.P.: Leaf fluxes of NO and NO_2 in four herbaceous plant species: The role of ascorbic acid, Atmospheric Environment **40**, 2235-2244, 2006.
- Teuber, M.: Nachweis, Lokalisation und Quantifizierung von autotrophen Nitrifizierern im Kronenraum der Fichte (*Picea abies* (L.) Karst.), Phd Thesis, Research Centre Karlsruhe, Albert-Ludwigs University Freiburg i. Brsg., Freiburg im Breisgau, Germany 253 p, 2003.
- Thoene, B., Schröder, P., Papen, H., Egger, A. and Rennenberg, H.: Absorption of atmospheric NO₂ by spruce (*Picea abies* L. Karst.) trees: I. NO₂ influx and its correlation with nitrate reduction, New Phytologist **117**, 575-585, 1991.
- Thoene, B., Rennenberg, H. and Weber, P.: Absorption of atmospheric NO₂ by spruce (*Picea abies*) trees: II. Parameterization of NO₂ fluxes by controlled dynamic chamber experiments, New Phytologist **134**, 257-266, 1996.

- Thomas, C. and Foken, T.: Flux contribution of coherent structures and its implications for the exchange of energy and matter in a tall spruce canopy, Boundary-Layer Meteorology **123**, 317-337, 2007.
- Tischner, R.: Nitrate uptake and reduction in higher and lower plants, Plant, Cell and Environment **23**, 1005-1024, 2000.
- Trebs, I., Bohn, B., Ammann, C., Rummel, U., Blumthaler, M., Koenigstedt, R., Meixner, F.X., Fan, S. and Andreae, M.O.: Relationship between the NO₂ photolysis frequency and the solar global irradiance, Atmospheric Measurement Techniques 2, 725-739, http://www.atmos-meas-tech.net/2/725/2009/, 2009.
- Vallano, D.M. and Sparks, J.P.: Quantifying foliar uptake of gaseous nitrogen dioxide using enriched foliar δ^{15} N values, New Phytologist **177**, 946-955, 2008.
- Walton, S., Gallagher, M.W., Choularton, T.W. and Duyzer, J.: Ozone and NO₂ exchange to fruit orchards, Atmospheric Environment **31**(17), 2767-2776, 1997.
- Wang, Y., Jacob, D.J. and Logan, A.: Global simulation of troposheric O₃-NO_x-hydrocarbon chemistry 1. Model formulation, Journal of Geophysical Research **103**(D9), 10713-10725, 1998.
- Warneck, P.: Chemistry of the Natural Atmosphere, San Diego, New York, Boston, Academic Press Inc, 1988.
- Weber, P., Nussbaum, S., Fuhrer, J., Gfeller, H., Schlunegger, U.P., Brunold, C. and Rennenberg, H.: Uptake of atmospheric ¹⁵NO₂ and its incorporation into free amino acids in wheat (*Triticum aestivum*), Physiologia Plantaru **94**, 71-77, 1995.
- Weber, P. and Rennenberg, H.: Dependency of nitrogen dioxide (NO₂) fluxes to wheat (*Triticum aestivum* L.) leaves from NO₂ concentration, light intensity, temperature and relative humidity determined from controlled dynamic chamber experiments, Atmospheric Environment **30**(17), 3001-3009, 1996a.
- Weber, P. and Rennenberg, H.: Exchange of NO and NO₂ between wheat canopy monoliths and the atmosphere, Plant and Soil **180**, 197-208, 1996b.
- Wellburn, A.R.: Why are atmospheric oxides of nitrogen usually phytotoxic and not alternative fertilizers, New Phytologist **115**, 395-429, 1990.
- Wendel, A., Stedman, D.H., Cantrell, C.A. and Damrauer, L.D.: Luminol-based nitrogen oxide detector. Analytical Chemistry **55**, 937-940, 1983.
- Wildt, J., Kley, D., Rockel, A., Rockel, P., Segschneider, H.J.: Emission of NO from several higher plant species, Journal of Geophysical Research **102**(D5), 5919-5927, 1997.
- Williams, E.J., Hutchinson, G.L. and Fehsenfeld, F.C.: NO_x and N₂O emissions from soil, Global Biogeochemical Cycles **6**, 351-388, 10.1029/92gb02124, 1992.

- Winer, A.M., Peters, J.W., Smith, J.P. and Pitts, J.N.: Response of Commercial Chemiluminescent NO-NO₂ Analyzers to other Nitrogen-Containing Compounds, Environmental Science & Technology **8**, 1118-1121, 1974.
- Yienger, J.J. and Levy II, H.: Empirical model of global soil-biogenic NO_x emissions, Journal of Geophysical Research **100**(D6), 11, 447–11, 464, 1995.
- Yoneyama, T., Ito, O., Engelaar, W.M.H.G.: Uptake, metabolism and distribution of nitrogen in crop plants traced by enriched and natural ¹⁵N: Progress over the last 30 years, Phytochemistry Reviews **2**, 121-132, 2003.
- York, D., Evensen, M., Lopez Martinez, M., De Basabe Delgado, J., Unified equations for the slope, intercept, and standard errors of the best straight line, American Journal of Physics **72**(3), 367-375, 2004.

CURRICULUM VITAE | 159