

**Proteomic Analysis
of Ocular Surface Components
by use of HPLC based
Mass Spectrometric Strategies**

Dissertation
zur Erlangung des Grades
„Doktor der Naturwissenschaften“
am Fachbereich Biologie der
Johannes Gutenberg-Universität
in Mainz

von
Sebastian Funke
geboren in Iserlohn

Mainz, 15.09.2014

Dekan:

1. Berichterstatter:

2. Berichterstatter:

Tag der mündlichen Prüfung: 20.04.2015

Annotation

Parts of the thesis have been published in international journals and/or presented on international conferences

Publications related to the thesis

Funke, S., Azimi, D., Wolers, D, Grus, F. H, Pfeiffer, N. 2012. Longitudinal analysis of taurine induced effects on the tear proteome of contact lens wearers and dry eye patients using a RP-RP-Capillary HPLC-MALDI TOF/TOF MS approach. *Journal of Proteomics* 75: 3177-3190

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Bell, K., Wilding, C., **Funke, S.**, Pfeiffer, N., Grus, F. H. 2014. Protective effect of 14-3-3 antibodies on stressed neuroretinal cells via the mitochondrial pathway. *BMC Neuroscience*, submitted

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Publications related to the thesis (in preparation)

Funke, S., Bell, K., Wilding, C., Wolters, D., Pfeiffer, N., Grus, F. H. Taurine effects on the conjunctival cell proteome.

Funke, S., Bechter, L., Perumal, N., Wolters, D., Pfeiffer, N., Grus, F. H. Proteomic analysis of glaucomatous human trabecular meshwork.

Funke, S., Perumal, N., Gabelt-Scheurich, S., Gramlich, O., Beck, S., Boehm, N., Wolters, D., Pfeiffer, N., Grus, F. H. Proteomic mass spectrometric analysis of the human glaucomatous retina.

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Perumal, N., **Funke, S.**, Wolters, D., Pfeiffer, N., Grus, F. H. Identification of human tear fluid biomarkers in different dry eye disease subgroups using label-free quantitative proteomics.

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Sicca-Förderpreis des Ressorts Trockenes Auge und Oberflächenerkrankungen im Berufsverband der Augenärzte Deutschlands 2013. Taurine induced effects on tear film and ocular surface revealed by mass spectrometric based proteomics. Berlin, Germany. 111th *DOG Excellent Vision-Vision of Excellence Conference 2013*

Acknowledgements

First, I would like to thank my supervisor for the opportunity to realize the thesis in the *Experimental Ophthalmology* lab of the University Medical Center Mainz and for his help, support and scientific experience he shared with me. I also thank my co-supervisor for his support.

Furthermore, I would like to thank my colleagues for their contribution to the realization of this work.

Special thanks go to my wife and families for their help and support over the past years.

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1 Purpose

The ocular surface is a complex functional unit, not completely understood, especially focusing on the proteome. Dry eye disease as a multifactorial disorder directly impacts the status of the ocular surface encircling tear film and cellular components, reflected by quantitative and qualitative changes in the proteome. Therefore, in this work, an intensive proteomic characterization of ocular surface components with special focus on tear film and ocular surface epithelial cells should be realized. Beside explorative analysis of the ocular surface proteome, two therapeutic strategies should be determined, encircling the influence of taurine eye drops on the tear film proteome of dry eye patients and contact lens wearers as well as the influence on the ocular surface of primary open angle glaucoma (POAG) patients switching from a common topical anti-glaucoma medication (Xalatan[®]) to a preservative-free formulation (Taflotan[®] sine) in terms of ocular surface complications. Accordingly, for “in depth” proteomic analysis of ocular surface components, HPLC based mass spectrometric workflows should be developed, optimized and compared. Results should provide a detailed proteomic view to the ocular surface and uncover underlying mechanism and potentials in the two focused therapeutic interventions, taurine and Taflotan[®] sine appliance.

2 Introduction

2.1 Mass spectrometry (MS) based proteomics

Proteomics first drafted by the groups of Wilkins and Wasinger (Wasinger *et al.*, 1995, Wilkins *et al.*, 1996a) referring to “PROTein complement of the genOME” (Apweiler *et al.*, 2009, Kraj and Silberring, 2008, Wilkins *et al.*, 1996b) describes the analysis of all proteins in living systems (Guerrera and Kleiner, 2005). Displaying high dynamics, (Kettman *et al.*, 2001) (Fig.1) a highlighted aim in proteomic science is the illumination of molecular changes in the course of a disease with the aim to find specific disease-related proteins indicating early diagnosis, molecular targeting or monitoring of therapeutic success (Etzioni *et al.*, 2003, Kraj and Silberring, 2008, Lescuyer *et al.*, 2007, Rifai *et al.*, 2006). MS evolved as the proteomic method of choice starting up from the 1980s demonstrating sensitivity and accuracy towards proteomic investigations (Mann *et al.*, 2001b). Generally, the first step of a proteomic workflow is to extract proteins or peptides from their sample source using various homogenization and extraction techniques (Chertov *et al.*, 2004, Cox and Emili, 2006, Ferro *et al.*, 2000, Goklen and Hatton, 1987, Holmquist and Carlson, 1977, Kushnirov, 2000, Molloy *et al.*, 1999, Naoe *et al.*, 1999, Naoe *et al.*, 1998, Selber *et al.*, 2004). Thus, for MS prior to protein/peptide ionization complexity reduction is mandatory to avoid masking effects by high abundant components (Granger *et al.*, 2005) and to reduce ion suppression provoked by salts, detergents and coextracted contaminants (Annesley, 2003, Hirabayashi *et al.*, 2007, Jessome and Volmer, 2006, Piwowar *et al.*, 2009, Remane *et al.*, 2010b, Wang *et al.*, 2003). Therefore, MS compatible fractionation (Issaq, 2001, Issaq *et al.*, 2005) or depletion (Bjorhall *et al.*, 2005, Magagnotti *et al.*, 2010) strategies have been developed. After sample processing, components are ionized and introduced to MS instruments showing a common fundamental design consisting of ion source, optics, mass analyzer and data processing electronics (Guerrera and Kleiner, 2005, Yates *et al.*, 2009). In case of proteomics soft ionization with low internal intensities leading to low ion fragmentation is usually coupled to mass analyzers (Daniel *et al.*, 2002, Pappin *et al.*, 1993). The two main soft ionization techniques are matrix assisted laser desorption ionization (MALDI) developed by Karas and Hillenkamp (Karas and Hillenkamp, 1988), in parallel by Tanaka and colleagues (Tanaka *et al.*, 1988) and electrospray ionization (ESI) invented by Fenn and colleagues (Fenn *et al.*, 1989, 1990) (Fig 2). MALDI features analyte co-crystallization with UV-absorbent small organic acids (Bird *et al.*, 2002, Canas *et al.*, 2007) and high vacuum pulsed laser evaporation leading to matrix and analyte desorption (Bahr *et al.*, 1992, Erra-Balsells and Nonami, 2003, Koubenakis *et al.*, 2004). The matrix transfers and receives protons and is believed to absorb energy as well as to isolate analytes from each other (Canas *et al.*, 2007). Usually, MALDI generates singly charged ions in the form $[M+H]^+$ or $[M-H]^+$ (Karas *et*

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al., 2000b, Yates *et al.*, 2009). In contrast ESI features ionization by application of high voltage (2-6kV) to a soluble analyte passing a narrow capillary which leads to the formation of an electrically charged spray, Taylor cone (Yates *et al.*, 2009) resulting in micro-drop evaporation and release of multiply charged analyte ions (Fenn *et al.*, 1990, Whitehouse *et al.*, 1985, Yates *et al.*, 2009). However, ESI and MALDI have different properties, ionization characteristics, sensitivity values and compatibilities. Importantly, MALDI is used in the off-line mode decoupling sample processing and MS analysis (Baessmann *et al.*, 2005, Mitulovic and Mechtler, 2006, Murray, 1997, Rejtar *et al.*, 2002, Stevenson and Loo, 1998) with respect to some exceptions (Brivio *et al.*, 2002, Musyimi *et al.*, 2005, Musyimi *et al.*, 2004, Preisler *et al.*, 1998). In contrast ESI is an on-line technique (He, 2000, Wang *et al.*, 2006, Xiang *et al.*, 1999). Pulsed MALDI sources are usually combined with time-of-flight (TOF) analyzers discriminating ions based on their time of flight in a field-free tube (Guerrera and Kleiner, 2005, Yates, 2000). In contrast continuous ESI sources are commonly coupled to ion traps (Guerrera and Kleiner, 2005). Due to their complementary character, MALDI and ESI are proposed to be used in combination (Bodnar *et al.*, 2003, Mo and Karger, 2002, Stapels and Barofsky, 2004, Stapels *et al.*, 2004). Regarding mass analyzers two fundamental instrument types exist: scanning/ion beam instruments [TOF, quadrupol (Q)] and traps [ion trap (IT), Orbitrap, *Fourier* transform ion cyclotron resonance (FT-ICR)] (Yates *et al.* 2009). Accordingly ion separation corresponds to time of flight, quadrupol electric field selection, selective ion ejection from a three-dimensional trapping field (Mann *et al.*, 2001a) and orbital ion trapping (Scigelova and Makarov, 2006). Generally two types of information are obtained by MS, accurate mass and amino acid sequence (Yates, 2000). To achieve information on the amino acid sequence, parent ions entering the mass analyzer can be further fragmented using different techniques, e.g. post source decay (PSD), collision induced decay (CID), high energy C-trap dissociation (HCD), electron capture dissociation (ECD) (Mo and Karger, 2002, Olsen *et al.*, 2007, Suckau *et al.*, 2003) or electron transfer dissociation (ETD) (Mikesh *et al.*, 2006, Syka *et al.*, 2004). The fragments can be measured with higher resolution e.g. in TOF/TOF analyzers (Medzihradszky *et al.*, 2000) or in hybrid instruments like QTOF or Orbitrap analyzers (Hardman and Makarov, 2003, Hu *et al.*, 2005). Method selection strongly depends on the technical features of a mass analyzer. PSD is a common technique in MALDI TOF/TOF MS/MS workflows (Medzihradszky *et al.*, 2000) featuring the metastable fragmentation of peptides by single or double cleavage of the peptide backbone (Suckau *et al.*, 2003). CID commonly features trap instruments inducing similar fragmentation at the N-O peptide bond leading to the generation of *b*- and *y*-ions with respect to amino acid side chains (Papayannopoulos, 1995, Schlosser and Lehmann, 2000) by collision with gas molecules such as argon or helium (Hoteling *et al.*, 2003). However, beside the heterogeneity of mass analyzers and fragmentation workflows, in this work special focus is

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laid on two instrument types: a MALDI-TOF/TOF and an ESI LTQ Orbitrap hybrid instrument. The application of both approaches towards the ocular surface proteome in association to dry eye related phenomena and potential therapeutic approaches were addressed in particular. Advantages, limitations and also the complementary character of both techniques were also examined.

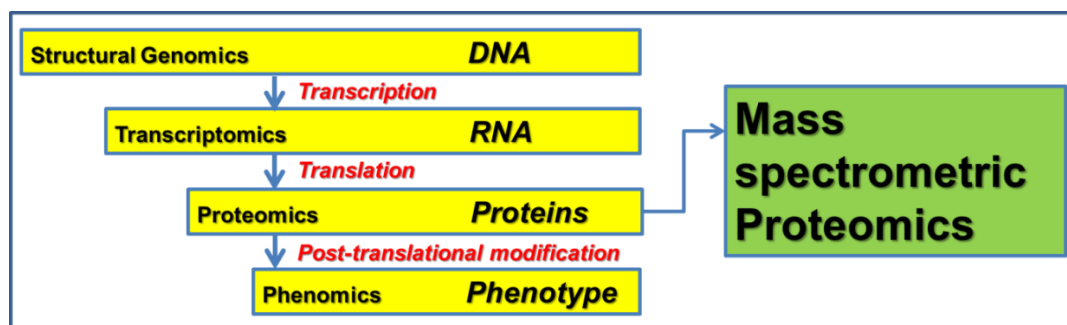


Fig 1 Hierarchical principle of proteomics science linked to mass spectrometry (illustrated according to Kraj and Silberring 2008).

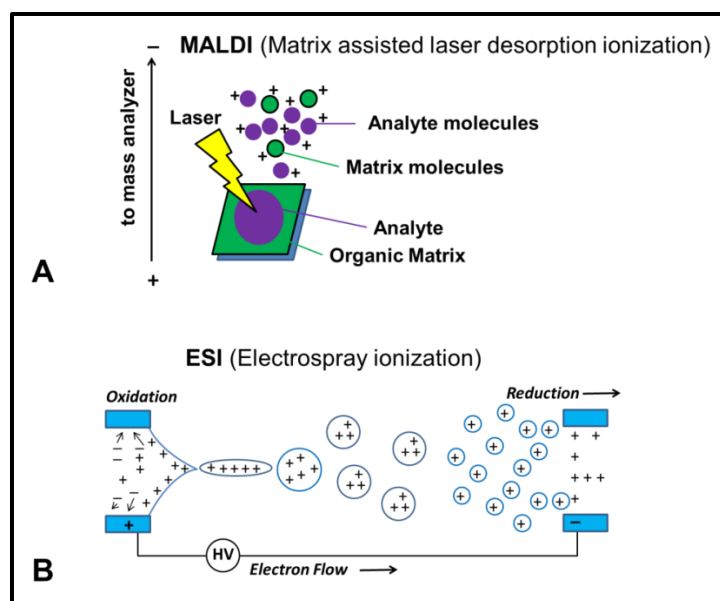


Fig 2 Soft ionization principles for MS based proteomics; A. MALDI: After analyte/matrix cocrystallization analyte molecules are ionized and vacuum evaporated by laser beam and enter the mass analyzer. B: ESI: Analyte solution passes an electric field leading to Taylor cone formation, charged droplet release and multiply charged analyte ion transfer to the mass analyzer (illustrated according to Kraj and Silberring 2008; www.intechopen.com).

2.2 Bottom Up (BU) vs. Top Down (TD)

Two major MS proteomic categories coexist: “Top Down” (TD) and “Bottom Up” (BU). TD means the MS introduction of sample ions derived from intact non-digested components (Reid and McLuckey, 2002), whereas in BU proteins are first enzymatic digested followed by MS analysis. Accordingly, the categoric definition refers to the entities introduced to an

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analyzer, which is in agreement with the majority of researchers (Capriotti *et al.*, 2011, Reid and McLuckey, 2002) based on the concept of McLafferty and colleagues (Kelleher *et al.*, 1999). However, there is growing confidence that a combination of both approaches will be most effective (Mo and Karger, 2002). Regarding TD a clear benefit of MALDI is the high analysis speed and contaminant robustness playing a predominant role in fast and direct experiments of crude sample materials and the possibility of storage and reanalysis (Frohlich and Arnold, 2006, Mukhopadhyay, 2005). In both approaches, there is a certain need of sample precleaning and purification to increase sensitivity, especially regarding low abundant components (Kraj and Silberring, 2008) desirable for their potential to predict health status or treatment success (Chertov *et al.*, 2005, Veenstra *et al.*, 2005). Regarding fractionation and separation methods applicable to recover low abundant intact peptides/proteins there is still a lack of development in the proteomics research field (Capriotti *et al.*, 2011). Therefore, several complexity reducing TD approaches have been addressed, e.g. solvent precipitation (Chertov *et al.*, 2004, Chertov *et al.*, 2005), filter size exclusion (Berven *et al.*, 2007, Wang *et al.*, 2003), subcellular fractionation (Cox and Emili, 2006, Huber *et al.*, 2003, Jiang *et al.*, 2004, Stasyk and Huber, 2004), liquid-liquid extraction (Du *et al.*, 2007, Kjellstrom and Jensen, 2003) or depletion of predominant proteins (Chromy *et al.*, 2004, Fratantoni *et al.*, 2010, Fu *et al.*, 2005, Yocum *et al.*, 2005). Regarding TD the special focus of the clinical proteomic biomarker research in the last decade was on solid phase extraction (SPE) methods, e.g. agarose/polyacrylamide bead based affinity chromatography (Cuatrecasas, 1970a, b, Nishikawa and Bailon, 1975), magnetic beads (Ketterlinus *et al.*, 2005, Rehm, 2006, van Hagen, 2008), which were successfully used to determine the human proteomic profiles of body fluids like serum (Baumann *et al.*, 2005, Jimenez *et al.*, 2007a), plasma (Zhang *et al.*, 2004b), saliva (Wu *et al.*, 2009b), urine (Fiedler *et al.*, 2007), cerebrospinal fluid (Jimenez *et al.*, 2007b) and tear fluid (Sekiyama *et al.*, 2008). A common TD strategy is surface enhanced laser desorption ionization (SELDI) ProteinChip[®] technology providing component enrichment on surface modified metal chips (Hutchens and Yip, 1993, Issaq *et al.*, 2002, Tang *et al.*, 2004). In spite of its promising use for proteomic analysis of body fluids (Issaq *et al.*, 2003, Jr *et al.*, 1999, Paweletz *et al.*, 2001, Petricoin *et al.*, 2004) SELDI was contrary discussed regarding reproducibility (Baggerly *et al.*, 2004, Bons *et al.*, 2006), robustness (Gast *et al.*, 2009) and sensitivity (Neuhoff *et al.*, 2004). Beyond technical aspects, high costs regarding single-use chips are disadvantageous (Rehm, 2006). In contrast, Callesen and colleagues reviewed the MALDI and SELDI platform regarding *m/z* peak biomarker candidate establishment in ovarian cancer and reported high reproducibility across different research groups and pre-fractionation methods (Callesen *et al.*, 2012, Callesen *et al.*, 2008) which gives new input to the debate of SELDI application to clinical TD proteomics. However, sensitivity and reproducibility of the mentioned approaches is very low

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compared to the abilities of high pressure liquid chromatography (HPLC). The use of LC MALDI for TD proteomic profiling is limited to a few publications. Wall *et al.* described a TD LC MALDI approach for the proteomic mapping of whole *E.coli* cell lysates whereby they first collected LC fractions in solution followed by a transfer to MALDI targets (Wall *et al.*, 1999). Koomen and colleagues investigated on the intact low molecular weight human serum proteome <5500 *m/z* by use of a LC MALDI TOF approach (Koomen *et al.*, 2005). Hu *et al.* used LC MALDI for oral cancer biomarker screening of saliva samples (Hu *et al.*, 2007). Thus, in contrast to bead or chip based approaches regarding ocular sample species there is still a lack of publications highlighting TD LC MALDI proteomics. Furthermore, most of the reported studies refer to in-solution fraction collection and direct on target collection approaches automated by robotic units have been attempted (Mukhopadhyay, 2005). Beside TD approaches to get a deeper view to the proteome BU workflows are a promising toolbox for clinical research. BU experiments focus on complex enzymatic peptide mixtures (Duncan *et al.*, 2010) challenging for MS performance due to contaminants, ion suppression and masking effects (Annesley, 2003, Gumerov *et al.*, 2002, Gustavsson *et al.*, 2001, Remane *et al.*, 2010a, Sjö Dahl *et al.*, 2005). Therefore, like in TD prefractionation, methods have to be integrated at protein level, at peptide level or at both stages. One usual strategy is to fractionate proteins in the first dimension by one-dimensional (1D) or two-dimensional (2D) gel electrophoresis followed by targeted digestion of molecular weight areas, gel lanes or single spots (Fountoulakis and Langen, 1997, Rosenfeld *et al.*, 1992). In contrast, multidimensional protein identification technology (Mudpit) highlights the gel-free digestion and fractionation of complex protein extracts by use of multidimensional HPLC systems (Chen *et al.*, 2006, Kislinger *et al.*, 2005, McDonald *et al.*, 2002). In fact, peptides generally cope better with chromatography and MS than proteins regarding solubility, ionization and fragmentation (Duncan *et al.*, 2010). Despite its lab intensive character, a certain advantage of gel-based systems is the effective removal of contaminants through electrophoresis (Lu and Zhu, 2005) taking advantage of the reproducibility of precast gels (Berkelman *et al.*, 2010, Herbert *et al.*, 2001). Furthermore, the gel technology itself allows quantification, for example realized by scanning densitometry (Claeys *et al.*, 1995, Ng *et al.*, 2000b) as well as control steps for method development. In summary, strategy selection, BU or TD strongly depends on research questions and sample type. The evaluation of both strategies for the analysis of ocular components is featured in this work.

2.3 MALDI-TOF-(TOF) MS (MS)

MALDI-TOF MS provides ion mass-to-charge ratio recording flight time in a field-free vacuum adjusted metal tubing of specific length (Domon and Aebersold, 2006) after source ionization considering faster travel of low *m/z* than high *m/z* molecules (Rehm, 2006). Resolution of

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TOF analyzers is limited but can be increased with tubing elongation (Schiller *et al.*, 2007). By coupling two TOF analyzers in tandem, PSD or CID fragments can be interpreted with higher resolution (Medzihradzky *et al.*, 2000). In these approaches a whole sample MS spectrum is generated in the first run, whereas in a follow-up run parent peaks corresponding to a mass list of the MS spectrum as well as their fragments are selected by a voltage dependent ion gating occurring in the precursor ion selector (PCIS) due to their flight time (Rehm, 2006, UltraflexTOF/TOFOperatorManual, 2001). Resolution is additionally increased by an “electrostatic mirror”, the reflector (Schiller *et al.*, 2007), which turns on ions compensating for small differences in their initial kinetic energies due to adsorbed initial laser energy (Aebersold and Mann, 2003, Rehm, 2006). This consequently results in higher resolution generating *Gaussian* shaped peak pattern allowing exact mass determination of isotopic peaks in MS/MS spectra (Suckau *et al.*, 2003). MS spectra can be used for simple peptide mass fingerprinting (PMF) via data bases, whereas MS/MS spectra give more additional information on the amino acid sequence in more complex samples (Kraj and Silberring, 2008, van Hagen, 2008). However, since the reflector works only for ions within a specific energy potential range, the analytic performance is usually limited to small components <4000 Da (Rehm, 2006) (UltraflexTOF/TOFOperatorManual, 2001). Decreased ion transmission to the detector in the reflector mode (Hortin, 2006) is the main reason for the analysis of intact peptides or proteins in the instruments linear mode without reflector. Accordingly, the drawback of lower resolution in the linear mode is balanced by its advantage of higher sensitivity. In this work, an Ultraflex II MALDI-TOF/TOF MS instrument was used (Fig. 3A). The instrument was developed by Suckau and coworkers and features a special “LIFT” technique (Fig. 3B) to select and fragment peptide ions overcoming the adjustment of different reflector potentials acquiring full fragment ion spectra within one single scan by kinetic energy adaption between parent ion and fragment ions (Rehm, 2006, Suckau *et al.*, 2003) (UltraflexTOF/TOFOperatorManual, 2001). First, parent ions and PSD fragments are selected by use of PCIS. Then, their kinetic energy levels are raised in the LIFT chamber, consisting of four electrodes building three chambers, by a two-step 19kV potential increase. Thereby, energy levels of parent ion and fragments are adapted to a difference of less than 30% and can for this reason be simultaneously reflected allowing their simultaneous detection in a single scan (Suckau *et al.*, 2003, UltraflexTOF/TOFOperatorManual, 2001). Whereas the Ultraflex series was successfully applied for protein profiling and/or identification in numerous proteomic studies of various cancer forms (de Noo *et al.*, 2006, Ebert *et al.*, 2006, El Ayed *et al.*, 2010, Feuerstein *et al.*, 2005, Freed *et al.*, 2008, Gamez-Pozo *et al.*, 2009, Groseclose *et al.*, 2008, Huang *et al.*, 2006, Kojima *et al.*, 2008, Lemaire *et al.*, 2007, Mustafa *et al.*, 2007, Oppenheimer *et al.*, 2010, Villanueva *et al.*, 2004), the implementation to ocular proteomic studies encircling pseudoexfoliation (Hardenborg *et al.*,

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2009, Joachim *et al.*, 2007a), glaucoma (Brust *et al.*, 2008, Grus *et al.*, 2008, Joachim *et al.*, 2007b, Joachim *et al.*, 2009, Joachim *et al.*, 2008, Joachim *et al.*, 2010) or eye surface diseases (Acera *et al.*, 2011, Kramann *et al.*, 2011) is still expandable. In this work the instrument is used as for TD as well as for BU analysis.

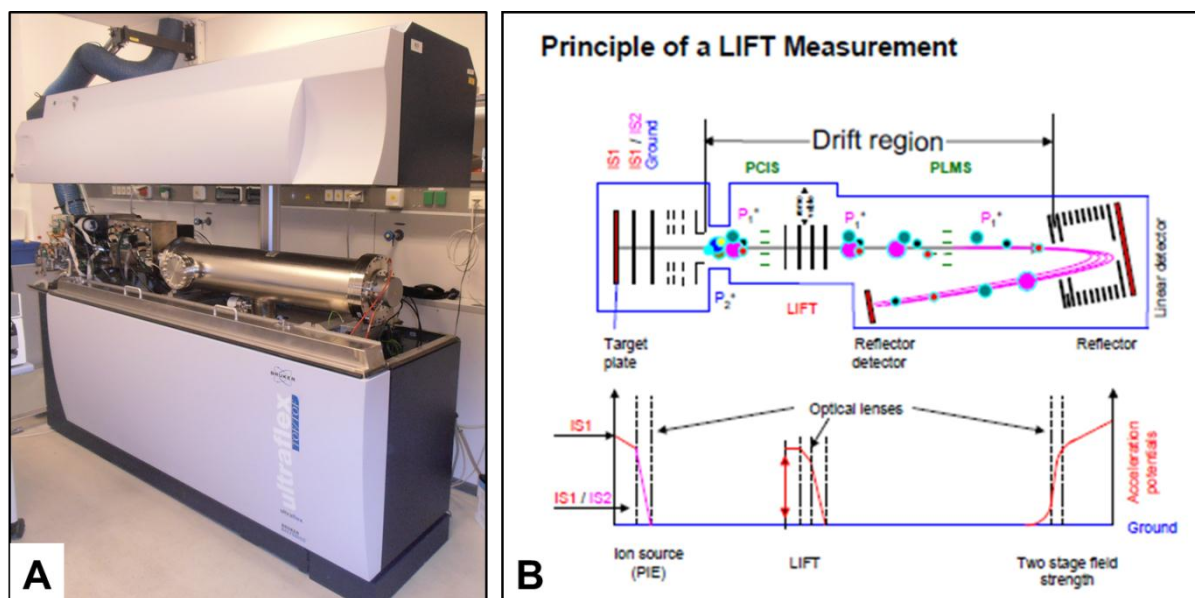


Fig 3 Principle of the Ultraflex II MALDI TOF-TOF MS instrument; A: Panorama view of the instruments interior containing MALDI source at the left side and flight tube on the right side of the machine; B: Ions passing the flight tube are compensated for kinetic energy differences by the reflector leading to increased resolution. Kinetic energy differences of precursor masses and corresponding fragment ions are approximated to each other in a LIFT chamber allowing their simultaneous detection.

2.4 LTQ-Orbitrap XL MS

The LTQ Orbitrap XL is a “hybrid” instrument combining a linear ion trap (LTQ) with an orbitrap analyzer (Fig. 4). Development and proof-of-principle of the orbitrap analyzer was realized by Makarov (Makarov, 2000) following establishment six years later by Makarov and colleagues (Makarov *et al.*, 2006a, Makarov *et al.*, 2006b). In the hybrid instrument the LTQ is used to accumulate ions and transfer them via a linked quadrupole c-shaped ion trap (C-trap) to the orbitrap analyzer (Hager, 2004, Scigelova and Makarov, 2006). The orbitrap itself consisting of an inner and outer electrode generates a quadro-logarithmic electrostatic potential to trap introduced ions, oscillating harmonically in axial direction while orbiting around a central electrode whereby the oscillating frequency (50-150 kHz for 200-2000 m/z) corresponds to their m/z values and can be determined after *Fourier* transformation (Makarov, 2000, Makarov *et al.*, 2006a, Makarov *et al.*, 2006b, Marshall and Hendrickson, 2008, Scigelova and Makarov, 2006, Yates *et al.*, 2009). Highlights of the orbitrap analyzer are its high resolution (max.150.000), high mass accuracy (2-5 ppm), high dynamic range

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(>103) and a m/z range of 6000 (Scigelova and Makarov, 2006, Yates *et al.*, 2009). Actually, instrumental mass deviation can be decreased to <1 ppm by the introduction of “lock-mass” background signals for internal calibration (Olsen *et al.*, 2005, Scheltema *et al.*, 2008). Usually, the LTQ Orbitrap workflow features ion fragmentation in the LTQ ion trap allowing multiple fragmentation levels (MS_n) and generates full MS scans in the orbitrap taking advantage of the speed (3-5 spectra/s) and sensitivity of the ion trap considering the mass accuracy and resolution of the orbitrap (Scigelova and Makarov, 2006, Yates *et al.*, 2009). Moreover, ion trap and orbitrap work in parallel since the initial orbitrap high resolution spectrum is used to select precursors for fragmentation in the ion trap while high resolution orbitrap measurement is still in progress (Scigelova and Makarov, 2006). For fragmentation CID, HCD, ECD or in modified machines ETD can be chosen and combined in experimental designs. Numerous studies in different proteomic fields, especially cancer proteomics (Kim *et al.*, 2012, Kosanam *et al.*, 2011, Lu *et al.*, 2009, Saratsis *et al.*, 2012, Whelan *et al.*, 2009, Zeng *et al.*, 2011) have been done since its commercial introduction in 2005. Nevertheless, the number of ophthalmological studies run on the LTQ Orbitrap XL is still limited (Bennett *et al.*, 2011, de Souza *et al.*, 2006, Galiacy *et al.*, 2011, Srinivasan *et al.*, 2012). In this work the instrument is used for a BU workflow for the analysis of the ocular surface.

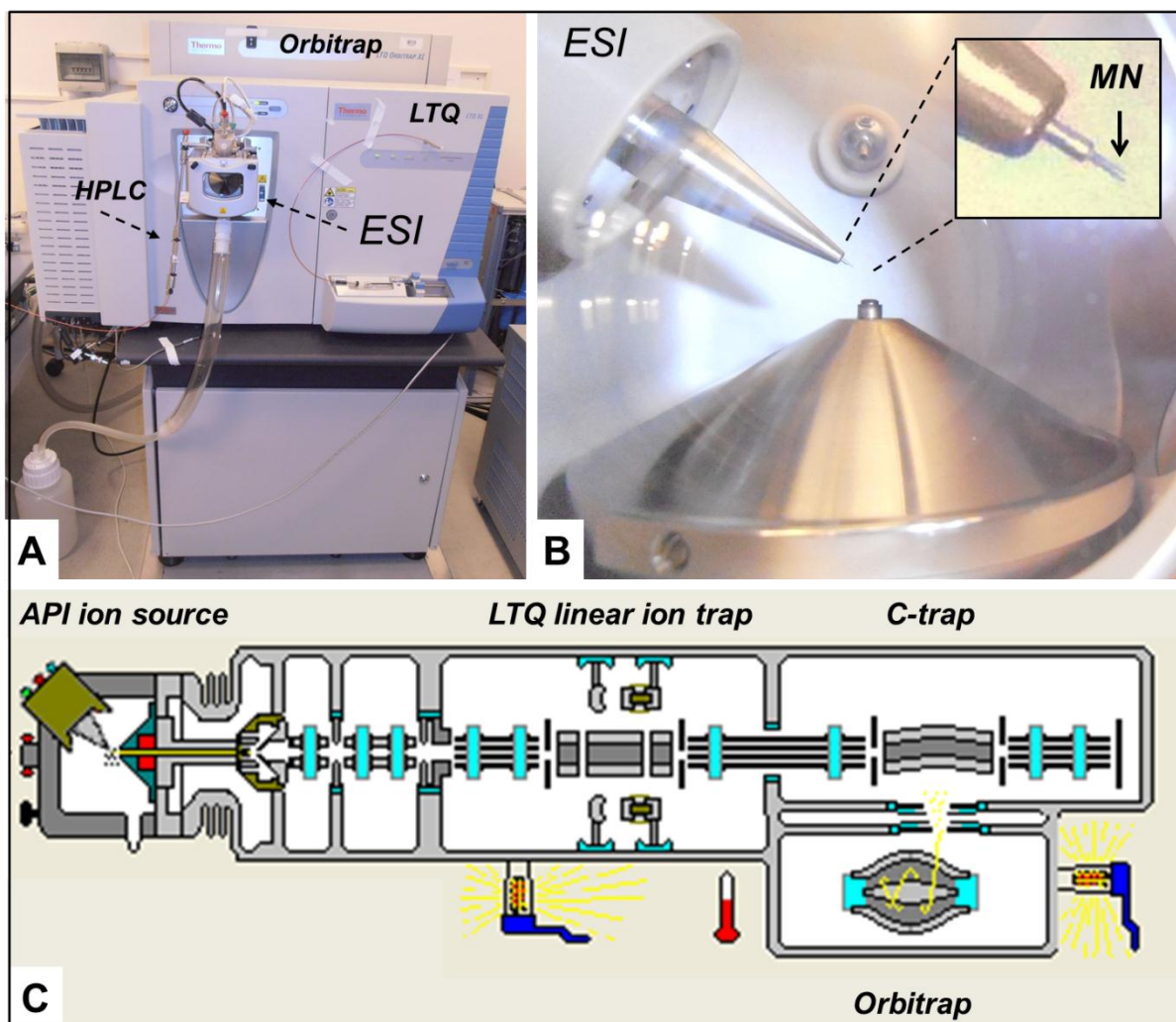


Fig 4 Principle of the LTQ Orbitrap XL instrument in LC ESI mode; A: Overview of the hybrid machine consisting of linear ion trap (LTQ) and orbitrap analyzer also indicating HPLC column system and ESI source; B: Detailed view of the ESI source featuring nozzle with integrated ESI metal needle (zoom view, arrow) and cone representing the heated capillary as mass analyzer entrance; C: Scheme of the hybrid instrument featuring ion trap for ion selection and fragmentation, C-trap for ion transfer and orbitrap for accurate mass analysis.

2.5 HPLC in mass spectrometry

HPLC approaches were developed to fractionate and purify samples by use of cylindrical tubing stationary phases encircling various retention modes (reversed phase, normal phase, size-exclusion, ion exchange, affinity) (Issaq, 2001, Mitulovic and Mechtler, 2006) based on porous, non-porous or superficial porous bead-shaped (Kirkland, 2000) or monolithic materials (Barroso *et al.*, 2003, Maruska *et al.*, 2007, Zou *et al.*, 2002a) prior to MS (Kraj and Silberring, 2008) (Examples of basic HPLC principles are given in Fig. 5). Reversed phase (RP) approaches separating molecules based on their hydrophobicity are very common in proteomics since most buffers are MS compatible (Yates *et al.*, 2009). Column parameters, e.g. inner diameter (ID), length, particle size, pressure, flow rate and capacity characterize the system directly affecting MS sensitivity (Rozing *et al.*, 2004). Column ID categories range

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from “preparative” (> 10 mm) over “narrow(small)-bore” ($4 \text{ mm} \geq \text{ID} \geq 2.1 \text{ mm}$) to “open tubular” ($< 25 \mu\text{m}$) (Roziing, 2003). However, in proteomics “microbore” ($2.1 \text{ mm} \geq \text{ID} \geq 1 \text{ mm}$), “capillary” ($1 \text{ mm} \geq \text{ID} \geq 100 \mu\text{m}$) and “nanobore” ($100 \mu\text{m} \geq \text{ID} \geq 25 \mu\text{m}$) are of common interest (Roziing, 2003) due to sensitivity increase by ID and flow rate scale-down (Frohlich and Arnold, 2006). Accordingly, microbore columns demonstrate a higher performance than “semi-preparative” columns (Bhown *et al.*, 1986, Bowermaster and Mcnair, 1983) and were early proposed for direct MS coupling (Garcia and Barcelo, 1993) allowing appropriate flow rates between 50 and 1000 $\mu\text{l}/\text{min}$ (Roziing, 2003). Nevertheless, in MALDI experiments ID and corresponding flow rates are not as crucial as in ESI workflows since fractions are collected off-line allowing complete solvent evaporation and analyte concentration on the target plate. In contrast, in on-line ESI approaches, column ID and flow rate will directly affect sample concentration introduced to the analyzer favoring down-scaled flow rates regarding ion formation (Karas *et al.*, 2000a). Despite increased sensitivity of nanobore systems (Mitulovic and Mechtler, 2006), disadvantages on the practical side addressing risk of column blocking, overloading, decreased column life times, consequent need for advanced sample pre-cleaning steps, extended analysis times due to nano flow rates, challenging pressure and dead volumes, advanced nano pump systems or split systems and higher costs have to be mentioned (Noga *et al.*, 2007). The use of HPLC techniques combined with gel-based or orthogonal chromatographic approaches linked to MS allowed the proteomic analysis of several ophthalmological relevant sample species like e.g. lens (Grey and Schey, 2009, Hoehenwarter *et al.*, 2005, Shearer *et al.*, 2008, Ueda *et al.*, 2002, Wilmarth *et al.*, 2009), optic nerve tissue (Bhattacharya *et al.*, 2006), optic nerve head astrocytes (Rogers *et al.*, 2012), microglia (Glanzer *et al.*, 2007), vitreous humour (Gao *et al.*, 2008, Kim *et al.*, 2007b, Ouchi *et al.*, 2005, Yu *et al.*, 2008), aqueous humor (Chowdhury *et al.*, 2010, Duan *et al.*, 2008, Funding *et al.*, 2005), sclera (Frost and Norton, 2007, 2012) and cornea (Dyrlund *et al.*, 2012, Galiacy *et al.*, 2011, Karring *et al.*, 2006, Karring *et al.*, 2005, Thompson *et al.*, 2003). Despite overall growing research attempts in ocular proteomics, there is still a high degree of necessity to use HPLC based methods for a more detailed basic appreciation of the eye surface realized in the present work. Accordingly, a brief introduction to achievements and limitations in ocular surface proteomics is given in the following chapters.

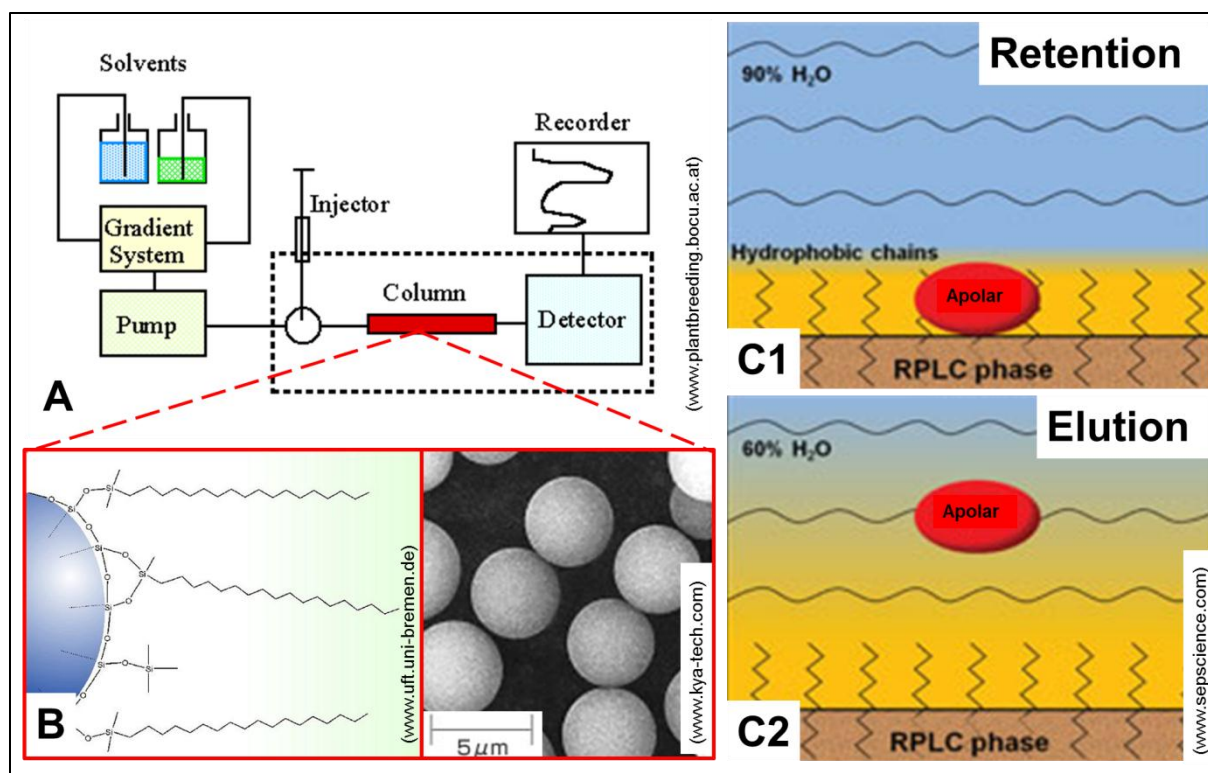


Fig 5 Principles of HPLC; A: Basic HPLC system buildup, in case of LC MS; detection and recording is realized by the MS instrument; B: Interior of a RP column system featuring the stationary phase consisting of microspheres with a functionalized alkyl chain surface for protein/peptide capture; C: Basic principle of RP retention facilitating component retention on the stationary phase under aqueous condition (C1) and component elution with increasing solvent portion of the mobile phase (C2).

2.6 Ocular surface proteomics

To realize the role of ocular surface proteomics, it is important to understand morphology and function of the eye surface encircling cornea, conjunctiva and tear film, as a direct environmental exposed organ, sensitive towards external and manipulative factors. The transparent, vessel-poor and immune-privileged six layer cornea (Dua *et al.*, 2013, Lang, 2008) contributes mainly to image focusing, whereas the conjunctiva is discriminated from the cornea by the *Limbus corneae* and represents a vessel and gland-rich mucous layer (Lang, 2008). Furthermore, stratified, wet surfaced corneal and conjunctival epithelia (Gipson and Argueso, 2003) are covered by the tear film (Kampik, 2002) protecting the integrity of sclera, cornea and conjunctiva as a functional unit, important for image focusing (Kampik, 2002, Tiffany, 2008) (Fig 6). The tear film consisting of three layers, outermost lipid layer, aqueous layer and mucus layer (Wolff, 1946, 1954) provides numerous functions: light refraction, eyelid lubrication, conjunctiva/cornea lubrication, nutrition, white blood cell access, outwash of foreign materials and antimicrobial defense (Davidson and Kuonen, 2004). The meibomian glands' secreted lipid layer (McCulley and Shine, 1997) is built by two sub layers [apolar lipid (meibomian oil layer) and a polar lipid sub layer (Shine and McCulley, 2003)],

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which are both associated to lipophil proteins like lipocalins (Glasgow *et al.*, 1995). Tear lipids are bound to proteins and protein-lipid interactions providing the functional integrity to the lipid layer protecting against fluid evaporation from the ocular surface (Bron *et al.*, 2004, Craig and Tomlinson, 1997). Tear mucins and also non-mucin proteins are known to lower tear surface tension assisting to spread the precorneal tear film (Lemp, 1973) for viscosity and stability control (Gouveia and Tiffany, 2005, Schoenwald *et al.*, 1997). The aqueous phase secreted by the lacrimal glands (Tiffany, 2008) includes electrolytes, amino acids, sugars, peptides and proteins (Dunson, 1970) highlighting gel-building mucins and glycoproteins (Holly and Lemp, 1977), which provide an optimal consistence of the tear fluid (Davidson and Kuonen, 2004). Beside pathogen defense by antimicrobial peptides (Haynes *et al.*, 1999), corneal nutrition is the predominant function of the aqueous layer (Chen *et al.*, 1997, Iwata, 1973). The glycocalyx consists mainly of glycoproteins and sol-building mucins, which are secreted by corneal epithelia while some mucin groups are membrane associated (Gipson, 1994, Gipson and Inatomi, 1998). Ocular glands, tear fluid and protein composition have been studied in several species like dog (Barrera *et al.*, 1992, Berger and King, 1998, de Freitas Campos *et al.*, 2008), cat (Petznick *et al.*, 2012, Petznick *et al.*, 2011), horse (Deeg *et al.*, 2006, Martin *et al.*, 1997, Parma *et al.*, 1987), camel (Chen *et al.*, 2011b, Shamsi *et al.*, 2011), rabbit (Zhou *et al.*, 2007), dolphin (Tarpley and Ridgway, 1991, Young and Dawson, 1992), crocodile (Dunson, 1970, Rehorek *et al.*, 2005, Schmidnielsen and Fange, 1958) and frog (Nowack and Wohrmann-Repenning, 2010). However, regarding the tear proteome the most intensive studies were applied on humans. Using MS, de Souza and colleagues reported 491 proteins encircling numerous proteases and protease-inhibitors (de Souza *et al.*, 2006), for example cystatin S demonstrated to inactivate cysteine proteinases (Barka *et al.*, 1991, Ghiso *et al.*, 1988). Zhou *et al.* could increase sensitivity, identifying 1543 tear proteins using a LC ESI Triple TOF 5600 system (Zhou *et al.*, 2012). Also, by targeted proteomic analysis of meibomian gland fluid 90 proteins have been revealed (Evans *et al.*, 2003, Tsai *et al.*, 2006). In fact, tear fluid is predominated by a small subset of high abundant proteins providing 70-80% of the whole protein content (Azzarolo *et al.*, 2004) including lysozyme, lactoferrin and lipocalin (Sack *et al.*, 2001). Furthermore, ceruloplasmin, Zinc- α -2-glycoprotein, secretory IgA, IgG, albumin and glycoproteins are detectable in moderate concentrations (Davidson and Kuonen, 2004, Green-Church and Nichols, 2008, Tiffany, 2008). In addition, tear proline-rich proteins have been characterized (Fung *et al.*, 2004). Most of the abundant tear proteins display antimicrobial activity like lysozyme (Jensen and Gluud, 1985, Selinger *et al.*, 1979), lipocalin (Fluckinger *et al.*, 2004), cystatine (Stoka *et al.*, 1995), secretory phospholipase A2 (Qu and Lehrer, 1998) and lactoferrin (Broekhuysse, 1974, Flanagan and Willcox, 2009, Jensen and Gluud, 1985, Selinger *et al.*, 1979). Deeper functions have been proposed for certain proteins, e.g. lipocalin including immunomodulation

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(Flower, 1996). A correlation between tear proteomic composition and the health status (Antoine *et al.*, 2010) has been demonstrated for ocular disorders, especially dry eye (Grus and Augustin, 1999, Grus *et al.*, 2005b, Li *et al.*, 2005, Versura *et al.*, 2010, Zhou *et al.*, 2009c), meibomian gland disease (Tong *et al.*, 2011), pterygium (Zhou *et al.*, 2009a) and glaucoma (Aho *et al.*, 2002, Pieragostino *et al.*, 2013, Pieragostino *et al.*, 2012, Roedl *et al.*, 2007, Roedl *et al.*, 2008), ocular affected Sjögren's syndrome (Caffery *et al.*, 2008, Tomosugi *et al.*, 2005b) or systemic diseases like cancer (Evans *et al.*, 2001, Lebrecht *et al.*, 2009, Streckfus *et al.*, 2006). There are many external factors that can influence the ocular surface including anti-glaucoma medication (Malvitte *et al.*, 2007, Wong *et al.*, 2011), use of contact lenses (Glasson *et al.*, 2006, Li *et al.*, 2005) and lens care solutions inducing proteomic changes (Green-Church and Nichols, 2008, Grus *et al.*, 2005a, Zhao *et al.*, 2009). In accordance, the tear proteome represents a rich source of markers to evaluate the ocular health status (von Thun Und Hohenstein-Blaul *et al.*, 2013) with special focus on cornea and conjunctiva (Macri and Pflugfelder, 2000, Pisella *et al.*, 2000). In terms of dry eye disease, several proteins are already proposed as "biomarkers". Despite numerous descriptions of the term (Hakansson, 2007, Strimbu and Tavel, 2010), a biomarker is commonly defined as "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention" (BiomarkersDefinitionWorkingGroup, 2001). Accordingly, biomarker proteins can be used in targeted monitoring to evaluate the ocular surface status (de Paiva *et al.*, 2013) encircling conjunctival impression cytology, immunocytological procedures, tear film glycomics, lipidomics and proteomics (Deschamps and Baudouin, 2013). In fact, tear proteins have been proposed as dry eye biomarkers including S 100A8, S100A9, lipocalin-1, secretory phospholipase A2, metalloproteinase 9, proline-rich protein 4, mammaglobin B and mucins as well as several cytokines and chemokines (Aluru *et al.*, 2012, Boehm *et al.*, 2013, Corrales *et al.*, 2011, Enriquez-de-Salamanca *et al.*, 2012, Kramann *et al.*, 2005, Zhou *et al.*, 2009b). However, there is still need for more investigation to explain tear proteomic changes related to ocular surface complications and to obtain a deeper view to the ocular surface proteome, especially under therapeutic condition. Therefore, in this work a HPLC based MS platform should be developed allowing in depth analysis of the ocular surface with special focus of topical appliance of taurine and Taflotan[®] sine.

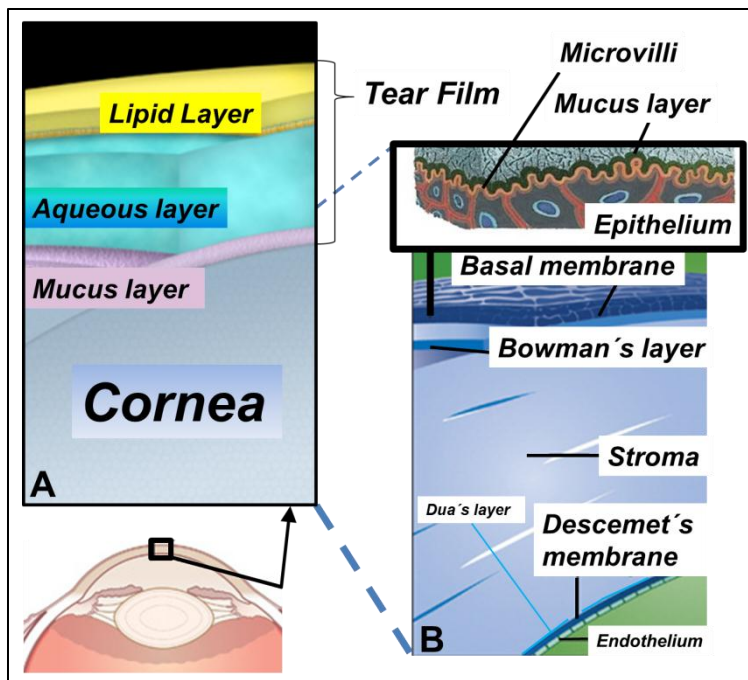


Fig 6 Constitution of A: Human tear film and B: Human cornea. The cornea epithelium containing microvilli entering the mucus layer are highlighted. (illustrated in accordance to Cornea research foundation of America: www.cornea.org; A. www.eyecalm.org, B. Morgan 2010, La Croix 2010)

2.7 Dry Eye: Causes and consequences

Directly affecting the ocular surface, dry eye disease also known as dry eye syndrome or keratoconjunctivitis sicca, is defined as a multifactorial disease complex in accordance to the International Dry Eye Workshop 2007 (Lemp and Foulks, 2008) (Fig. 7) with prevalences ranging from 7% (USA) over 11% (Spain) and 21% (China) to 33% (Taiwan, Japan) (Gayton, 2009, Jie *et al.*, 2009, Viso *et al.*, 2009). Dry Eye disease is characterized by various symptoms like ocular fatigue (Toda *et al.*, 1993), foreign body sensation, photophobia, burning (Lemp, 1995), irritation and blurred vision (Tutt *et al.*, 2000). Symptoms are correlated with tear film dysfunction, decreased cornea/conjunctiva lubrication (Rolando and Zierhut, 2001), tear film instability and hyperosmolarity resulting in a damage of the ocular surface (Lemp, 2008, Peng *et al.*, 2013, Sweeney *et al.*, 2013, Tomlinson *et al.*, 2006, Tomlinson *et al.*, 2010). Moreover, inflammatory processes are common dry eye features (Baudouin, 2001, Lemp, 2008). Regarding the manifestation of dry eye disease cornea, conjunctiva, accessory lacrimal glands, meibomian gland, tear film, lacrimal gland and nervous innervations build a functional unit and dysfunction in one or more components lead to dysbalances resulting in inflammatory processes and ocular surface damage (Stern *et al.*, 1998, Stern *et al.*, 2004) The tear film is one functional trigger point since excessive tear evaporation and/or reduced tear (Johnson and Murphy, 2004). Consequently, the disease manifests in a decline of image quality and vision (Goto *et al.*, 2002, Tutt *et al.*, 2000) and

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affects patients daily life quality (Deschamps *et al.*, 2013, Kim *et al.*, 2011, Li *et al.*, 2012, Miljanovic *et al.*, 2007, Miljanovic *et al.*, 2004, Nelson *et al.*, 2000, van Landingham *et al.*, 2013). Several risk factors have been documented, e.g. arthritis (McCarty *et al.*, 1998, Moss *et al.*, 2000), diabetes (Barcaroli *et al.*, 1997, Seifart and Stempel, 1994) (Barcaroli *et al.*, 1997, Kaiserman *et al.*, 2005), caffeine consumption, high-density lipoprotein cholesterol ratio (Moss *et al.*, 2000), smoking (Thomas *et al.*, 2012), use of video display terminals (Blehm *et al.*, 2005, Schlote *et al.*, 2004, Tsubota and Nakamori, 1993, Uchino *et al.*, 2013, Von Stroh, 1993) and general diet (Miljanovic *et al.*, 2005). Thereby, female gender, progressed age (Schaumberg *et al.*, 2003) as well as an ethnic bias have been reported as prevalences (Lee *et al.*, 2002) (Foong *et al.*, 2007, Lavanya *et al.*, 2009) (Guo *et al.*, 2010, Tran *et al.*, 2013). Furthermore, contact lens wearing was documented to induce dry eye with controversial findings regarding gender prevalence (Chalmers and Begley, 2006, Fonn *et al.*, 1995, Hikichi *et al.*, 1995, Mackie, 1985, Nichols and Sinnott, 2006, Young *et al.*, 2011). Hormonal changes (Rocha *et al.*, 2013, Sullivan, 2004, Sullivan *et al.*, 2002) and viral infections (Alves *et al.*, 2013, Kamoi and Mochizuki, 2012, Lemp, 2008, Lestari *et al.*, 2013) are validated as further risk factors. Moreover, clinical surgery like LASIK (laser in situ keratomileusis) (Donnenfeld *et al.*, 2003, Huang *et al.*, 2012, Toda, 2007, Toda *et al.*, 2001) can lead to dry eye and other eye diseases like macular edema or diabetic retinopathy (Manaviat *et al.*, 2008, Najafi *et al.*, 2013) were proposed for prevalence. Regarding ocular disorders topical medications represent risk factors for ocular surface complications since ophthalmic formulations commonly contain antimicrobial preservatives (Barkman *et al.*, 1969, Messmer, 2012, Olson and White, 1990, Tu *et al.*, 2013), also able to realize intraocular penetration (Messmer, 2012). Various preservatives are in use (Herrero Vanrell, 2007) displaying toxic qualities (Gasset *et al.*, 1974). Former used preservatives like neoplycine, cocaine and phospholine iodid lead to morphological changes of the ocular surface indicated by loss of peripheral microvilli and complete corneal layers (Pfister and Burstein, 1976). Quaternary ammoniums (Lapiere and Thunus, 1967), displaying a wide range of antimicrobial properties (Caillier *et al.*, 2009) are common in ophthalmic solutions (Herrero Vanrell, 2007). Among these chemicals, benzalkonium chloride (BAC) (Fig. 8) is widely distributed in eye drop formulations (Messmer, 2012). Especially, topical application of BAC containing intraocular pressure (IOP) reducing agents in anti-glaucoma therapy (Fig. 8) is known to elevate the risk of ocular surface complications in POAG patients (Arici *et al.*, 2000, Dejong *et al.*, 1994, Erb *et al.*, 2008, Jandrovic *et al.*, 2013, Noecker, 2001, Nordmann *et al.*, 2003, Steven and Cursiefen, 2013) with a prevalence of 42% (Stewart *et al.*, 2011) leading to “medication induced dry eye” (Fraunfelder *et al.*, 2012). De facto, long-term BAC utilization had been reported to alter the ocular surface (Arici *et al.*, 2000, Cho *et al.*, 2011, Yalvac *et al.*, 1995) supported by dose-dependent BAC toxicity on epithelia demonstrated by

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in-vitro and in-vivo studies (Baudouin *et al.*, 2007, Becquet *et al.*, 1998, Liang *et al.*, 2008, Whitson *et al.*, 2006, Yee *et al.*, 2006). In fact, BAC was found to lead to membrane damage of cornea epithelia while also rupturing tight junctions (Pfister and Burstein, 1976) and increasing corneal permeability (Ramselaar *et al.*, 1988). Tripathi and coworkers could demonstrate that 0.01% BAC caused cessation of cell movement, cytokinesis and mitosis followed by complete cell degeneration within 2-8 h (Tripathi *et al.*, 1992). Apoptotic processes were supported for lower BAC concentrations, whereas higher concentrations were reported to induce necrosis (Debbasch *et al.*, 2001). In accordance, BAC associated morphological and anti-proliferative changes on cells have been reported for glaucoma therapeutics (Seibold *et al.*, 2013). Moreover, in vivo studies showed negative BAC induced effects on tear fluid production (Kuppens *et al.*, 1995) and stability (Baudouin and de Lunardo, 1998, Wilson *et al.*, 1975). Most toxic effects are due to the chemical nature of BAC displaying detergent-like features, binding and altering the cellular membrane, inducing toxic processes in cells, e.g. through interaction with cellular proteins and precipitation of enzymes (Furrer *et al.*, 2002, Grant and Acosta, 1996, Herrero Vanrell, 2007). BAC induced declined membrane integrity leads to decreased viability in rabbit corneal cells (Georgiev, 2011). Strong BAC lipid interactions could be monitored and also perturbation of the tear film lipid layer has been observed (Georgiev, 2011). Besides, BAC was found to display inflammatory potential beyond toxicity, reflected by tolerance breakdown (Galletti *et al.*, 2013) and sensitizer and histamine releaser activity in allergic reactions (Weston and Assem, 1994). Accordingly, latanoprost containing BAC caused increase in eosinophiles in glaucoma patients (Costagliola *et al.*, 2001). In confidence inflammation was found to occur on conjunctival epithelial cells administered in the course of topical antiglaucoma monotherapy using latanoprost, betaxolol and timolol (Cvenkel and Ihan, 2002). However, there is still a need to investigate on proteomic ocular surface changes in the course of glaucoma medication. Monitoring of proteomic changes of the tear film can help to objectively estimate the health status of the ocular surface in POAG sicca patients building a fundament for new therapeutic strategies. Tear proteomic changes in POAG sicca, non-glaucoma sicca and contact lens wearers should be focused on in this work. Therefore, as a toolbox HPLC based MS methods should be developed for the detailed investigations on the ocular surface proteome.

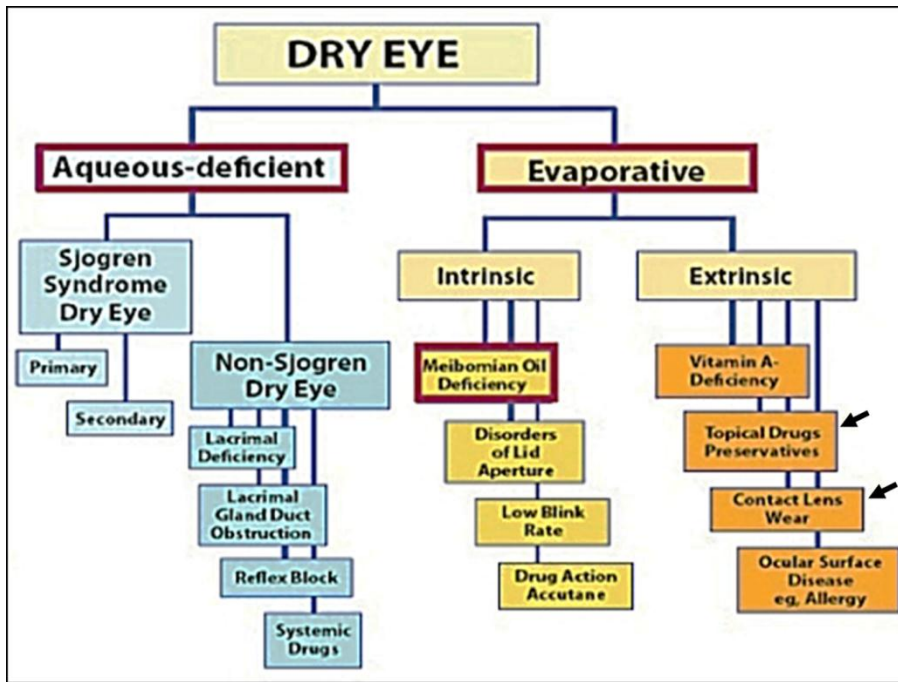


Fig 7 Classification of dry eye syndrome pathologies based on the concept of the 2007 Dry Eye Workshop (DEWS) (Lemp 2007). The arrow highlighted dry eye forms are especially focused on in the present work.

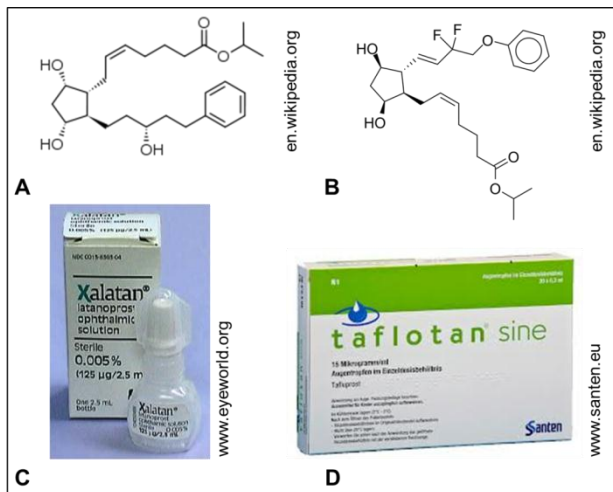


Fig 8 Prostaglandine related IOP reducing agents and commercial formulations used in POAG glaucoma therapy; A: Latanoprost; B: Tafluprost; C: Xalatan® (Pfizer) containing 0.005% latanoprost and 0.02% BAC; D: Preservative-free Taflotan® sine (Santen) containing 0.0015% tafluprost.

2.8 Therapeutic management with special focus on Taflotan® sine and taurine

According to the complex character of dry eye disease, new therapeutic strategies have to be addressed. Thereby, special focus is on “medication induced” dry eye (Fraunfelder *et al.*, 2012) in POAG therapy. Consequently, therapeutic key features should address preservative, especially BAC reduction, in topical formulation. Horsley & Kahook reported beneficial effects on the ocular surface in glaucoma patients switching from latanoprost (0.02% BAC) to sofZIA™ medication indicated by an amendment in ocular surface disease

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index (OSDI), corneal staining and tear break-up time (TBUT) (Horsley and Kahook, 2009). Because BAC displays toxic potential in very low concentrations (Furrer *et al.*, 2002), a complete avoidance of this preservative most likely is the aim for long term application. Nevertheless, common prostaglandine analogue formulations contain preservatives, e.g. latanoprost, travoprost or bimatoprost (Liu and Mao, 2013). In contrast tafluprost (AFP-168), the latest prostaglandin $F_{2\alpha}$ derivate (Takagi *et al.*, 2004), as the first IOP lowering agent available in a preservative-free formulation, evidenced promising results regarding effectivity and tolerability (Hamacher *et al.*, 2008, Hommer *et al.*, 2010, Hos *et al.*, 2013). In comparison to preservative-containing formulations tafluprost exhibited very low or no apoptotic/necrotic/oxidative effects revealed in IOBA-NHC cells (Brasnu *et al.*, 2008). Local tolerability was evaluated to improve up to 86% after a medical switch from preservative-based medication to tafluprost in a large patient cohort of 2123 patients (Erb *et al.*, 2011). Uusitalo and colleagues reported that tafluprost compared to latanoprost showed comparable results revealed from a study encircling 533 glaucoma patients (Uusitalo *et al.*, 2010). In confidence Janulevičienė and coworkers showed an equal IOP lowering effect but a higher ocular tolerability indicated by lower tear osmolarity levels for tafluprost compared with BAC containing latanoprost (Januleviciene *et al.*, 2012). Tafluprost available under the trade name Taflotan[®] sine (Santen Pharmaceutical Co., Ltd.), is in use in glaucoma therapy, but objective indicators for the effect of Taflotan[®] sine on the ocular surface are still missing and no detailed proteomic examination has been realized so far. Despite the preservative focused strategy in “medication induced dry eye” additional therapeutic strategies generally addressing dry eye had been proposed including tear secretagogues (Koh *et al.*, 2013, Nakamura *et al.*, 2012, Nichols *et al.*, 2004), anti-inflammatory topical therapeutics (McCabe and Narayanan, 2009, Mrukwa-Kominek *et al.*, 2007, Pflugfelder, 2003, 2004) like cyclosporine, tetracycline, doxycycline, corticosteroids, vitamin A topical utilization (Calonge, 2001, De Paiva *et al.*, 2006, Kim *et al.*, 2009a, Kobayashi *et al.*, 1997, Perry *et al.*, 2008, Yang *et al.*, 2006), cytokines (Pflugfelder *et al.*, 2013), anakinra (Vijmasi *et al.*, 2013), coumarin 18 (Govek *et al.*, 2010) and the utilization of artificial tears (Sindt and Foulks, 2013). However, there is still need to discover new drugs and to obtain a deeper understanding of proposed agents in dry eye therapy (Aquavella, 2013, Dogru *et al.*, 2013, Gadaria-Rathod *et al.*, 2013, Skalicky *et al.*, 2013). Thereby, topical or oral appliance of small receptor interacting molecules like CF101 (Avni *et al.*, 2010), essential fatty acids (Kokke *et al.*, 2008, Rashid *et al.*, 2008), peptides (Li *et al.*, 2013, Peral *et al.*, 2008, Sosne *et al.*, 2012) amino acids (Aragona *et al.*, 2013, Peral *et al.*, 2008) are among the future components for dry eye management. Regarding amino acids special focus is on taurine (Fig. 9), which was proposed as a promising agent in preventive medicine (Birdsall, 1998, Chesney, 1985, Kendler, 1989) and is still under investigation for its therapeutic potential facing ocular disorders (Ripps and

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Shen, 2012). Taurine is a multifunctional zwitterionic, sulfonated, non-proteinaceous β -amino acid (Huxtable, 1992, Redmond *et al.*, 1998, Wright *et al.*, 1986) derived from methionine and cysteine metabolism (Brosnan and Brosnan, 2006, Redmond *et al.*, 1998) and is present in high concentration in tissues of algae and animals (Jacobsen and Smith, 1968). Also, in mammalian species taurine has been documented for various tissues (Awapara *et al.*, 1950), with high concentrations in ocular tissues (Heinamaki *et al.*, 1986) including lens, iris ciliar body (Gupta and Mathur, 1983, Heinamaki *et al.*, 1986, Reddy, 1967, Reddy, 1970) and retina (Lake and Verdonessmith, 1989, Orr *et al.*, 1976, Pow *et al.*, 2002, Rassin *et al.*, 1978, Sturman *et al.*, 1978, Voaden *et al.*, 1977). Especially, on the ocular surface, high taurine abundance had been reported encircling cornea, conjunctiva and tear film (Nakatsukasa *et al.*, 2011, Pescosolido *et al.*, 2009, Reddy, 1967). Beyond metabolic functions including nutrition (Tesseraud *et al.*, 2009, Wu, 2009), more specialized features are associated to taurine, e.g. vitamin transport (Petrosian and Haroutounian, 2000) and osmoregulation (Gupta and Mathur, 1983, Lange, 1963, Thurston *et al.*, 1980, Uchida *et al.*, 1991). De facto, taurine was found to display anti-toxic cytoprotective features in host tissues (Dawson *et al.*, 2002, Lerdweeraphon *et al.*, 2013, Pasantes-Morales and Cruz, 1985, Saad and Al-Rikabi, 2002, Timbrell *et al.*, 1995, Venkatesan *et al.*, 1997, Waterfield *et al.*, 1994, Wettstein and Haussinger, 1997). Most of these effects are due to the ability of taurine to prevent oxidative stress and peroxidative damage (Cozzi *et al.*, 1995, Endo *et al.*, 2002, Mahalakshmi *et al.*, 2003, Nakamori *et al.*, 1993a, Pasantes-Morales and Cruz, 1984) by its membrane interacting and stabilizing features (Koyama *et al.*, 1996, Lopez-Colome and Pasantes-Morales, 1981, Pasantes-Morales and Cruz, 1985). Confidently, the amino acid has been demonstrated to protect plasma/mitochondrial and ER membranes under stress condition (Ye *et al.*, 2013) facilitating the interaction with lipids and proteins (Sebring and Huxtable, 1986), stabilizing proteins, especially membrane proteins from denaturation (Arakawa and Timasheff, 1985, Nandhini and Anuradha, 2003, Ye *et al.*, 2013). Interestingly, osmolytes like taurine have been lately proven to protect lysozyme as a major tear protein representative (Bruzdziaik *et al.*, 2013). Antioxidative effects of taurine have been intensively studied (Nakamori *et al.*, 1993a), whereby protective effects exceed effectivity of vitamin C and E (Rodriguez-Martinez *et al.*, 2004). Regulation of Ca^{2+} uptake, Ca^{2+} ATPase, generation of peroxide and superoxide anions in mitochondria (Chang *et al.*, 2004) as well as inhibition of consequent oxidative damage could be demonstrated for various tissues and organisms, e.g. for rat spermatozoa (Alvarez and Storey, 1983), rat liver (Kerai *et al.*, 1998), rat mesangial cells (Trachtman *et al.*, 1993), rat liver cells (Kerai *et al.*, 1998), plasma, liver and aorta of rabbits (Balkan *et al.*, 2002), hamster bronchioles (Gordon *et al.*, 1986) and human lymphoblastoid cells (Pasantesmorales *et al.*, 1984). Regarding ocular tissues, taurine was documented to prevent lipid peroxidation in rod outer segments (Pasantes-Morales, 1982,

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Pasantes-Morales and Cruz, 1985), photoreceptors (Keys and Zimmerman, 1999) and diabetic precataractous lens (Kilic *et al.*, 1999, Mitton *et al.*, 1999, Obrosova and Stevens, 1999). Beside the ability of stabilizing proteins and membranes, taurine prevents uncontrolled protein modification, e.g. nonenzymatic glycosylation which plays a role for a proteomic functional decrease in aging and disease (Rattan, 1996, Soskic *et al.*, 2008). Szwergold could demonstrate that taurine has the ability to prevent generation of a sugar-protein “Schiffbase”, which represents the initial step in glycation (Szwergold, 2005, 2006). Confidently, taurine was demonstrated to inhibit glycation of lens proteins (Devamanoharan *et al.*, 1997), to counterstrike glucose-induced apoptosis (Di Wu *et al.*, 1999), to prevent sugar induced cataracts in rabbits (Malone *et al.*, 1993) and retinal glial cell apoptosis in rats (Zeng *et al.*, 2010). In fact, anti-apoptotic effects of taurine have been intensively studied (Verzola *et al.*, 2002) evidencing taurine to inhibit the formation of the Apaf-1/Caspase-9 apoptosome (Takatani *et al.*, 2004), to inhibit the *m*-calpain and caspase-3 mediated cascade (Sun and Xu, 2008) and to reduce *Bcl-2* (Wu *et al.*, 2009a). Furthermore, taurine can modulate cellular pathways by modulation of protein phosphorylation (Lombardini, 1985, Lombardini *et al.*, 1996, Mollerup and Lambert, 1996) and receptor binding, e.g. triggering GABA (Bureau and Olsen, 1991, Louzada *et al.*, 2004) and insulin receptors (Maturo and Kulakowski, 1988) reflecting the wide range of molecular taurine trigger points. Moreover, immunomodulative functions (Kim, 2006, Marcinkiewicz and Kontny, 2012, Redmond *et al.*, 1998, Schuller-Levis and Park, 2003) due to chloramine generation (Marcinkiewicz *et al.*, 1995, Verdrengh and Tarkowski, 2005) have been reported for the molecule. In fact, taurine chloramine release by activated granulocytes and monocytes during oxidative burst (Arnitz *et al.*, 2006) has been widely demonstrated (Schuller-Levis and Park, 2004). Taurine levels decrease in ocular tissues in the course of aging (Baskin *et al.*, 1977) explaining a correlation between aging, oxidative damage, taurine availability and disease (Malone *et al.*, 1990, Militante and Lombardini, 2004, Yildirim *et al.*, 2007). In confidence with these findings protective effects of taurine are known for various disorders including arterial calcinosis (Yamauchitakahara *et al.*, 1986), pancreatitis (Akay *et al.*, 2013), closed head injury (Sun *et al.*, 2013), diabetes (Kim *et al.*, 2007a), heart diseases (Franconi *et al.*, 1985, Wojcik *et al.*, 2010), renal injuries (Chiba *et al.*, 2002, Wang *et al.*, 2008), including bladder and kidney injuries (Sener *et al.*, 2005) and neuronal injuries like Alzheimer’s disease (Louzada *et al.*, 2004). Special interest was on ocular disorders indicating taurine’s regenerative potential in goldfish retina (Lima *et al.*, 1993), its role in retinal ganglion cell survival in glaucoma models (Froger *et al.*, 2012, Gaucher *et al.*, 2012) and its beneficial performance regarding diabetic cataracts (Hsu *et al.*, 2012, Kilic *et al.*, 1999). Actually taurine showed high performance in epithelial woundhealing (Degim *et al.*, 2002, Dincer *et al.*, 1996) and derivatives like taurine bromamine have been found effective for the management of epithelial inflammation

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(Marcinkiewicz, 2009, 2010, Marcinkiewicz and Kontny, 2012), which is promising for ocular surface disorder management. Taurine application has been found beneficial for the prevention of oxidative damage of the ocular surface (Koyama *et al.*, 1996, Nakamori *et al.*, 1993b) and topical *N*-chlorotaurine performed well in bacterial conjunctivitis therapy (Nagl *et al.*, 2000, Teuchner *et al.*, 2005). Regenerative features of taurine have been proposed for the formation of protective polymer-based ocular surface films (Kedik *et al.*, 2011) and for the application in eye drops in case of ocular surface damage (Koyama *et al.*, 1996). Up to date, taurine has been included in food supplemental oral spray, numerous eye drop formulations and in AMO Complete Moisture Plus[®] contact lens cleaning solution indicating a regenerative effect on the tear film of contact lens wearers referring to MS tear protein profiles shifting towards profiles of healthy control subjects (Grus *et al.*, 2005a). This finding could be validated for conjunctival cells incubated with different contact lens solutions, whereby cells incubated with AMO Complete Moisture Plus[®] have been found to show less signs of apoptosis, necrosis and proteomic alterations (Bell *et al.*, 2012a). Since taurine is the main ingredient of this lens care solution taking advantage of the molecules ability to increase the buffer capacity towards alkaline influence (Hansen *et al.*, 2010, Watanabe *et al.*, 2006), it is proposed to be responsible for the observed beneficial effects on the ocular surface (Grus *et al.*, 2005a). However, no study of longitudinal topical effects of taurine in terms of ocular surface disorders have been realized so far and little is known about the influence on the ocular surface proteome. Accordingly, the identity of taurine target proteins need to be investigated on, which is part of the present work. In summary, two therapeutic strategies and their impact on the ocular surface should be contemplated in this work by use of LC MS proteomics: the topical use of taurine and Taflotan[®] sine.

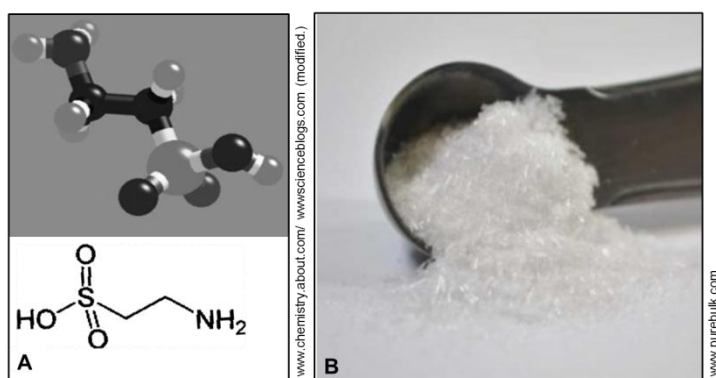


Fig 9 A: Taurine (2-aminoethanesulfon acid, C₂H₇NO₃S) molecule; B: Purified, crystallized taurine

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3.1 Training samples

For LC MS method development RGC5 neuroretinal mouse (*Mus musculus*) cells (Van Bergen *et al.*, 2009, Wood *et al.*, 2010) (Fig. 10), established in the *Experimental Ophthalmology* lab were used. Despite their discussed biological status the cell line was intensively characterized (Crabb *et al.*, 2010, Kim *et al.*, 2009b, Yang and Tezel, 2005) providing a proper proteomic reference for method development purposes. Moreover, material can be easily obtained in the research group. Cell culture was realized due to a standardized protocol of the *Experimental Ophthalmology* including transfer of N₂ frozen cells to DMEM/FCS [10%], penicillin/streptomycin [1%], L-alanin-L-glutamine [4%] at 5% CO₂/37°C, thawing and incubation (2 days/37°C). Cells were monitored in a *Neubauer* chamber by use of a Nikon Eclipse TS 100 phase contrast microscope and 45000 cells were transferred for cultivation. Cell pellets were recovered by dissolving (DPBS/2 mM EDTA/Protease inhibitor) and centrifugation (300 g/4°C/ 20 min). Washed pellets were stored at -80°C. Cell lysis buffer volume was adjusted to the cell pellet weight with a reference ratio of 0.06 g pellet to 100 µl lysis buffer (0.1% *n*-dodecyl-β-maltoside). Cells were disrupted by freeze/thawing (5x, 20°C vs -80°C), lysis buffer and sonication (ice bath/1 min). Resulting homogenates were centrifuged (1500 g/4°C/10 min). Pellets were repeatedly lysed. Finally, supernatants (S1, S2) were combined and filtered through an Acrodisc GHP filter (13 mm x 0.45 µm). 10 µl were used for protein concentration assay while 30 µl filtrate aliquots were stored at -80°C. As substitute simillary processed extracts of bovine retinae (*Bos taurus*) obtained from the local slaughterhouse (Robert-Bosch-Str. 23, 55232 Alzey, Germany) were used.

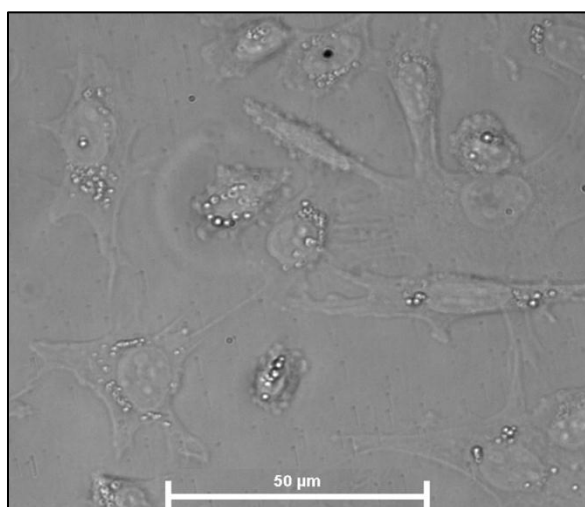


Fig 10 Microscope image of RGC5 cells used in this work for method development

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3.2 Study samples

For proteomic characterization and taurine treatment studies, non-transfected spontaneous immortalized IOBA conjunctival cells (IOBA-NHC; IOBA-University of Valladolid, Spain) derived from human conjunctiva were used as ocular surface epithelia model (Fig. 11). Cells have been characterized as a human conjunctiva model at the *University Institute of Applied Ophthalmobiology* (IOBA) and were defined as preserved epithelial cells (Diebold *et al.*, 2003). Gene expression was documented to differ from primary conjunctival cells, but was found to be more suitable for e.g. drug and cosmetic ocular surface studies than alternative conjunctival cell lines, e.g. ChWK (Tong *et al.*, 2009) explaining their frequent use in ocular surface studies (Benito *et al.*, 2013, Brasnu *et al.*, 2008, Li *et al.*, 2009). However, there is still a lack of sensitive proteomic investigations on NHC-IOBA cells. The cells were maintained in DMEM/HAM's F12 1:1, 10% FCS, insulin [1 µg/ml], EGF [1 µg/ml], hydrocortisone [5 µg/ml]). Preparation workflow corresponds to the described RGC5 workflow.

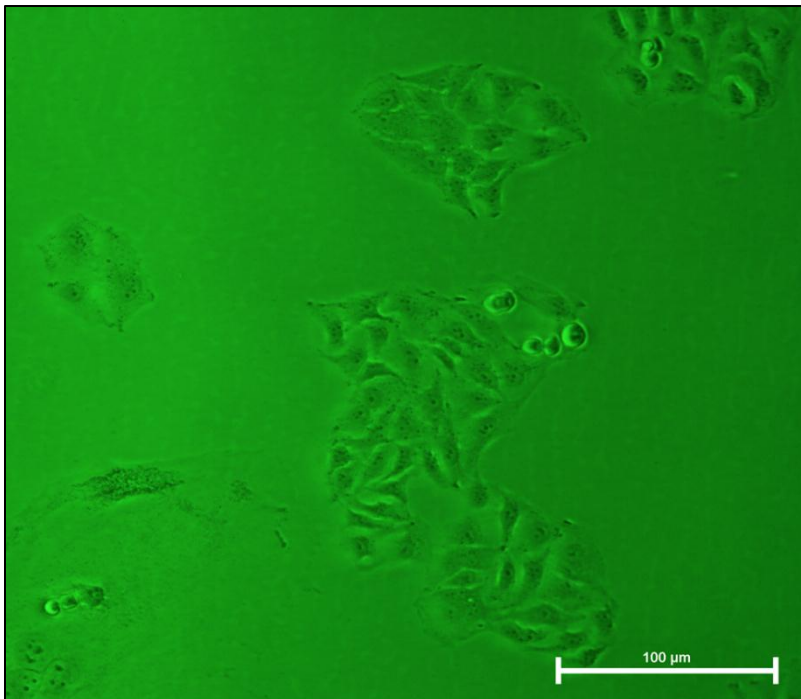


Fig 11 Microscope image of NHC-IOBA cells, human immortalized conjunctival cells used as an ocular surface culture model.

3.2.1 Tear fluid

Tear fluid was sampled by two methods, glass micro capillary tubes and Schirmer strips (Wright and Meger, 1962). Due to their diagnostic purpose strips are common in clinical routine (Whitcher, 1987). Regarding strip collection in this work Tear Test ophthalmic strips were adjusted to the eye lid without cornea anaesthesia and left for 5 min in the eye. For

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sample analysis, each strip was soaked in 300 μ l DPBS and tear proteins were extracted overnight on a KS250 basic Intellimixer at 4°C. Regarding capillary tears, tear fluid was collected by use of Ringcaps® Duran® disposable micro pipettes with ring mark. In both methods resulting samples were stored at -80°C until use. Prior to use, samples had been adjusted to -20°C and then to 4°C to allow a gentle adjustment of proteins to avoid freeze/thawing protein loss effects as much as possible. Taflotan® sine study sample collection and taurine study sample collection were supported by members of the lab staff.

3.3 Sample preparation

3.3.1 Intact sample processing

Protein amount estimation was realized by use of the Pierce® BCA Protein assay and a Multiscan Ascent scanner according to the protocol of the manufacturer. Freshly thawed sample aliquots for TD LC MALDI analysis were adjusted to 3 μ g/ μ l final concentration due to 15 μ g maximum column capacity and 5 μ l injection volume (3 μ g/ μ l x 5 μ l = 15 μ g) with running buffer A (4% ACN, 0.1% TFA). 40 μ l sample volume were prepared for each LC run. The samples were transferred to V-bottom 96 well plates for automated sample injection.

3.3.2 SDS-PAGE

Since a direct MS analysis is crucial in BU proteomic approaches due to high complexity, 1D SDS PAGE invented by Lämmli (Laemmli, 1970) was used to fractionate proteins corresponding to their molecular weight prior to LC MS analysis. Usually, form, charge and mass influence migration of a protein through a polyacrylamide gel network (Schagger and von Jagow, 1987). Anionic SDS detergent binds to hydrophobic regions (Löffler, 2003) provides negative charges and facilitates denaturation. Consequently, electrophoretic field migration through the gel meshwork is directly proportional to the protein mass, whereby smaller proteins migrate faster than higher mass proteins (Löffler, 2003). However, the qualitative output depends on various factors, e.g. the maximum gel loading capacity (van Hagen, 2008). Therefore, protein load should not impede migration, but consider MS sensitivity. Based on RGC5 extracts, a protein load of 80 μ g was estimated optimal for ESI and MALDI experiments. Above 80 μ g protein lanes clearly smeared due to overloading effects. In contrast, if available, at minimum 20 μ g protein should be separated per lane to fit the MS dynamic range. Samples encircling RGC5 or IOBA-NHC lysates and tear fluid were mixed with NuPAGE LDS sample buffer (+/- reducing agent). Corresponding samples were heated (10 min/90°C) and run on NuPage 12% Bis-Tris, 10-well prepared minigels (100 V/10 min - 150 V) in the XCell SureLock™ system using MES or MOPs buffers. SeabluePlus 2 prestained protein standard was used as molecular weight reference. Reproducibility was

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tested using replicate aliquots of test samples. Gels were fixed and stained overnight using the Novex Colloidal Blue staining kit according to the manual of the manufacturer. For documentation gels were recorded by use of a DCP-9042 CDN benchtop scanner with a resolution of 1200 x 1200 dpi as *tif*-data assisted by Photoshop C53 extended version.

3.3.3 In-gel digestion

The procedure was based on a modified protocol of Shevchenko *et al.* (Shevchenko *et al.*, 2006) considering ESI/MALDI optimizations. In case of LC MALDI experiments, a gel lane of interest was divided into 15 slices, which allows one lane to be positioned on a 386 spot target. In case of LC ESI a gel lane was divided into 17 slices. The lane division was decided corresponding to the CBB protein signal abundance. Mass area slices were cut into equally small pieces, destained (50% ACN, 50% 100 mM Ambi, 10 min) and dehydrated (100% ACN). Dried gel pieces were saturated with trypsin solution (200 μ l reaction buffer, 1300 μ l 20% ACN/10 mM Ambi) ([15 ng/ μ l], 60-100 μ l/slice) and incubated over night (37°C). Digestion was quenched by a pH shift by addition of 10 μ l 0.1% TFA. Supernatants were transferred to fresh 0.5 ml tubes and peptides were extracted with appropriate amount of extraction buffers on a Unimax 1010 shaker at 700 rpm. In case of LC MALDI, prior to analysis, combined supernatants and peptide extracts of each slice were filtered through GHP Acrodisc 13 mm x 0.45 μ m pre-HPLC syringe filters followed by vacuum concentration (30°C) in a Concentrator 5301 instrument. LC MALDI destinated samples were equilibrated with 20 μ l running buffer A (1% MeOH, 1% ACN 0.05% TFA), whereas LC ESI samples were equilibrated with 20 μ l 0.1% TFA. In case of LC ESI analysis, sonication resolubilized peptides were SPE purified once again to reduce background signals due to the less contaminant robust character of ESI systems (Kraj and Silberring, 2008).

3.3.4 Solid phase extraction (SPE)

Because SPE tips had been demonstrated to generate more accurate results than magnetic beads (Tiss *et al.* 2007) ZIPTIP[®] C18 pipette tips (15 μ m, 200 Å) were used. 10 μ l peptides were bound by 20 binding cycles on the C18 resin followed by a 0.1% TFA wash step followed by 10 μ l duplicate elution (50% ACN, 0.1% TFA). The tip was once again rinsed with pure ACN, equilibrated with 0.1% TFA and binding cycles were repeated. Resulting aliquots were combined leading to a 40 μ l peptide sample corresponding to each slice digest. Duplicate ZIPTIP[®] purification was used to exceed the resin saturation limit of 5 μ g protein (UserGuideforRPZipTipPipetteTipsforSamplePreparation, 2007) since approximately 4-10 μ g protein load (1 lane = 80 μ g/17 slices = 4.7 μ g/slice; ~4-10 μ g/slice) was provided per gel slice with respect to protein distribution pattern. Lyophilized peptides were stored at -80°C until use. For LC ESI analysis peptides were resolubilized (20 μ l 0.1% TFA, ice

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bath/sonication/10 min) followed by transfer to V-bottom 96 well plates for automated sample injection.

3.3.5 Microbore-RP-HPLC for MALDI Top Down (TD) analysis

For direct MALDI TD ocular sample analysis employing the Ultraflex II linear mode, a microbore column (0.8-2 mm Id) (Kraj and Silberring, 2008) was chosen achieving a higher robustness in comparison to capillary and nano approaches (0.05-0.8 mm Id), but exceeding performance of common analytical (2-10 mm Id) and semipreparative columns (10-100 mm Id) (Kraj and Silberring, 2008). However, care had to be taken for column blocking regarding direct analysis of crude samples. Accordingly, microbore should balance robustness and sensitivity. For gradient elution, a Rheos Allegro pump system was used. Furthermore a PAL HTS robot was programmed for automated sample injection and on target fractionation (Fig. 13). Eluting peptides were deposited on 386 MTP polished/matt steel MALDI targets by use of 75 µm ID fused silica tubing. A Prosphere P-HR 4 µm microbore column (150 x 1 mm) was used as analytical column directly coupled to a Security GuardTM cartridge system with an RP1 (4x2 mm) filter inlet (Fig. 12). The guard column should allow direct injection of crude "dirty" samples. Even if the cartridge was developed for columns with IDs >2 mm and was widely used with good suitability for these systems (Halquist and Karnes, 2011, Ireland *et al.*, 2006, Lichtenthaler *et al.*, 2005, Woodward *et al.*, 2009), it was not used in a TD LC MALDI workflow. In fact, the guard column was documented to work well as a trapping column (Boernsen, 2000) in terms of column-switching system and was actually found to perform excellent in direct combination with the 1 mm ID system in this work. The split-free flow rate was adjusted to 20 µl/min and 15 s injection times enabled a 5 µl injection volume. A start delay of 14.25 min. was set prior to fraction deposition. The spotting frequency was 3 s for each fraction approximating 1 µl/spot. Spots were dried and 2 µl matrix (18 mg CHCA, 60% ACN, 2% TFA) was applied on each spot. The matrix composition and spotting technique was found optimal considering homogen crystal formation and signal generation. Matrix spotting automation was realized on a Sun Collect MALDI spotter. Since the manufacturer's spotting needle (50 µm ID) was found to exhibit a low degree of robustness, a custom-built adapter in combination with a Tecan steel teflon-coated spotting tip was adjusted to the robot showing reproducible and robust performance resulting in homogeneity spots and low carry-over effects (Fig. 14). The HPLC fractionation method was optimized by use of retina lysates from 240 over 120 to finally 92 fractions collected for each sample run. Running buffer A were prepared as follows: A= 4% ACN/0.1% TFA, B= 96% ACN/0.1% TFA). The gradient was programmed as follows: 0-12 min: 90-30% A, 12-16: 30-0% A, 16-21: 0% A, 21-25: 0-100% A, 25-35:100% A. Optimizations and monitoring were realized by the 2D-gel view generator Survey Viewer version 1.1 tool. Finally, the reproducibility of the system was

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determined by comparing elution behavior of reporter m/z peaks monitored between replicate runs as well as comparison of fraction profiles between sample runs. To assess system sensitivity, the number of detected WarpLC compounds was ascertained and compared to the literature. Also unfractionated samples were measured in parallel and resulting profiles were compared to HPLC processed samples. The microbore HPLC method should allow the analysis of ocular cell species and should finally be transferred to tear samples.

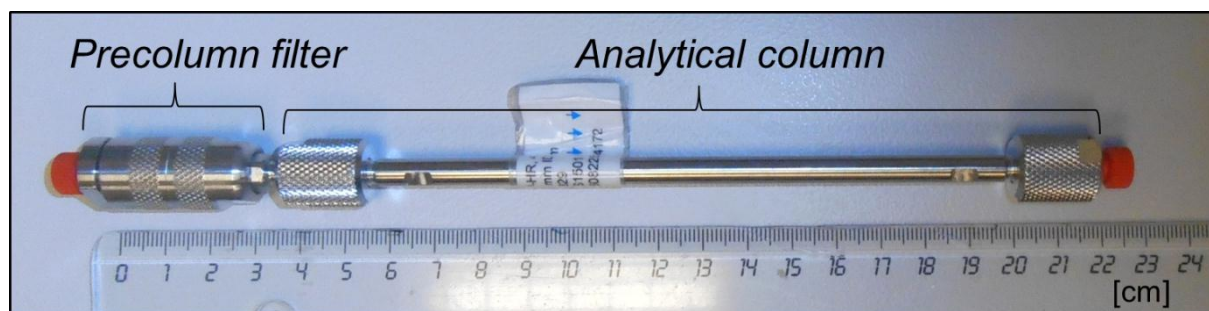


Fig 12 Microbore column system for TD LC MALDI consisting of a Security Guard™ cartridge with a RP1 filter (4x2 mm) connected to a microbore analytical Prosphere P-HR 4µm column (150 x 1 mm)

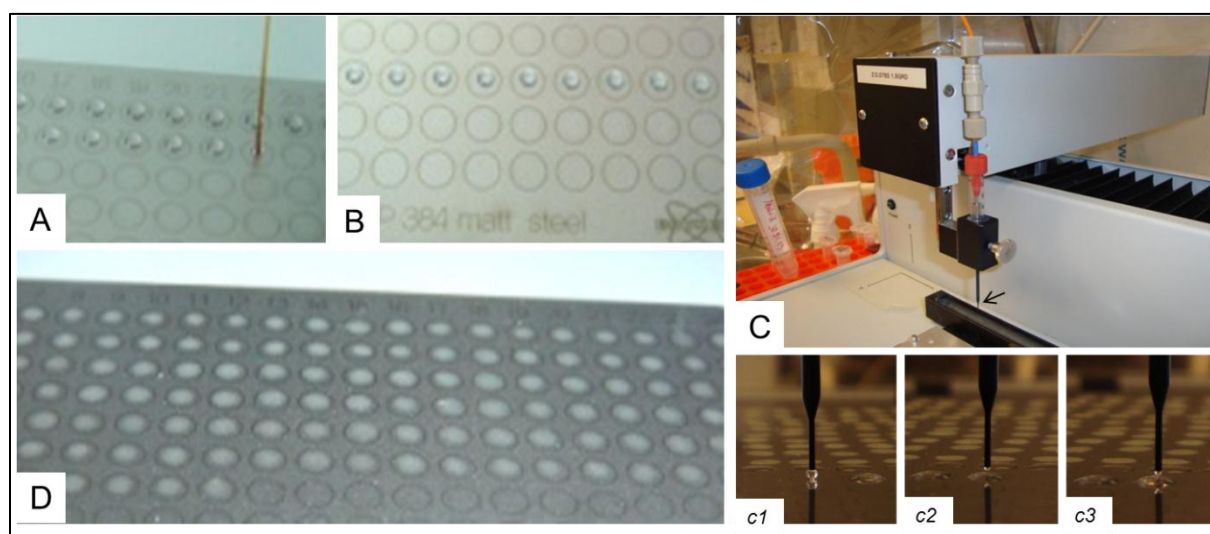


Fig 13 TD LC MALDI Sample preparation; A: Direct on target sample fractionation realized by a PAL HTS robot connected to the column system leading to B: accurate fraction deposits; C: Matrix spotting by use of a customized Tecan Teflon coated needle unit (arrow) integrated on a Sun Collect robot station allowing accurate matrix deposition on dried analyte fractions (c1-c3) leading to D: homogen analyte/matrix spots.

3.3.6 Capillary-RP-RP-HPLC (for MALDI)

For BU MALDI analysis the Ultraflex II and the Rheos Allegro Pump system were employed. The system performs optimal gradient mixing in a flow rate range between 20 µl to 1000 µl/min according to the manual of the manufacturer. Accordingly, flow rates <20 µl/min lead to declined reproducibility of the gradient. Another disadvantage is the extended time period

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the gradient needs to reach the column head, probably causing diffusion effects and decreased performance. For optimal conditions, a pre-column flow splitting strategy was established. Thereby, the flow split should consider the pump's optimal gradient range and should leave the analytical column with low flow rate. As mechanical split system an M-472 graduated micro-split valve was used. The micro-split valve was selected due to a pressure resistance >200 bar. The challenge in flow splitting refers to the column system, which creates high pressure. Consequently, without appropriate waste port backpressure all solvent will flow out of the waste port failing to pass the split port and without entering the column. Therefore, appropriate backpressure adjustment is a challenging step. Accordingly, for optimal pump flow splitting different splitter graduation settings and pump flow rates have been tested. In consequence, the adjustment of the split flow rate exclusively by the splitter graduation was found to be unsuitable for accurate fine regulation of the split stream. Hence, reproducible adjustment was successfully achieved only in a small setting window confident with manual splitter inspection, which supports the splitter to work preliminary as a shut-off valve with distinct limitation in split ratio adjustment. Consequently, as a counteracting splitting strategy, peek tubings with varying IDs were connected to the waste port to generate appropriate backpressure at a fixed graduation setting which was also documented for comparable splitter models (Kozerski *et al.*, 2003, Rogatsky *et al.*, 2009). The use of peek tubings for backpressure adjustment is an economic strategy to down-scale common HPLC systems to capillary or nano dimension (Aguilar, 2004, Moritz, 2007, Profrock, 2010, Yi *et al.*, 2003, Zhang *et al.*, 2004a). Therefore, variations of peek tubing length and pump flow rates were combinatorial determined for backpressure generation and capillary flow rate setting. A combination of a 1/16" OD x 0.10" ID x 5 ft PEEKTM tubing under a pump flow rate of 300 μ l/min was found to work optimal resulting in a split flowrate of 7.8 μ l/min, matching the capillary range. Hence, 300 μ l/min pump flow had been documented for most systems to guarantee suitable pump function and solvent portioning (Chervet *et al.*, 1996). Split flow rate reproducibility was tested by time assays monitoring different mobile phase compositions passing through a 20 μ l defined capillary tube. The splitting system was connected to the injection valve by combination of 1/16" OD x 0.0025" ID and 1/16" OD x 0.007" PEEKTM tubings connected by use of a stainless steel NanoTightTM union (10-32 to 10-32, 0.007" thru-hole) to allow an easy reinstallation of the microbore system or other systems (Fig. 14). All tubings and connectors were capillary compatible to avoid dead volumes in the system. Accordingly, the 6-port valve outlet was connected to the column system by use of a 1/16"OD x 0.005 ID PEEKTM tubing. Tubings were connected by use of 10-32 PEEKTM fingertight fittings. Since the present instrumentation did not allow column switching and peptide trapping, the design of cationic/anionic exchange and reversed phase as common 2D-HPLC strategies for BU analysis (Link, 2002) was not possible. To counteract this

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drawback the idea was to develop a column system which displays slightly different retention properties achieving proper orthogonality under identical RP solvent conditions in an online-system. Thereby, the combination of C18 with a phenyl stationary phase was the key point of the concept since there is growing evidence for the efficiency of combinatory RP-RP strategies in BU proteomics (McQueen and Krokhin, 2012). In fact, Gilar and colleagues showed that a RP-RP combination under different pH conditions performs higher orthogonality than a SCX-RP combination (Gilar *et al.*, 2005). Actually, alkyl residues, especially C18, are commonly used to retain peptides, whereas RP materials like cyanopropyl ligands and phenyl materials offer additional selectivity (Aguilar, 2004). Consequently, the idea was to combine RP materials taking advantage of slightly selectivity differences for peptides. To realize this concept a RP-RP-capillary column system was developed, which consisted of a 30 x 0.5 mm BioBasic[®] C18 precolumn connected to a 100 x 0.5 mm BioBasic[®] Phenyl analytical column combined with a 150 x 0.5 mm Jupiter[®] 4 μ Proteo analytical column. To avoid gel particles and contaminants to enter the column system the whole system was protected by an A 316 0.5 μ m online precolumn filter. All system parts were directly connected via Onyx[®] column couplers to minimize dead volumes and rediffusion (Fig.15). Consequently, the column phases including the C18 pre-column refer to RP but differ in selectivity properties. Regarding the gradient conditions running buffer A consisted of 98% water, 1.94% ACN, 0.06% MeOH and 0.05% TFA. Running buffer B consisted of 95% ACN, 3 % MeOH, 2% water and 0.05% TFA. For each gel slice a 60 min gradient was run (0-5 min: 100% A, 5-45 min: 0-80% D, 45-47 min: 80-100% D, 47-49 min: 100% D, 49-58 min: 0% A-100% A, 58-60 min: 100% A). The addition of MeOH was found to improve the selectivity of the phenyl phase towards aromatic residues (Yang *et al.*, 2005)(www.restec.com). Elimination of sample carry over effects, which play a critical role in LC-MS based experiments (Burton *et al.*, 2008, Mitulovic and Mechtler, 2006, Vu *et al.*, 2013), was achieved by running equilibration gradients of 12 min between samples by 80% ACN injection also maintaining sample loop and valve port clearance. For automated sample injection and on target fractionation, the PAL HTS robot was programmed. Eluting peptides were deposited with a frequency of 40s/spot on 386 MTP polished steel MALDI targets. Thereby, each gel slice extract was fractionated over 24 spots on the target. Accordingly, one target corresponded to one sample lane consisting of 15 gel slices. CHCA based matrix was applied on dried fractions as already described. Representative “reporter ions” eluting in replicate runs were monitored to evaluate retention time (RT) reproducibility. Finally, the method should be used for the sensitive analysis of tear samples.

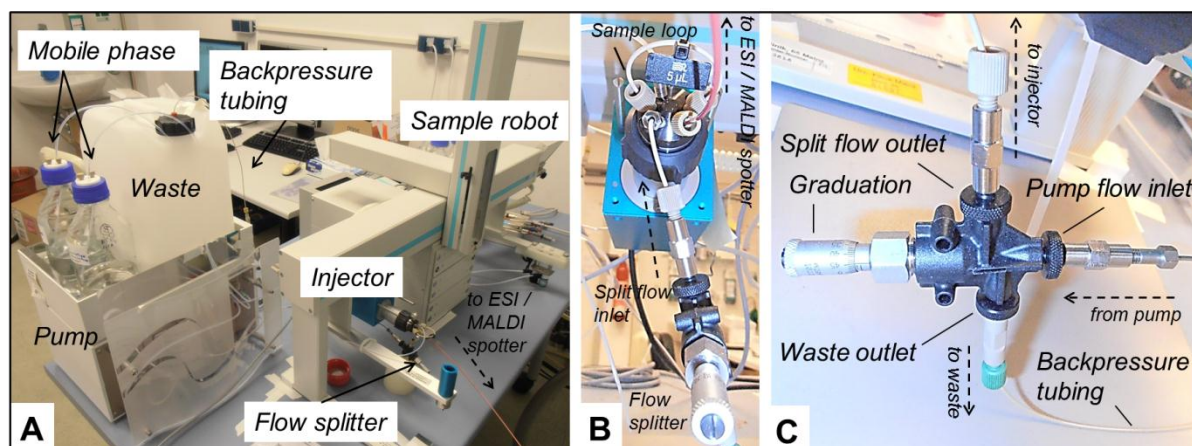


Fig 14 A: Overview of the capillary HPLC system developed for the ESI/MALDI BU application consisting of an Rheos Allegro HPLC pump and PAL HTS sample robot, which can be used also to fractionate eluting samples direct on MALDI targets; B: Precolumn split system using a M-472 (for MALDI) or a P-470 (for ESI, shown) microsplitting valve to downscale high pump flow rates to capillary flow rates between 7-10 $\mu\text{l}/\text{min}$; C: Detailed view of the microsplit system.

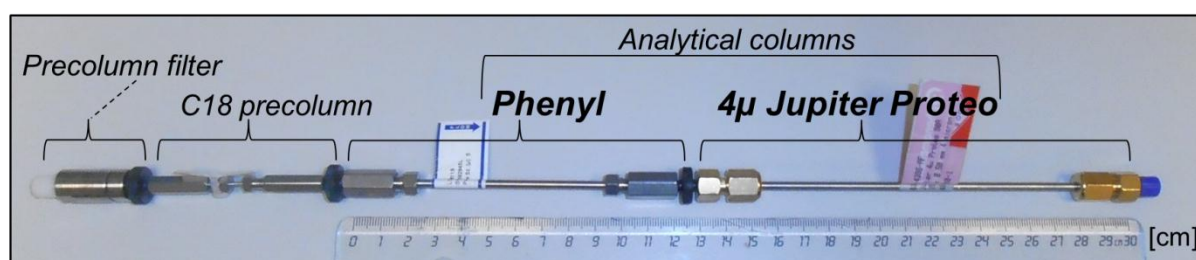


Fig 15 RP-RP column system developed for MALDI BU analysis of tear fluid. The system consists of two online coupled stationary capillary (0.5 mm ID) RP phases, phenyl and 4 μ Jupiter Proteo material, which provide slight differences in retention properties. Actually the C18 precolumn additionally contributes to orthogonal phase selectivity.

3.3.7 Capillary-RP-HPLC (for ESI)

For ESI experiments the identical split system described for the MALDI system was used with slight modifications, instead of the M-472 the P-470 splitter was used which comes with a 10-32 split port instead of a 6-32 split port, allowing easier installation. As column system only a 1D HPLC strategy was used consisting of a 30 x 0.5 mm BioBasic[®] C18 precolumn connected to a 150 x 0.5 mm BioBasic[®] C18 analytical. The whole system was protected by use of an A 316 0.5 μm online precolumn filter (Fig. 16). Like in the MALDI setup, all components were directly connected via Onyx[®] column couplers and also mobile phase composition was identical to the LC MALDI approach. Since the flow rate was influenced by different backpressure condition due to the ESI needle, system settings could not be directly transferred from the MALDI setup. The pump flow rate was reduced from 300 to 200 $\mu\text{l}/\text{min}$ resulting in a flow rate of $\sim 6.7 \pm 0.03 \mu\text{l}/\text{min}$ ($N= 20$ measurements), whereby different gradient settings were tested. The challenge was to find a compromise between

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sensitivity and analysis time which should allow lab routine integration. Therefore, a first workflow using 80 min gradient per slice corresponding to 17 gel slices per lane was developed determining reproducibility and sensitivity running RGC5 lysates ($N= 8$). Quantitative (number of identified proteins) as well as qualitative reproducibility (protein identification congruency) were determined. During a sequence after each slice run 30 min washing runs injecting 80% ACN were used to counterstrike carryover effects. However, washing runs were eliminated to reduce total analysis time without affecting the performance remaining washing runs between different sample subjects (eg. control, treatment). In the end of a sequence the system was equilibrated to 80% solvent for column storage. To economize analysis time, the 80 min peptide elution pattern was inspected using two different gradient optimization strategies. At first, a proportional economization of the gradient was tested for its sensitivity; secondly, a targeted reduction of the elution area was applied taking main peptide elution areas of chromatograms into consideration. For optimization, RGC5 gel mass area replicates (1/17 lane) were used. In parallel, a different strategy was tested to economize analysis time decreasing the number of slices per gel lane in various steps. To determine the reproducibility of the LC ESI workflow, as already mentioned, quantitative and qualitative parameters were analyzed. Reproducibility of sensitivity was realized by quantitative comparative component analysis including number of CID precursor masses, CID fragment ions and resulting protein hits, considering two different scoring strategies (relaxed= MASCOT ion score $p<0.05$, stringent= peptide score <30). Qualitative comparative analysis revealed overlap in CID precursors, fragments and proteins focusing on technical replicates ($n= 6$) corresponding to representative RGC5 gel mass areas ($N= 8$). Moreover, the RT reproducibility was examined monitoring the RT of representative reporter ions observed in all replicates of a certain gel mass area. Finally, the LC ESI approach should be applied to ocular surface samples encircling tear fluid and conjunctival cells.

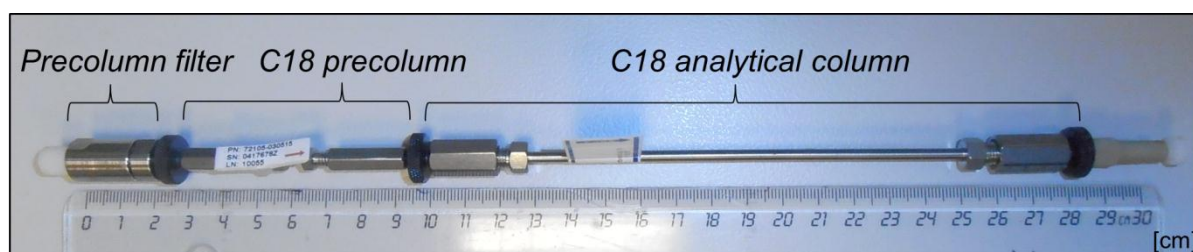


Fig 16 HPLC capillary column system (0.5 mm ID) employed for ESI BU analysis of ocular surface sample species.

3.3.8 MALDI-TOF/TOF MS/MS

3.3.8.1 Linear mode for Top Down (TD) MS (microbore HPLC)

For TD LC MALDI analysis, the Ultraflex II instrument was used. The instrument was equipped with a nitrogen laser under control of MarathonControl software. Samples were measured in the MS linear mode controlled by Flex Control/Flex Analysis under priority of WARP-LC software. For Flex Control adjustment, the pulsed ion extraction was set to 100 ns and the linear detector gain was adjusted 38x with 0.10 sample rate and highest electronic gain. Matrix clusters were suppressed up to 1000 Da implementing a blast of 10 shots with 75% laser power (LP) for background suppression. For signal recording 600 shots in 60 shot steps with 25% LP were found to give the best results balancing intensity and resolution. Laser shots were fired without fuzzy logic on each spot using a hexagon raster focusing on masses between 1000 and 17000 Da ($S/N > 3$; resolution > 100). The system was calibrated using the protein calibration standard I for reference.

3.3.8.2 Reflector mode for Bottom Up (BU) MS (capillary RP-RP HPLC)

For fraction analysis the Ultraflex II instrument was used taking advantage of the instrument's TOF/TOF MS/MS concept using the described control software platform. The instrument was run scanning a range of 900-3700 m/z to record precursor masses. Thereby, 500 shots in 50 shot steps from ten different spot positions were applied for each fraction without fuzzy logic. MS peaks were detected using the *SNAP* (Sophisticated Numerical Annotation Procedure) algorithm, which detects monoisotopic masses in raw reflector MS spectra recognizing a component's isotopic distribution by a non-linear fit function (FlexAnalysis2.4OperatorManual, 2005). Based on detected peaks a WARP-LC compound list was generated. Thereby, the WARP-LC algorithm represents an intelligent concept linking MS to chromatography (WARP-LC1.1UserManual, 2006), whereby peptides that had once been fragmented are excluded for further fragmentation, which increases the probability of low abundant peaks to be selected for analysis. The strategy economizes shots, sample and time while providing higher sensitivity. It is important to note that each laser shot during fragmentation physically eliminates sample components, which means that multiple fragmentation cycles corresponding to a particular peptide reduce the chance to identify low abundant coeluting peptides in a fraction. Confidently, chromatographic effects like peak shifting and smearing are taken into consideration (Berger, 1993, Castells and Castells, 1998, Conder, 1982, Fraga *et al.*, 2001). Furthermore, compound list settings are used as MS/MS precursor selection filter. To account for chromatographic blur, peaks of similar m/z value considering 100 ppm mass tolerance were defined as identical compound if they were separated by less than 3 fractions ($S/N > 10$, intensity > 700), which provided best results for

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efficient overall fragmentation in preexperiments. Compounds occurring in more than 90% of an analysis were defined as background and excluded from precursor selection. A maximum of 70 MS/MS measurements was allowed for each spot during a WARP-LC run. The system was calibrated using the peptide calibration standard for reference and internally by autodigestive trypsin signals. Limits of the system were also considered by monitoring the accuracy of peak detection and fragmentation efficiency.

3.3.9 ESI LTQ Orbitrap XL MS

3.3.9.1 Spray system

For spray generation, a 75 μm ID silica-fused tubing was used, which displayed fast performance decline. Microscopic inspection of the tubing tip indicated corrosive surface damage leading to spray inaccuracy, instability and electrical circuits (Fig. 17). Corresponding spray fluctuations were found to lead to nozzle droplet adhesion causing a whole breakdown of the spray and analysis. Therefore, the needle system was substituted by a low flow metal needle for API 2 probes appropriate for the capillary flow range. Actually, by use of the metal needle a small sensitivity decline was recognized, however stability and reproducibility of the system was distinctly improved which represents the main argumentation for installation of the metal needle. Thereby, the spray voltage was ~ 4.2 kV, spray current ~ 0.9 μA , capillary voltage ~ 44 V, capillary temperature $\sim 275^\circ\text{C}$ and tube lens was ~ 125 V. The ESI probe coordinates were adjusted to position B defining the depth of the probe nozzle in the probe chamber and graduation 1.1 which describes the accurate distance of the needle to the ESI cone. However, events of needle blocking in case of the metal needle have been recognized by corrosive particles of unknown origin, which brings need for periodical maintenance of the needle system (Fig 17). Also the heated ion transfer capillary was frequently cleaned by use of methanol and sonication.

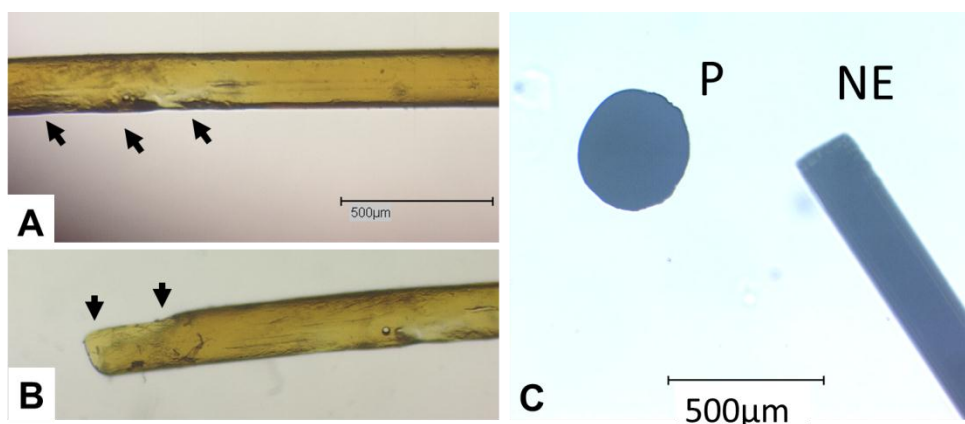


Fig 17 A/B: Corrosive modifications at the ESI fused silica tubing outlet after several measurements (indicated by arrows) leading to spray instability; C.: Corrosive particle (P) blocking the inlet of the ESI metal needle end (NE) resulting in decreased analysis performance.

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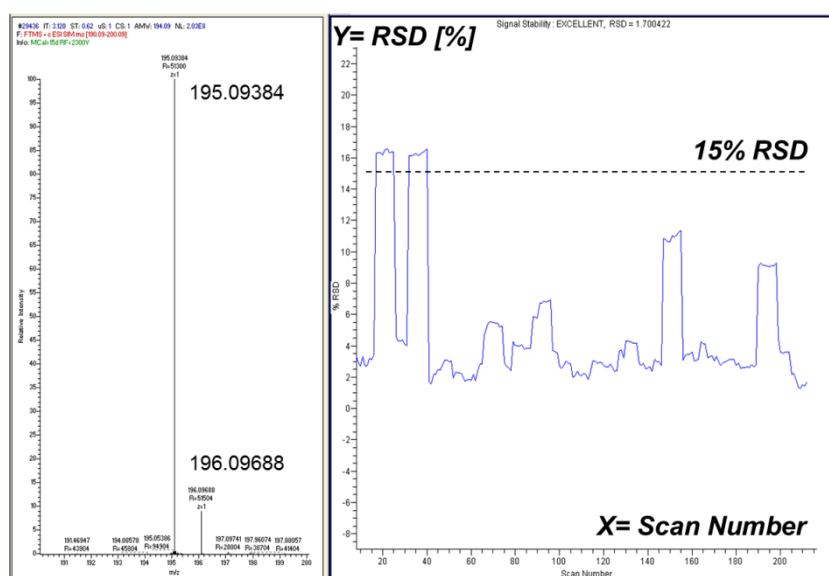


Fig 18 Spray stability test indicating high stability with $RSD < 10\%$ during test measurements focusing on reporter masses 195.094 and 196.097 m/z after installation of a low flow metal ESI needle.

3.3.9.2 MS settings

The coordination between MS, sample robot and HPLC was realized using XCalibur[®] related contact closure signaling. For Orbitrap MS a range of 300–2000 m/z in the instruments positive mode was focused on. The maximum injection times were set to 50 ms for ion trap (LTQ/ITMS) and 500 ms for orbitrap (FTMS) analysis. For LTQ peptide fragmentation, CID was used with helium as collision gas (normalized collision energy= 35, activation time= 30ms, activation Q= 0.25). The top five centroid detected monoisotopic peaks (charge state=2, intensity \geq 500) were selected within an isolation width of 2 m/z for fragmentation in each scan event. Data acquiring time was adapted to the length of the gradient (finally 50min). The complete instrument was semi-automatically calibrated weekly using Pierce[®] LTQ ESI positive ion calibration mix corresponding to five components: caffeine (195 m/z), MRFA (524 m/z) and Ultramark components (1322, 1422 and 1522 m/z). For internal real time calibration a 445.120025 m/z ambient air background peak corresponding to polydimethylcyclosiloxane (PCM) was used according to the mass lock calibration strategy (Lee *et al.*, 2011, Olsen *et al.*, 2005). Optimal LTQ settings for dynamic exclusion and resolution were determined using RGC5 lysate replicates (1/17 lane), referring to number of identified proteins and score values leading to 90 s dynamic and 30 s repeat duration. Since a resolution >30000 was shown to result in a protein identification rate decline, 30000 was set for analysis. Also the dynamic range of the system was tested by dilution of RGC5 (80

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μg) standard digest (Fig. 19). Finally, optimal MS settings corresponding to 50 min HPLC gradients were used for the analysis of tear fluid and conjunctival cell studies.

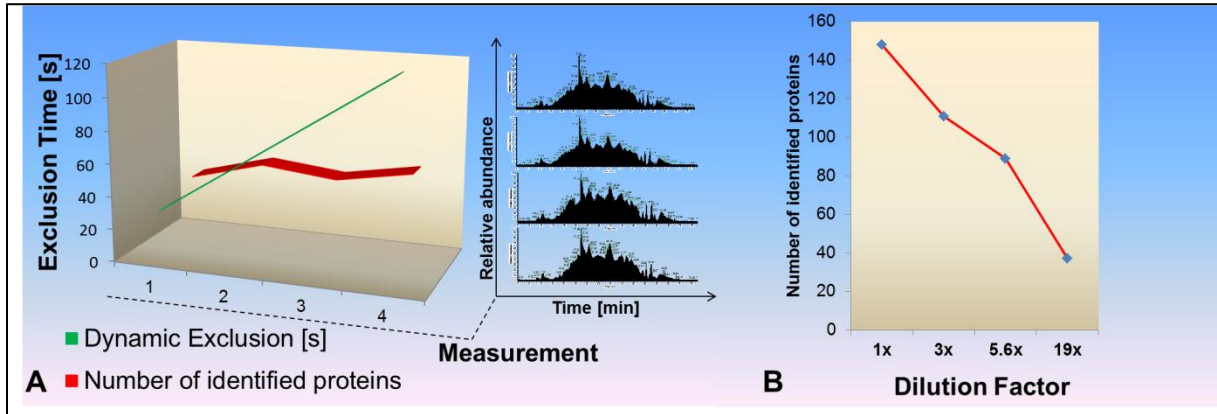


Fig 19 A: Influence of dynamic exclusion duration on the number of identified proteins showing a slight positive correlation; B: Correlation between sample dilution and sensitivity

3.3.10 Data processing & Protein identification

3.3.10.1 Linear Microbore LC-MALDI Top Down (TD) analysis

Raw MS spectra were manually inspected and optimal processing and peak detection was realized in FlexAnalysis. Spectra were *Savitzky Golay* smoothed (Gorry, 1990, Savitzky and Golay, 1964) with an optimum width of 30 and 3 cycles balancing noise reduction and sensitivity (Fig. 20) followed by *Convex Hull* baseline subtraction (Fung and Enderwick, 2002, Ilina *et al.*, 2010) and *Centroid* peak detection ($S/N \geq 17$, m/z width = 9, 80% percent height, 0.2% rel. intensity) (Blais and Rioux, 1986, Yang *et al.*, 2009) (Fig. 21). Optimal processing and peak detection were automated using a FlexAnalysis method script in the batch processing mode (Fig. 22).

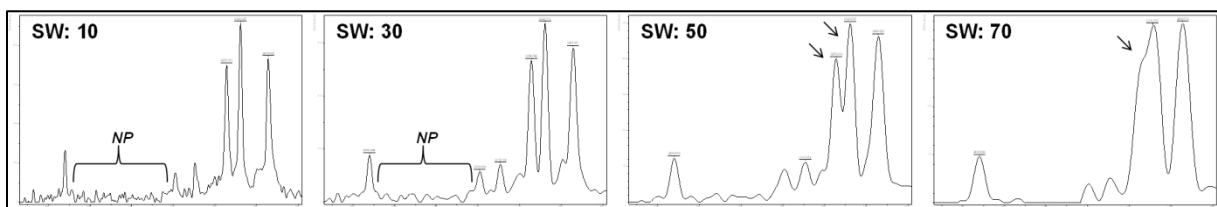


Fig 20 Influence of smoothing width (SW) settings on spectrum quality in linear MALDI MS spectra; whereas noise peaks (NP) are suppressed at SW 70 also distinct peaks (arrows) merged and could not be resolved. An optimal balance between noise reduction and resolution was found at SW 30.

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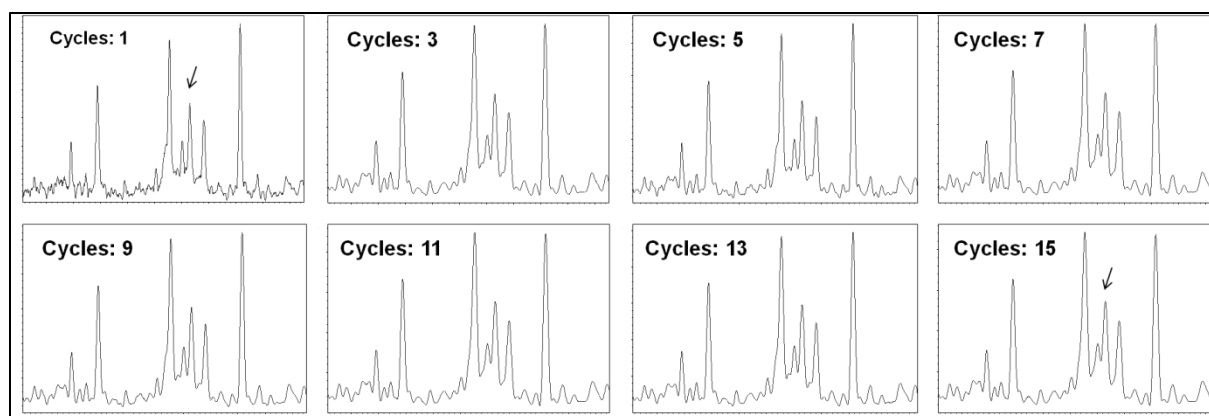


Fig 21 Influence of smoothing cycles on linear MALDI MS spectra. No clear changes on distinct peaks (example indicated by arrow) were found after 3 smoothing cycles.

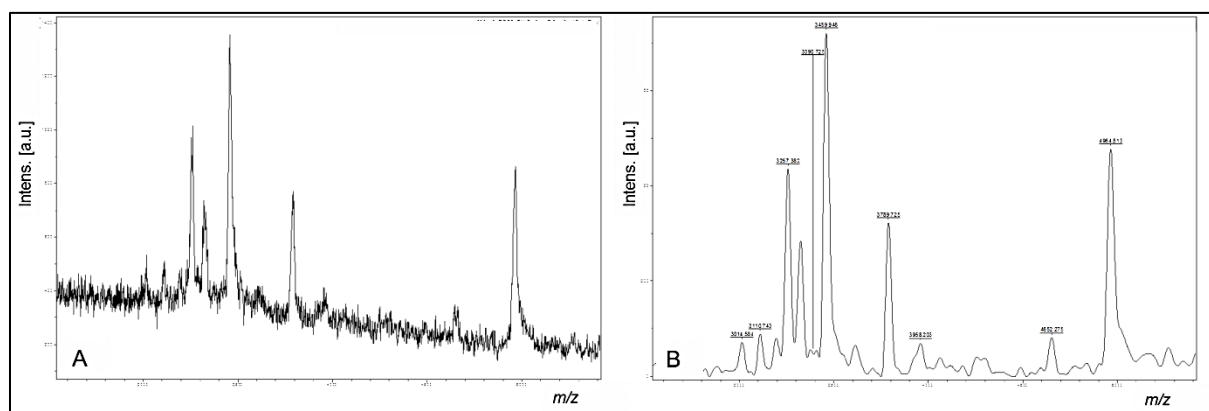


Fig 22 Automated batch mode processing featuring noise reduction and peak detection with implemented optimal settings; A: Raw linear MALDI MS spectrum; B: Processed spectrum allowing accurate peak detection.

3.3.10.2 Reflector Capillary LC-MALDI Bottom Up (BU) analysis

For protein identification, combined MS/MS spectra corresponding to a particular experiment were transferred to MASCOT as *mgf* files and searched against the SwissProt version 57.8 database (SwisProt_111101, 519348 sequences; 183273162 residues). Depending on the sample origin *Homo sapiens* (tears, NHC-IOBA) (20359 sequences) or *Mus musculus* (RGC5) (16409 sequences) were set as taxonomy, trypsin as enzyme and one missed cleavage was allowed. Charge state was adjusted to 1+ and mass tolerance was set 100 ppm for MS precursors and 0.8 Da for MS/MS fragments. To achieve most stringent results protein queries were run either under *MUDPIT* scoring conditions (if queries/entries>0.001; after any taxonomy filter, www.matrixscience.com) with a significance threshold of $p < 0.05$ for protein identification. Additionally, only those proteins that were based on unique peptide sets were documented to avoid redundant protein entries.

3.3.10.3 Capillary LC-ESI Bottom Up (BU) analysis

All MS/MS spectra of a particular experiment were combined by use of the Proteome Discoverer[®] version 1.1. The resulting *msf* file containing all experimental data was MASCOT searched against the SwissProt as described for MALDI. ESI protein identification settings were realized in a Proteome Discoverer[®] script featuring a precursor mass range of 150 to 2000 Da (charge= 1-4, S/N ≥6), trypsin as enzyme (1 allowed missed cleavage), 20 ppm mass tolerance for precursors and 0.8 Da for fragments. Moreover, ammonia-loss (N-term C), oxidation (H/W, M) and phosphorylation (S/T, Y) were selected for variable modifications and carbamidomethyl (C) was set as fixed modification.

3.3.11 Protein quantification and statistics

In case of LC MALDI BU analysis semi quantitative protein level estimation was carried out by use of a modified version of the in-house developed *P2M* software platform (Tschoerner, 2008), whereby MS and MS/MS spectra were imported directly from Flex Analysis following normalization of MS spectra according to the mean total ion current (TIC) ($\text{Intensity}_{\text{MS}} \times \text{meanTIC}_{\Sigma\text{MS}}/\text{TIC}_{\text{MS}}$). Spectra with a minimum factor of 0.1 or maximum factor of 10 were categorized as “low quality” and excluded from analysis. To allow comparison of protein levels between all samples, a protein that failed to be identified in some of the samples was extrapolated regarding the intensity value at the corresponding *m/z* position with a tolerance of 0.3 Da. Since proteins can smear over several gel slices, a “Blur” function considering drifting proteins in gels was implicated with a tolerance of +/- 1 slice. All sub experiment files were merged in one Excel *csv* summary file and transferred to Statistica version 8/10. Moreover all normalized peptide levels, which contributed to the identification of a particular protein, were summarized for the protein. Protein levels were then used for statistical protein level determination. In case of LC ESI experiments, a further optimized *P2M* workflow was established (Wolters, 2011, Wolters, 2012) featuring *raw* files and Proteome Discoverer[®] result *mgf* file containing protein and peptide information for TIC normalization. Thereby, each particular peptide corresponding to an identified protein is searched with a tolerance between 0.02-0.05 Da considering experimental quality in all samples of the experiment that should be compared. Moreover a RT shift algorithm had been implemented in the software, which scans spectra for peptides within the RT range, where the peptide was primarily identified. Normalized peptides were summarized for each particular identified protein and transferred to a final Excel *csv* file for statistical analysis of protein levels in Statistica.

3.3.12 Protein functional annotation analysis

To determine origin, function and implementation of ocular surface proteins two different software solutions were used. Cytoscape version 2.8.3 with integrated BINGO 2.44 plugin

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(www.cytoscape.org), which enables gene ontology annotation of proteins, was used for the taurine tear study, whereas for the Taflotan[®] sine study additionally the Ingenuity[®] (www.ingenuity.com) IPA[®] tool system was used. Beside software based analysis, data has also been inspected manually by literature search.

3.3.13 Ocular surface studies

3.3.13.1 Taurine tear study design

The study was realized by use of the LC MALDI BU workflow focusing on capillary tears monitoring three age-matched groups ($N= 4$ individuals/group): two contact lens wearer groups (soft daily lens use >2 years), one applying 0.05% taurine eye drops (CL_Taurine, 25.5 ± 1.29 years); the other for control physiological NaCl solution (CL_NaCl, 25 ± 0.82 years) and a third taurine drops using a group of sicca patients (S_Taurine, 24.75 ± 3.86 years). Dry eye was diagnosed in the clinical ambulance by use of a Schirmer test II (BST <10 mm) and subjective dry eye ocular discomfort. Study exclusion criteria for probands were: Diabetes mellitus and/or eye diseases/medication probably influencing inflammatory processes or tear production. Accordingly, all participants were asked to avoid topical therapeutics during the study period and informed consent was obtained prior to the study. All protocols were approved by the institutional ethics committee and were conformed to the provisions of the 1964 Declaration of Helsinki. Individual tear samples were collected once a week (at study onset; after 3 days; after 1, 2, 4 and 5 weeks) from the left or right eye. Furthermore, tears of an additional group encircling healthy non-contact lens wearers ($N= 8$) were examined for verification. Individual tear samples of each group and time point were adjusted to 12 μg of total protein and pooled referring to time points. Accordingly, tear pools of each experimental group per time point (e.g. S_Taurine_3days) contained 48 μg total protein. Four gels were run in parallel (100 V, MES) encircling 18 lanes (18 time points/group pools) and one additional lane referring to the healthy non-contact lens wearer pool divided each in 15 slices/lane. Gel fixation, staining, in-gel digestion, peptide extraction and LC MALDI sample preprocessing were carried out as already described for LC MALDI BU analysis. Accordingly, tryptic peptides were fractionated by use of the developed capillary RP-RP LC MALDI system. Since each slice extract was fractionated over 24 spots on a MALDI target, one target corresponded to one sample lane (group tear pool per time point). In summary 19 study targets were prepared (3 groups \times 6 time points + 1 reference group) and analyzed following the described LC MALDI BU protocol. Protein identification was realized by creation of a *mgf* file containing MS/MS spectra of all 19 sub experiments (Fig. 23). Followed by the described *Homo sapiens* MASCOT/SwissProt search protocol with the exception that no fixed modification was selected since samples have not been reduced, also no variable modification was allowed considering MUDPIT scoring and $p<0.05$ for protein

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identification. After *P2M* transfer, normalized protein sum intensities were analyzed over the study period in Statistica using a linear regression model for the three experimental groups.. Proteins (scores \geq 35) were selected by their R^2 values. R^2 values \geq 0.5 in at least one of the groups qualifies the focused protein for further examination. Moreover, non-linear effects were determined by use of *Kruskal-Wallis* analysis regarding protein level medians over the whole study period of a particular protein. Therefore, regarding a particular protein, intensity levels of all time points of the taurine treatment groups (CL_Taurine, S_Taurine) were normalized to the start point level of the control group (CL_NaCl) to eliminate initial group specific differences not associated with the treatment. Normalized levels were used for *Kruskal-Wallis* followed by *post hoc* analysis. Moreover, a functional annotation analysis using Cytoscape was realized with candidate proteins. In addition, data were compared to the literature.

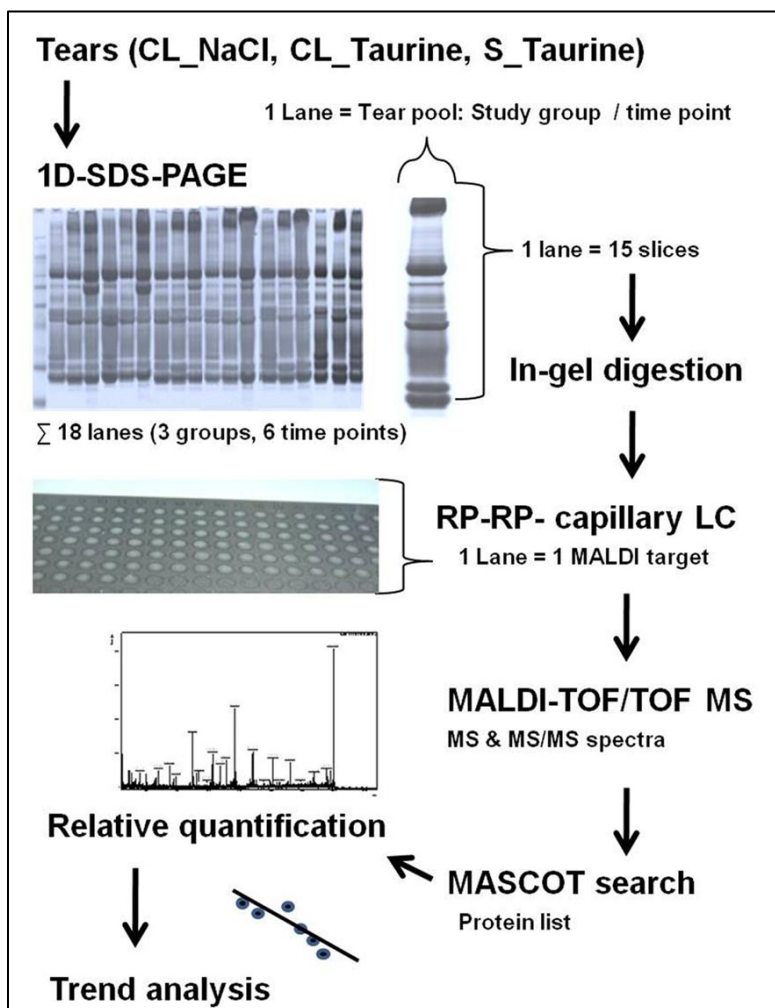


Fig 23 Study design of the longitudinal taurine tear study illustrating tear gels of 3 study groups and 6 time points (the additional reference sample, lane #19 is not shown). Samples were analyzed by the developed RP-RP LC MALDI BU workflow (Funke *et al.*, 2012).

3.3.13.2 Taurine conjunctival cell study design

To monitor taurine effects on stressed conjunctival cells (IOBA-NHC), cells were prepared in 24well plates for two stress conditions: staurosporine (30000 cells, 0.25 μ M/5 h) and H₂O₂ (40000 cells, 20 mM/1 h). Cell viability was examined by crystal violet assay and reactive oxygen species (ROS) level by DCFH-DA ROS assay, both performed in duplet for each taurine concentration. Moreover, each experiment was repeated four times. For LC ESI MS analysis 50000 cells/dish were seeded to pool for proper protein amount (taurine= 3xdishes/concentration; control= 3xdishes). Taurine solutions were freshly prepared in cell buffer featuring 3h cell incubation for each concentration: 100, 50, 20, 4, 2, 1, 0.5, 0.2, 0.05 mM inferred from the literature (Das *et al.*, 2011, Franconi *et al.*, 1985, Jong *et al.*, 2012, Muhling *et al.*, 2002, Pan *et al.*, 2010, Pan *et al.*, 2012, Pasantes-Morales, 1982, Weiss *et al.*, 1982, Wu *et al.*, 1999) followed by cell harvest, lysis and protein extraction, BCA assay, SDS PAGE and LC ESI MS. For the first experiment 6 gel lanes (50, 20, 4 mM; 3x control) were run, for the second experiment two duplicate lanes (2x 0.05 mM, 2x control) were run in parallel to monitor gel reproducibility. For the first experiment all lanes ($N= 6$), for the second experiment one taurine (0.05mM) and one control lane were prepared for LC ESI MS. Also, all samples were group randomized and measured ($\Sigma N= 6$) in the first experiment. Corresponding *raw* files of three samples (4 mM, 50 mM taurine; control) were combined for quantification. For the second experiment, each sample lane was measured in duplicate to additionally counteract technical bias, whereby *raw* files of all runs were combined. SwissProt corresponding normalized proteins were quantified using the described *P2M/Statistica* workflow. Statistical analysis was realized using a minimum 2fold level alteration threshold in confidence with the literature (Blagoev *et al.*, 2004, Choe *et al.*, 2005, Friedman *et al.*, 2004, Leal *et al.*, 2012, Mann and Kelleher, 2008, Molloy *et al.*, 2003, Ohlmeier *et al.*, 2004, Old *et al.*, 2005, Wilkins, 2006) for potential taurine responding protein candidates. Conjunctival proteome characterization was done by Cytoscape and literature screening. The complete workflow is depicted below (Fig. 24).

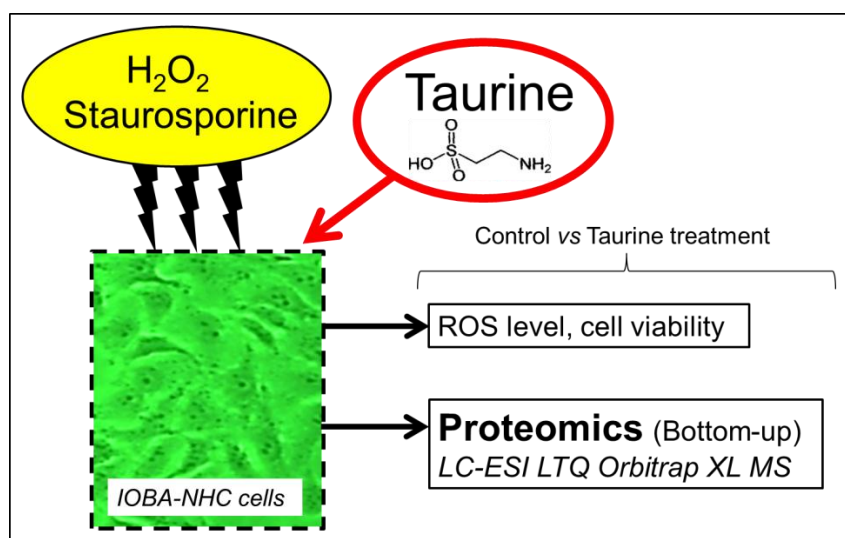


Fig 24 Study design of the taurine conjunctival cell study: IOBA-NHC cells were stressed with staurosporine or peroxide and incubated with different taurine concentrations followed by ROS and cell viability estimation and proteomic analysis of taurine treated cells by the LC ESI BU approach.

3.3.13.3 Taflotan[®] sine tear study design

The study was realized by use of the developed LC ESI workflow. Thereby, a group of POAG patients, who switched from the topical use of preservative-containing latanoprost (Xalatan[®], application \geq 6 month) to preservative-free tafluprost (Taflotan[®] sine) because of ocular surface discomfort symptoms, were focused on. Patients were diagnosed POAG in the clinical ambulance due to optic disc glaucomatous appearance and visual field. Moreover, only patients who displayed $IOP_{baseline} \leq 22$ mmHg and clinical dry eye parameters (BST $<$ 10 mm, TBUT $<$ 10 s) reporting subjective ocular surface discomfort, e.g. irritation or burning, have been included in the study. Informed consent was taken from all study participants prior to study onset and all protocols had been approved by the local ethics committee corresponding to the 1964 Declaration of Helsinki. Also, clinical parameters were recorded longitudinally including TBUT, BST, visual acuity and *superficial punctate keratitis* (SPK). Moreover, subjective life quality parameters referring to the *ocular surface disease index* (OSDI) (Schiffman, 2001) were documented. Regarding tear proteomic analysis at first longitudinal pattern changes should be monitored after the switch in a pilot POAG patient cohort by LC ESI MS BU explorative determination followed by microarray validation of MS derived candidate proteins and preselected dry eye markers in comparison to healthy subjects. For explorative LC ESI MS BU analysis Schirmer proteins were concentrated by 3K MWC spin filters recovering proteins ≥ 3 kDa by centrifugation (14000 g/30 min/4°C). To estimate filter effects, raw and filtered aliquots of a test tear pool ($N= 20$) were compared and filtrates were inspected for the occurrence of components ≤ 3 kDa by SDS PAGE, moreover evaluating dynamic range and suitability for MS analysis. A protein amount of 45 μ g/lane was found optimal addressing MS sensitivity. A first batch of patients ($N= 7$) was recruited for

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explorative MS analysis. However, samples corresponding to 4 weeks were missing in the sample set for two patients and a high protein content variance was found challenging for representative pooling and follow-up microarray (MA) analysis. Moreover, only three patients of the first batch were found to contribute enough protein over the remaining time points. Consequently, exclusively three patients ($N= 3$; 2 females, 1 male, 66 +/- 16 years) were monitored over four time points (0, 2, 12, 24 weeks +/- 1 week) in the explorative MS analysis. Accordingly, 15 μg protein/patient at a particular time point was pooled resulting in a sample pool of 45 μg /time point. Time point corresponding pools were analyzed by the developed LC ESI MS BU workflow. Peptide extracts corresponding to time point samples were run in duplet during LC ESI analysis. The “original run” SwissProt protein list was completed by proteins exclusively reported in the “replicate run” list ($p < 0.05$ MASCOT ion score, $\text{score} \geq 30$) in a combined *mgf* file for *P2M* quantification. Finally, normalized protein levels were transferred to Statistica for time pattern analysis averaging duplicate time point intensities for each identified protein analyzing dynamic intensity pattern by linear regression at first instance. To screen for protein candidates, which potentially respond in the course of time, only those proteins were selected as candidates, which significantly fit the linear model ($p < 0.05$) or display $R^2 \geq 0.9$. Thus, non-linear effects were examined by polynomial regression analysis. Thereby, only proteins displaying a significant fit to the polynomial model were selected as candidates. Additionally, a level increase or decrease was taken into consideration. Consequently, proteins which describe a distinct concave or convex shape in their time pattern without a clear slope direction were excluded from further analysis. Additionally, a fold change cluster method according to the “optimal matching” algorithm (Gabadinho *et al.*, 2011) was applied to the data to record non-linear approximately time dependent regulation pattern of proteins. Thereby, the *TramineR* package of the *R-project* was used. For pattern alignment three categories have been generated. The category “decrease” defined a minimum fold decrease of 0.5 of the protein intensity from a time point intensity value to the intensity value of the previous time point, whereas “increase” defined a minimum fold increase of 2 in the same manner and “equal” defined no change. The 2fold change value has been selected as significance threshold in consistence to the literature as already mentioned. Accordingly, 4 clusters have been calculated for the whole protein list. In a final approach the protein list was screened for already documented dry eye marker proteins and the time pattern of those “a priori” defined markers was screened with a lower linear regression threshold of $R^2 \geq 0.5$ to ascertain regulation tendencies in suspective dry eye proteins. Accordingly, linear, polynomial, fold change clustering and “a priori” selection were used to generate a final candidate list for Ingenuity[®] analysis and manual literature screening. In the second part of the study, MS observed tear dynamic pattern changes were validated by MA analysis using a higher number of subjects and comparing POAG patients

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($N= 16$, 70 \pm 10 years, 11 females, 5 males) with age-matched healthy controls without any ocular disorders ($N= 15$, 11 females, 4 males, 51 \pm 10 years) over five time points (0, 2, 4, 12, 24 weeks \pm 1 week) for POAG and over three time points (0, 4, 24 weeks \pm 1 week) for healthy subjects. Thereby, commercial available antibodies against selected representative MS derived candidate proteins, dry eye associated preselected markers and cytokines were immobilized on nitrocellulose slices and incubated with fluorescent labeled longitudinal tear protein samples. For analysis, a sciFLEXARRAYER MA spotter equipped with a piezo driven spotting unit (100-500 pl/drop) and a 428TM array laser scanner equipped with a green and red laser for fluorophor excitation (532/635 nm) were employed. MA preparation followed a laboratory-established workflow (Boehm et al. 2011, 2012; Bohm et al. 2011). Accordingly, 1 nl commercial antibodies was applied per spot on Oncyte Avid 6.5mm x 6.5mm nitrocellulose slides in triplicate and incubated with 7.5 μ g of fluorescent labeled (Cy5 Alexa Fluor 647) tear protein of each individual and time point to generate a longitudinal intensity pattern for each subject. Proteins, which could not be spotted properly, were excluded from further analysis. Spots were scanned in preview with 20 and 50 μ m and for quantification with 10 μ m resolution. Quantification was realized by use of ImaGene[®] version 5.5 exporting median spot and background signals to Statistica and difference was calculated between both signal median values. Negative difference values were excluded from the list, which contained triplicate antibody response intensities for each patient. Mean intensities encircling all group individuals (POAG vs Control) were calculated for each time point and each candidate. For regression analysis, only tendencies were focused on. To obtain a more accurate impression, if the mean intensity level of a certain protein converges from POAG to control, a time point comparison was operated. For that purpose, the mean of all time point intensities of the Control group was compared to the mean intensity of the initial and final time point of the POAG group estimating the confidential degree of the "POAG to Control level approximation" by t-test. A highly confidential approximation should therefore show significant intensity differences between the initial time point of the POAG group compared to the mean value encircling all time points of the Control group, but displaying non-significant differences between the final time points of the POAG group compared to the overall control group level. A brief overview on the study workflow is illustrated in Fig. 25.

Material & Methods

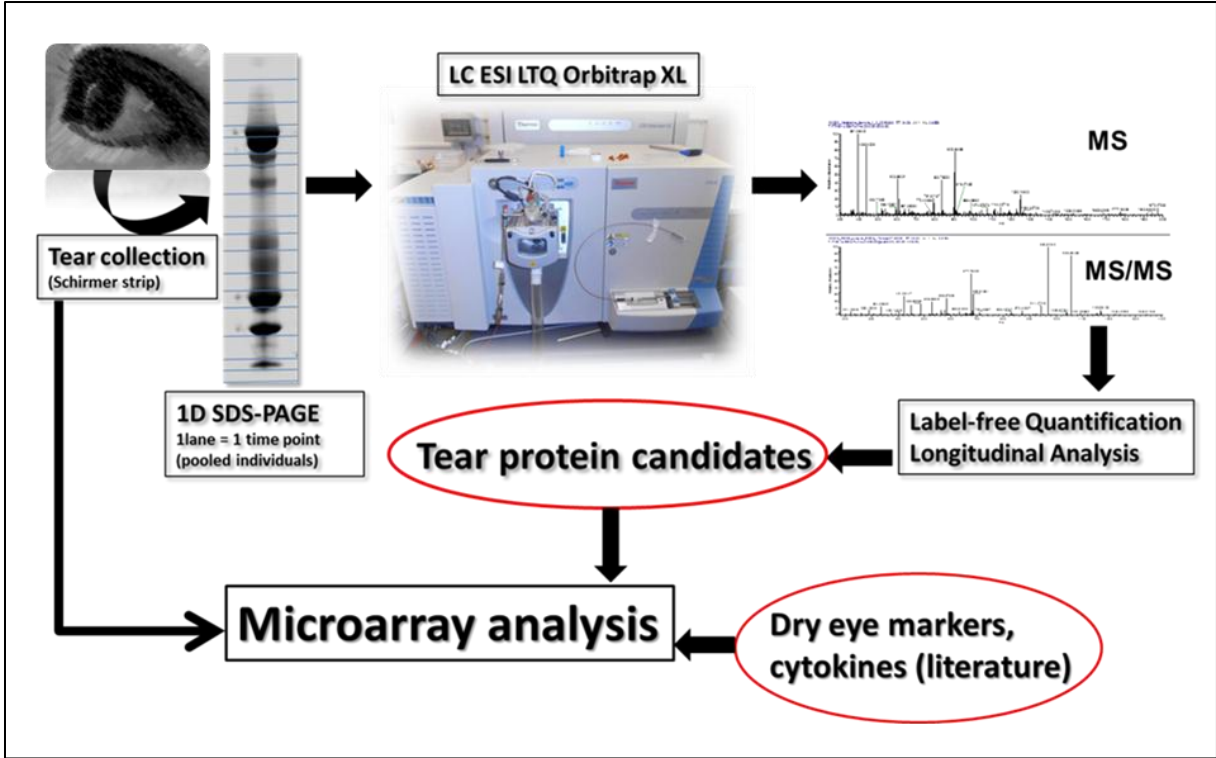


Fig 25 Study design of the Taflotan[®] sine tear film study: Schirmer tear samples from POAG patients were processed by the LC ESI BU workflow following statistical analysis of longitudinal protein pattern dynamics. Results were validated by use of MA analysis in a larger patient cohort including POAG patients and healthy controls. (Funke *et al.*, 2014)

4 Results

4.1 Microbore-RP-HPLC (for MALDI)

The developed column system was found appropriate for direct fractionation of crude samples without further sample pretreatment indicated by successful retaining of contaminant particles by the Security Guard™ cartridge (Fig. 26). Defined isolated fractions could be deposited on MALDI targets displaying low sample carry-over and homogeneous crystallization (Fig. 27). Regarding MS output, the method generates “high quality” linear MS spectra regarding m/z peak pattern of selected LC fractions (Fig. 28). Furthermore, high separation efficiency of the LC system could be achieved (Fig. 29). Compared to non-fractionated samples, an elaborative purification benefit could be registered for the HPLC prepared samples which resulted in a distinct increase of peak intensities (Fig. 30). Regarding training samples, the method showed a high level of run-to-run reproducibility (Fig. 31, 32). Retention deviation was estimated <12 s on average for a 3 s fraction deposition frequency monitored for selected reporter peaks 2481, 4962 and 6275 m/z based on first component detection (Fig. 33). The spotting time could be adjusted to a high degree of speed counting 4 min 36 s for each LC run. During training set runs, the HPLC running time for each run was 35 min and the complete HPLC running time of 24 samples including equilibration runs and automated matrix spotting could be recorded ($92 \times 24 \times 35 = 7782$ min + 24×10 Dummy runs = 240 min + $92 \times 24 \times 10$ s matrix spotting/spot = 368 min) <6 days (139.8 h). The linear MS measuring rate using 50 Hz laser frequency was set to approximately 44 s/spot. Accordingly, the study encircling analysis of all 6 study targets could be finished in approximately 27 h. In summary, the total analysis time including HPLC fractionation, matrix spotting and MALDI TOF MS did not exceed 7 days for 24 samples. Therefore, high quality MS pattern could be generated and the overall sensitivity of the approach prior to normalization was estimated by 856 RGC5 WARP-LC compounds corresponding to a mean number of 960 peaks detected in each LC run ($N = 24$). However, identification of exemplary reporter components 3789, 4279, 6648, 7208 and 8567 m/z detected in all samples was challenging. Direct fractionation of masses >4000 Da is critical in MALDI MS (Nilsen *et al.*, 2011) and even direct fractionation of peaks below this threshold was found problematic due to decreased reflector sensitivity. Only 3789 m/z could be detected by reflector analysis (Fig. 34), however, failing to reach threshold abundance for successful fragmentation. For this reason, an alternative strategy in reference to a comparable workflow (Boehm *et al.*, 2013) was chosen to clarify protein identities including ZIPTIP® C18 fractionation (2-40% ACN in 2% steps) of intact lysates, 1D SDS PAGE, intact protein extraction from the gel mass range of interest, MALDI linear reanalysis and in-gel digestion of the corresponding gel mass range for MALDI/LC ESI MS/MS analysis (Fig. 35, 36A). Despite reporters 3279, 4279 and 8567 m/z could be

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redetected in linear MS spectra corresponding to ZIPTIP[®] fractions (Fig. 35) only 8567 *m/z* could be successfully gel-extracted in its intact form, reprofiled in the MALDI linear mode (Fig. 36B) most likely corresponding to ubiquitin (gel area B: ESI: polyubiquitin B; score= 114; gel area C: ESI: polyubiquitin B; score= 365; MALDI: ubiquitin-40s ribosomal protein S27a; score= 134; polyubiquitin B; score= 102). Confidently, literature screening supported the observed correspondence of 8567 *m/z* to ubiquitin in MALDI (Niu *et al.*, 1998, Sanders *et al.*, 2008) and ESI spectra (Ebeling *et al.*, 2000, Scalf *et al.*, 2000). Regarding CNS, ubiquitin was identified as a peak at 8565 *m/z* in brain tissue sections (Pierson *et al.*, 2004) and at 8588 *m/z* in R28 cells by SELDI-TOF-MS (Brust *et al.*, 2008) supporting the finding. The reporter 6648 *m/z* was found to refer most likely to cytochrome C oxidase, which was detected as the predominant protein in corresponding gel areas (area A: ESI: cytochrome C oxidase subunit 5a; score= 132; area D: ESI: cytochrome C oxidase subunit 7a2; score= 69; area A: MALDI: cytochrome C oxidase subunit 7c; score= 58). Despite the failure for intact linear reprofiling, the attribution of the component to cytochrome C oxidase was supported by the literature according to MALDI brain studies (Bruand *et al.*, 2011, Hanrieder, 2011, Pierson *et al.*, 2004). Despite two clarified reporter compounds, several other proteins could be identified in the focused mass range including profilin, 10 kDa heat shock protein, thioredoxin, galectin, 40s ribosomal protein S21/28, protein S100 A6 and ATP synthase protein 8 (gel areas A, B, C, D) and keratines (gel areas E, F). For the remaining reporter compounds only literature matching attempts could be realized attributing 4279 *m/z* most likely to a β -amyloid polypeptide form exhibiting molecular weights of 4.2/4.3 kDa producing signals in the range of 4-5 kDa (LeBlanc, 1995, Lippa, 1993, McGowan, 2005, Rohner, 2005). The candidate marker at 3789 *m/z* most likely indicates calcitonin gene related protein neuropeptide (Goodman and Iversen, 1986, Jonhagen *et al.*, 2006, Kuwayama and Stone, 1987, Simon *et al.*, 2003), highly abundant in rodent CNS (Silverman and Kruger, 1989, Skofitsch and Jacobowitz, 1985) and also reported for tear fluid (Tsuji, 2012) as a peak of 3785 *m/z* (Mulvenna *et al.*, 2000). In summary, reporter mass identification was challenging leading to likely identification of ubiquitin and cytochrome C oxidase and critical records for β -amyloid peptide and calcitonin gene-related protein. Despite low protein identification performance, the LC microbore method was employed for the analysis of tear fluid, however with low success yielding clearly less intact signals compared to cell lysates, most likely due to the less complex nature of tear fluid in the small to medium proteome range compared to cell samples. Consequently, TD approach was found to enable fast, reproducible and robust analysis of crude sample materials, e.g. cell materials, but contrary it was hampered by insufficient sensitivity towards tear samples and performance was also decreased by difficult, intensive follow-up steps considering the risk of uncertain protein identification.

Results

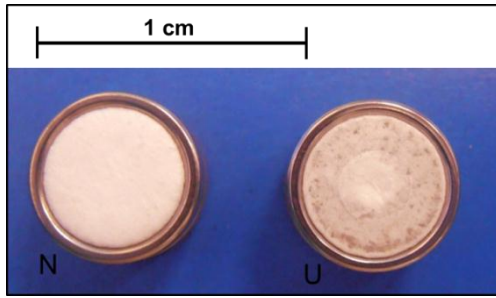


Fig 26 Successful retainment of contaminant particles of crude RGC5 test lysates by the direct coupled Security Guard™ cartridge indicated by contaminant spots on the filter RP1 resin (right) in comparison to a fresh filter (left).

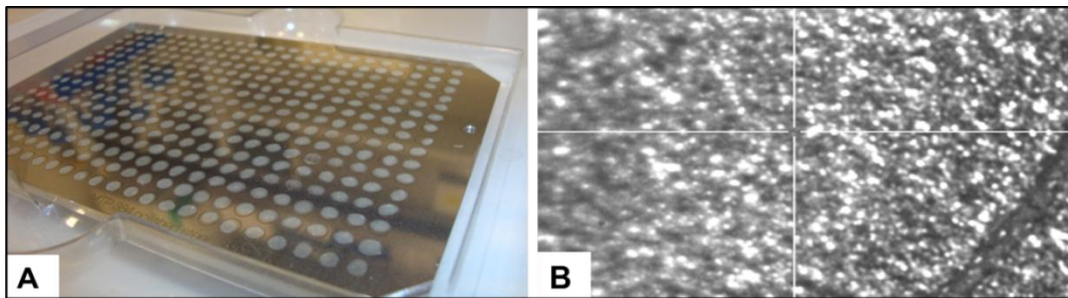


Fig 27 Microbore fractions after automated matrix application resulting in A: distinct separated spots displaying B: homogeny crystal distribution allowing accurate representative measurements.

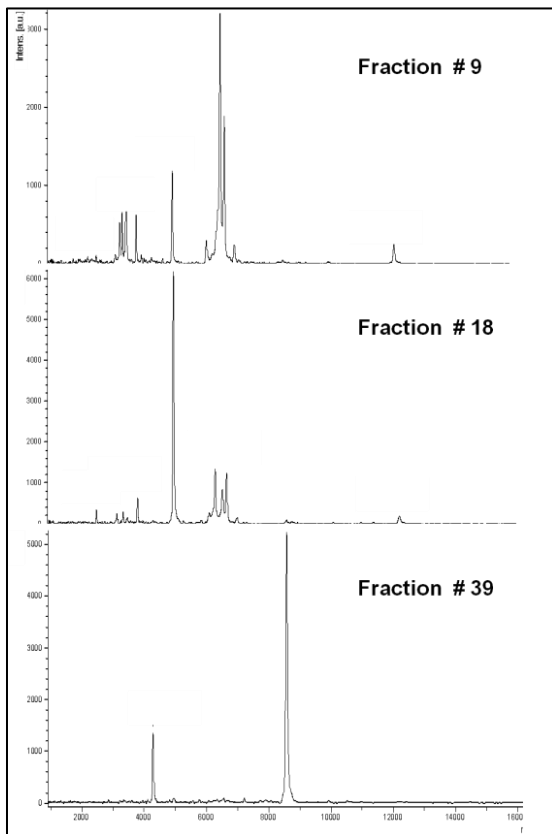


Fig 28 Exemplary high qualitative linear microbore MS spectra corresponding to selected microbore LC MALDI TD fractions of crude RGC5 test sample lysates.

Results

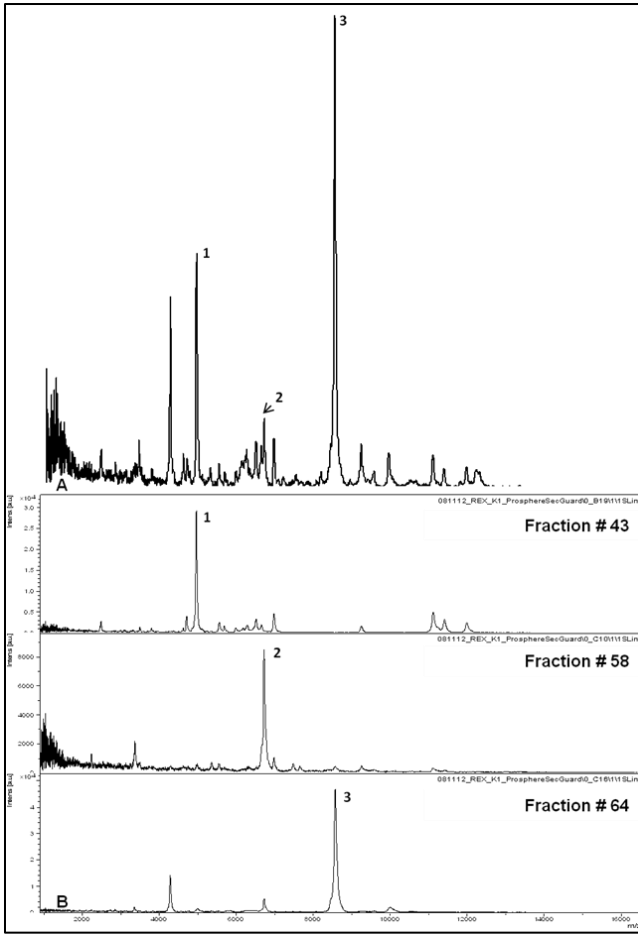


Fig 29 Separation efficiency of the microbore LC MALDI TD approach indicated by three reporter peaks; A: Purified RGC5 sample without fractionation showing the three reporter peaks with noise background; B: Efficient separation of the three reporter peaks in three defined fractions by microbore HPLC.

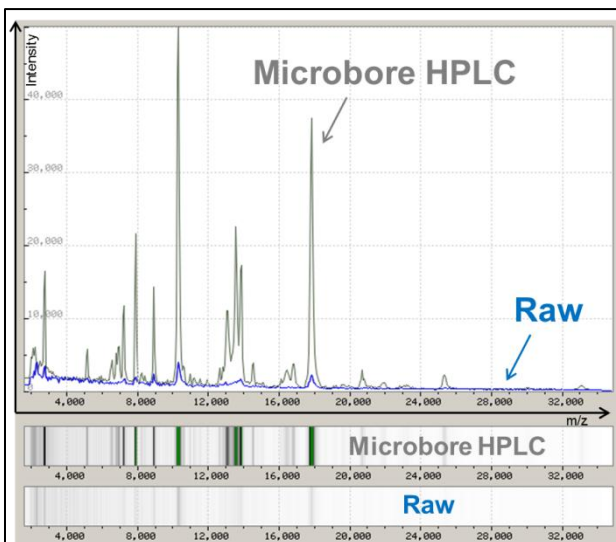


Fig 30 Linear MALDI sensitivity increase obtained by microbore HPLC: Whereas the analysis of raw unfractionated RGC5 test samples showed only low intense peaks (blue MS spectrum) by use of microbore LC high intensity values for peak species could be obtained indicated in a merged HPLC run spectrum corresponding to 92 fractions (grey). The difference is also illustrated in the gel-like view below.

Results

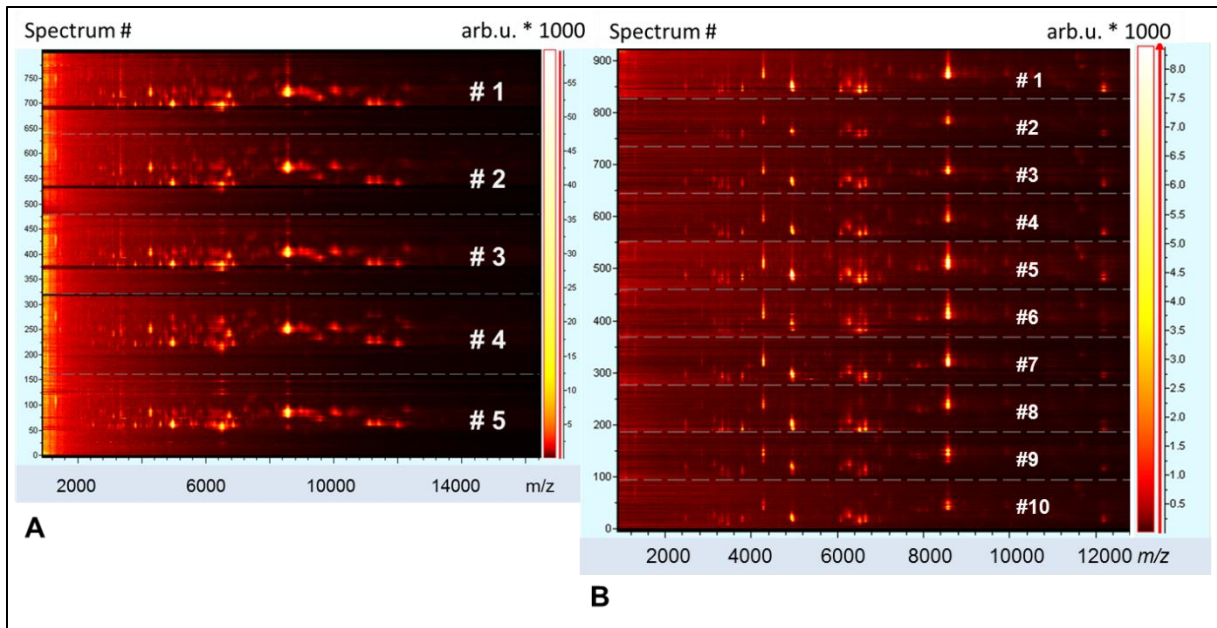


Fig 31 Survey View of run-to-run reproducibility corresponding to A: 120 fractions/sample (retina sample replicates; $N= 5$) and to B: 92 fractions/sample (RGC5 samples; $N= 10$) showing high congruency in signal pattern.

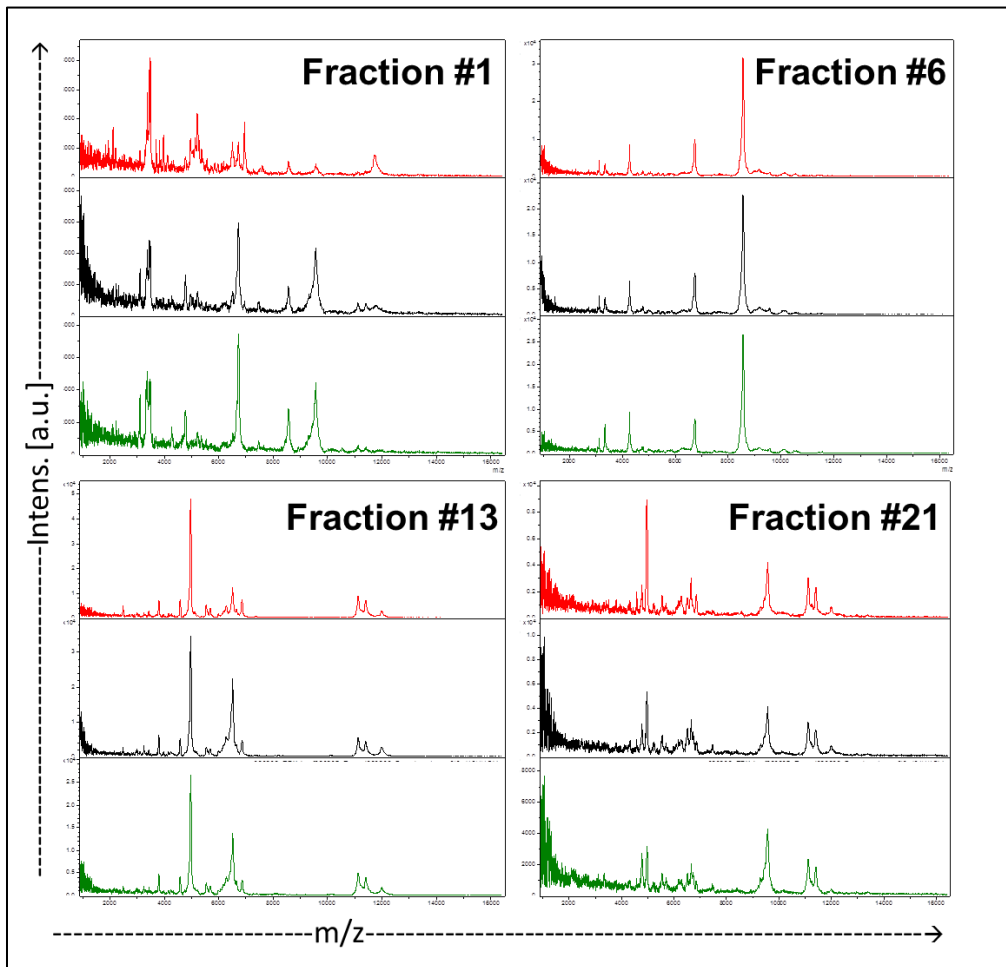


Fig 32 Run-to-run reproducibility indicated by linear MS spectra corresponding to exemplary fractions of three similar processed retina samples showing high congruency in peak pattern.

Results

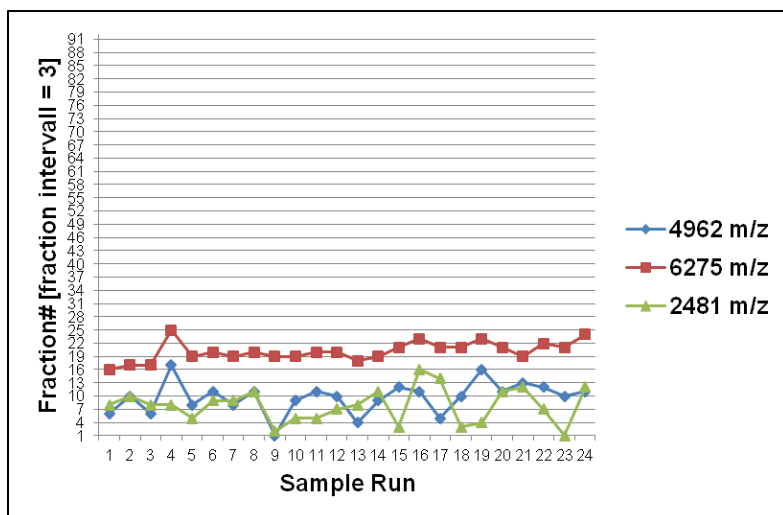


Fig 33 Retention time (RT) reproducibility indicated by three reporter peaks detectable in RGC5 test sample runs (N= 24) showing a low degree of fraction shift (<12 s for 3 s/fraction).

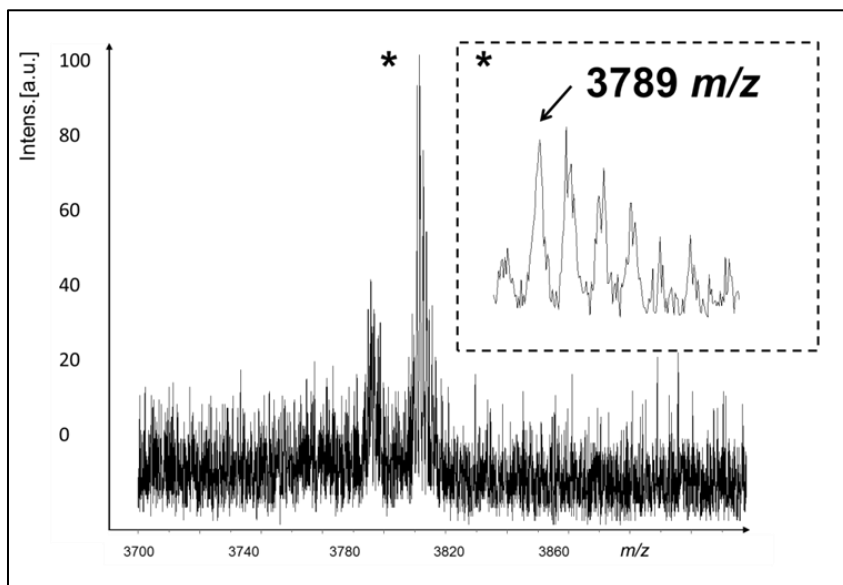


Fig 34 Redetection of reporter peak 3789 m/z in the reflector mode: Despite the successful monoisotopic peak recognition (asterisk), the intensity was too low for fragmentation and MS/MS related identification.

Results

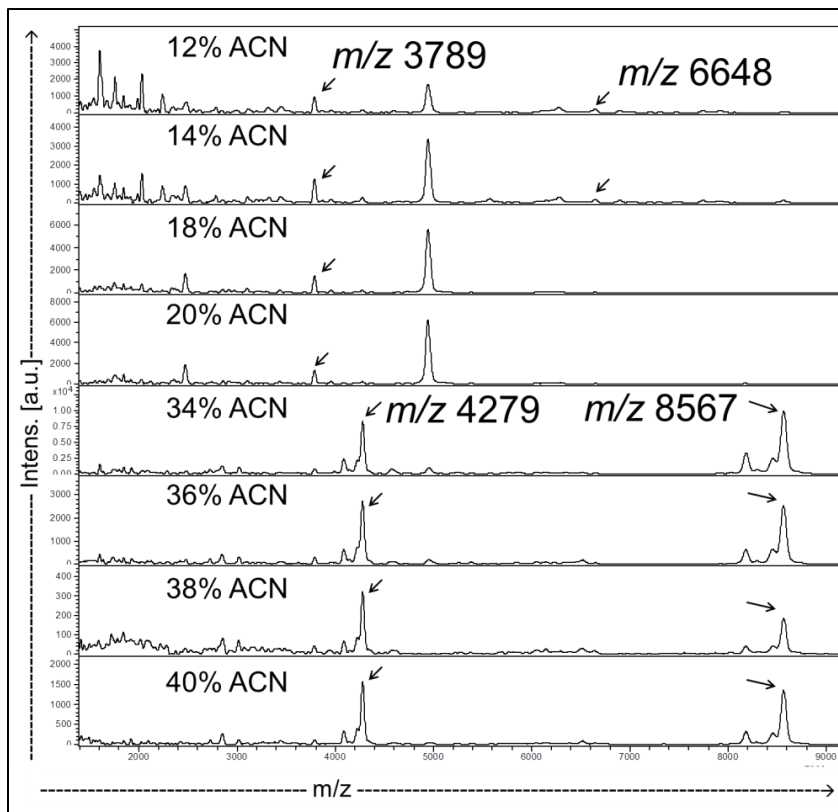


Fig 35 Purification of four exemplary reporter masses by use of ZIPTIP® C18 2% ACN step purification for follow-up identification. The four components could be captured between 12-40% ACN.

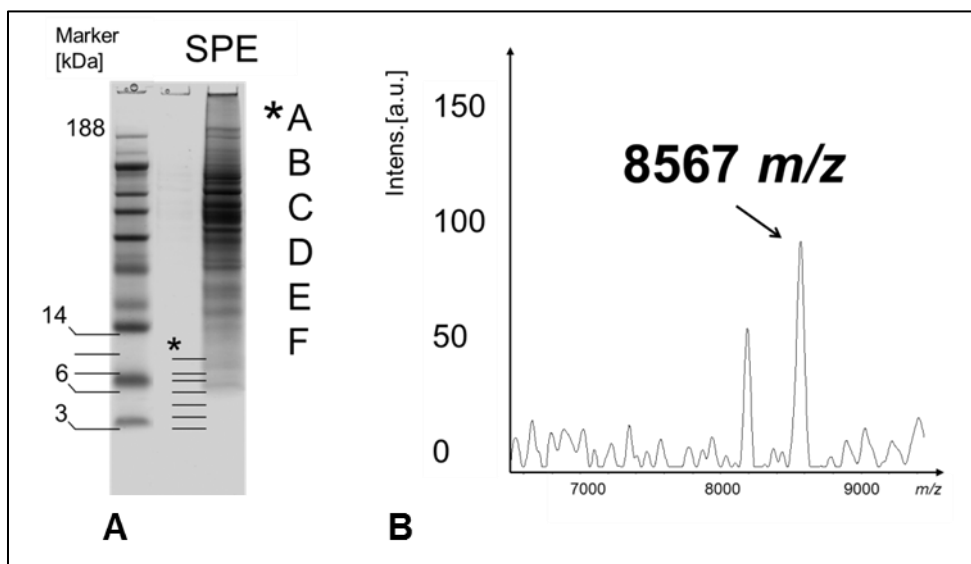


Fig 36 Reprofile and follow-up identification attempts of selected reporter peaks; A: ZIPTIP C18 purified 12-40% fraction (SPE) with focused mass regions (asterisk: A-F) between 14 and 3 kDa for reporter reprofile by intact extraction, linear MALDI reprofile, in-gel-digestion and identification by MALDI/ESI MS/MS analysis; B: Only reporter 8567 m/z could be successfully reprofiled with low intensity after gel intact extraction in the MALDI linear mode.

Results

4.2 Capillary-RP-RP-HPLC (for MALDI)

For initially performance evaluation of the workflow, training samples were focused on. In case of cell lysates ($N= 12$), the combination of two sample pools lead to the detection of 13246 tryptic WARP-LC compounds. Thereby, 8180 MS/MS measurements lead to the identification of 294 proteins under stringent *MUDPIT* scoring conditions using SwissProt database. Compared to common proteomic strategies focusing on RGC5 cells used as test reference in this work, the sensitivity of the developed gel-based LC MALDI workflow was found comparable to the literature regarding approximately 300 (Yang and Tezel, 2005), 400 (Kim *et al.*, 2006, Tezel *et al.*, 2005, Zhang *et al.*, 2008) and 1000 detectable 2D gel protein signals (Kanamoto *et al.*, 2009). Using 2D gels followed by MALDI TOF MS Kim *et al.* reported 318 significant protein matches in RGC5 lysates (Kim *et al.*, 2009b). Also a 2D-nano-HPLC MALDI TOF/TOF MS/MS shotgun workflow focusing on lysates generated from in vivo rat retinal ganglion cells resulted in the identification of 268 proteins (Crabb *et al.*, 2010). In confidence with these studies, the present workflow showed an appropriate sensitivity also taking the stringent scoring strategy into consideration. Moreover the approach was proven suitable for lower amounts of material since 1D instead of 2D SDS PAGE was used as first fractionation dimension, which also simplified protein selection from the gel since instead of single spots whole mass regions were selected. Therefore, the workflow was also more economic regarding analysis time and laborative effort and was found suitable for the analysis of study tear samples. Accordingly, regarding capillary tear fluid by use of the developed workflow, a combinatory database search including 5535 MS/MS spectra obtained from two capillary sample pools including individual tears ($N= 8$), lead to the detection of 5640 peptide compounds and 90 significantly identified proteins using stringent *MUDPIT* scoring conditions. Inspection of raw data showed high qualitative MS spectra corresponding to LC fractions and also high qualitative MS/MS fragment spectra could be recorded (Fig. 37). Regarding the output of a large tear sample set, the taurine study was focused on. Single gel lanes in the tear study showed 3425 WARP-LC compounds, which were detected on average across all experiments ($N= 19$ lanes) leading to the identification of 267 significant unique tear proteins under stringent scoring condition (Fig. 38). In comparison, using 2D SDS PAGE 63 (Ananthi *et al.*, 2008) and 243 (Herber *et al.*, 2001) capillary tear protein spots have been detected which fits well with the results of the present study. Using SELDI-TOF MS 56 protein peaks in a range of 1500-20000 m/z (Tomosugi *et al.*, 2005a) and >1000 below 50000 m/z (Grus *et al.*, 2005b) have been described. Referring to protein identifications, only few works based on gel or MALDI techniques provide comparable results. For example, 30 capillary tear proteins have been identified by N-terminal Edman sequencing from 2D gels (Molloy *et al.*, 1997). Moreover high reproducibility of the HPLC system monitoring elution behavior of three different mass region

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reporter peaks detected across investigated samples could be documented. Accordingly, a fraction shift of ± 0.8 fractions focused on a reporter ion 1792.92 m/z found in HPLC runs of the slice #9, a fraction shift of ± 0.9 fractions referring to ion 1400.73 m/z found in chromatographic runs corresponding to slice #13 and a shift of ± 0.8 fractions related to ion 2849.88 m/z detected in runs of slice #3 could be documented. Moreover, peptides showed a high degree of purity since CBB could be retained from MS analysis. In fact, an additional aromatic selectivity of the system could be supported by the effective retention of CBB traces originated from roughly destained test samples during method development. Thereby, CBB signals indicated by a deep blue staining of the spot matrix surface were detected in the final fractions of RGC5 test samples corresponding to higher solvent condition and reflecting a strong retention of the component on the stationary phase (Fig. 39). Regarding the measurement protocol, several limitations of the MALDI system could be revealed. Differences in sample and matrix spotting could lead to crystal caldera formations challenging homogeneity analysis leading to quality differences in the MS output. Another drawback is the high number of unassigned MS/MS spectra in database searches also reported in the literature (Kapp *et al.*, 2005, Katz *et al.*, 2007, Ning *et al.*, 2010), which could be due to database records or to analysis limitations, encircling spectrum quality and peak detection tenacity. Regarding the SNAP algorithm, it was found in some cases, that a misleading peak in near neighborhood of the real monoisotopic peak was selected as parent mass precursor, which can lead to inaccuracies in fragmentation and MS/MS pattern assignment. Besides, post-processing of MS/MS spectra regarding smoothing and noise reduction increased the MS/MS annotation rate, thereby increasing also the number of scores of already identified hits. However, this approach was evaluated as highly time consuming and therefore not suitable for large datasets.

Results

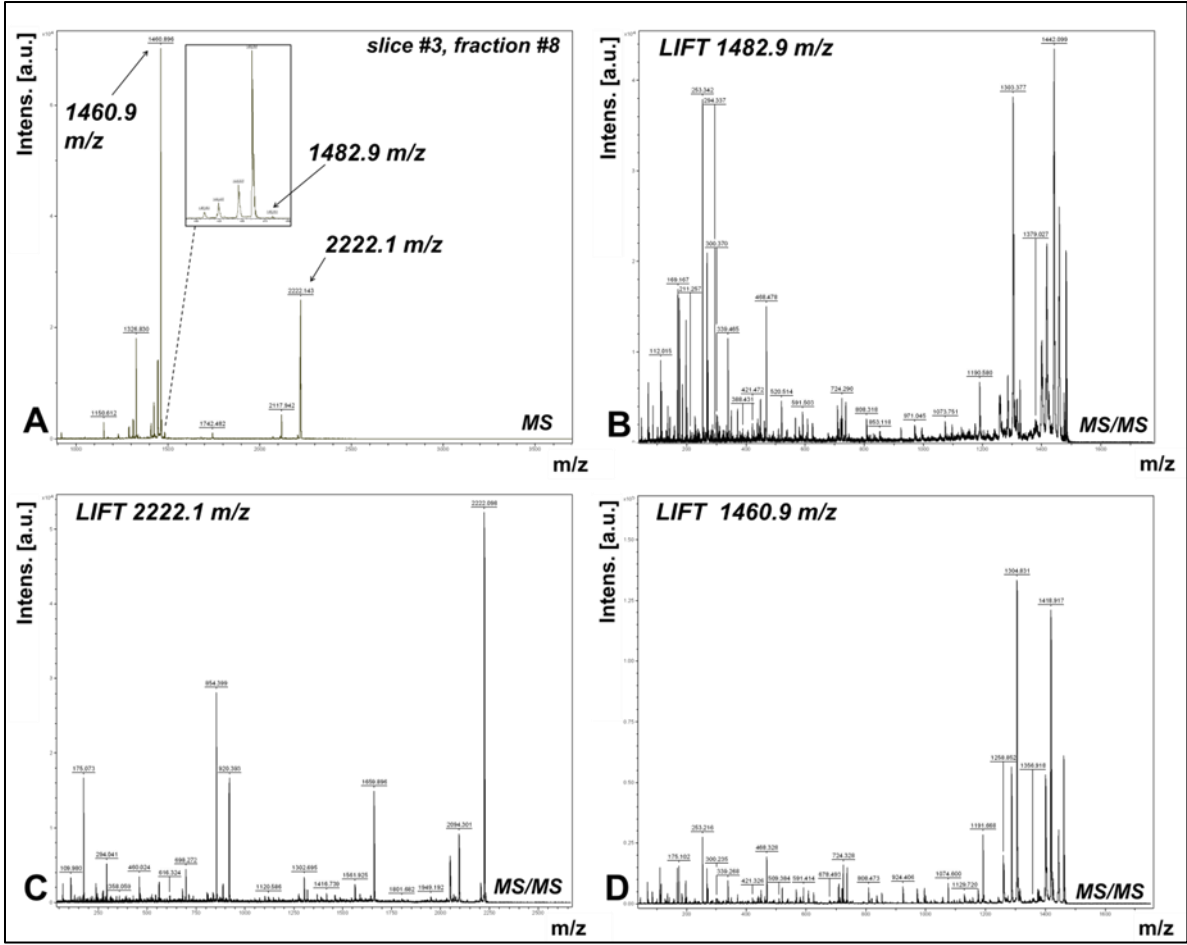


Fig 37 Performance of the capillary RP RP LC MALDI BU approach; A: Detection and resolution of three exemplary masses showing low (1482.9 *m/z*, zoom view), medium (2222.1 *m/z*) and high intensity (1460.9 *m/z*); leading to B-D: high qualitative fragment MS/MS spectra.

Results

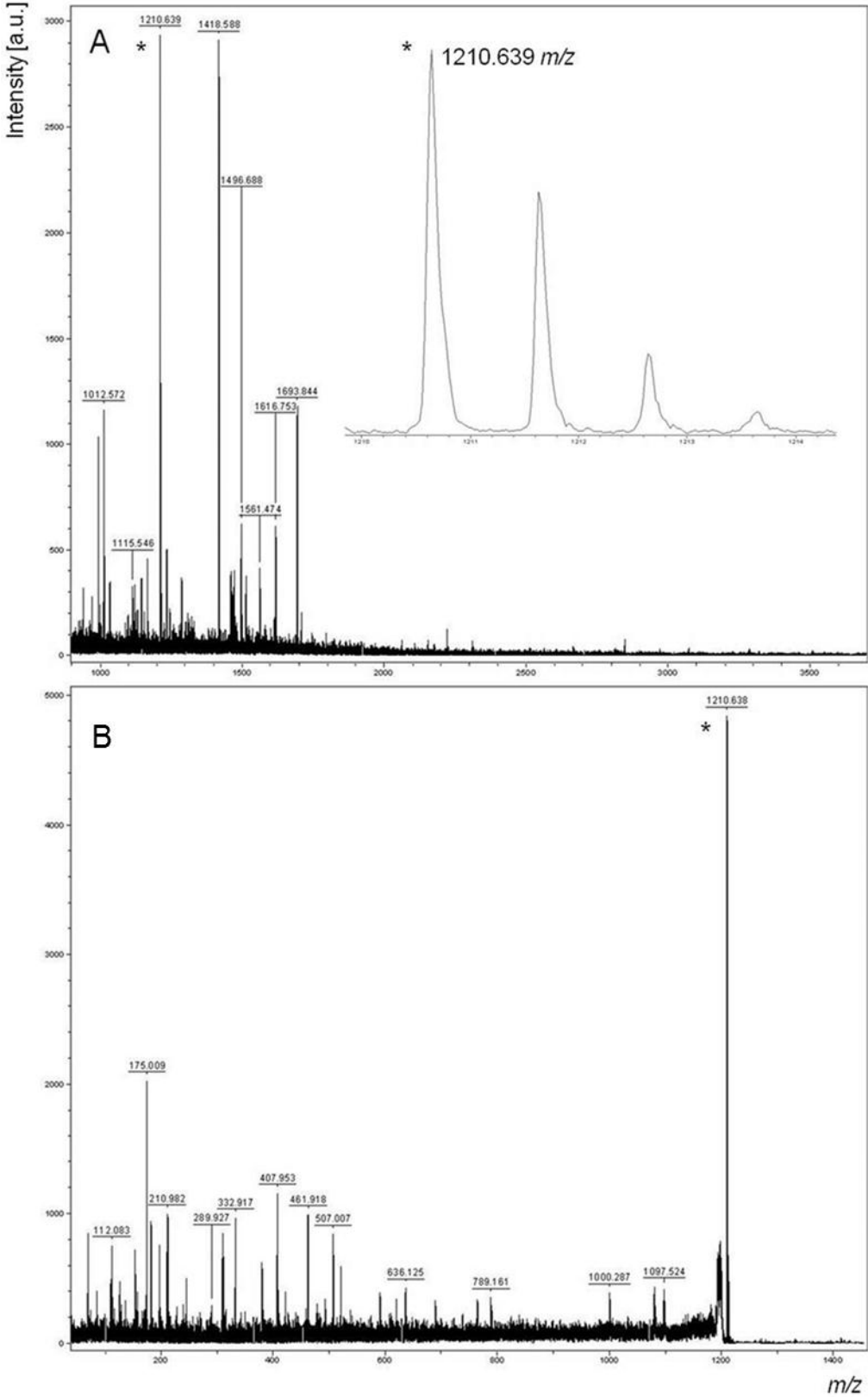


Fig 38 Exemplary succesful protein identification for proline-rich protein 4 (PRP4) by fragmentation of A: high resolution precursor 1210.639 m/z (asterisk, zoom view) leading to B: high qualitative MS/MS spectrum allowing identification of PRP4 (Funke *et al.*, 2012)

Results

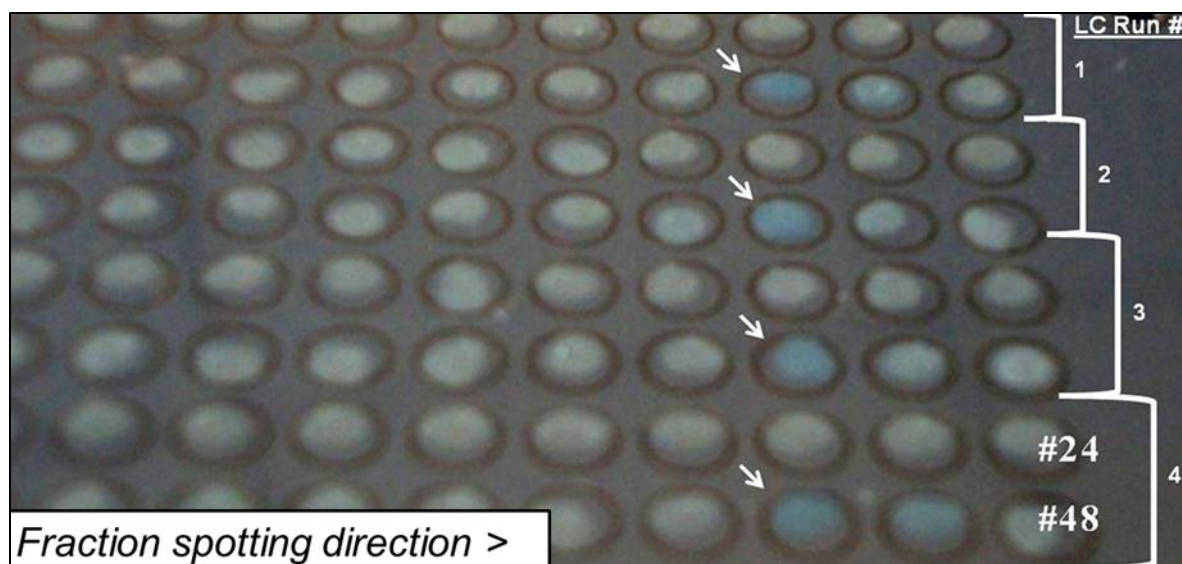


Fig 39 Strong CBB retention indicated by strong reproducible blue crystal staining in final fractions (indicated by arrows) of 48 fraction training sample test runs supporting additional aromatic selectivity of the developed capillary RP RP HPLC system (Funke *et al.*, 2012)

4.3 Capillary-RP-HPLC (for ESI)

Employing the 80 min LC ESI gradient, the total processing period for one gel lane corresponding to one sample was calculated to ~1.3 days (17 x 80 min gradient + 17 x 30 min washing run = 1870 min = 31.17 h). To reduce the analysis time, at first washing runs have been eliminated. De facto, after chromatographic inspections, no distinct signs of carryover were found. The intensive chromatographic inspection of representative replicate RGC5 slice digests (1/17 lane, mass area #2, #4) (Fig. 40) showed, that predominantly peptides eluting from 20 to 40 min of the 80 min gradient area contributed to significant protein identification (Fig. 41). Taking a system dead volume of ~1.6 min (tubing: 0.005" ID, 0.127 $\mu\text{l}/\text{cm}$, 86 cm length; flow rate 6.7 $\mu\text{l}/\text{min}$ = 0.127 x 86 = 10.922 μl ; 10.922 $\mu\text{l}/6.7 \mu\text{l}/\text{min}$ = 1.6 min) into consideration, most likely the elution profile reflected the observation, that mainly hydrophobic peptides favored for ionization in ESI MS, which is in confidence with the literature (Stapels *et al.*, 2004). Adjustment of the starting mobile phase composition to 15% B saved analysis time and increased sensitivity in case of the 80 min gradient. An optimization in 5 min steps referring to the main elution area between 20 and 40 min of the reference 80 min gradient (Fig. 42) showed a distinct sensitivity decrease below 35 min, whereas from 80 to 35 min a slight decline could be observed. In accordance, the range between 40 and 80 min was focused on for further optimization (Fig. 42). The proportional adaption of this "optimum gradient range" in 10 min-steps resulted in a significantly linear increasing number of protein identifications for both, relaxed and stringent scoring strategies (MASCOT ion score $p < 0.05$: $R^2 = 0.98$, $p = 0.0112$; peptide score ≥ 30 : $R^2 = 0.99$, $p = 0.0052$). Consequently, as a compromise between analysis time and sensitivity, the optimized 50 min gradient was selected as study setting which lead to a total analysis time of ~0.6 days per

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sample (lane) (17x50 min [analytical gradient] + 1x 30 min [washing run] = 880 min = 14.67 h) representing a 54% reduction of analysis time. The final optimized 50 min LC ESI workflow showed a reproducible efficient peptide elution pattern with the majority of masses observed between 10 and 40 min RT (Fig. 44) and a high chromatographic congruency of technical replicates of corresponding gel mass areas indicated by inspection of TIC chromatograms (Fig. 45). A high RT reproducibility inferred from time monitoring of reporter ions ($n=20$) observable in all technical replicates ($N=6$) of investigated gel mass areas ($N=8$, M1-8) (Fig. 43) could be demonstrated. Thereby, the mean CV was 0.62% ($SD=0.57\%$) with a minimum CV value of 0.15% for reporter peak 1067.00 m/z observed at 34.02 min mean RT in all replicates of gel mass area M6 (Fig. 46A) and a maximum CV value of 2.37% for reporter peak 544.33 m/z observed at 16.65 min mean RT in all replicates of gel mass area M2 (Fig. 46B). The reproducibility values of the system fits well with observations reported by the literature for comparable systems reporting a range of 0.17-4.5% *RSD* (Duan et al. 2009) and 0.1-2.6% *RSD* (Dunn et al., 2008). Furthermore, high analysis accuracy observing most of high scoring masses with an error of approximately 2-4 ppm (Fig. 47) could be achieved. Also, a reproducibility of quantitative sensitivity was determined with a CV of 1.94% regarding the number of CID fragmented precursor ions, which means that in replicate runs nearly identical numbers of peptides have been detected. Thereby, the qualitative overlap of CID precursors was 21% in all replicates, after outlier extraction increasing to 57% in 4/6 of the samples. The analysis of CID fragment congruency lead to a heterogenic image of fragmentation since the overlap of investigated precursors showed a mean value of 22% with a *SD* of 28% with a maximum value of 64% and minimum value of 0% depending on estimated precursor ions (Fig. 48). These observations conclude a relatively high congruency in precursor detection and selection but a lower degree of precursor fragmentation control. Resulting consequences for protein characterization were investigated on in the next steps. The sensitivity of the initial 80 min workflow towards RGC5 samples ($N=8$) was estimated with a mean of 1521 +/- 128 proteins (MASCOT ion score, $p<0.05$) and 633 +/- 72 proteins using the more stringent scoring strategy (peptide ion score >30) decreasing only slightly in the optimized 50 min gradient workflow (Fig. 49, 50). Regarding the qualitative reproducibility analysis, a congruency of 33% in all samples to 76% corresponding to half of the sample set was observed in cell samples under relaxed scoring condition. Additionally, a congruency of 49% in all samples to 88% in half of the sample set using more stringent condition could be determined regarding the different cell samples run on different days on different gels focusing on the 80 min gradient method. The optimized 50 min gradient workflow resulted in a mean overlap of 52% in all technical replicates ($N=6$) to 76% overlap in 4/6 of replicates encircling all investigated gel mass areas ($N=8$) under relaxed scoring condition. By use of more stringent scoring the overlap was increased to 63% in all replicates and 88%

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congruency observed in 4/6 of the replicates (Fig. 51). Thereby, the observed reproducibility values were in confidence to the values inferred from the literature (Crabb personal communication 2013, Mezhoud *et al.*, 2008, Tabb *et al.*, 2010, Zhang *et al.*, 2005). In summary, the results emphasize the robust performance potential of the developed workflow with drawbacks in fragmentation control. Compared with the literature the optimized gel-based LC ESI workflow showed a high degree of sensitivity and was found to distinctly exceed gel-based as well as MS related protein detection approaches and was therefore employed for the examination of conjunctival cells and tear fluid studies.

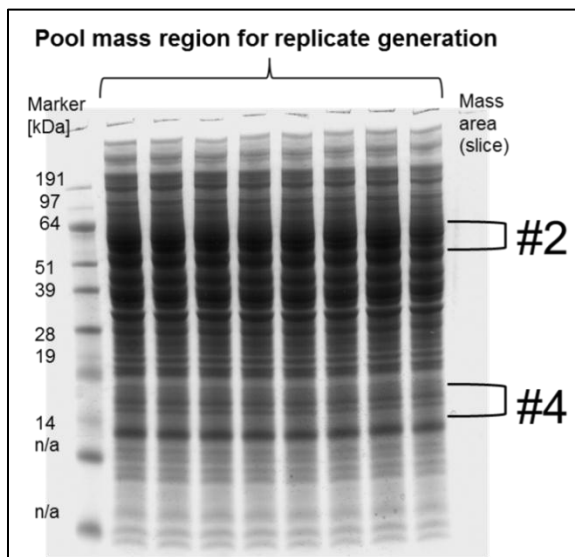


Fig 40 Training sample replicate gel for method development containing 80 μ g RGC5 lysate/lane; mass area #2 and #4 were depicted pooled, digested and processed and split to identical replicates immediately before LC ESI MS analysis.

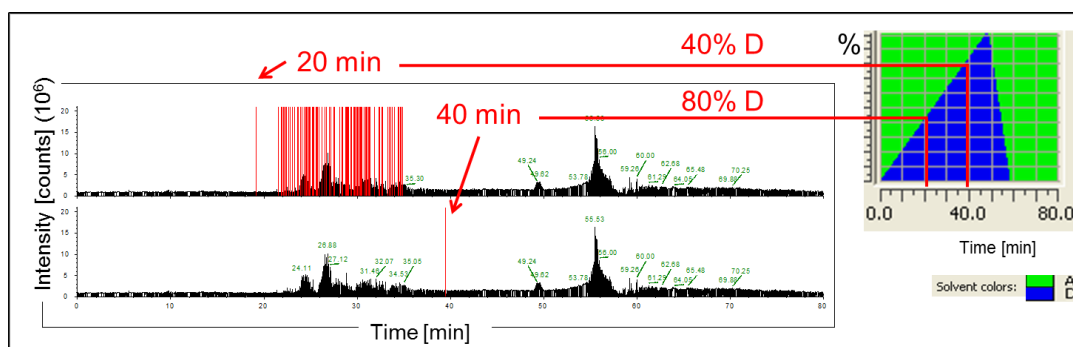


Fig 41 Chromatographic view of the initially developed 80 min HPLC gradient indicating the RT range containing spectra, which contributed to successful protein identification. Proteins were identified between 20 and 40 min. RT, which represents 40-80% running buffer B. The chromatographic inspection was used as reference for follow-up gradient development.

Results

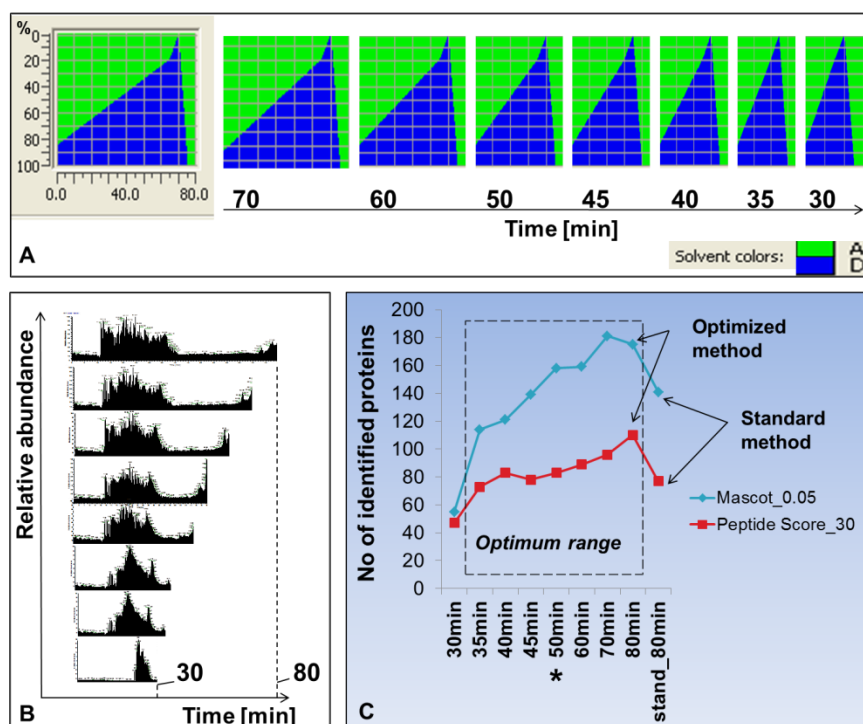


Fig 42 LC ESI gradient development; A: Stepwise gradient adaption (5 min steps) to reduce analysis time; B: chromatograms corresponding to adapted gradients showing a distinct elution signal decline correlating with decreased gradient time; C: Sensitivity obtained in the course of gradient adaption and optimization, whereby an optimized 50 min gradient was selected as final optimum method (asterisk) due to a compromise between sensitivity and analysis time.

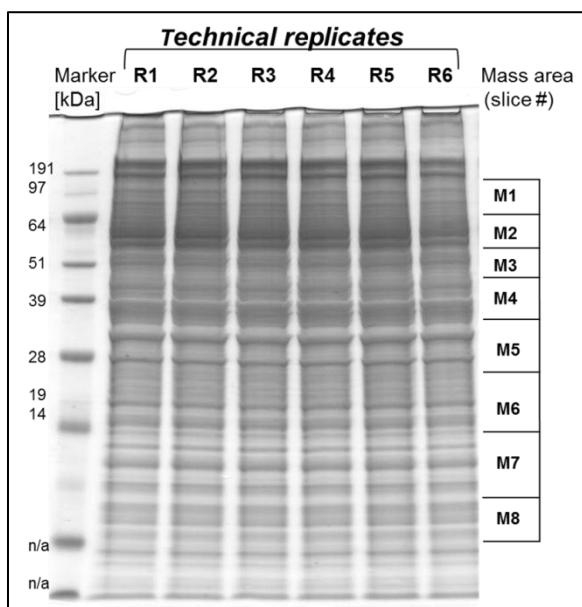


Fig 43 Training sample replicate gel for reproducibility testing off the developed LC ESI MS BU approach. Technical replicates for MS analysis were selected from RGC5 replicates (80 µg/lane) focusing on 8 mass regions (M1-M8). Mass area replicates (e.g. M1, R1-R6) were selected, pooled, in-gel digested ZIPTIP® purified and split to identical aliquots for LC ESI MS analysis.

Results

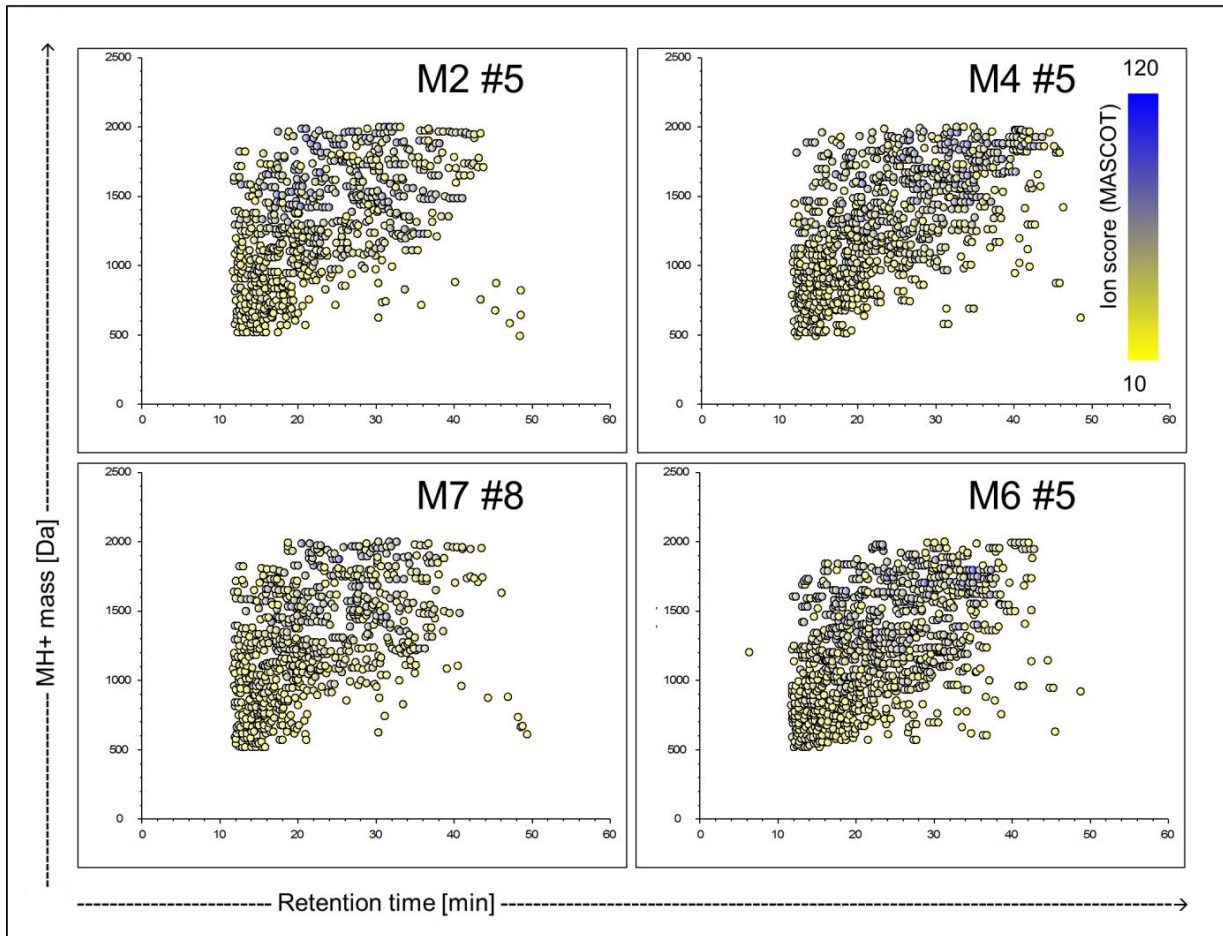


Fig 44 Exemplary replicate elution profiles of identified peptide masses corresponding to the developed 50 min gradient showing high congruency and optimal fit to the gradient time range.

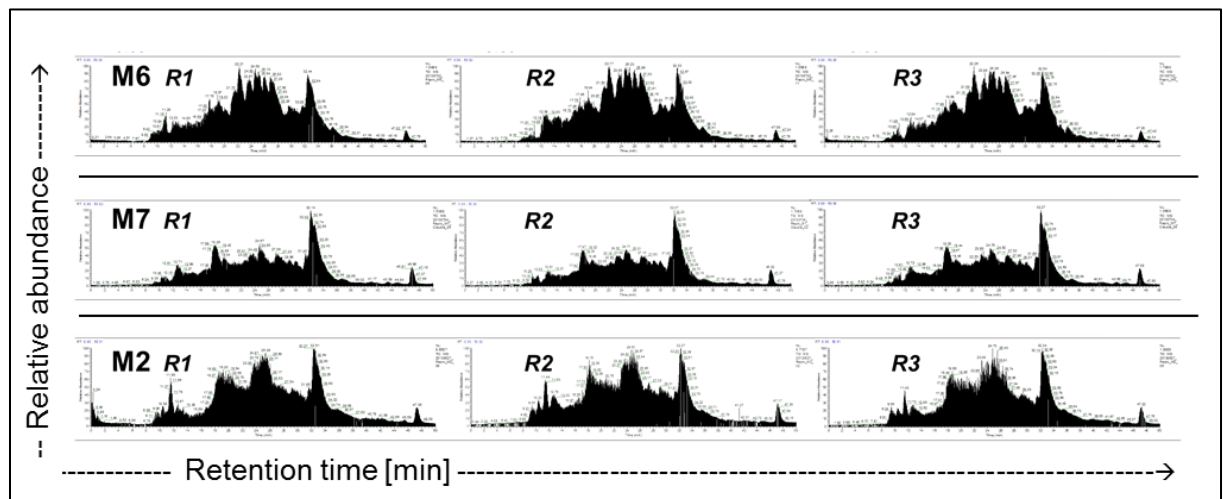


Fig 45 Exemplary TIC chromatograms corresponding to three exemplary replicate runs for three exemplary mass regions (M6/7/2) showing high elution pattern congruency indicating high technical reproducibility of the HPLC system using the 50 min gradient method.

Results

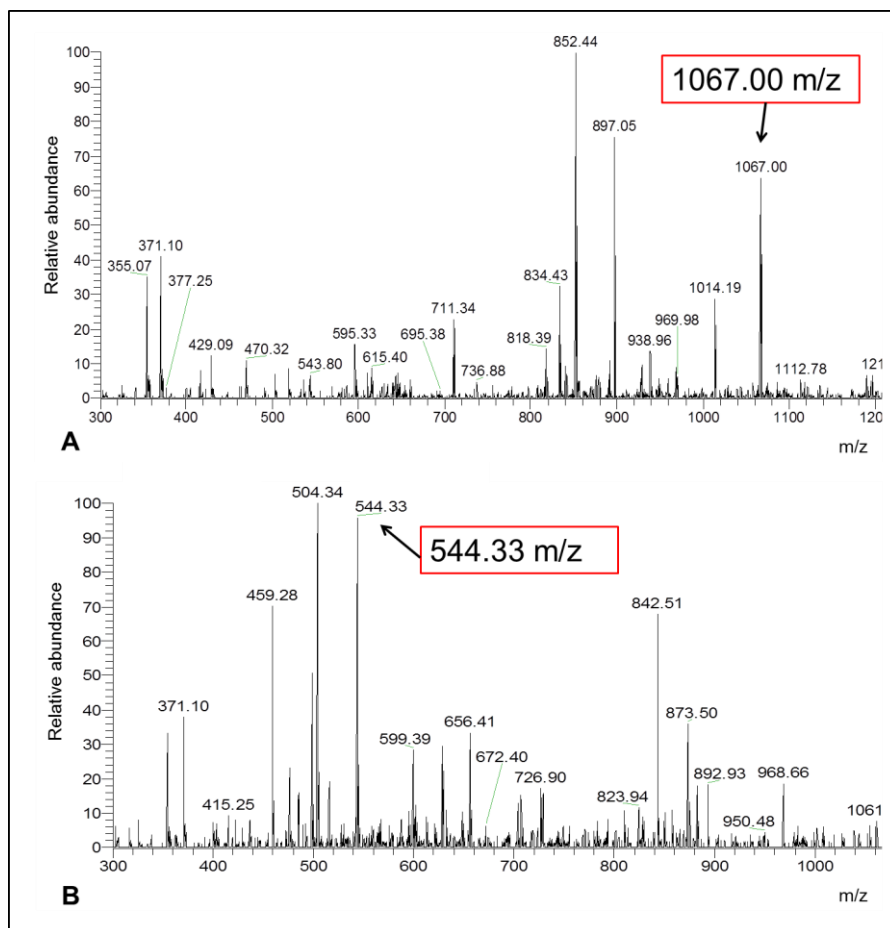


Fig 46 Two reporter ions representing minimum and maximum RT reproducibility values inferred from RT monitoring of A: 1067.00 *m/z* observed at mean RT 34.02 min in all replicates of M6 and showing a 0.15% CV; B: 544.33 *m/z* observed at mean RT 16.65 min in all replicates of M2 showing a 2.37% CV.

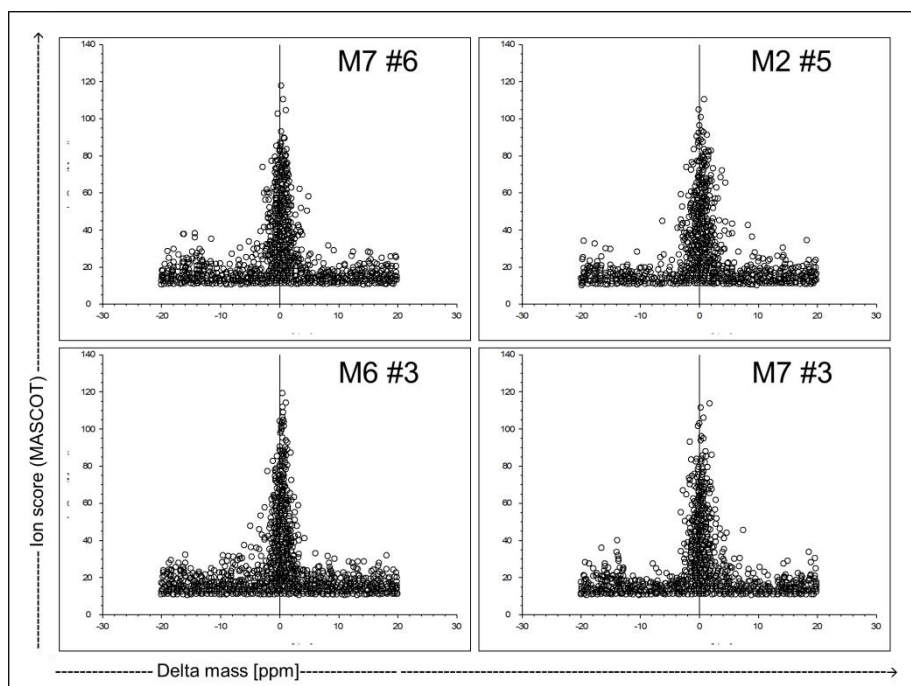


Fig 47 Mass accuracy achieved during 50 min gradient LC ESI MS analysis exemplary shown for exemplary mass region corresponding technical replicates displaying the majority of high scoring masses detected with 2-4 ppm deviation.

Results

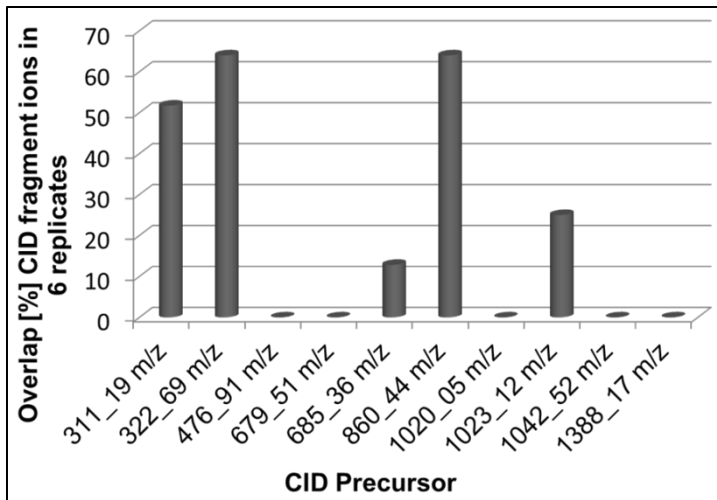


Fig 48 CID fragmentation observed for reporter precursors ($N= 10$) displaying a highly heterogenic pattern; whereas some reporter ions show high qualitative congruency in obtained CID fragments regarding technical replicates (e.g. 322.69 m/z) several reporter precursors (e.g. 476.91 m/z) should no overlap in fragment contingents.

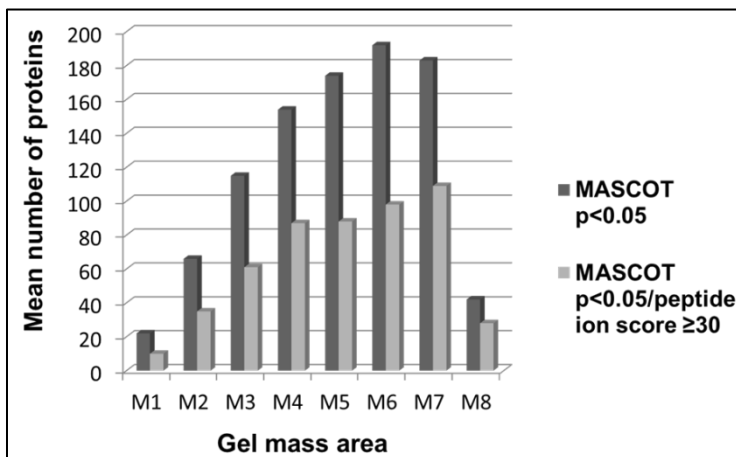


Fig 49 Mean protein output regarding test mass areas (M1-8) by use of the 50 min gradient LC ESI MS method.

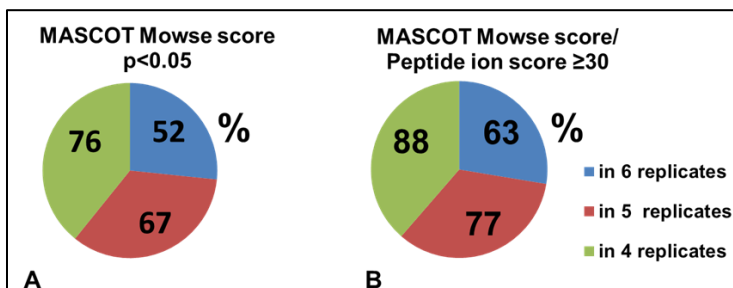


Fig 50 Qualitative reproducibility of the 50 min gradient LC ESI MS BU approach illustrated by identified proteins with two scoring strategies; A: relaxed; B: stringent.

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4.4 Proteomic characterization of ocular surface sample species

4.4.1 Capillary tears

Gel images provided a clear pattern of capillary tear proteins with distinct areas of protein migration with respect to protein reducing and protein amount (Fig. 50). The LC MALDI method (15 mass areas/lane) as well as the LC ESI workflow (17 mass areas/lane) (Fig. 51) provided a deep view to the tear proteome in a mass range between 3-188 kDa. Moreover, as already shown for training samples, SDS PAGE showed a high degree of reproducibility inferred from pattern congruency based on pool tears of healthy subjects ($N= 6$) (Fig. 52). In summary, >1000 proteins could be documented with both approaches. Actually, the MALDI approach yielded in 267 proteins from non-reduced capillary tear samples (appendix 1), which was significantly less sensitive despite consuming a higher number of sample lanes ($N= 19$) in comparison to the LC ESI approach, which provided a significant higher sensitivity resulting from the analysis of only few sample lanes ($N= 3$). Also, the LC ESI approach screening the Swissprot database and implementing an additional Mascot *MOWSE* score threshold value of 35 yielded in the identification of 1427 tear proteins (appendix 2). The output clearly exceeded the results of former “in depth” mapping approaches, identifying 491 (de Souza *et al.*, 2006) and 589 (Zhang *et al.*, 2009) human tear proteins and was comparable to the sensitivity obtained with a high speed Triple TOF workflow leading to the identification of 1543 human tear proteins (Zhou *et al.*, 2012). Regarding the protein ranking in the present work, the 10 top scoring proteins in ESI analyzed capillary tears were lactotransferrin (score 1982), complement C3 precursor (score 1680), basement membrane-specific heparan sulfate proteoglycan core protein (score 1669), serum albumin precursor (score 1448), keratin, type II cytoskeletal 1 (score 1389), titin (score 1300), zinc- α -2-glycoprotein (score 1091), mesothelin (score 845), keratin, type I cytoskeletal 9 (score 742) and polymeric immunoglobulin receptor (score 741). In case of MALDI results the top three prominent tear proteins identified with distinct higher scores than in ESI experiments were lactotransferrin (score 16010), lipocalin-1 (score 15876) and proline-rich 4 protein (score 15116). MALDI identified proteins ranking below these top scoring tear proteins exhibited a rapid score decrease up to 32% like e.g. observed for lysozyme C (score 4800), albumin (score 4415) and polymeric immunoglobuline receptor (score 4283), whereas lipocalin-1 (score 704) was found among the top 20 scoring proteins. In contrast to the MALDI results, proline-rich protein 4 was found as protein number 51 in the scoring rank (score 271) in the ESI analysis. Another example is zinc- α -2 glycoprotein (score 2514), which was found among the top 10 scoring ESI detected proteins, whereas it was found in MALDI experiments at a distinct lower score rank (rank 17). A high portion of proteins was congruently detected using MALDI and ESI ionization techniques, e.g. clusterin, α -2-macroglobin, mammaglobin B, ceruloplasmin, annexin A1, haptoglobin or serotransferrin. Despite several proteins

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overlapped with relatively high scores in both, the MALDI and ESI approach, some proteins, e.g. ribosome-binding protein 1 or neutrophil gelatinase-associated lipocalin were found with comparable low scores in result lists of both workflows supporting their existence in tear fluid. Whereas the ESI workflow provided higher sensitivity leading to an increase in protein identifications and identifying hundreds of proteins, that failed to be detected by the MALDI workflow, several proteins like transthyretin (score 241), apoptosis-inducing factor 2 (score 145), fibrinogen (score 37), eukaryotic translation initiation factor 2A (score 152), complement C4 A (score 50) or Nck-associated protein 5 (score 53) could exclusively be detected by LC MALDI. Since both workflows depend on a comparable gel-based fractionation strategy and moreover the MALDI approach depends on a more sophisticated LC technique highlighting a two phase column system, these findings most likely reflect the influence of the used ionization method on the output of the tear fluid analysis. Regarding the 267 capillary tear proteins, which could be identified in the taurine study by MALDI the majority referred to intracellular proteins (Fig. 53A), most likely originated from ocular surface epithelia, whereby most of the detected cellular tear proteins were found to be localized in the cytoplasm, whereas only a minority, were found to be derived from the membrane. Actually, the majority of organelle associated proteins were related to the nucleus. Moreover, a major part of MALDI identified tear proteins was found to be linked to the immune system (Fig. 53B). In accordance, woundhealing related proteins could be documented with high portion (Fig. 53B). Regarding the congruency of the MALDI protein list with the literature an overlap of 23% with the list of de Souza et al. (de Souza *et al.*, 2006), of 50% with the list of Li et al. (Li *et al.*, 2005), of nearly 50% with the lists of Zhou et al (Zhou *et al.*, 2009b, Zhou *et al.*, 2006), of 74% with results of Green-Church et al. (Green-Church *et al.*, 2008) and of 84% with the list of the Cojokaru group (Cojokaru *et al.*, 2011) could be documented. In particular, many of the detected proteins could be annotated to multiple functions, e.g. deleted in malignant brain tumors 1 (score 1305), which had been characterized as a cancer-related multifunctional protein (Mollenhauer *et al.*, 2001) or phospholipase A2 (PLA2), as a membrane traffic involved protein (Brown *et al.*, 2003). Beside PLA2, several other lipid-binding proteins could be detected, e.g. lipocalin-1, apolipoprotein A I (score 425), a reverse cholesterol transport plasma protein (Frank & Marcel 2000) or zinc- α -2-glycoprotein (score 2514) playing a key role in lipid recruitment and immune response (Hassan *et al.*, 2008). Ceruloplasmin (score 39) was reported as a secretory protein from the human lens epithelial cells (Harned *et al.*, 2006), which makes it interesting to be detected in tear fluid, however, it could only be reported with low score. Regarding 1427 capillary tear proteins, identified by LC ESI MS, the list showed a constant high congruency level to the literature. Regarding gene ontology annotation analysis, 73% of all protein hits could be annotated to a cellular component. Of annotated proteins, 88% were intracellular, whereas only 12% were

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extracellular originated, which fits well with the results from the MALDI analysis (Fig. 54A) and reflects the proposed involvement of ocular surface epithelia as tear protein source (Gipson, 2007). Like MALDI, also ESI analysis revealed several lipid-binding proteins like apolipoprotein A I (score 398) or phospholipid transfer protein (score 412), which has already been documented for human tear fluid and has been supposed to play a key role in the formation of the tear film with special regards to the lipid layer (Jauhiainen *et al.*, 2005). Transcobalamin-I (score 416), a vitamin B12 binding protein (Hall, 1975), could be detected in tear fluid for the first time by LC ESI MS. Regarding intracellular proteins, two groups could be generated: cytoplasmic and membrane proteins. Thereby, 40% of intracellular proteins were found to be originated from membranes (Fig.54B), which to some degree is contrary to MALDI experiments. According to protein localization, most of the intracellular proteins were found to be associated to the nucleus (44%) (Fig.54C), which is in confidence with MALDI annotation results. Nuclear pore membrane glycoprotein 210-like (score 131), heterogeneous nuclear ribonucleoprotein U (score 48), serine/threonine-protein kinase ICK (score 84) or nuclear receptor-interacting protein 2 (score 75), nucleobindin 2 (score 324) were typically nucleus associated proteins that could be documented in capillary tears by LC ESI MS. Thereby, nesprin 2 (score 250) is a multi-isomeric protein that binds lamin at the nuclear envelope building a subcellular network (Libotte *et al.*, 2005, Padmakumar *et al.*, 2005) and was detected in this work in tears for the first time inferred from literature screening. Prominent intracellular enzymes like glyceraldehyde-3-phosphate dehydrogenase (score 246), α -enolase (score 334) or E3 ubiquitin protein ligase RBBP6 (score 134) could be described for tears and are most likely originated from ocular surface epithelia. Confidently, several epithelia derived protein markers most likely originated from the cornea or conjunctiva could be detected in the tear samples, e.g. periplakin (score 94), desmoplakin (score 263), symplekin (score 94), cadherin 23 (score 83), cadherin EGF LAG seven-pass G-type receptor 1 (score 65), cadherin-like protein 26 (score 62), protocadherin 16 (score 51), protocadherin γ -A11 (score 51), catenin- α 1 (score 48), galectin 3 binding protein (score 447), and various cytokeratines by LC ESI MS. The occurrences of these proteins reflect the strong relationship between epithelia and tear film and could be used to estimate the integrity of cornea and conjunctiva. In contrast, the MALDI analysis did not show any of the ESI detected epithelial marker despite a variety of cytokeratines. An antimicrobial membrane corneal epithelial protein marker, mucin 16 (score 245) (Laurie, 2008, Spurr-Michaud *et al.*, 2007) could be detected in capillary tears in this study by the LC ESI workflow. However, by use of both approaches, proteins reported to be associated to ocular surface dryness could be revealed in capillary tear fluid, e.g. lysozyme C, mammaglobin B, protein S100 A 8/A9 and proline-rich protein 4 (Aluru *et al.*, 2012, Boehm *et al.*, 2013). In fact, despite overlapping proteins, a distinct difference between MALDI and ESI maps exists. Nevertheless, a deep

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insight view to the capillary tear proteome could be achieved by use of both techniques, while the LC ESI workflow was shown to provide clearly higher sensitivity, analysis speed and lab performance lacking only few proteins, which had been revealed by the MALDI approach. On the other hand the MALDI workflow performed excellently for the detailed analysis of defined abundant tear proteins referring to high scores achieved for tear proteins like lactotransferrin, lipocalin-1 or proline-rich 4 protein. To complete the proteomic view of the fluid component of ocular surface, in the next step, Schirmer collected tears have been examined.

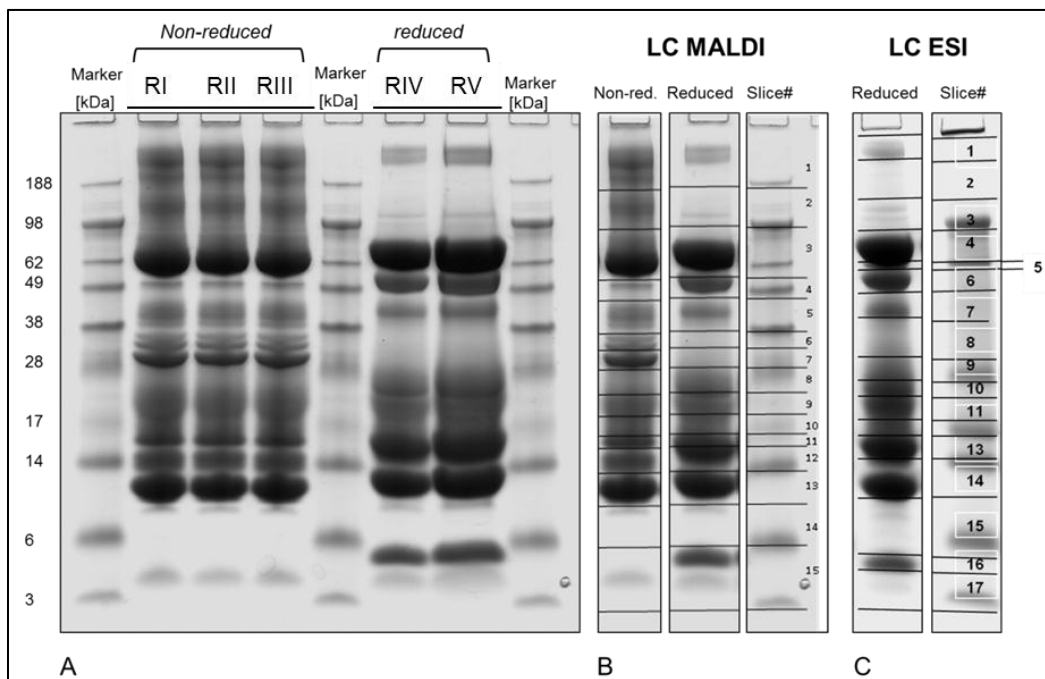


Fig 51 SDS PAGE migration pattern of microcapillary collected tear proteins and selection for LC MALDI/LC ESI BU analysis; A: Replicates of a representative tear pool ($N=6$) showing distinct pattern (RI-IV: 50 μg ; RV: 80 μg) and differences due to sample reducing; B: Mass area selection for LC MALDI (15 slices/lane); C: Mass area selection for LC ESI (17 slices/lane).

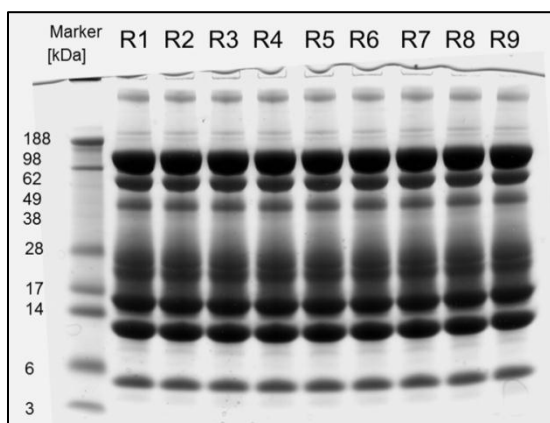


Fig 52 Tear pool ($N=6$) replicate (R1-9; 50 μg /lane) runs: The runs show a high degree of pattern congruency indicating high reproducibility of the SDS PAGE system justifying its use as first dimension for MS BU analysis.

Results

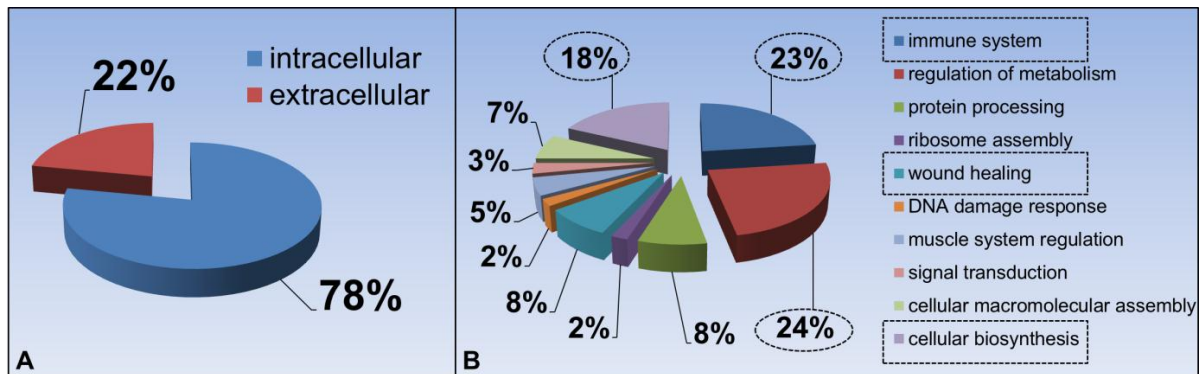


Fig 53 LC MALDI BU characterization of the capillary tear proteome by Cytoscape GO annotation; A: the majority of capillary tear proteins were found to be intracellular proteins; B: Predominant tear protein contingents refer to immune system, woundhealing and cellular biosynthesis.

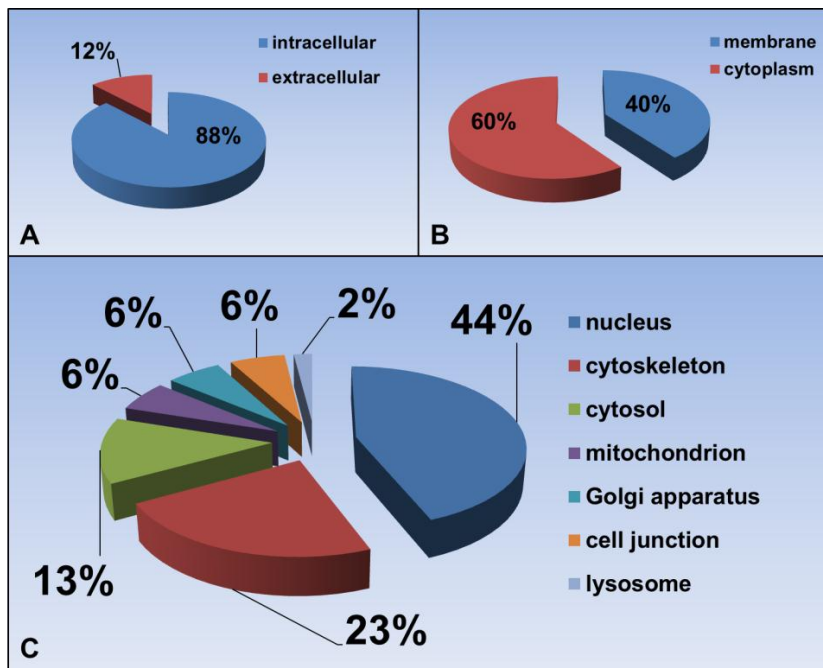


Fig 54 LC ESI BU characterization of the capillary tear proteome by use of Cytoscape GO annotation; A: Mostly intracellular proteins were detectable; with B: a predominant portion of cytoplasmic proteins; C: Most cytoplasmic proteins were originated from organelles with a predominant nucleus protein contingent.

4.4.2 Schirmer strip eluted tears

As a first step in Schirmer tear analysis, prior to the Taflotan[®] sine study various tear protein concentrations had been tested to estimate the optimal sample condition and to ensure representative pooling of individual tear samples taking fluctuations of tear protein amount between patients and time points of sample collection into consideration. Gel pattern showed linearity in a range from 2 to 80 µg total protein referring to CBB abundance. A 9 µg protein

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gel load lead to visualization of characteristic tear protein migration areas, whereby nearly all bands could be recognized at 15 µg (Fig. 55). Therefore, 15 µg were found to be suitable to generate a representative tear proteomic snapshot of each patient contributing to the pool sample. Furthermore, there was no difference in the gel pattern comparing 3 kDa MWC filter concentrated Schirmer eluates to crude Schirmer strip eluates which vindicated the use of MWC filters for protein purification and concentration allowing follow-up future studies on the samples, in which salts and small molecular compounds have to be removed, e.g. binding assays. Filtrates of test Schirmer eluates were furthermore inspected by SDS PAGE for the occurrence of proteins <3 kDa probably indicating filter membrane damage or inaccuracies. In fact, no protein gel signals could be detected referring to the filtrates and protein patterns were not found to be affected by the filtering process in the gel-related mass range, resulting in highly dense bands, allowing tear proteome distribution in 17 representative mass areas for LC ESI MS analysis (Fig. 56). Regarding overall protein identification in tear concentrates, after normalization 1039 proteins ($p < 0.05$, $\text{score} \geq 30$) (appendix 3) resulted from combinatory SwissProt database search of all LC ESI measurements and have been focused on for quantification and proteome description, whereby 950 proteins showed scores ≥ 35 . Replicates showed a high consistence emphasized by regression pattern comparison between technical replicate runs referring to qualitative, reproducible analysis. Regarding mapped tear proteins, there was a distinct overlap with tear proteome characterization in the literature so far (de Souza *et al.*, 2006, Karnati *et al.*, 2013, Zhou *et al.*, 2012). A predominant portion of proteins was found to be cellular originated, whereby only a minority of annotated proteins referred to the cytosol (Fig 57A/B), which had also been documented for capillary tears by use of both, MALDI and ESI workflows. Most of the membrane proteins could be associated to organelles (Fig. 57C) with predominant roles in protein binding and enzymatic activities (Fig. 58). Most likely the majority of these proteins were originated from epithelial cells corresponding to the ocular surface. Thereby, ocular surface leakage in the course of POAG therapy and with increasing age could explain the distinct occurrence of intracellular proteins in the tear film (Fukuda *et al.*, 1996, Nielsen *et al.*, 1981, van Haeringen and Glasius, 1976, vanHaeringen, 1997). Nevertheless, also capillary tears including healthy subjects showed high portions of intracellular proteins revealed by LC ESI and MALDI MS. De facto, MS analysis indicated the existence of a large fraction of intracellular proteins in the tear film. In any case, the levels of these proteins are likely to be affected under certain circumstances and need focused investigation. In accordance, some of the top protein hits refer to extracellular tear proteins, but also to cellular proteins. Indicated by protein scores, the most abundant proteins were lactotransferrin (score 2196), serum albumin precursor (score 1721), keratin, type II cytoskeletal 1 (score 1525), keratin, type I cytoskeletal 10 (score 1365), α -enolase (score 1134), zinc- α -2-glycoprotein (score 1075), heparan sulfate

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proteoglycan core protein (score 1073) and retinal dehydrogenase 1 (score 1029). Also anti- α -trypsin (score 886) and pyruvate kinase isozymes M1/M2 (score 666) were detected among the top 30 protein hits (Fig. 59), which was comparable to the capillary tear results of prior LC ESI/MALDI experiments. Proline-rich protein 4, which was found among the most prominent proteins, revealed by MALDI MS, was also detected in ESI analysis of capillary tears, only in the medium score rank (score 167, rank 98), probably reflecting its chemical preference for MALDI ionization. Like in capillary tears several immunoglobulin fractions could be detected with high scores, e.g. Ig α -1 chain C region (score 588), Ig α -2 chain C region (score 525) or Ig κ chain V-III region WOL (score 381). Also, several retina characteristic proteins could be documented with high scores including retinal dehydrogenase 1 (score 1029), peroxiredoxin 1 (score 654) and heat shock protein β 1 (score 646), which indicates distant ocular regions as additional tear protein source. Actually, scores of detected proteins decline dramatically at a value of 251, whereby after this value also the correlation between score and number of identified peptides for each protein fluctuates to a certain degree. Most peptides were identified for lactotransferrin (7530 peptides), lipocalin-1 (3987 peptides), lysozyme C (3675 peptides) and proline-rich protein 4 (1671 peptides), which were also documented among the predominant MALDI detected proteins. Mammaglobin B (1021 peptides) and secretoglobin family 1D member 1 (550 peptides), both reported as dry eye markers (Boehm *et al.*, 2013, Kramann *et al.*, 2011), could also be characterized with high peptide numbers. Like in capillary tears, also epithelial proteins, most likely derived from conjunctiva or cornea, e.g. desmoplakin (score 273), desmoglein 2 (score 69), desmoglein 4 (score 47), dermatan-sulfate epimerase-like protein (score 47), multiple epidermal growth factor-like domains protein 8 (score 38), protocadherin-9 (score 35) and γ -A12 (score 38), cadherin-5 (score 79), plectin (score 231) as well as various cytokeratine species, also documented for capillary tear fluid, could be identified. In summary, even consuming a smaller protein amount than in capillary tear analysis the LC ESI MS strategy provided a detailed image of the tear proteome emphasizing the high sensitivity of the developed ESI approach. Moreover, a high degree of congruency between capillary and Schirmer tear proteome could be documented with 95% of Schirmer eluted proteins also detected in capillary tears by the developed LC ESI MS workflow, which most likely shows that different results are unlikely to be a matter of the tear sample species, but are more likely to depend on the sensitivity of the method. In addition, retinal proteins; retinal dehydrogenase 1 (score 1029) and retinal guanylyl cyclase 1 (score 48) could be exclusively detected in the Schirmer eluted tear samples. Also, other highlighting proteins like leukocyte elastase inhibitor (score 747), aldehyde dehydrogenase (dimeric NADP-preferring) (score 540), 14-3-3 protein Σ (score 373) or alcohol dehydrogenase (score 343) could be exclusively identified with relatively high scores in Schirmer tears. In summary, an in-depth

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view to the Schirmer tear proteome could be achieved revealing beside characteristic secretory tear proteins, proteins originated from more distant ocular regions, e.g. retina. Especially, the predominant class of intracellular proteins could be focused on to obtain an impression on the ocular surface status considering epithelia integrity. The high number of identified enzymes and binding proteins furthermore builds a fundament for future activity assays and functional studies.

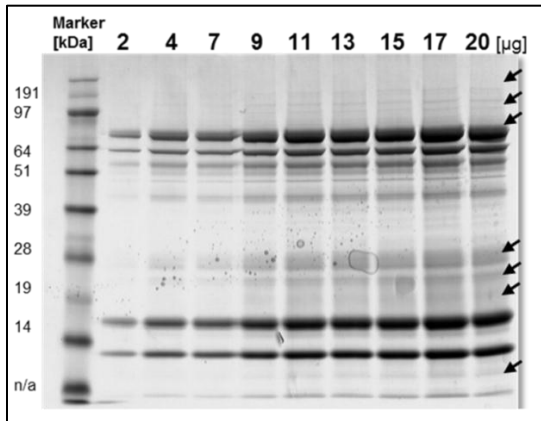


Fig 55 Dynamic range of the Schirmer tear proteome revealed by 1D SDS PAGE: 9 µg protein tear load generated complete tear proteomic gel pattern (low abundant protein mass areas are indicated by arrow), whereby at nearly 15 µg protein load signal saturation could be observed.

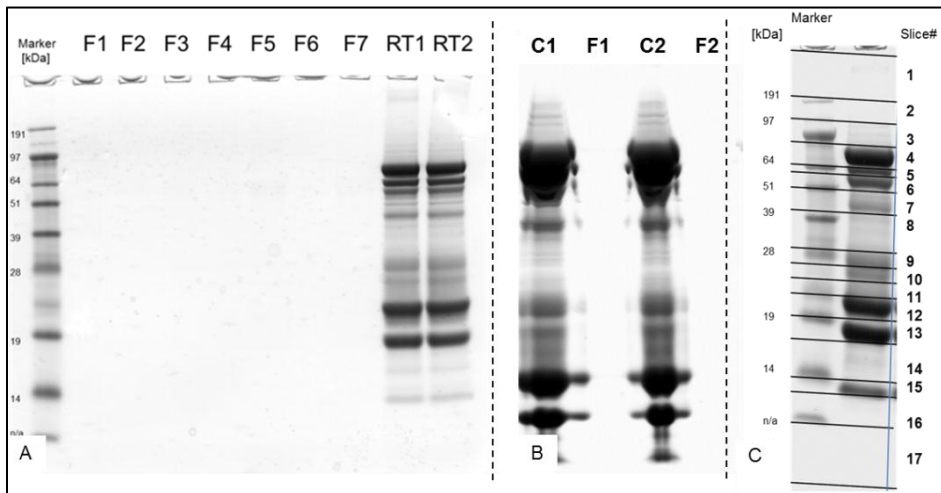


Fig 56 Performance of 3K MWC spin filters (3 kDa>concentrate) to concentrate Schirmer tear samples prior to LC ESI MS analysis; A: MWC tear filtrates (F1-7) showed no filter leaking proteins >3 kDa indicating high protein recovery (RT1/2= tear control samples); B: Highly concentrated tear samples (C1/2) and corresponding filtrates (F1/2); C: Mass area selection of Schirmer tear proteins for LC ESI MS analysis (17 mass area slices/lane).

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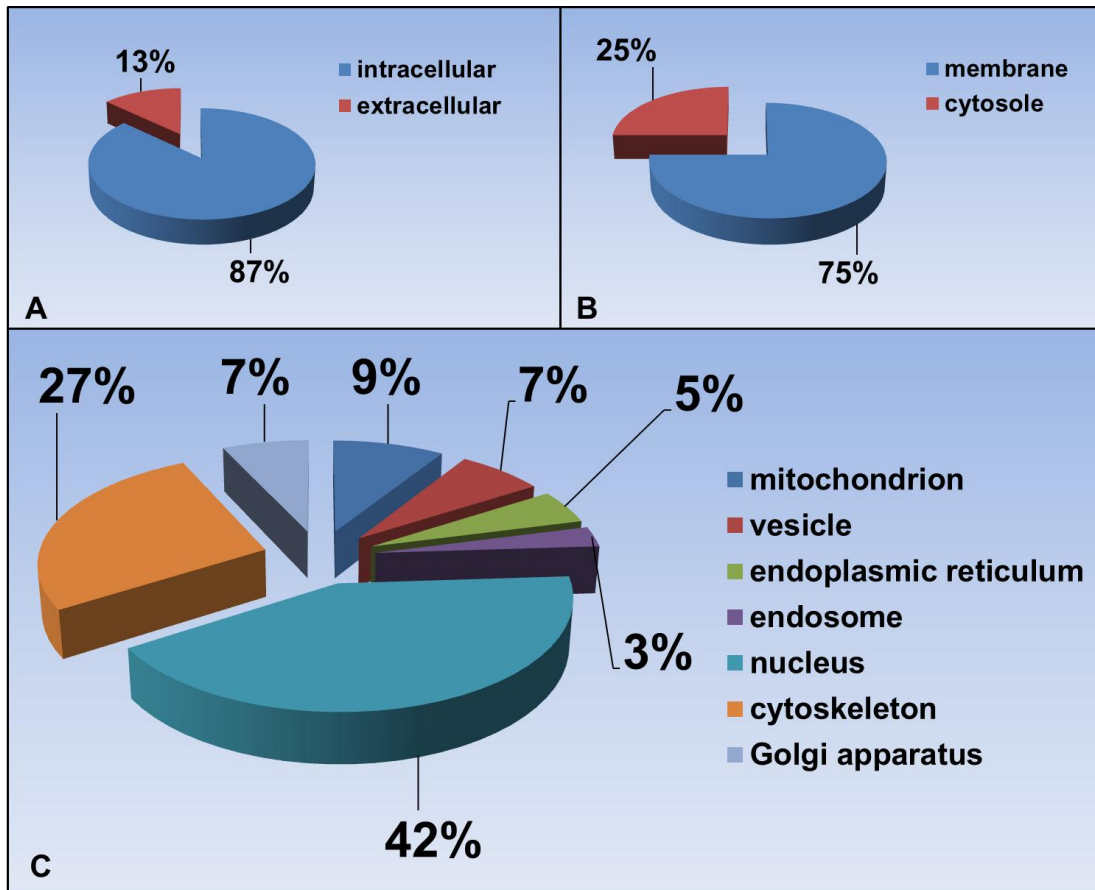


Fig 57 Functional analysis of the Schirmer tear proteome by Cytoscape GO annotation; A: The majority of LC ESI identified Schirmer tear proteins were associated to cells; B: A predominant portion of cellular proteins were related to membranes; C: The highest protein contingent was associated to the nucleus followed by cytoskeleton.

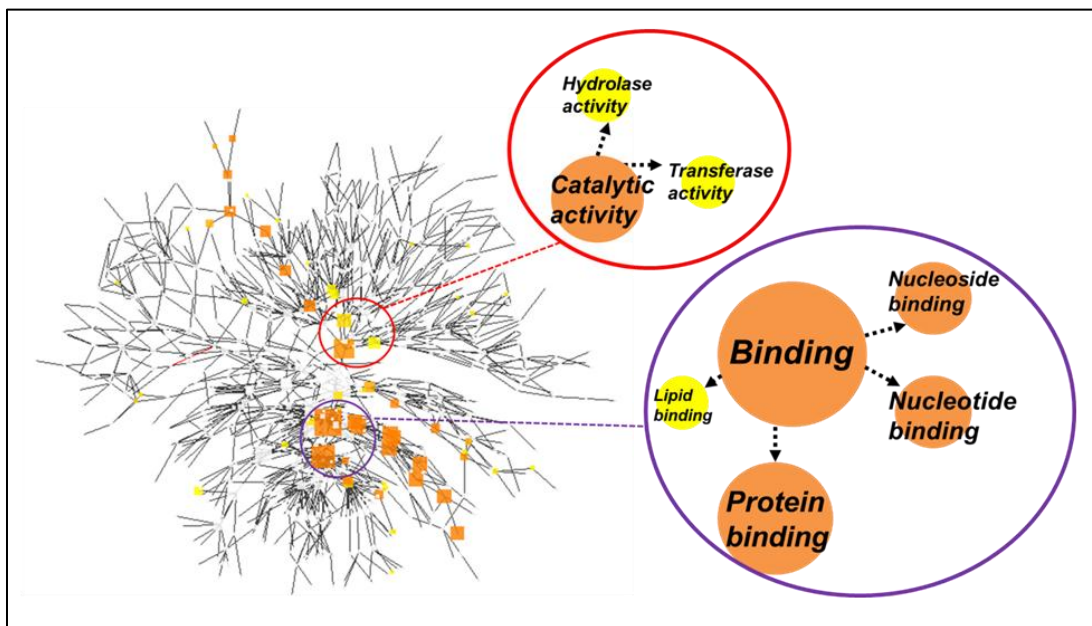


Fig 58 Molecular function of LC ESI MS identified Schirmer tear proteins by Cytoscape GO annotation indicating predominant tear protein contingents to be involved in molecule binding and enzymatic functioning.

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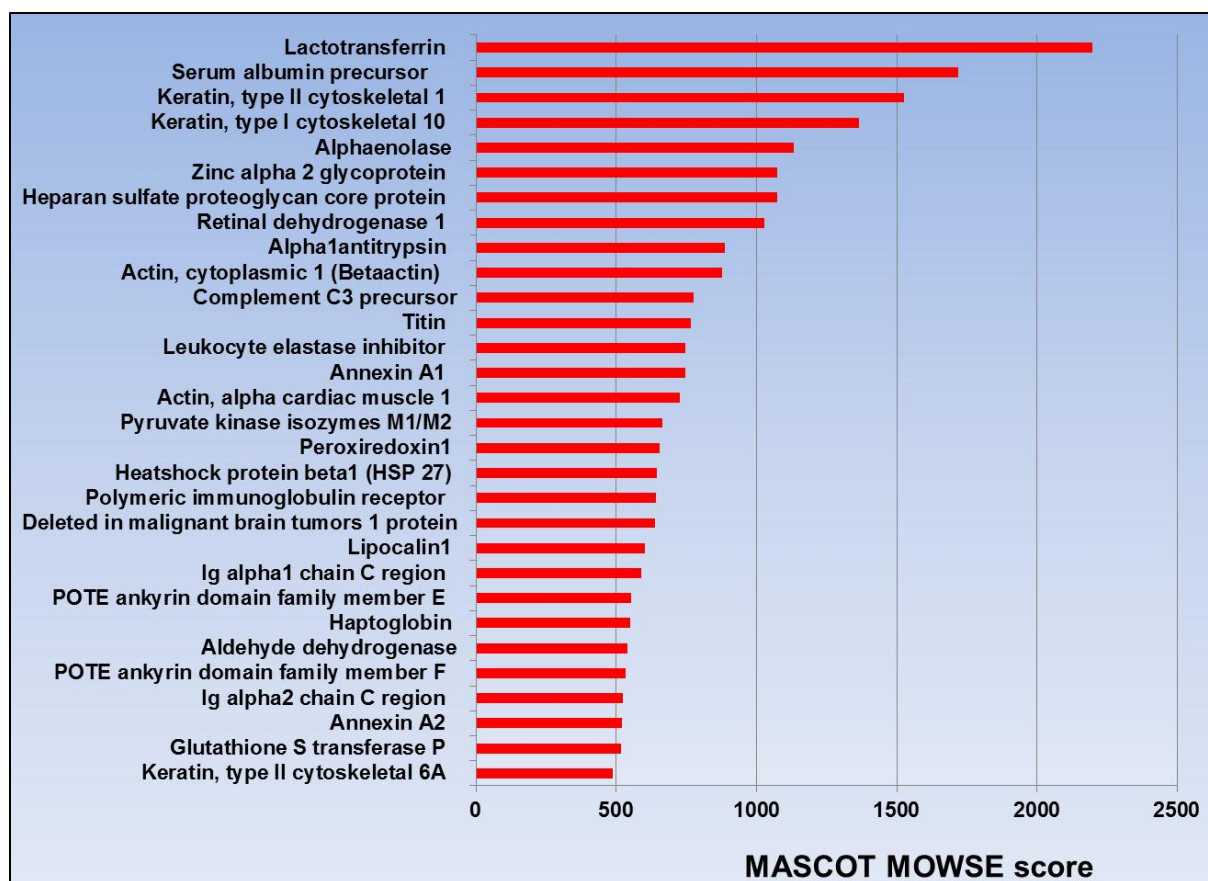


Fig 59 Top 30 LC ESI MS identified Schirmer tear proteins indicated by the MASCOT *Mowse* Score.

4.4.3 General tear results

Taking all LC ESI experiments on capillary and Schirmer tears together, a score independent frequency analysis using Statistica showed that lactotransferrin, lipocalin-1 and secretoglobulin family 1D member 1 were the predominant frequent proteins in tear fluid, followed by Ig α -2 chain C region, extracellular glycoprotein lacritin, proline-rich protein 4, uncharacterized protein C1orf127, Ig κ chain C region, zinc- α -2-glycoprotein, polymeric Ig receptor, Ig α -1 chain C region, lysozyme C precursor, prolactin-inducible protein, cystatin S precursor (cystatin 4), mammaglobin B and Ig J chain. Interestingly, beside prominent tear proteins like proline-rich protein 4 and mammaglobin B, a low abundant protein; forkhead-associated domain-containing protein 1 could be detected in all experiments and was not documented for tear fluid before. Also, lymphoid-specific helicase, a nucleus residing DNA-binding protein (Delmas *et al.*, 1993) could be detected in most of the experiments and could be documented for tear fluid for the first time. Centaurin- γ 1 was detected in most of the samples for the first time for tear fluid. Also, cytosolic 5'-nucleotidase III-like protein was documented for tear fluid for the first time detectable with high frequencies. Further prominent proteins detected in the majority of experiments were serum albumin precursor, β -2-microglobulin, melanotransferrin, putative uncharacterized protein LOC401522, cytoplasmic protein NCK2

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and myozenin 2. Transmembrane channel-like protein 8, a multi-pass membrane protein (www.uniprot.org) and myozenin 2 could also be reported for the first time for tear fluid in the majority of experiments. Mesothelin, Ig λ -2 chain C regions, proline-rich protein-1, protein S100-A6, glutathione S-transferase P, α -1-antitrypsin and protein S100-A9 were detectable in 58% of the experiments in confidence with the literature (Karnati *et al.*, 2013, Zhou *et al.*, 2012). Another protein detected in more than 50% of the experiments was MHD domain-containing death-inducing protein, not detected in tear fluid before. Interestingly mucins, especially mucin 5AC or mucin 16, were documented for their presence in tear fluid (Argueso *et al.*, 2003, Jumblatt *et al.*, 1999, Spurr-Michaud *et al.*, 2007, Zhao *et al.*, 2001) and could be identified in few of the experiments. Tear proteins, that were high frequently detected in ESI experiments, even with low scores, have also been detected in most cases in MALDI experiments, e.g. lactotransferrin, lipocalin-1, proline-rich protein 4, lysozyme C, mammaglobin B, proline-rich protein 1, cytoplasmic protein NCK2 or protein S100-A9. Also, several of the proteins identified in approximately 50% of the ESI experiments were detectable by MALDI analysis, e.g. neutrophil gelatinase-associated lipocalin or γ -enolase. Indeed, also proteins that were rarely detected in ESI experiments could be documented in MALDI experiments, e.g. ribosome-binding protein-1, POTE ankyrin domain family member I, α -1-acid glycoprotein 1, 2 or centrosomal protein of 164 kDa supporting their existence in tear fluid. A highlighted group of proteins were lipid associated proteins, which could be detected by ESI and/or MALDI analysis. Beside lipocalin-1, apolipoprotein AI and neutrophil gelatinase-associated lipocalin, phospholipase A2 and phospholipid transfer protein numerous other lipid-associated proteins could be frequently observed including liprin α 2/3, lipoxygenase homology domain containing protein 1, patatin-like phospholipase domain containing protein 7, lipid phosphate phosphor hydrolase 3, non-specific lipid transfer protein, StAR related lipid transfer protein 13, phospholipase A2 activating protein, lysophospholipase-like protein 1, putative lipocalin1-like protein 1, apolipoprotein O, Prolow-density lipoprotein receptor-related protein 1, low density lipoprotein receptor-related protein 4, inactive phospholipase C-like protein 2 and arachidonat 5-lipoxygenase. Most likely these proteins are associated to the tear lipid layer and are potential targets for future investigations regarding evaporative dry eye disease. In summary, by use of both, LC ESI and MALDI approaches encircling a huge number of experiments, an in-depth view of the tear proteome could be obtained, whereby the presence of numerous proteins could be supported independently from their scores by their frequencies in ESI experiments as well as by their independently detection by the LC MALDI analysis. In consequence, a sensitive proteomic image of the tear film as the key extracellular ocular surface component could be provided.

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4.4.4 Conjunctival cells

To achieve a more detailed proteomic image of the ocular surface beyond the fluidic compartment encasing the cellular region, NHC-IOBA conjunctival cells have been examined. By use of the developed LC ESI workflow initially 1412 proteins followed by 1481 proteins in a second experiment could be catalogued (MASCOT ion score $p < 0.05$). Thereby 1044 hits of the first experiment and all proteins of the second experiment were used for quantification assessed by a protein score ≥ 35 . For descriptive proteome characterization a combined list of both experiments was used (for overview protein list see appendix 4; top 30 scoring proteins are illustrated in Fig. 60). Several proteins originated from cytoplasm as well as membrane-associated proteins could be documented. Regarding cytoscape GO annotation analysis 74% of all proteins could be annotated to certain cellular compartments. Actually, 94% of the annotated hits refer to intracellular proteins (Fig. 61A). Several extracellular proteins were detected with high scores and were mainly found to represent secreted proteins. De facto, numerous proteins were intracellular located, but also secreted to the cellular surface like annexin A2 (score 12539), 14-3-3 protein Σ (score 1518), macrophage migration inhibitory factor (score 534), apolipoprotein A-I-binding protein (score 286), peroxiredoxin 4 (score 3295) or calreticulin (score 2542) (www.uniprot.org). Actually, interleukin 17 (score 44)/ 18 (score 280)/ 25 (UPF0556 protein C19orf10) (score 758), serpin B5 (score 1343) spondin-1 (score 89) and mucin-5B (score 242) were confirmed as exclusively secreted proteins (www.uniprot.org). RNA-transcripts for interleukines and other cytokines have been documented in lysates of conjunctival tissue lysates (Pflugfelder *et al.*, 1999) and cytokine induction was indirectly analyzed by ELISA in conjunctival tissue (Solomon *et al.*, 2001) supporting the secretory activity of these cells. The detection of mucin 5B as a characteristic epithelial secreted protein and also the identification of EGF-like module containing mucin-like hormone receptor-like 2 (score 133) is in confidence with the detection of mucin RNA transcripts (MUC2, MUC5AC) (McKenzie *et al.*, 2000) and mucin proteins (MUC16) (Argueso *et al.*, 2003, Gipson and Argueso, 2003) in human conjunctiva and indicates the functional activity of NHC-IOBA cells as an ocular surface model (Tong *et al.*, 2009). Also, numerous cytokeratines, e.g. cytoskeletal keratin type I 13, 15, 17, 19 and 24 (scores 244/333/345/189/96) as well as cuticular keratine Ha1 (score 85) have been reported as characteristic human conjunctiva I proteins (Krenzer and Freddo, 1997) also documented for NHC-IOBA cells (Diebold *et al.*, 2003). Other epithelial marker proteins like desmoglein 1, 2 (score 336/296) (Wan *et al.*, 2003) and epiplakin (score 113) (Spazierer *et al.*, 2006) could be identified in the cell lysates. Furthermore, despite the majority of proteins were found to be cytoplasmic proteins, 30% of all proteins were membrane associated (Fig. 61B). Transmembrane proteins like transmembrane protein 33 (score 566), 109 (score 411), coiled-coil domain containing protein (score 166) or emp24 domain containing protein 10

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(score 581) could be recovered by LC ESI. Numerous receptors, e.g. transferrin receptor protein 1 (score 1908), insulin-receptor related protein (score 166) and membrane associated progesterone receptor component 1 (193) could also be revealed. Interestingly, also two neurotransmitter receptors were identified, however, with low scores; dopamine receptor (score 45) and GABA receptor subunit delta (score 39). Also a chemokine receptor, C-C chemokine receptor type 9 has been determined (score 95) fitting well with the interleukine secretion activity of the cells. Identification of vinculin, a high scoring periphere membrane protein (score 1327) associated to cell junctions (www.uniprot.org) supported the functionality of the cell system. GPI transamidase component PIG-T (score 107) residing at the ER membrane (Ohishi et al. 2001) and subunits of the guanine-nuclotide-binding protein, a palmitoylated membrane G protein (www.uniprot.org) are further examples of successful membrane protein recovery. In addition, several membrane-located amino acid transporters have been identified, e.g. sodium-coupled neutral amino acid transporter 2 (score 82), large neutral amino acids transporter small subunit 1 (score 534) or neutral amino acid transporter B(0) (score 504). Furthermore, a cation transporter, probable cation-transporting ATPase 13 A5 (score 131) could be identified. Regarding the annotation of proteins to cellular organelles, 82% of all annotated proteins were found to be associated to organelles. Despite the portion of plasma membrane and cytosolic proteins, predominantly nucleus proteins could be documented in cell lysates followed by mitochondrial and ER proteins (Fig. 61C). Cofilin 1 (score 2800), carboxy-terminal domain RNA polymerase II polypeptide A small phosphatase 2 (score 280), proteasome subunit beta type-5 (score 788), heterogenous nuclear ribonucleoprotein U (score 214) were exemplary detectable nucleus proteins observed in NHC-IOBA lysates. Besides, numerous histone species could be documented. Several proteins like dynamin 1-like protein (score 289) were annotated to ER, Golgi and mitochondria (www.uniprot.org). Furthermore, among nucleus associated proteins oncogene signalling proteins like Ras GTPase-activating-like protein IQGAP1 (score 5207) (Shen *et al.*, 2006) could be detected with high abundance. Up-regulated during skeletal muscle growth protein 5 (score 254), mitochondrial GrpE protein homolog 1 (score 128) and cytochrome b-c1 complex subunit 8 (score 235) represent exemplary catalogued mitochondrial proteins. Most of the found proteins were typical cellular proteins; however, only few of them have been identified for NHC-IOBA cells, so far. Map kinases were reported for NHC-IOBA cells (Liu *et al.*, 2007) and two representatives could also be supported by this work; NHCJNK1/MAPK8-associated membrane protein (score 75) and MAP3K12-binding inhibitory protein 1 (score 64). Despite transglutaminase 2 (protein-glutamine gamma-glutamyltransferase 2) (score 201) could be proven, the prior reported enzyme, conjunctival associated transglutaminase 1 (Nakamura *et al.*, 2001), could not been detected in this work. In fact, there was a certain overlap with the tear proteome indicating the cross-relation

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between tear film and ocular surface epithelia, supporting the cellular origin of numerous tear proteins. 12% of all conjunctival cell proteins could be revealed in both types of tear fluid, capillary and Schirmer tears, encircling several intracellular enzyme-linked proteins (scores are indicated for NHC-IOBA), e.g. L-lactate dehydrogenase A chain (score 5121), protein arginine N-methyltransferase 5 (score 131), phosphoglycerate kinase 1 (score 7648), fructose-bisphosphate aldolase A (score 3229), γ -enolase (score 4082), pyruvate kinase isozymes M1/M2 (score 14932) (Fig. 62), glucose-6-phosphate isomerase (score 5578), glutathione S-transferase P (score 5286) and hexokinase 1 (score 73). Characteristical tear film proteins like calreticulin (score 2542), cystatin B (score 1056), POTE ankyrin domain family member F (score 6266) and the proteins S100-A6/A8/A9 (scores 988/69/40) could be documented for NHC-IOBA cells. Additionally, S100-A10 (score 673), A13 (score 396) and A16 (score 236) could be exclusively identified in cell lysates. Moreover, there was overlapping finding in desmoglein, annexin, heat shock protein, myosines and histones in cells and tears. Various members of the 14-3-3 protein family were detected in tears and cells. These findings support the hypothesis, that a majority of tear proteins is originated from ocular surface epithelia. Only a minority of these proteins could be clarified to be actively secreted by the conjunctival cells, e.g. cathepsin D, macrophage migration inhibitory factor, serpin B5, galectin 1, galectin 3 or galectin 3-binding protein (www.uniprot.de). For some of the proteins, it remains unclear, if the protein is secreted or originated from cellular leakage since proteins like annexin A5 are located in the cytoplasm, but are also located at the cell periphery (www.uniprot.org) vulnerable to extracellular release. Several other proteins like galectin 7 or thioredoxin exhibit traffic between nucleus, cytoplasm and extracellular space (www.uniprot.org). Numerous proteins, e.g. 14-3-3 proteins, calreticulin, glucose-6-phosphate isomerase and peroxiredoxin 4 are documented to be as well located in the cytoplasm as to be secreted by the cells (Chang *et al.*, 2006) (www.uniprot.org). Also, inflammation-associated complement decay-accelerating factor (score 192) could be detected in the cells. In summary, by use of the LC ESI MS workflow, a detailed view of the conjunctival cell proteome as a cellular constituent of the ocular surface could be obtained. The proteomic overlap of conjunctival cells with tear fluid as well as the occurrence of characteristic secretory proteins and epithelial marker proteins emphasized the potential of the cell line as a model system for ocular surface research. In summary, a deep proteomic view could be obtained encircling the fluidic and cellular dimension of the ocular surface building a fundament for ocular surface studies regarding Taflotan[®] sine and taurine.

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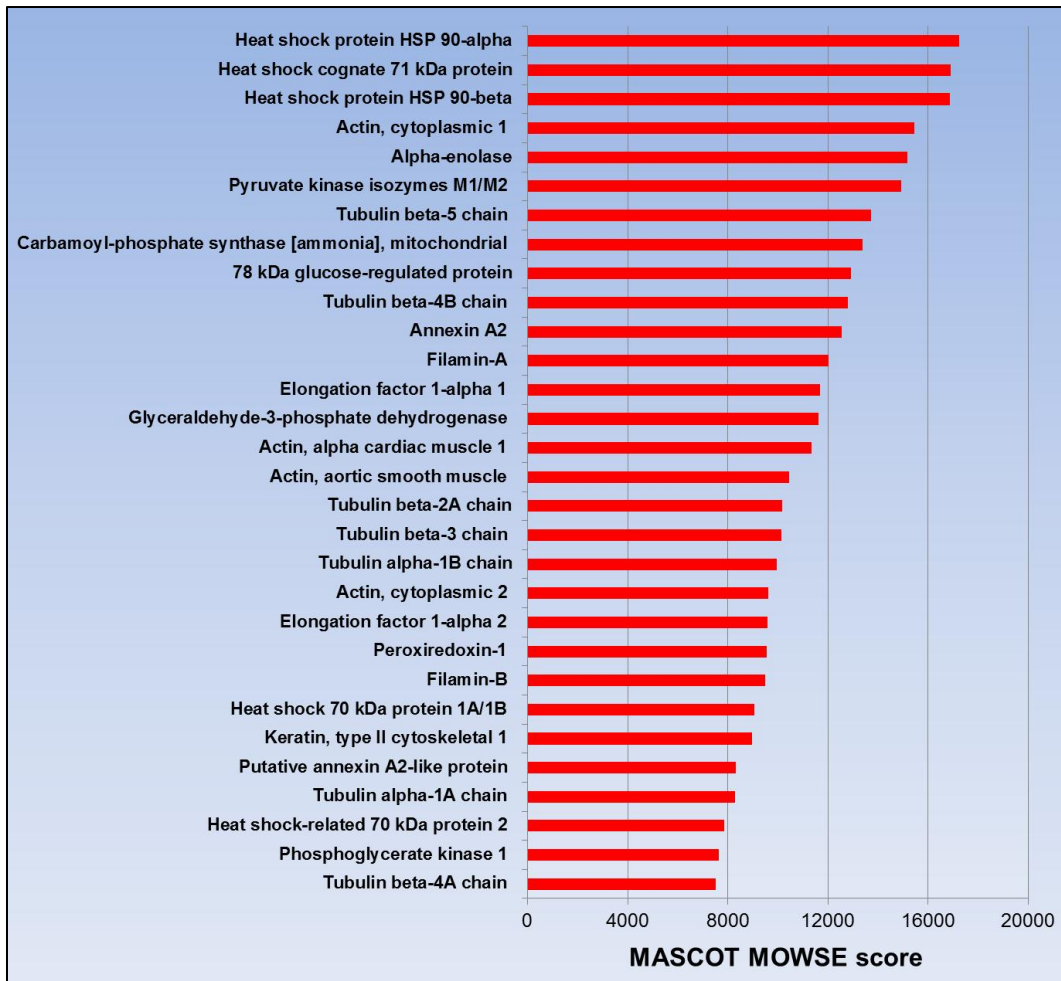


Fig 60 Top 30 scoring conjunctival cell proteins identified in NHC-IOBA 0.1% DDM lysates by use of the LC ESI MS BU workflow.

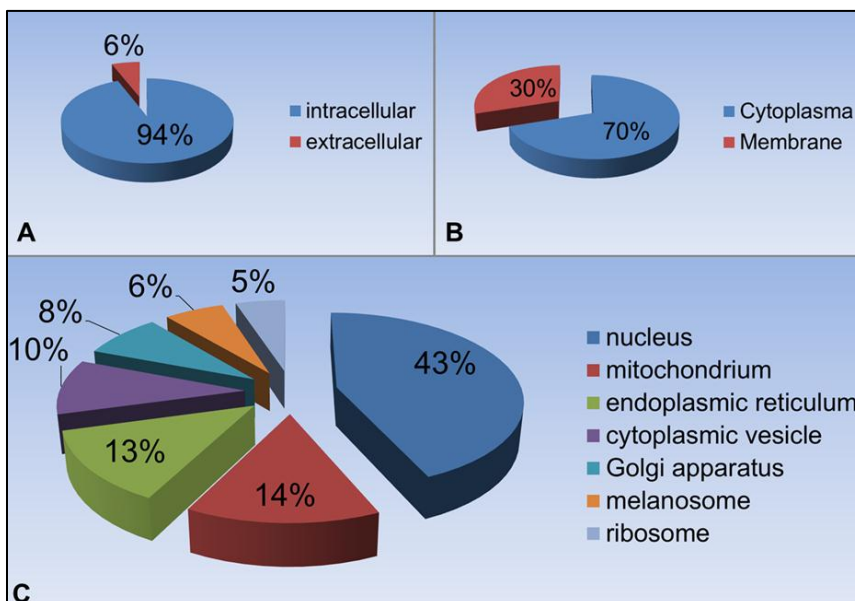


Fig 61 Functional analysis of the conjunctival cell proteome by Cytoscape GO annotation; A: Most proteins were intracellular protein species with a high contingent of B: Cytoplasmic proteins; C: The predominant protein portion was found to be associated to nucleus followed by mitochondrion and ER.

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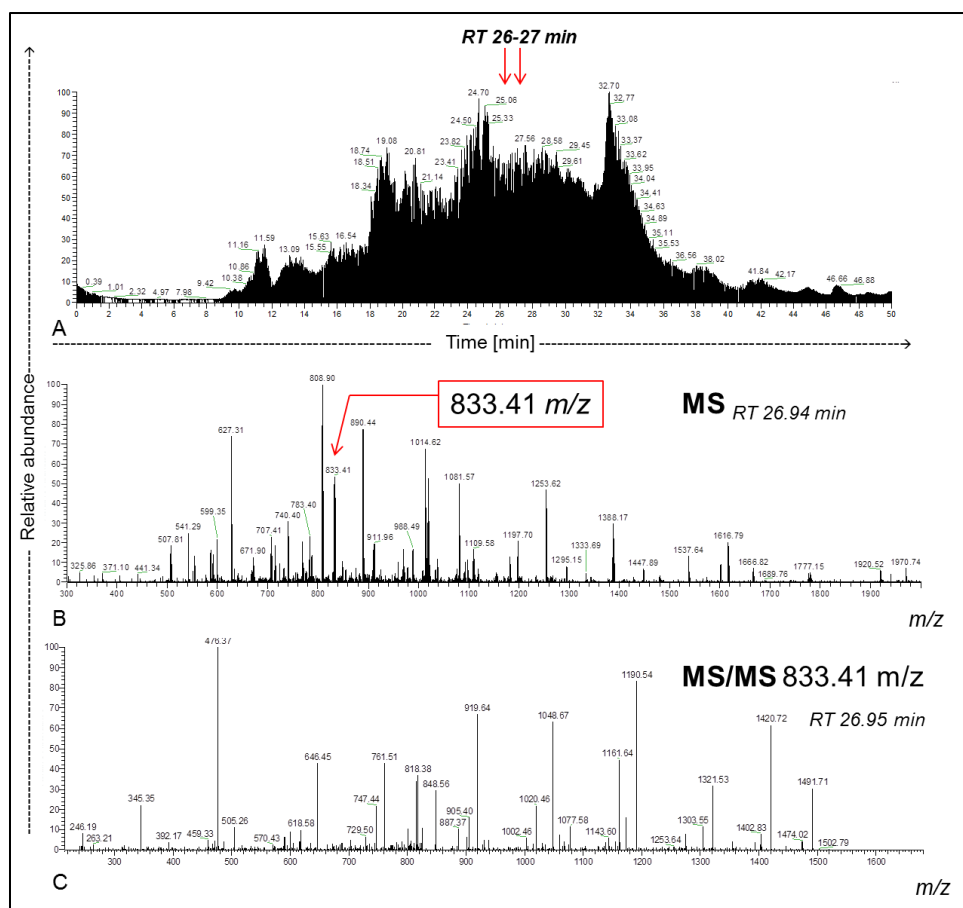


Fig 62 Exemplary identification of high scoring pyruvate kinase isozymes M1/M2 in conjunctival cells; A: Exemplary TIC chromatogram indicating the elution area for a high scoring PKM precursor between 26/27 min RT; B: PKM precursor 833.41 m/z selected at 26.94 min for fragmentation; C: ITMS fragment spectrum corresponding to precursor 833.41 m/z contributing to PKM identification.

4.5 Influence of Taflotan[®] sine on the ocular surface of POAG patients

Regarding regression analysis of the studied tear proteome, 12% of all identified proteins showed a distinct level change. Thereby, 5% of responding proteins displayed a significant linear level change ($p < 0.05$), 3% showed a distinct tendency of linear level regulation ($R^2 < 0.9$) and 4% significantly corresponded to a polynomial pattern ($p < 0.05$) (Fig. 63). Related to linear regression, several dry eye related proteins showed significant level decline. Serotransferrin ($R^2 = 0.9384$, $p = 0.0313$, score 444), galectin 7 ($R^2 = 0.9298$, $p = 0.0357$, score 64), annexin A11 ($R^2 = 0.9085$, $p = 0.0469$, score 51), cadherin 5 ($R^2 = 0.9598$, $p = 0.0402$, score 80) and myosin-10 ($R^2 = 0.9118$, $p = 0.0451$, score 92) were found to show a significant linear level alleviation (Fig. 64). Interestingly, other myosin family members, myosin 6, 7, 9 and 11 also showed a regulation diminishment, however, with non-significant R^2 values. Regarding regression slopes of linear responding proteins the majority of proteins (69%) showed negative values reflecting a longitudinal level decrease, whereby most members (83%) were slightly decreased over time indicated by moderate slope values ($\text{slope} < 0 \leq -50$). Comparable results were obtained from positively responding proteins,

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whereby also the majority of proteins (78%) showed slight level increase ($\text{slope} > 0 \leq 50$) (Fig. 65). Regarding non-linear time progression, several dry eye related proteins displayed a significant polynomial decrease over time like e.g. low density lipoprotein receptor-related protein 4 ($p = 0.0207$, score 59) or manifested a significant increase like ankyrin-3 ($p = 0.0432$, score 54). Beside polynomial regression fit, the fold change “optimal match” cluster analysis revealed a group of non-linear responding candidate proteins. The analysis calculated four clusters, whereby 56% of all proteins were categorized as non-regulated, showing an “equal”, “equal”, “equal”-level alteration time pattern grouped in cluster #1. Despite proteins that show high fluctuating time pattern in cluster #4 encircling 14% of all proteins, 68% of cluster members showed a distinct fold drop over time (“equal”, “decrease”, “equal”). 45% of the proteins in cluster #3 including 17% of all proteins showed a level increase pattern over time (“equal”, “increase”, “equal”). Moreover, related to all identified proteins the portion of proteins displaying a longitudinal fold decline was 21% higher than the increasing protein contingent. Furthermore, manual screening for longitudinal protein responses focusing on proteins documented as dry eye marker proteins, provided most proteins showing a linear descending tendency ($R^2 \geq 0.5$) like secretoglobin 1 family member D1 ($R^2 = 0.5680$, score 245), lactotransferrin ($R^2 = 0.6940$, score 2196), heat shock related 70 kDa protein 2 ($R^2 = 0.6415$, score 46) or heat shock cognate 71 kDa protein ($R^2 = 0.5842$, score 60), whereas some proteins exhibit tendencies of linear ascending like e.g. α -1-anti-trypsin ($R^2 = 0.8007$, score 886), interleukine-12-receptor β -2-chain precursor ($R^2 = 0.7400$, score 41) or tumor necrosis factor α induced protein 3 ($R^2 = 0.5951$). However, known dry eye marker proteins like S100A4, 6, 8 and 9 failed for response. Also proline-rich protein 4 (score 167) and proline-rich protein 1 (score 261) were found unaffected. Among the proline-rich proteins only the large proline-rich protein BAT2 showed a slight tendency of linear decrease ($R^2 = 0.6256$, score 78). Ingenuity[®] protein allocation analysis classified only 10% of the selected candidate proteins as extracellular proteins, whereas the remaining 90% were depicted to be originated from cellular compartments. Most of these proteins appertain to cytoplasm followed by nucleus related proteins (Fig. 66). The functional process analysis revealed 18 functional process hits above the adjusted protein allocation threshold of 25. Thereby processes like tissue development, cellular development as well as cell survival and cell death including apoptosis were highlighted (Fig. 67). In summary, a high number of cellular proteins could be detected in the tear film of POAG patients most likely reflecting ocular surface epithelial cell leakage events. It could be observed, that in first instance overall cellular processes were influenced by the medical switch since predominantly intracellular proteins most likely originated from ocular surface epithelial cells were found in the tear film and showed a decline over the investigative period after the medical switch. Several marker proteins also showed alterations in their regulation pattern, which indicates a changing health status of the

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ocular surface after switching. MA analysis of selected marker candidates emphasized the level decrease of several characteristic proteins highlighted by MS analysis. “A priori” dry eye associated MA tested proteins were: mucin 6, aquaporine 5, proline-rich protein 4, lactotransferrin, lipocalin, hemopexin, mammaglobin B, α -2-macroglobin, cystatin, S100A8, S100A9, lysozyme C, anti-trypsin, complement C3b and cytokines: TGF- β 1, TGF- β 2, TNF- α , IL1a, IL1b, IL2, IL5, IL8, IL9, IL10, IL12, IL13, IL17a, IL23a, mcp1/ccl2, MIP1a/ccl3. For proteomic result validation commercial antibodies against the following marker candidates have been selected: serotransferrin, pyruvate kinase isozymes M1/M2, plectin, annexin A5, annexin A11, myosin 10, galectin 7, tight junction protein ZO1 and cadherin 5. 46% of all tested proteins showed distinct tendencies of linear approximation of the POAG group to the control group in the course of the study (Fig. 68). Thereby, the following “a priori” dry eye associated marker proteins were found to be affected by the Taflotan[®] sine appliance: aquaporin 5, mammaglobin B, anti-trypsin and complement factor C3/C3b. Interestingly numerous cytokines were found to shift to the control level including the interleukines, IL1a, IL2, IL2b, IL13, IL17, IL23a, and the chemokines MIP1a/ccl3 and TGFB1 (Fig. 69). This finding clearly reflects a diminishment of the inflammatory status of the ocular surface in the course of the study. Focusing on MS derived marker candidates, several proteins showed tendencies of linear level diminution in direction to the healthy control level and furthermore a significant approximation response revealed by t-test (p-values are denoted in brackets; $p_1 = \text{POAG}_{\text{timepoint1}}$ compared to $\text{Control}_{\text{mean level}}$, $p_2 = \text{POAG}_{\text{timepoint5}}$ compared to $\text{Control}_{\text{mean level}}$). Thereby, pyruvate kinase isozymes M1/M2 ($p_1 = 0.0012$, $p_2 = 0.0740$), annexin A11 ($p_1 = 0.0009$, $p_2 = 0.2026$), cadherin 5 ($p_1 = 0.0084$, $p_2 = 0.0712$), serotransferrin ($p_1 = 0.0013$, $p_2 = 0.0700$) and kinectin ($p_1 = 0.0007$, $p_2 = 0.1320$) were found to converge significantly to the healthy control level in the course of Taflotan[®] sine exertion since at the start of the study protein levels for the highlighted proteins were significantly different between both groups. At the end of the study, no significant level difference could be observed anymore predicting an alignment of protein levels between the POAG and the healthy control group (examples are illustrated in Fig. 70). Beside MS derived candidates, this was also demonstrated for “a priori” MA selected dry eye associated proteins lipocalin ($p_1 = 0.0005$, $p_2 = 0.0641$) and complement factor c3/c3b ($p_1 = 0.0001$, $p_2 = 0.0846$). Moreover, the level decline of cytokines IL1b ($p_1 = 0.0009$, $p_2 = 0.2794$), IL2 ($p_1 = 0.0003$, $p_2 = 0.1620$) and IL23a ($p_1 = 0.0021$, $p_2 = 0.0804$) could be evaluated as a significant drawing to the healthy stage. Accordingly, microarray results clearly supported a beneficial effect of Taflotan[®] sine application reflected by cytokines and MS derived protein candidates. Moreover, objective clinical parameters represented by TBUT and BST strengthen beneficial effects of therapeutic switching since both parameters exhibited significant alterations revealed by t-test “start to endpoint” comparison (Fig. 71). In summary, a strong evidence for ocular surface regeneration in the

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course of preservative-free tafluprost appliance in POAG patients could be revealed by use of the developed workflow.

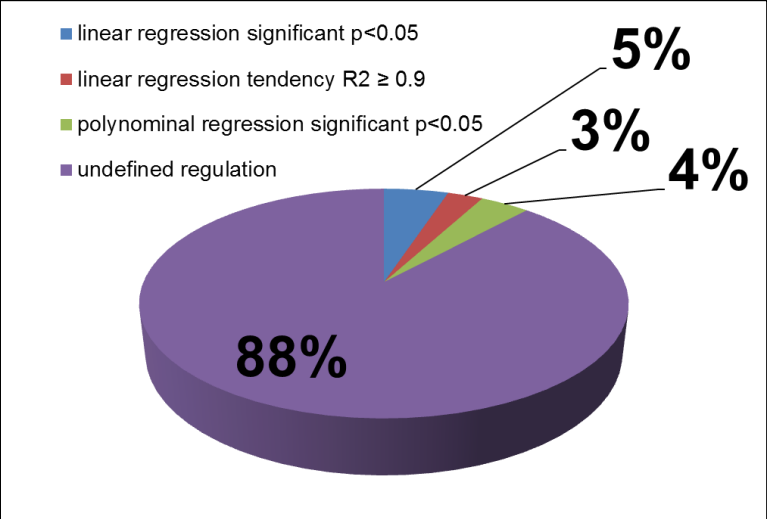


Fig 63 Distribution of longitudinal tear proteome dynamics observed in POAG patients switching from Xalatan[®] to preservative-free Taflotan[®] sine.

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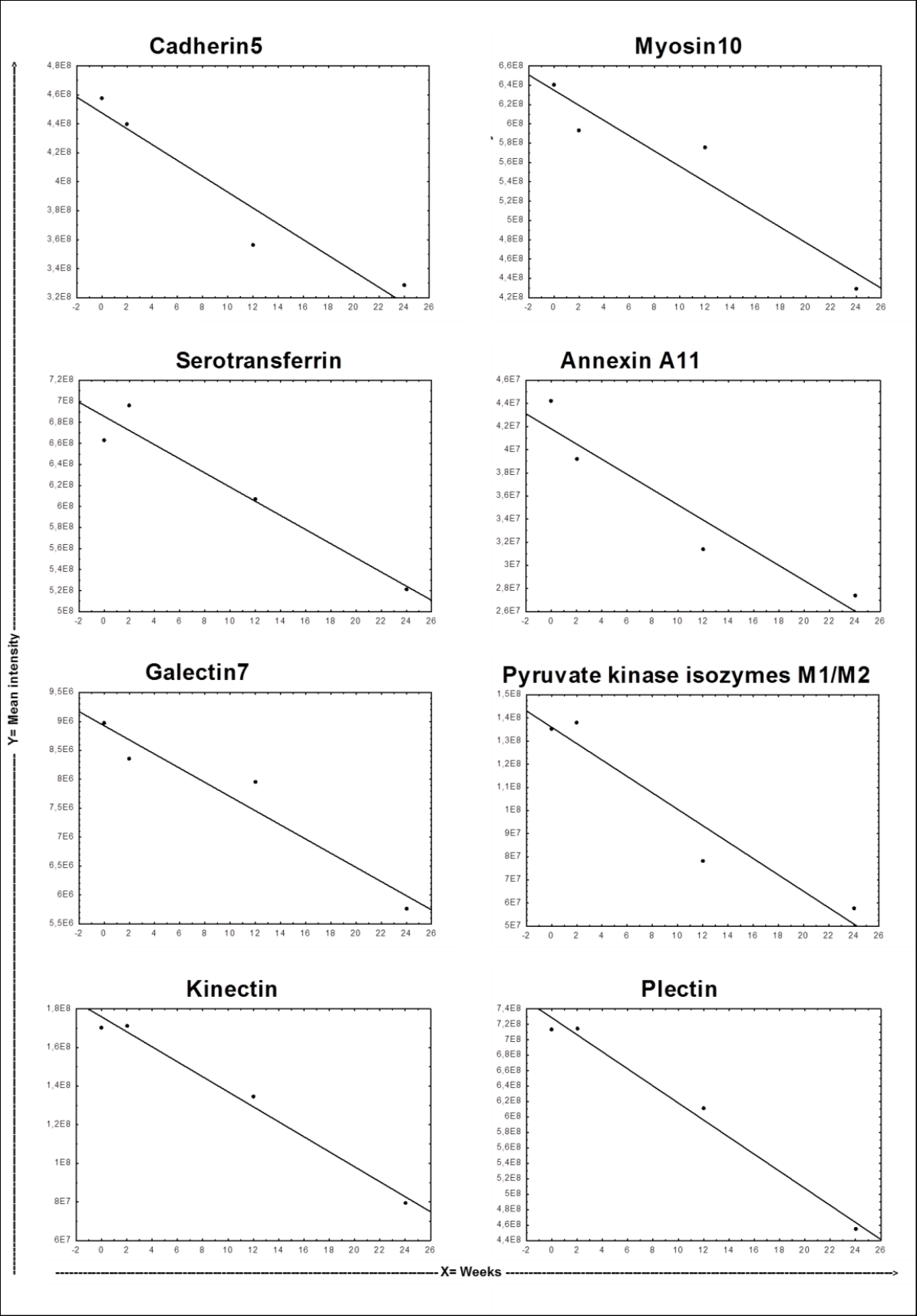


Fig 64 Longitudinal pattern changes exemplary shown for selected tear proteins displaying a significant linear level decline in POAG patients in the course of Taflotan[®] sine application after swichting from Xalatan[®] medication. (Funke *et al.*, 2014)

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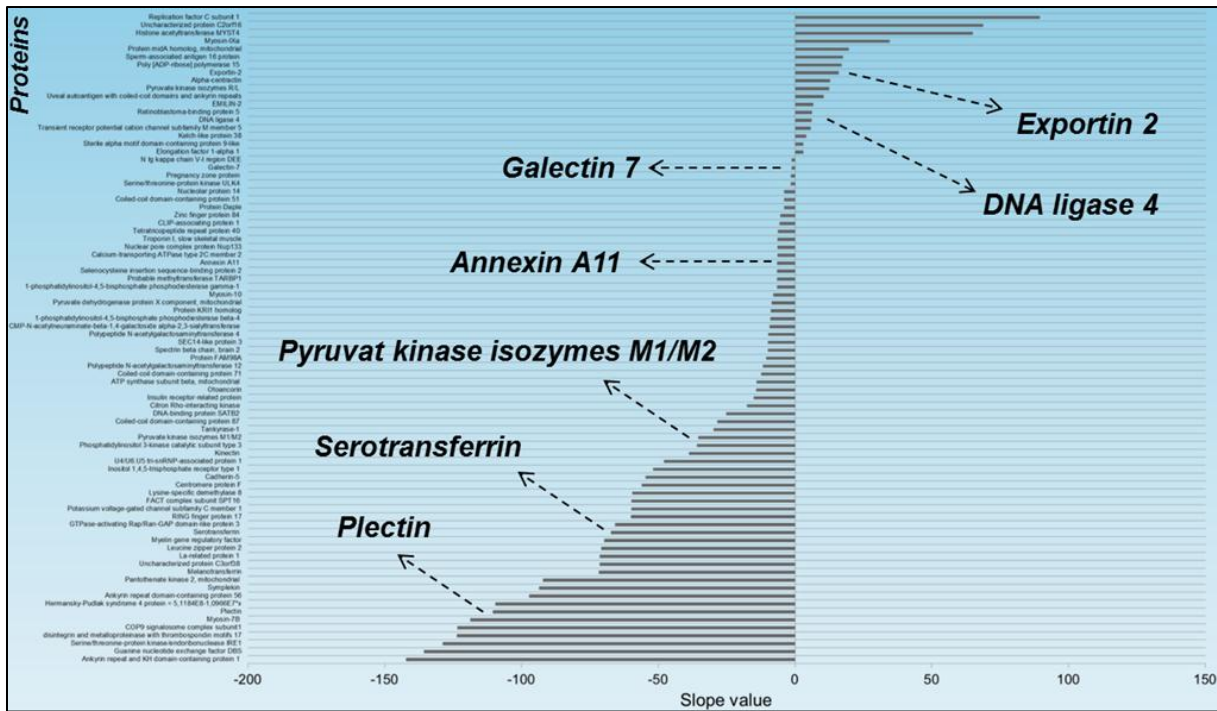


Fig 65 Slope distribution of linear responding tear proteins in the course of Taflotan[®] sine medication: The predominant protein contingent showed negative slope values. Modified from (Funke *et al.*, 2014)

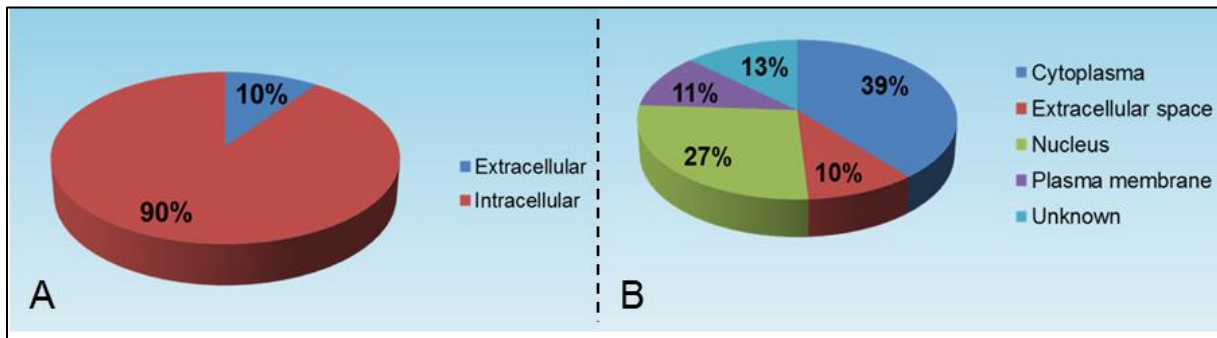


Fig 66 Ingenuity[®] functional analysis of regulated candidate tear proteins; A: Most candidate proteins responding in the course of Taflotan[®] sine application were found to be intracellular proteins; B: The predominant portion of intracellular protein referred to cytoplasm and nucleus. Modified from (Funke *et al.*, 2014)

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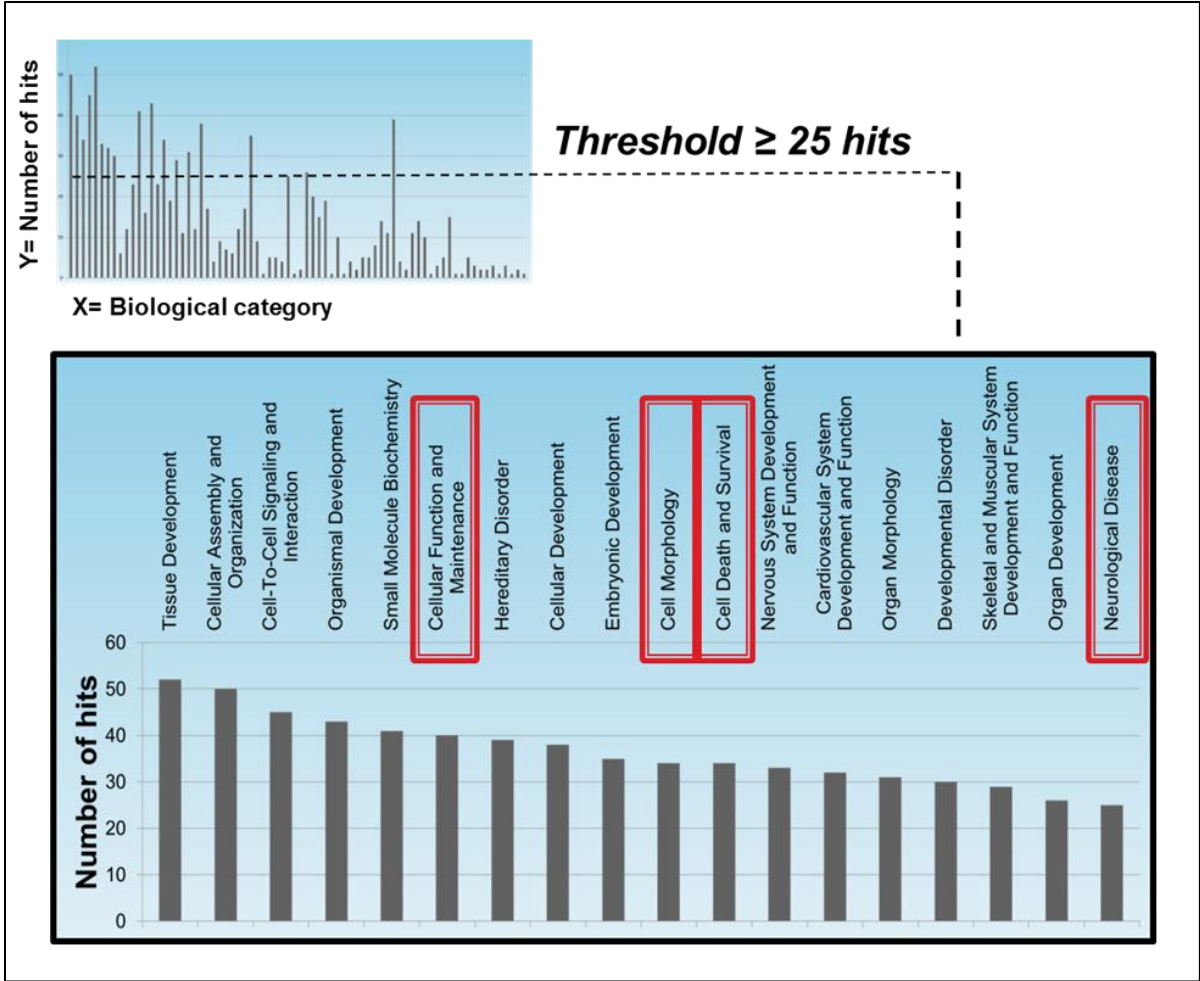


Fig 67 Ingenuity® functional analysis of candidate tear proteins responding after the medicative switch: Numerous candidates were found to be involved in cell maintenance, survival and death; interestingly also several candidates could be annotated to neurological disease considering glaucoma.

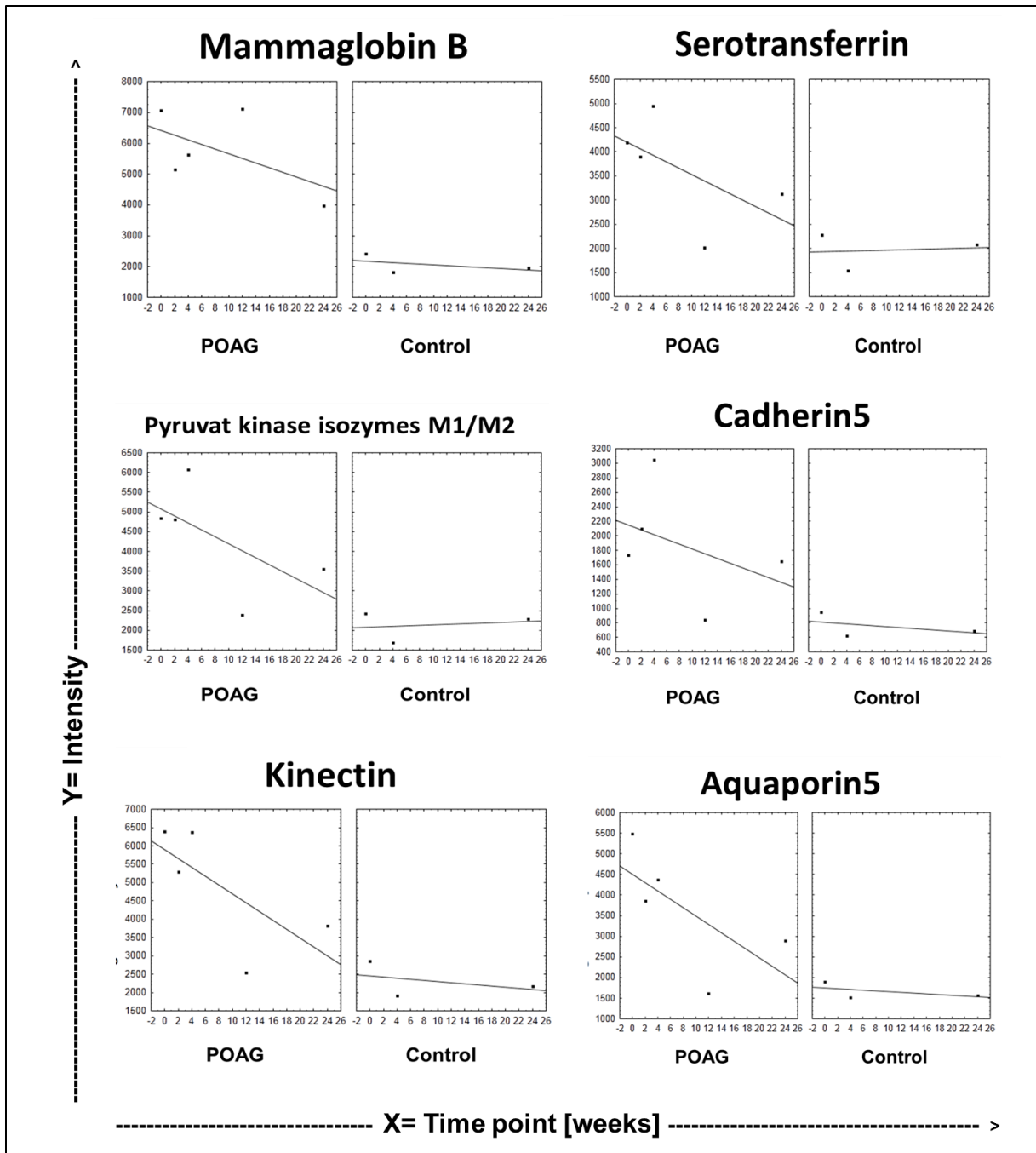


Fig 68 Microarray (MA) validation of selected MS established candidate tear proteins in a larger subject cohort: The illustrated candidates show a distinct level approximation in the POAG group to the control group in the course of Taflotan[®] sine appliance indicating a recovery of the ocular surface. (Funke *et al.*, 2014)

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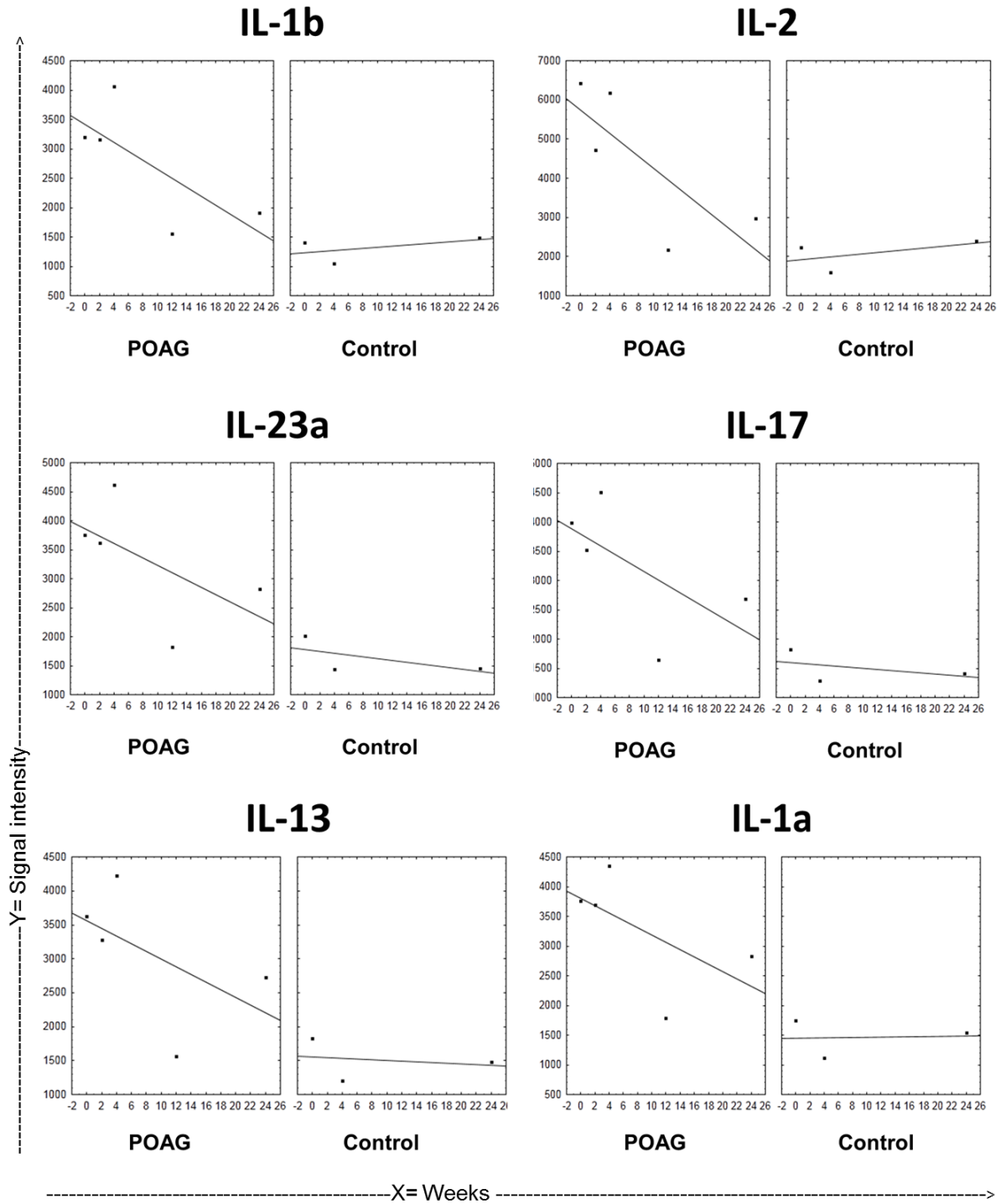


Fig 69 Microarray (MA) validation tear cytokines: A distinct cytokine level approximation in the POAG group to the control group in the course of Taflotan[®] sine appliance could be revealed indicating a decline in inflammatory processes on the ocular surface. (Funke *et al.*, 2014)

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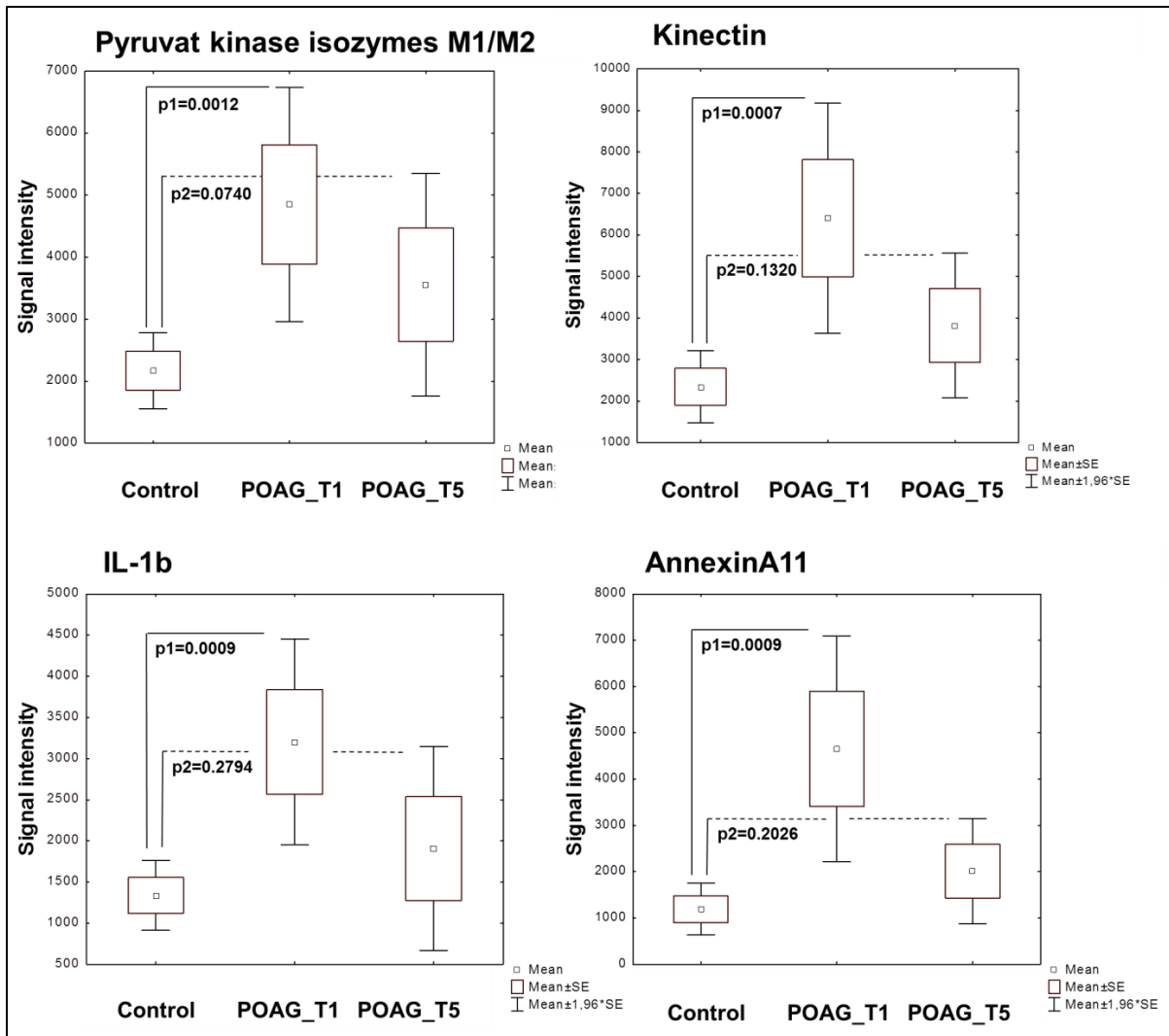


Fig 70 Statistical estimation of level approximation in the course of Taflotan[®] sine application comparing control group to POAG group mean levels of exemplary candidate tear proteins (T1= study start, T5= study end): E.g. PKM showed an elevated significant different mean level to control at T1 and approximated to the control level at the end of the study (T5) displaying no significant mean level difference between POAG and control anymore. (Funke *et al.*, 2014)

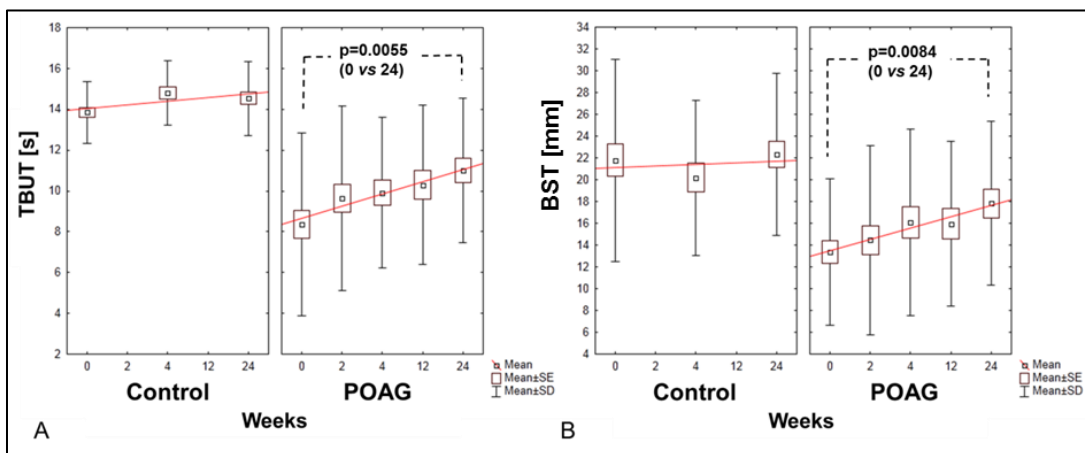


Fig 71 Significant improvement of clinical parameters, TBUT (A) and BST (B) observed after the therapeutic switch from Xalatan[®] to Taflotan[®] sine indicating ocular surface recovery. (Funke *et al.*, 2014)

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4.6 Influence of taurine on the ocular surface

4.6.1 Taurine effects on the tear proteome of contact lens wearers and sicca patients

Regarding 267 LC MALDI identified capillary tear proteins, longitudinal effects of taurine application were found as distinct potential responses concluded from linear regression over the 5 week study period in 9 proteins supported by R^2 values ≥ 0.5 (illustrated for exemplary tear proteins in Fig. 72). All of these corresponding proteins showed negative regression slopes representing a longitudinal protein level decrease as a potential response (Fig. 73). Linear taurine responses were predominantly observed in the contact lens group, whereas in the sicca group no distinct effects could be documented. Interestingly, the main portion of the responding candidate proteins refer to the immune system, e.g. complement C3 (score 383), Ig γ -4 chain C region (score 51), Ig λ -7 chain C region (score 826), secretoglobin 1D1 (score 680), α -2-macroglobulin (score 507) and haptoglobin-related protein (score 125). Thereby α -2-macroglobulin showed the most distinct level decrease towards taurine application reflected by its regression slope value (Fig. 73). Also, secretoglobin 1D1 and Ig λ -7 chain C region showed distinct negative slopes. Revealing non-linear taurine responses the *Kruskal-Wallis* analysis documented additional, prevalent inflammatory proteins like several Ig κ -related subunits. In contrast to linear responding candidates, these proteins showed a connatural bias in both taurine treatment groups; contact lens and sicca. Interestingly, the non-linear responding proteins showed a longitudinal level decline in compliance to the linear trend analysis. Ig κ VIII region HAH (score 107) and Ig κ chain C region (score 4135) (Fig. 74) showed significant effects towards taurine application ($p < 0.05$). Post-hoc analysis revealed significant differences between the S_Taurine and CL_NaCl group for these two proteins. Regarding distinct tendencies, protein AMBP (score 39), putative beta-actin-like protein 3 (score 241) and transcription elongation factor B polypeptide 3 (score 129) were found to be downregulated by taurine treatment ($p < 0.08$) in the sicca and in the contact lens group. Taurine responding candidate proteins were furthermore verified by comparing initial tear protein levels of contact lens wearers and sicca patients with those of healthy non-contact lens wearers. Distinct changes (≥ 2 fold; ≤ -2 fold change) in protein levels of both groups; sicca and contact lens compared to those of healthy non-contact lens subjects could be demonstrated (Fig. 75). Focusing on the subset of taurine down-regulated candidates from both, linear and non-linear analysis, levels of all proteins were found to be increased in tears of dry eye and contact lens wearers compared to the healthy non-contact lens wearer tear pool. Ig λ -7 chain C region (score 826) showed a highly consistent strong level increase in both groups, sicca and contact lens compared to the healthy reference, whereas Ig γ -4 chain C was found to be upregulated in the sicca group with a higher level than in the contact

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lens group. Most of the other proteins showed moderate differences between contact lens and dry eye, but significant alterations compared to the healthy reference.

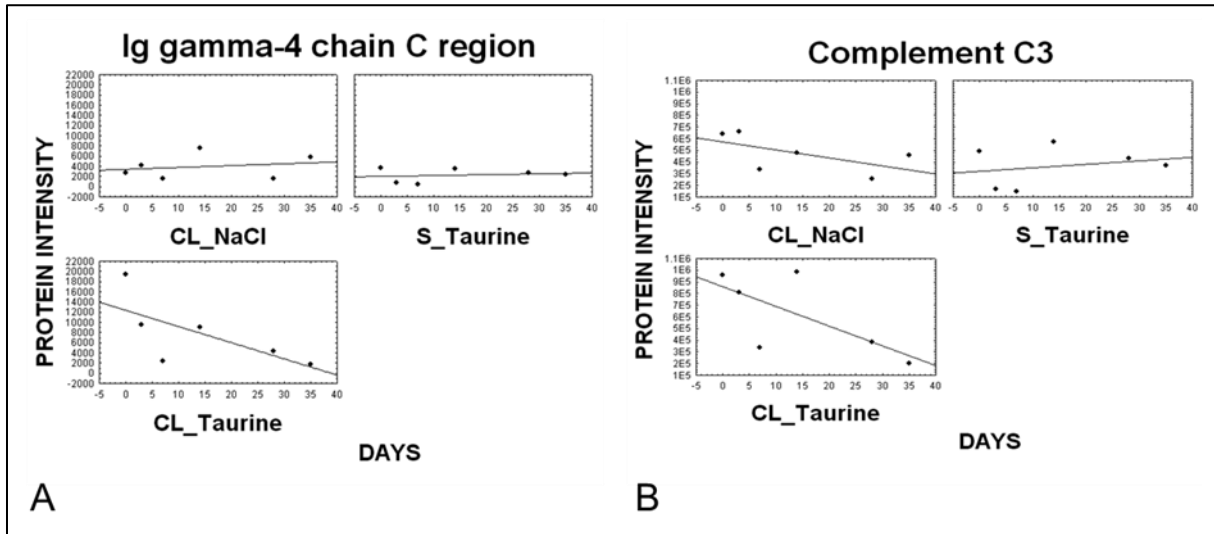


Fig 72 Exemplary tear proteins responding to taurine application in the course of the study in the contact lens group (CL_Taurine) with linear level decline indicated by $R^2 \geq 0.5$. (Funke *et al.*, 2012)

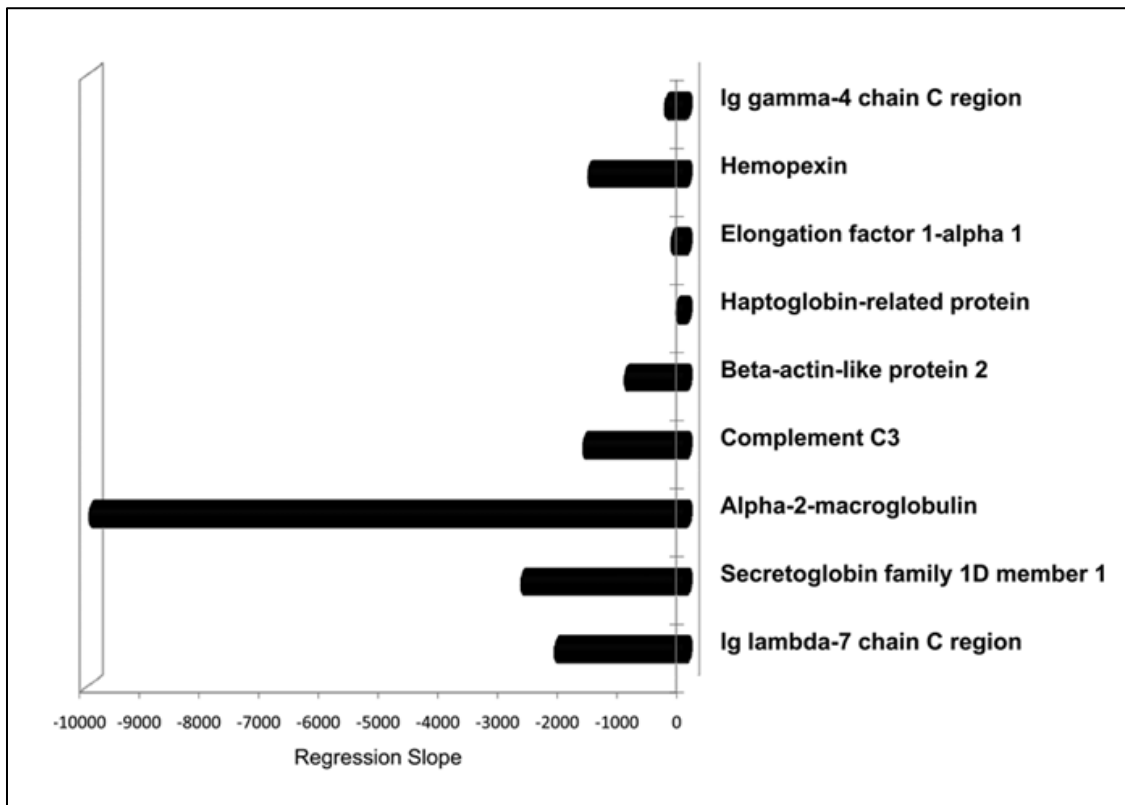


Fig 73 Slope values illustrated for distinctly linear taurine responding tear proteins: α -2-macroglobulin displayed the highest level decrease towards taurin application. (Funke *et al.*, 2012)

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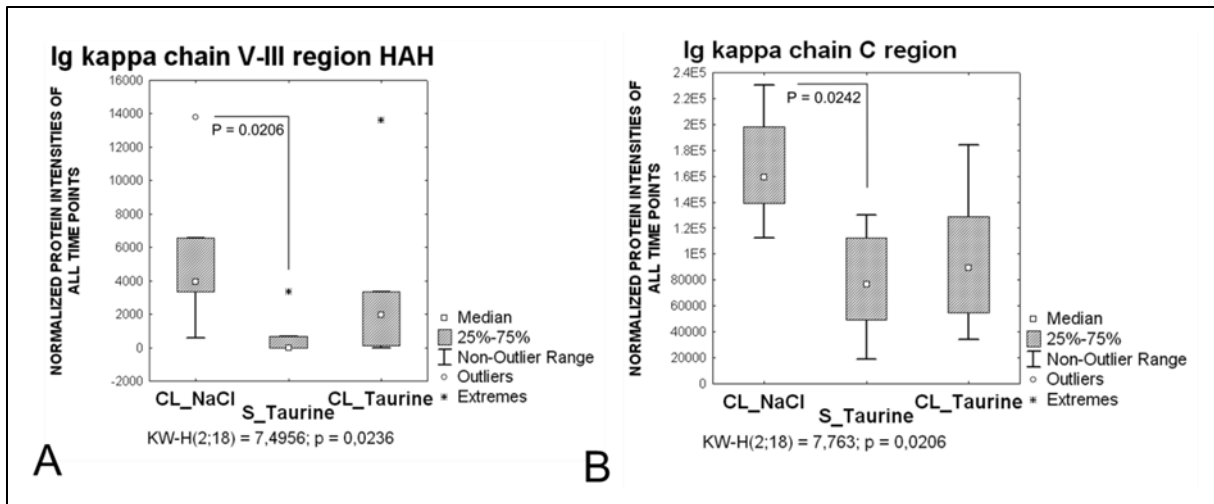


Fig 74 Non-linear taurine responses revealed by *Kruskal-Wallis* analysis shown for two exemplary tear proteins, whereby both proteins showed significant differences between sicca and control group revealed by post hoc analysis. (Funke *et al.*, 2012)

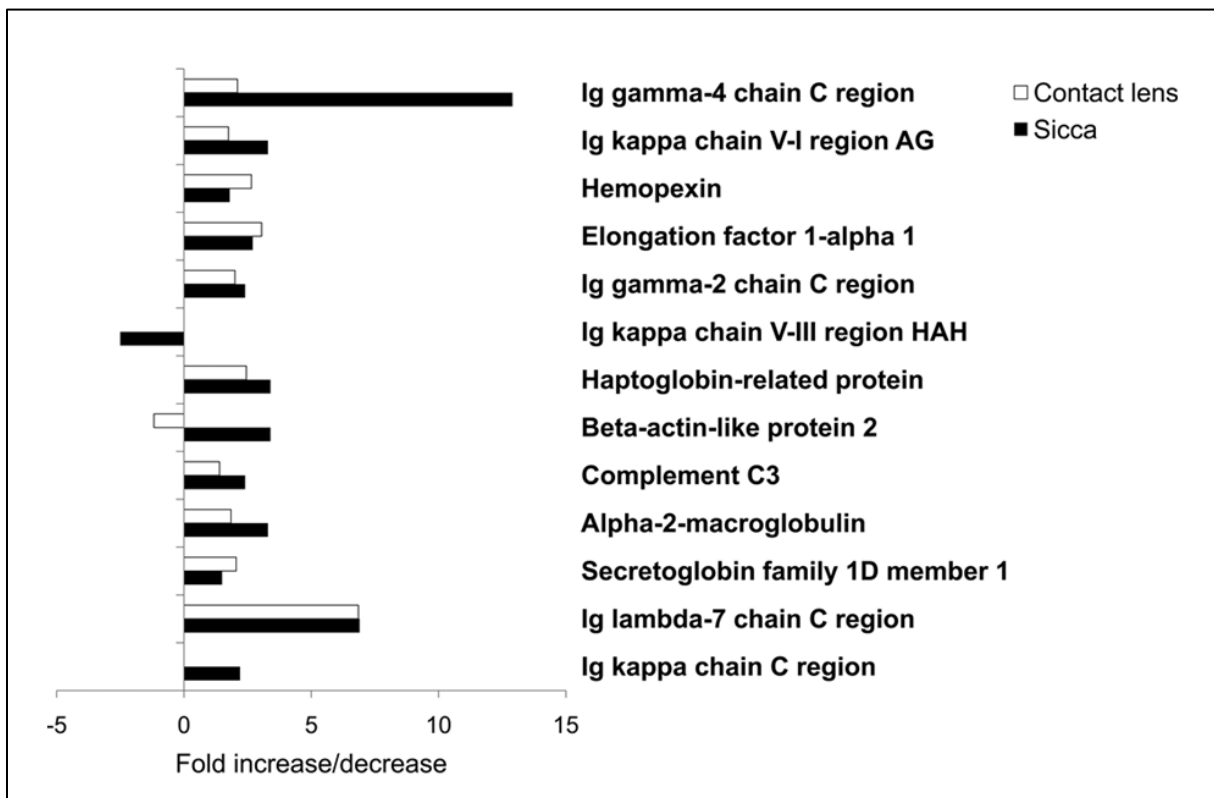


Fig 75 Level changes between sicca patients and contact lens wearers in reference to a healthy non-contact lens wearer group ($N=8$) as zero x-axis at study onset. (Funke *et al.*, 2012)

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4.6.2 Taurine effects on the conjunctival proteome

Regarding the oxidative stress level of H₂O₂ stressed NHC-IOBA cells, it could be demonstrated that incubation with tested taurine concentrations (0.05, 0.2, 0.5, 1, 2 mM) lead to a distinct decline in ROS production compared to stressed control cells, which were not taurine treated. Accordingly, application of 0.05 mM taurine lead to a ROS reduction of 26% ($p= 0.0356$), incubation with 0.2 mM to a decrease of 29% ($p= 0.0153$) and treatment with 1 mM taurine showed a decline of 32% ($p= 0.0016$). Also, the other tested concentrations showed distinct ROS decrease responses, however, not significant reactions (Fig. 76A). Despite the obvious effects on ROS production, no significant effects were observed for taurine treatment of H₂O₂ stressed cells regarding cell viability. Furthermore, staurosporine stressed cells also failed to show significant viability effects and showed only weak tendencies of increased viability, reflected by a viability increase of 7% in cells incubated with 0.05 mM taurine. In contrast incubation of unstressed cells with taurine lead to a significant increase in viability observed for 0.05 mM (26%, $p= 0.0055$) and 0.2 mM (19%, $p= 0.0181$). Whereas also 0.5 mM taurine showed tendency of increased viability (15%, $p= 0.0681$), concentrations above this value showed no effect or negative responses of cell viability towards taurine treatment (Fig. 76B). Incubation of cells with taurine in a 20-100 mM range concluded in a significant loss of cells up to a percentage of 20% (100 mM, $p= 0.0010$) (Fig. 77B). Accordingly, in the presence of staurosporine for these critical concentrations additive negative effects could be observed with a maximum of 32% viability decline at 100 mM taurine ($p= 0.0001$). Moreover, ROS levels were found to be increased significantly from taurine concentrations above 20 mM with a maximum of 102% supplement of ROS production at 100 mM taurine ($p= 0.0005$). Thereby, incubation of stressed cells with taurine showed no significant counteraction towards ROS production displaying a maximum ROS level of 82% at 100 mM taurine ($p= 0.000002$). Contrary to ROS results of low taurine concentration, slight ROS elevations could also been observed in the initial concentrations, but were not revealed as significant and clearly differing from higher concentrations above 20 mM (Fig. 77A). In summary, these findings give evidence of taurine dose dependent effects regarding cell viability and ROS production. Whereas taurine incubation performed beneficially on cell viability and ROS generation in a 0.05-0.5 mM range, concentrations >0.5 mM displayed negative effects. For BU proteomic analysis on cellular taurine effects cell lysates obtained from taurine treated unstressed NHC-IOBA cells displayed high qualitative SDS PAGE migration pattern for both study parts corresponding to “elevated” and to “low” taurine concentrations (Fig. 78). Focusing on proteomic results, LC ESI analysis of unstressed conjunctival cells incubated with critical “low” taurine concentrations (4, 20, 50 mM) revealed several taurine target protein candidates. Thereby, taurine affected cellular proteins could be revealed. Interestingly, exclusively significant elevations reflected by a

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minimum 2fold level increase in comparison to the control mean level could be observed. 30 proteins showed significantly increased levels in minimum one of the taurine concentration groups (4, 20, 50 mM) and a tendency of increase in the remaining groups (Fig. 79). Among these proteins, phosphoglycerate kinase 2 (score 3317), protein SET (score 2713), ubiquitin 60s ribosomal protein L40 (score 2235), protein S100 P (score 927), enoyl-CoA delta isomerase 1 (score 561) were found as the top 5 scoring proteins. Moreover, several of the taurine responding proteins revealed by fold change displayed a significant linear response according to an increasing taurine concentration, which makes them highly suspicious for direct or indirect taurine interaction, e.g. glutamate dehydrogenase 2 ($R^2= 0.978$, $p= 0.011$) and glutaredoxin 1 ($R^2= 0.959$, $p= 0.021$). Next to the affected glutaredoxin 1 (score 214), which is indicative for cellular stress (Anathy *et al.*, 2009, Enoksson *et al.*, 2005, Murata *et al.*, 2003, Voehringer, 1999), numerous further altered proteins were found to be involved in cellular stress and apoptotic processes, e.g. cytochrome C oxidase subunit 2 (score 69) (Buron *et al.*, 2006, Green and Reed, 1998, Kadenbach *et al.*, 2004), which showed the highest increase level of all affected proteins (12.4 fold) or septin 2 (score 185) (Garcia-Fernandez *et al.*, 2010, Kinoshita, 2003, Longtine *et al.*, 1996). Also, an interaction with thimet oligopeptidase (score 143), a cytoplasm located metalloprotease (www.uniprot.org) could be proposed since taurine was also demonstrated to interact with matrix metalloproteinase 9 (Park *et al.*, 2000). Also sodium/nucleoside cotransporter 2 (score 37), nevertheless detected with a low score, could be proposed to interact with taurine since taurine was found to be released from cells by treatment with adenosine transport inhibitors (Hada *et al.*, 1996). In summary, the level increases of the mentioned proteins most likely indicate a direct cellular response towards elevated taurine concentrations which fits well with the observed negative taurine effects for these concentrations, demonstrated by ROS and cell viability assays. Additionally, the detected apoptotic and stress response proteins reflect the declined viability of the cells incubated with critical high taurine concentrations. In accordance, these findings support the functionality of the LC ESI MS workflow since apoptotic events on the cellular level could be reflected on a proteomic level. Thus, a huge contingent of responding proteins was not associated to apoptosis, conclusively highly interesting to be modulated by taurine in alternative pathways. Encircling both apoptotic associated and non-associated proteins, among the responding proteins several candidates were found to interact in a direct or indirect manner with taurine inferred from the literature, e.g. phosphoglycerate kinase (Midwinter *et al.*, 2004, Nandhini *et al.*, 2005, Peskin and Winterbourn, 2006), cytochrome C oxidase (Millett *et al.*, 1982), protein S100 P (Reymond *et al.*, 1996) and glutamate dehydrogenase 2 (Schaffer *et al.*, 2000). Another interesting protein that was found upregulated by taurine treatment was sepiapterin reductase (score 97), which was suggested to play a crucial role in lipid metabolism (de Roos *et al.*, 2005). Upregulation

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of proteolipid protein 2 (score 235), also gave evidence for taurine interaction. Since hydrophobic surfactant proteins had been already detected for tear fluid and/or the lacrimal gland system and had been proposed to play a key role for tear film stability (Brauer *et al.*, 2007a, Brauer *et al.*, 2007b) it is likely to suggest a comparable function for proteolipid 2 and moreover worth to mention, that it responds to taurine treatment. Since the membrane modifying potential of taurine is well documented (Huxtable, 1990, Llew *et al.*, 1992) proteolipid 2 increase could be a consequence of taurine-membrane interactions. Confidently, transmembrane protein 109 (Mitsugumin 23) (score 411), an ER/nucleus membrane protein (Venturi *et al.*, 2011), detected for the first time for the ocular surface in this study, was found to display a level increase by taurine treatment, which emphasizes the influence of taurine on membrane systems. Regarding response reactions towards taurine treatment, several proteins showed no distinct fold changes, but 87 proteins were found to show a significant linear response correlating with taurine concentration increase, probably indicating a weak taurine response. Among them, the top 5 scoring candidates were heterogeneous carbamoyl-phosphate synthase (score 13385), elongation factor 1- α 2 (score 9574), neuroblast differentiation-associated protein (score 6396), L-lactate dehydrogenase B chain (score 6193) and multifunctional protein ADE2 (score 1718). Candidate related proteins displaying a clear fold change were depicted by the linear analysis. For example, importin-9 (score 131) was found to be significantly elevated, whereas importin-5 (score 815) was found to show a linear increase (slope: 3405, $R^2= 0.959$, $p= 0.021$) below the fold change threshold. De facto, 47% of potential apoptosis marker proteins, that have been detected in a study of hepatic stellate cells incubated with taurine could also be detected in this study with relatively high scores, e.g. annexin A1 (score 6596), peroxiredoxin 2 (score 4946) or protein DJ-1 (PARK7, Parkinson disease protein 7) (score 633) (Deng *et al.*, 2010). Protein DJ-1 is known to directly interact with EF-hand calcium-binding domain-containing protein 6 (score 150) (www.uniprot.org), a nucleus protein, which was found to be significantly upregulated by taurine treatment in the present study. Since taurine is well documented for its role in calcium translocation and homeostasis (Dolara *et al.*, 1978, Pasantes-Morales, 1982), it is very likely that it interacts with calcium-binding proteins. However, only translationally-controlled tumor protein (score 698), as one of these proteins, showed a significant linear level increase towards taurine treatment in confidence with the results of the hepatic stellate cell study. Interestingly, several proteins associated to oxidative stress response have been also found to respond to taurine treatment in a linear manner, e.g. thioredoxin 1 (score 121) (Holmgren, 2000, Yildirim *et al.*, 2007). In summary, 87 proteins were found to show a linear pattern change with increasing taurine concentration encircling protein up-and downregulations. However, in most cases intensities were below the fold change threshold, but are likely to indicate a regulation prognosis. Regarding

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potential taurine protein targets obtained from LC ESI experiments, focusing on cells incubated with 0.05 mM taurine as preselected “beneficial” concentration, numerous responses could be recorded. Accordingly, 41% of the conjunctival cell proteins were found to respond to taurine treatment demonstrated by a minimum 2fold intensity change, whereby a small portion of proteins showed high level alterations towards taurine. Analysis of molecular function showed that the majority of the 2fold regulated proteins are involved in catalytic and molecule binding processes, whereby most of the proteins mediate protein-protein interactions (Fig. 80). In addition, whereas among the group of 2 to 5fold regulated proteins up and downregulation could be documented, the group of “highly regulated” proteins (>10fold) displayed exclusively downregulations, whereby like documented for the “mild regulated” proteins the majority of the “high regulated” proteins were involved in protein and nucleic acid binding (an overview of regulation distribution is given in Fig. 81). Thereby calmodulin, the taurine transporter control major player (Huang *et al.*, 2001, Ramamoorthy *et al.*, 1994), was found to be extremely decreased (24fold). Also, calmodulin-like protein 3 (score 95) was found to be extremely diminished (6fold). Additionally, another calmodulin related protein, calcium/calmodulin-dependent protein kinase type 1 (score 37) showed a slight decrease (2.3fold). ER calcium ATPase 2 (score 64) another calcium depending protein was also found to be weakly downregulated (2fold). These findings support a response of the calcium/calmodulin apparatus towards taurine treatment (Fig. 82). Furthermore a complete class of proteins, the 14-3-3 proteins (α , β , Σ , γ , η , ϵ , δ) (scores: 1207-1435) exhibited slight level diminishments (2.2-2.5fold). Beside intracellular proteins, 7% of candidate proteins were found to be associated to the extracellular milieu, whereby the secretory protein galectin-1 (score 203) was slightly decreased (2.5fold) and galectin 3-binding protein (score 94), was found to be highly diminished (14fold). Also, a signalling protein, interleukin enhancer-binding factor 2 showed a distinct decline (10fold). The response of secretory proteins indicates the impact of taurine on cellular interactions with the extracellular environment. Focusing on “highly regulated” candidates, a predominance of downregulated histone species could be observed (Fig. 83). Also, the level of histone binding protein RBBP4 was decreased (10fold). Furthermore, a decline of certain DNA interacting proteins like DNA binding protein A, Y-box protein 2, representative fragments of DNA primase/polymerase/lyase could be documented indicating gene expression changes. Downregulation of F-box only protein 22 is indicative for anti-apoptic processes, due to ubiquitin-dependent proteolysis and apoptosis mediation (Patton *et al.*, 1998, Winston *et al.*, 1999). High abundant stress and apoptose associated proteins, e.g. peroxiredoxin 4 (score 1047), 6 (score 1724), glutamate dehydrogenase 2 (score 73), cytochrome C (score 48), cytochrome C oxidase subunit 2 (score 43), thioredoxin (score 207), glutathione S transferase κ 1 (score 129) showed level declines between 2 and 3fold (Fig. 84). Additionally,

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glutathione S-transferase Ω (score 687), superoxide dismutase [Cu-Zn] (score 1419) and microsomal glutathione S-transferase 1 (score 92) exhibited distinct to extreme level diminishments (6fold, 7fold, 12fold). In contrast, several of related oxidative stress proteins, e.g. glutaredoxin 1, thioredoxin 1, glutamate dehydrogenase 2 and peroxiredoxin 2 were found to be upregulated in cells treated with critical taurine concentration >20 mM, which emphasizes the beneficial potential of lower taurine concentrations on cells. In summary, cell viability assays demonstrated a dose-dependent taurine agency on oxidative stress level and viability of NHC-IOBA cells. Distinct proteomic changes could be documented for adverse and beneficial taurine concentrations supporting the impact of taurine on conjunctival cells, thereby revealing numerous potential taurine protein targets.

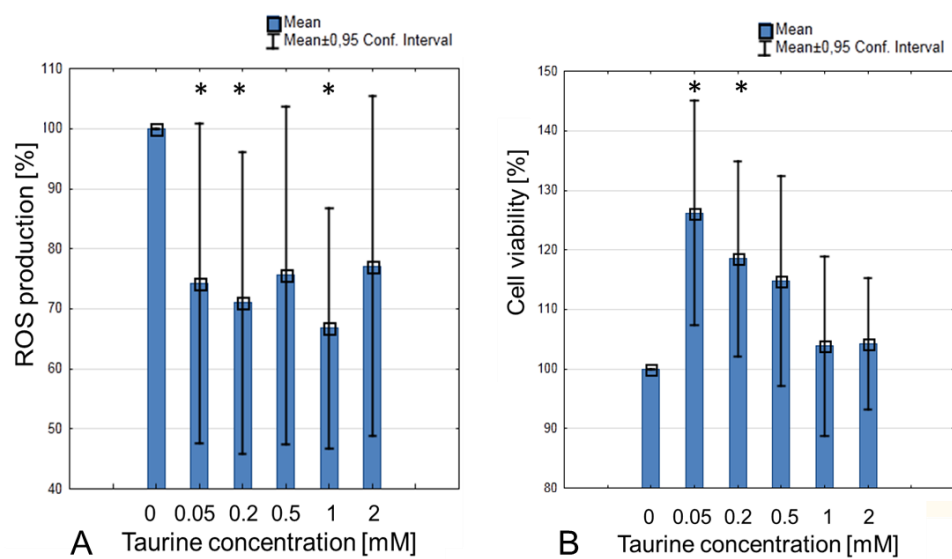


Fig 76 Influence of different low taurine concentrations on conjunctival cells; A: ROS production is distinctly decreased compared to untreated control cells; B: Viability increased by incubation with 0.05-0.5 mM taurine (significant responses $p < 0.05$ are indicated by asterisk).

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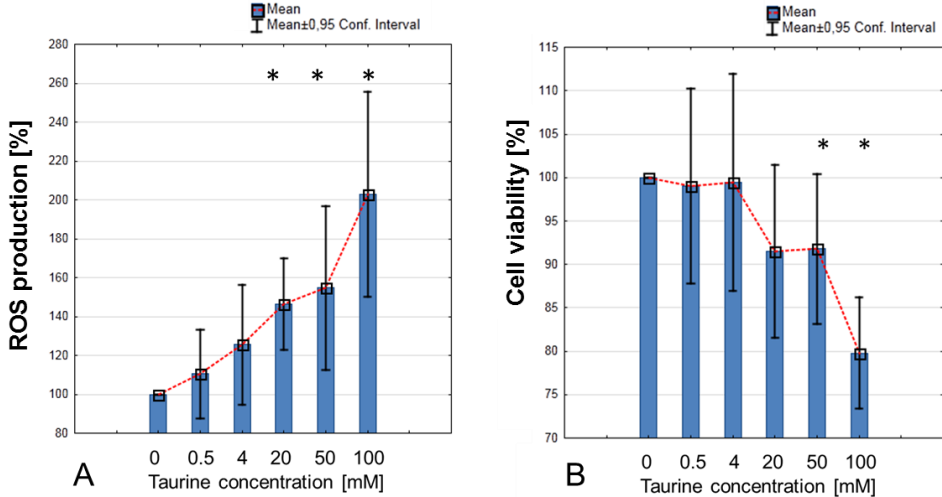


Fig 77 Influence of different elevated taurine concentrations on conjunctival cells indicating dose-dependent taurine toxicity in elevated levels; A: ROS production levels show an increase towards taurine treatment with significant differences from 20-100 mM taurine; B: Cell viability was found to decline from 20-100 mM. (significant responses p<0.05 are highlighted by asterisk)

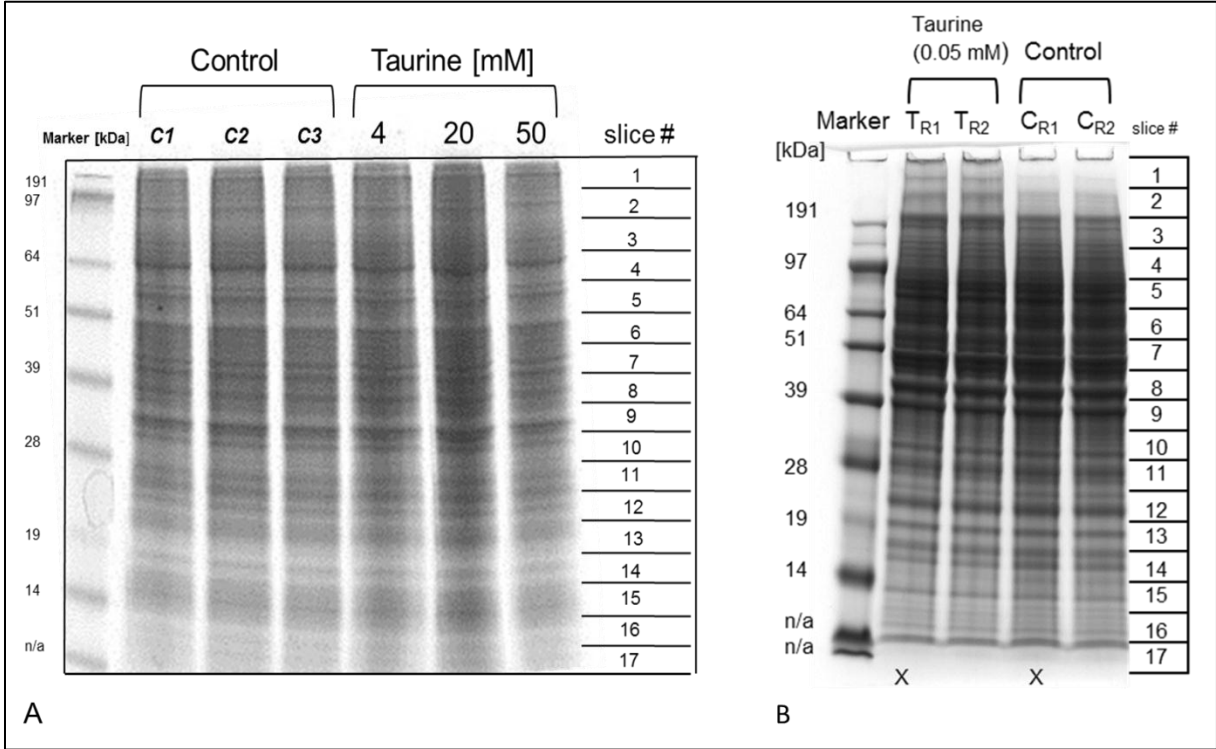


Fig 78 SDS PAGE migration pattern of lysates corresponding to pooled conjunctival cells treated with different taurine concentrations destined for LC ESI MS analysis; A: "Elevated" taurine concentration range (C1-C3= untreated control cell lysates, all lanes were selected for MS analysis); B: "Low" taurine concentration range (R1/R2= technical replicates, lanes highlighted by x were selected for MS analysis).

Results

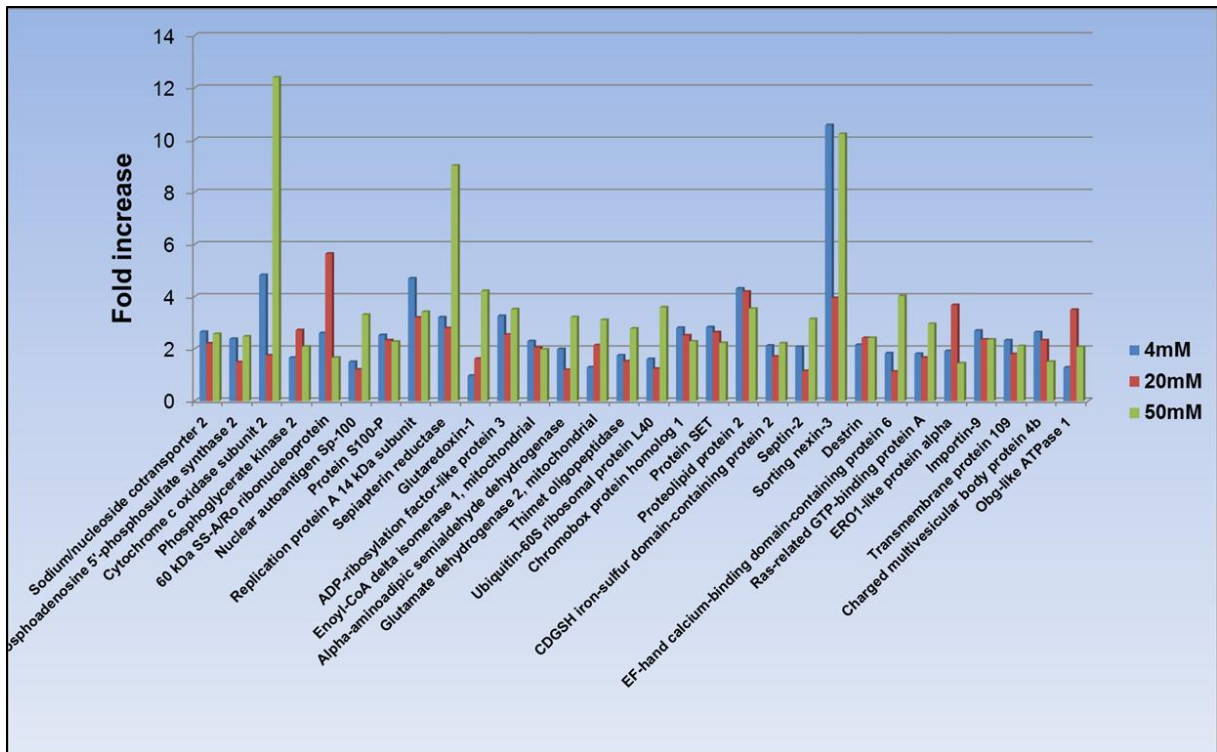


Fig 79 Conjunctival cell proteins responding to incubation with different “elevated” taurine concentrations inferred from level alteration referring to untreated controls (x-axis: zero-line). High upregulations could be e.g. observed for cytochrome C oxidase subunit 2, septipaterin reductase, glutaredoxin 1, sortin nexin 3 which are involved in cellular stress.

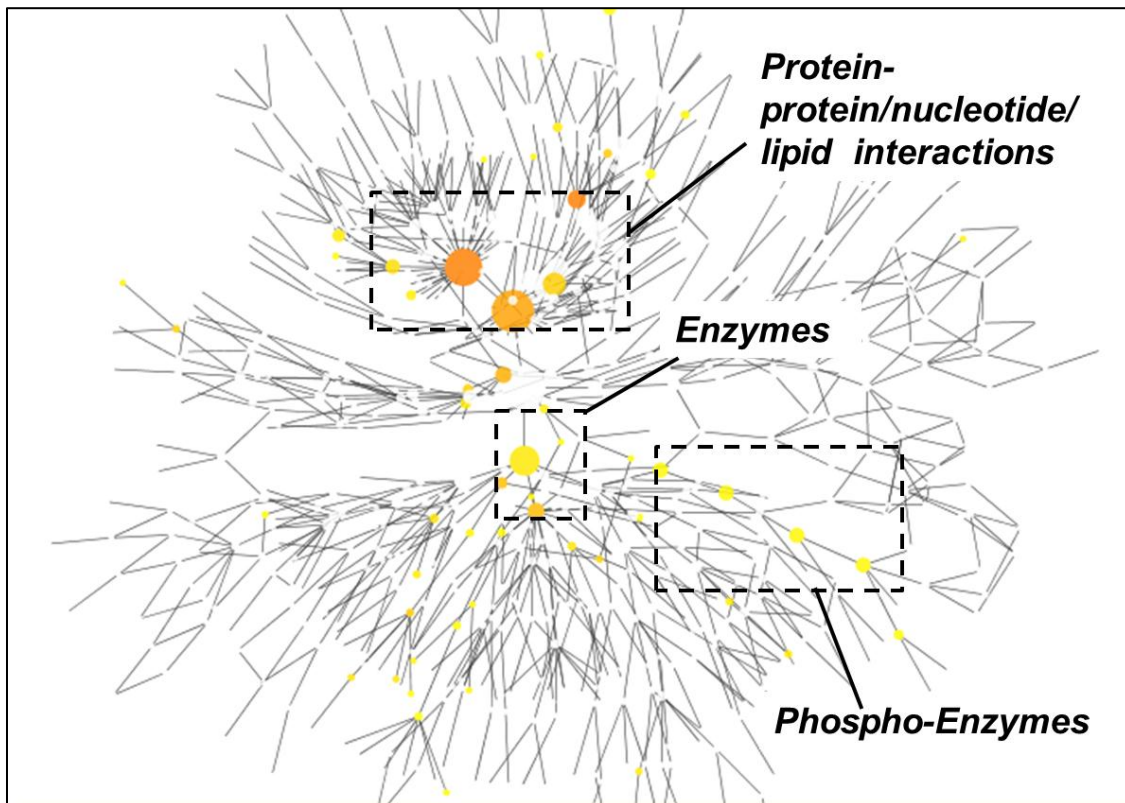


Fig 80 Cytoscape GO analysis of conjunctival cell proteins responding to incubation with 0.05 mM taurine indicating a predominant contingent of enzymes and binding proteins.

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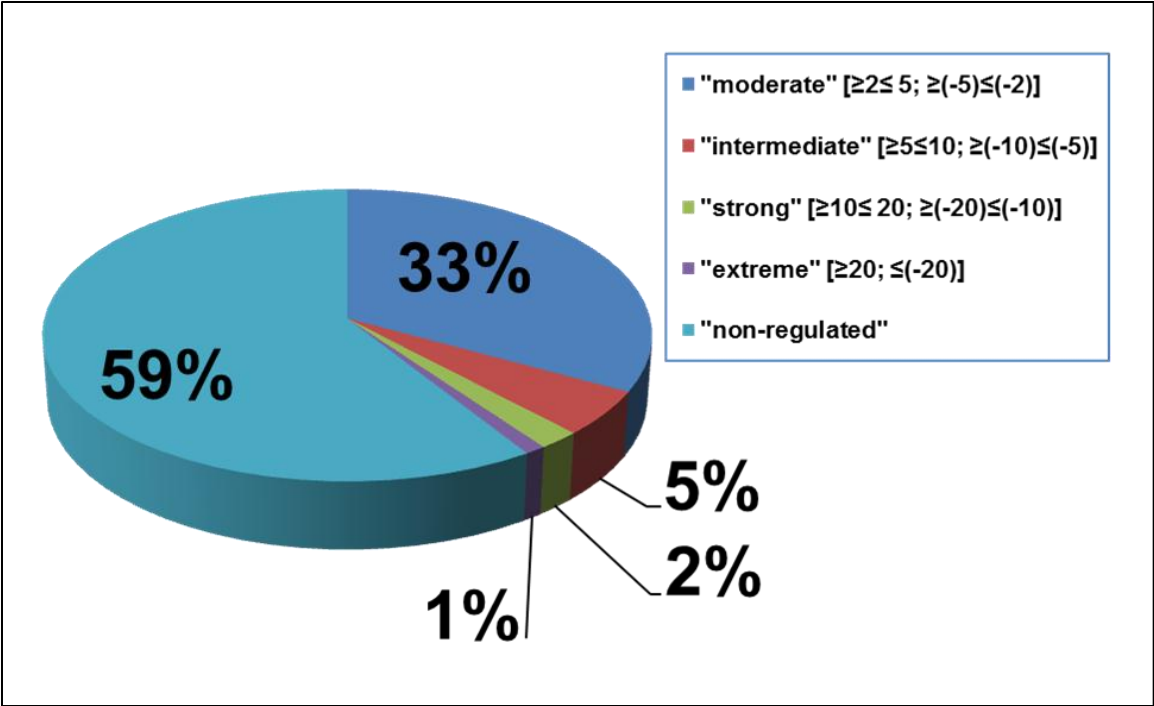


Fig 81 Classification and distribution of regulation forms towards incubation of conjunctival cells with 0.05 mM taurine.

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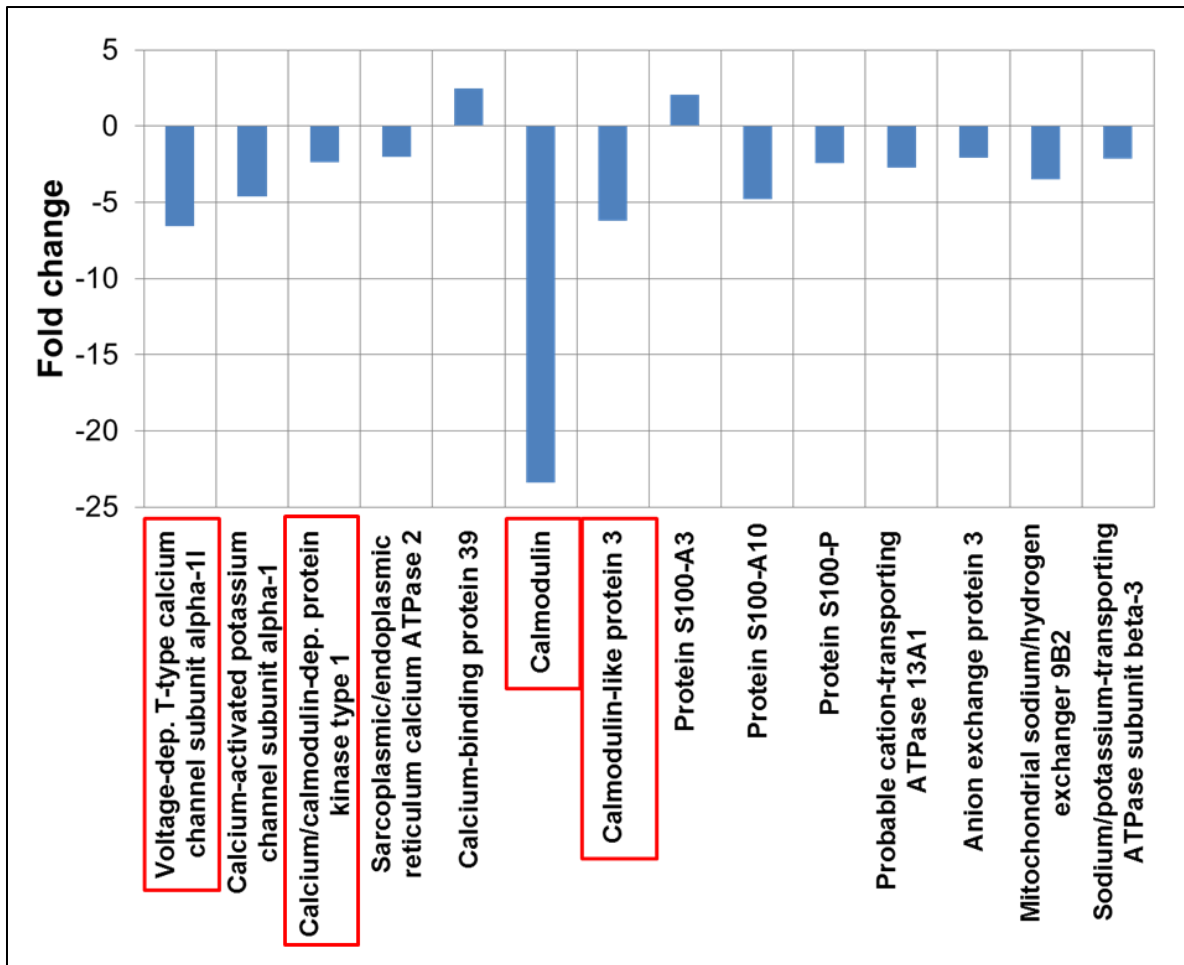


Fig 82 Exemplary taurine affected proteins revealed by LC ESI MS BU analysis: Members of the calcium/calmodulin system (highlighted in red frames) were distinctly downregulated after incubation with 0.05 mM taurine referring to an untreated control (x-axis, zero line) indicating changes in taurin traffic.

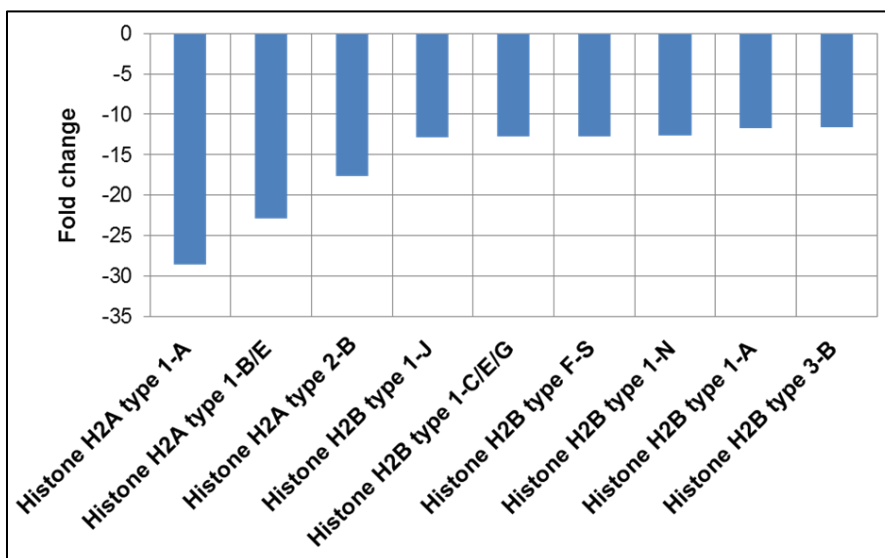


Fig 83 Distinctly decreased histone species levels revealed by LC ESI MS BU analysis observed in conjunctival cells incubated with 0.05 mM taurine in reference to an untreated control (x-axis, zero line).

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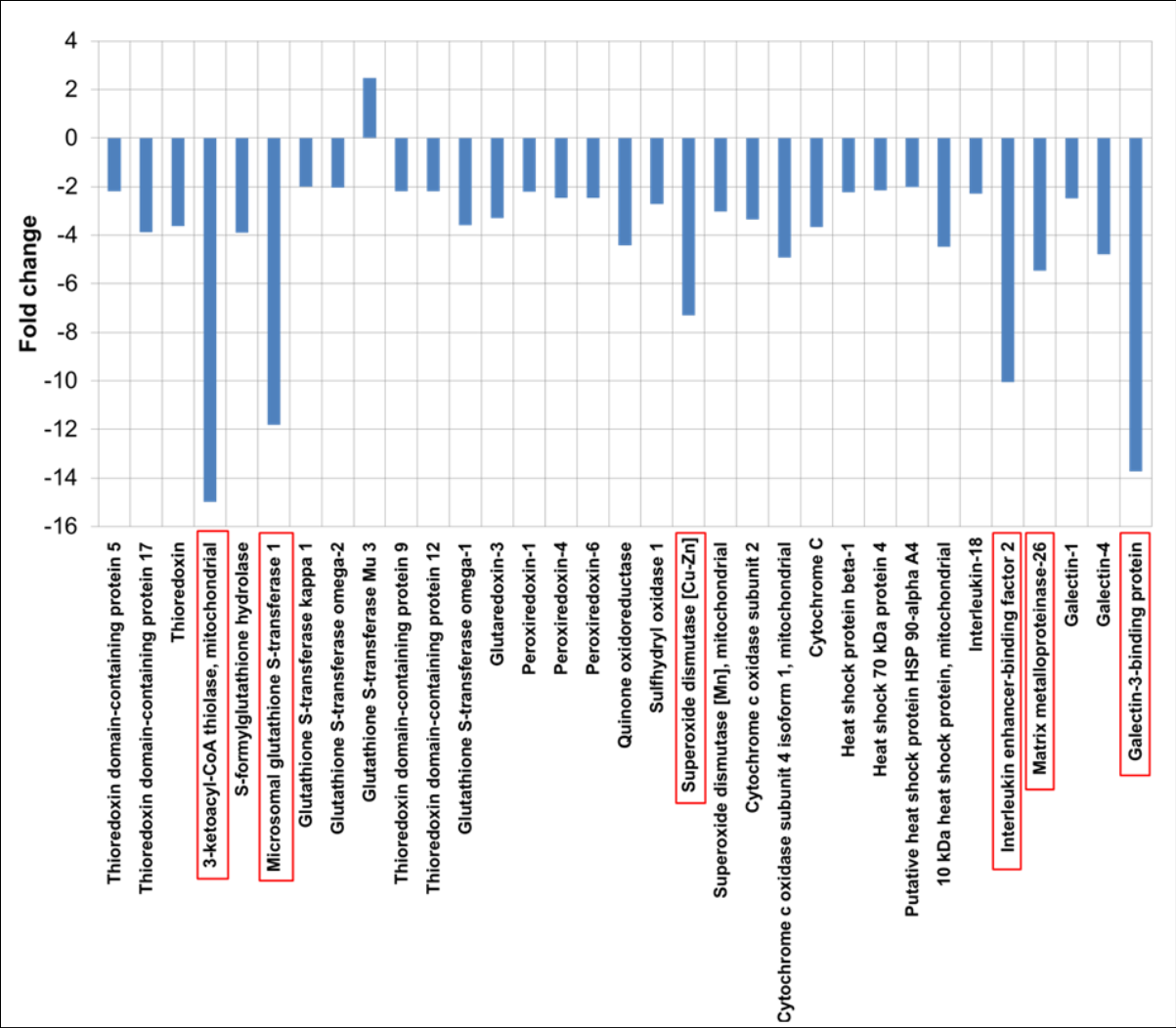


Fig 84 Cellular stress related proteins affected by 0.05 mM taurine incubation referring to a healthy control (x-axis, zero line) (distinct regulated protein candidates are highlighted in red frames).

5 Discussion

5.1 Top Down (TD) vs. Bottom Up (BU)

Referring to the data, BU analysis showed several advantages over the TD setup. In terms of the advantages of the TD workflow high analysis speed, minimal sample processing, robustness towards crude sample species, suitability for highthroughput studies, sensitivity towards the medium mass range of cellular materials, low costs and easy lab routine integration are highlighted. Besides, the method robustness allowed a highly pure view to the medium mass cell proteome. Like in SELDI, the main drawbacks of the engineered LC MALDI TD approach were attributed in the difficulties to directly identify intact protein (Nilsen *et al.*, 2011). Even by use of more sophisticated instrumentation like quadrupol analyzers, a direct fragmentation >6000 Da is still challenging (Peng *et al.*, 2009). Moreover, the diminished reflector detection sensitivity made it hard to select targets which already displayed low abundance in the linear mode. In fact, proteins of interest have to be enriched and purified for successful identification (Seibert *et al.*, 2004). Accordingly, follow-up strategies for recapturing the component of interest were found time-consuming and difficult, encircling enrichment of the target component by use of SPE, visualization by SDS-PAGE, alignment of the mass area, in-gel digestion and finally alignment to the literature. Time consuming and laborious accumulation of these TD components can also be realized by antibody pull-down experiments with subsequent MS redetection (De Bock *et al.*, 2010) or by use of size-exclusion chromatography combined with SDS-PAGE and reprofiling of the components of interest (Guo *et al.*, 2005). Nilsen and coworkers reported a more promising follow-up strategy for protein reprofiling recommending “on-chip elution” (Nilsen *et al.*, 2011). Nevertheless, in context of protein identification, initial TD benefits were decreased by the drawbacks of the follow-up steps. Furthermore, the low performance of the method to analyze sample species of less protein content, like tears, was found also disadvantageous. However, for fast and reproducible analysis of cellular sample species as demonstrated for RGC5 training samples, the workflow is highly suitable and can also be used for the analysis of ocular surface epithelia lysates to monitor overall proteomic effects that may indicate cellular responses, e.g. in therapeutic studies. Such proteomic patterns have been successfully used in a TD study to monitor the influence of contact lens cleaning solutions on conjunctival cells (Bell *et al.*, 2012a), to evaluate the influence of glaucoma patient serum, antibodies (Bell *et al.*, 2012b) and cyclooxygenase inhibitors (Brust *et al.*, 2006) on retinal cells and to characterize tumor cells (Cazares *et al.*, 2002, Gretzer *et al.*, 2004, Yanagisawa *et al.*, 2003, Zhukov *et al.*, 2003). Another approach could be to address small protein/peptide components (Jimenez *et al.*, 2007a, Kutz *et al.*, 2004, Villanueva *et al.*, 2006) or low molecular components <1000 Da (van Kampen *et al.*, 2011), e.g. ocular surface

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degradative fragments (Abalain *et al.*, 2000, Wheater *et al.*, 1999) or profiling of known masses. To overcome the lack of identification, the method can also be suited for the analysis of tryptic digests initially facilitating statistics followed by fragmentation of tryptic peptides of interest (Dekker *et al.*, 2005, Neubert *et al.*, 2008). Nevertheless, in this work for in depth analysis of the ocular surface, encircling conjunctival cells and tear fluid, the BU strategy has turned out the more effective approach characterizing proteins >4000 Da. Also, SDS PAGE as a first fractionation dimension allows visual inspection and densitometric evaluation if needed (Neuhoff *et al.*, 1990). Regarding in-gel digests it is possible to directly monitor protein markers in a wide mass range obtaining knowledge of their identity. In contrast, in TD approaches a high risk remains that significant markers stay unidentified resulting in a costly loss of biological information and making targeted follow-up studies on candidate proteins impossible. In fact, by use of the BU approach “in depth” analysis of human tear fluid and cell samples could be realized providing >1000 proteins per sample type, whereby proteins could be annotated to biological functions and subcellular localizations and could be further reconciled with the literature for disease and clinical acquisition. Nevertheless, also several limitations have to be mentioned. First of all, the approach is more costly, time and lab intensive, since proteins have to be fractionated by SDS-PAGE, in-gel digested and in case of LC ESI purified by SPE prior to analysis. Considering sample preprocessing steps and the more profound analysis protocol the method is less easy to implement to the daily lab routine as the TD approach. Further drawbacks are the use of pool samples to reduce costs and analysis time, especially in labs with a high project contingent. Accordingly, there exists a challenge in statistical evaluation and the need for time consuming and costly validation experiments, which can be realized by antibody-based microarray analysis of MS derived marker candidates. In fact, the use of pool samples is widely practiced in proteomic analysis (Castano *et al.*, 2006, Neubauer *et al.*, 2006, Tonge *et al.*, 2001) addressing costs and limitation of resources, and was found as a valid but considerable strategy in terms of reduction of biological variance and comparison between pool and mean protein expression (Diz *et al.*, 2009). Thus, despite sample pooling has been proposed to “mute unrelated alterations by dilution” (Weinkauff *et al.*, 2006) it seems to exist a sample-dependent ratio between technical and biological variation that has to be considered for the setup of experimental designs (Karp and Lilley, 2009). De facto, an “in depth” analysis of a large number of individual samples is not realistic in case of the developed BU strategy considering costs, effort, analysis time and realization of numerous parallel running projects. However, sensitivity and the possibility to identify large numbers of proteins allowing orthogonal follow up research on identified candidate proteins and the possible establishment of large protein maps with knowledge on protein localization and function achieved by the BU approach surmount the possibilities of the developed TD

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approach. However, there is a high potential in combining both developed strategies in the analysis of cellular ocular surface sample species, BU studying proteins >4 kDa and TD monitoring statistical changes in small protein fragments. As an outlook, the fast TD screening for known signal peptides in digests which have already been proven for particular proteins by use of the intensive BU proteomic strategy or orthogonal methods may be a promising future direction to combine benefits of both strategies. For this reason, the protein maps established in this work could be screened for signature peptides that uniquely correspond to a particular protein of interest and their fast reprofiling in large sample cohorts by use of the TD approach, probably allows less costly, lab-intensive and deeper statistical understanding of ocular surface disease processes. However, such signature peptides have to be proven in TD analysis in a first step, which is the challenge in such a combinatory strategy with respect to different ionization mechanism in ESI and MALDI systems.

5.2 Bottom-up (BU) analysis: LC-MALDI vs LC-ESI workflow

The obtained results support the high reproducibility for both approaches. However, there are distinct differences between MALDI and ESI workflows regarding the developed BU approaches. The RT reproducibility was excellent in the ESI workflow with low precursor mass shifting additionally supported by CV values <1% in confidence with the literature reporting values between 0.1 to 4.5% observed for comparable systems (Duan *et al.*, 2009, Dunn *et al.*, 2008). In comparison, also the MALDI approach showed a high RT reproducibility. An important factor which impacts on LC separation efficiency in LC MALDI experiments is the fraction collection performance and sample space limitation regarding target plates. In contrast, separation efficiency in continuous LC ESI workflows is predominantly limited by the column system and flow rate. However for both approaches there is a need to compromise between efficiency, lab effort, costs and analysis time regarding analysis of multiple sample sets settling pro and contra between both strategies. Focusing on the effort/output ratio for analysis of complex sample types, the LC ESI workflow presented superior over LC MALDI. That is why the sensitivity was found to be clearly higher in the ESI method regarding severe complex sample species like tear fluid to more complex sample species like cell lysates. Without a doubt, this is not explicable by the ionization technique but due to the coupled hybrid MS system providing a higher dynamic range (Makarov *et al.*, 2006b) for higher performance than TOF systems (Zhang *et al.*, 2000). De facto, the main reason for the high accuracy is based on the high resolution achieved by the orbitrap system clearly exceeding TOF systems thereby exceeding a critical value which allows the discrimination between coeluting components (van der Heeft *et al.*, 2009). In comparison with the LC ESI based highly accurate hybrid system, the MALDI platform cannot go strong with its beneficial attributes like high salt and contaminant tolerance, small

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sample consumption and sample storage for reanalysis (van Kampen *et al.*, 2011, Zhang *et al.*, 2004a). As a matter of fact, regarding practical aspects of the preionization sample process fraction, deposition complications due to abrasion of the robot tip in the MALDI workflow balanced with spray incursions due to thermal damages of the spray needle in the ESI approach. Nevertheless, whereas the MALDI approach saved time in sample pre-processing due to higher contaminant tolerance it lost speed in fraction deposition and matrix spotting compared to the ESI approach. A further interesting difference was observed by tear film analysis, whereby it was shown that the MALDI approach yielded in extremely high scores for a few prominent tear proteins, whereas the ESI approach identified a higher number of proteins but did not accomplish the absolute score values resulting from the MALDI analysis for selected predominant tear proteins, e.g. proline-rich protein 4 or lysozyme. Most likely this observation reflects adverse of the TOF analyzer resolution performance for complex protein mixtures in comparison to the orbitrap analyzer. Another possibility is the concurrence for ionization and ion suppression effects (Annesley, 2003, Jessome and Volmer, 2006, Muller *et al.*, 2002, Wang *et al.*, 2004), which may lead to a positive discrimination of abundant peptide ionization in the MALDI approach. Preference of high intensive peptides is a well-known phenomenon in MALDI experiments and counteracting strategies like replicate runs considering precursor ion exclusion lists (Chen *et al.*, 2005) have been developed. Furthermore, fragmentation strategies in both instruments are different, whereas the MALDI approach was prepared for PSD, the ESI method was tuned for CID fragmentation leading to different fragment pattern (Hoteling and Owens, 2004, Li *et al.*, 1999). Although, the possibility of reanalysis and optimization of the measuring protocol of identical sample sets is an important advantage of MALDI over ESI technology (Ishihama, 2005), it is hampered by the need for analysis time. Actually, the fragmentation speed of the TOF/TOF systems was found to clearly underlie the speed of the LTQ Orbitrap instrument. A problem observed in both developed BU workflows was the high portion of fragment spectra, which failed to successfully contribute in probability-based protein identification in data bases (Perkins *et al.*, 1999). Accordingly, the generation of large amounts of redundant data, whereby a high percentage of high quality MS/MS spectra fail to contribute to protein identification, is a known phenomenon (Cox *et al.*, 2008, Karty *et al.*, 2002, Nesvizhskii, 2010, Neuhauser *et al.*, 2012, Xu *et al.*, 2005). Consequently, strategies are on the way to exploit biological information out of unassigned spectra, whereby e.g. the use of “de novo sequencing” algorithm was proposed by numerous authors (Ning *et al.*, 2010, Seidler *et al.*, 2010). From a practical point of view the MALDI method is more lab intensive, since fractions have to be deposited and matrix has to be applied by pipetting robotic units, which can be sources of error. On the other hand it is more robust towards contaminants than ESI, which allowed a direct HPLC analysis without SPE purification of gel

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digests prior to LC MALDI. However, the more obvious advantage is the decoupling of sample preparation and measuring in the MALDI approach, allowing storage and reanalysis of samples (Kuzyk *et al.*, 2009), whereas in case of system errors samples are lost in the ESI approach. In these studies, during test runs, this happened in case of robot error, column blocking or needle damage. However, in the developed LC ESI workflow each sample provided enough material for one reanalysis. Thereby the storing was a more critical factor since freezing of remaining sample runs in sample buffer or rehydrophilization of samples was found to impact sample quality. Nevertheless, the complex MALDI software platform was found to be an important source of error in the course of the studies which was not the case in LC ESI. Further differences could be attributed to the complementarity of ionization principles (Bodnar *et al.*, 2003, Stapels and Barofsky, 2004, Stapels *et al.*, 2004, Yang *et al.*, 2007). In up to date literature, it was reported that more hydrophilic peptides had been obtained in MALDI (Yang *et al.*, 2007), whereas ESI favors more hydrophobic species (Cech and Enke, 2000). Moreover, whereas ESI was supported to favor ionization of peptides ending with lysine over those ending with arginine, the opposite was observed for MALDI ionization (Stapels and Barofsky, 2004). The complementary ionization character had been earlier exploited to analyze HDL apolipoproteins (Bondarenko *et al.*, 2002). In confidence, this work showed the differential character of both ionization techniques combined with two different analyzer strategies, which balance advantages and limitations. In conclusion, both approaches have to be used as an amendatory proteomic platform. Whereas the ESI approach exhibited high performance in getting an encompassing view of the ocular surface proteome, the MALDI method achieved high accomplishment in the characterization of particular tear proteins allowing targeted study designs. Both approaches were successfully used in longitudinal study designs, which are of growing interest in proteomics science according to the highly dynamic and complex character of the proteome (Cohen *et al.*, 2008, Corthals *et al.*, 2000, Flood-Nichols *et al.*, 2013, Hawkridge and Muddiman, 2009, Hawkridge *et al.*, 2010, Pratt *et al.*, 2002, Zabel *et al.*, 2012).

5.3 Characterization of the ocular surface proteome

By use of the developed workflows an in depth view to tear film and conjunctiva proteome could be obtained. Despite the intensive study of the corneal proteome by Dyrland and coworkers, who described 3250 proteins associated to different cornea regions encircling epithelium, endothelium and stromal cells proteins by use of a sophisticated TripleTOF 5600 system (Dyrland *et al.*, 2012), only few explorative investigations had been realized examining conjunctival cells. Actually, Zhou and co-workers identified 600 proteins in NHC-IOBA cells by MS (Zhou *et al.*, 2011). Nevertheless, by use of the developed LC ESI workflow >1000 conjunctival proteins could be catalogued. Furthermore, a rough proteomic

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based classification of the IOBA-NHC cells could be achieved in the present work. Conjunctival cells are classified in five groups, whereas only two groups, goblet cells and stratified squamous cells had been successfully biochemical defined so far (Dartt, 2002). Despite the difficulties to characterize conjunctival cell lines, Tong and coworkers found a high degree of congruency between NHC-IOBA cells and primary cultures proposing NHC-IOBA cells as a valid substitute model for the conjunctival epithelium, which had been realized by use of gene expression monitoring (Tong *et al.*, 2009). In confidence with the literature numerous epithelial key proteins encircling cytokeratines, desmogleins, epiplakin (Fukuoka *et al.*, 2012) and others could be identified by use of the developed LC ESI BU workflow, which justified once more the use of NHC-IOBA as an ocular surface epithelia model, e.g. allowing investigation on cellular integrity focusing on cell-cell contact sites. Moreover, several secretory proteins, e.g. interleukines, mucins and other classes of exclusively or secondary secretory proteins have been identified, which additionally emphasizes the active functionality of NHC-IOBA cells shifting them in direction of goblet cells since these cells have been defined for their high mucin secretion activity to the tear film (Dartt, 2002, Dartt *et al.*, 2000, de la Fuente *et al.*, 2010, Watanabe *et al.*, 1995). Commensurate to this Inatomi, Gipson and colleagues recorded expression of mucin species, especially mucin 5 in the human conjunctival epithelia, (Gipson *et al.*, 2003, Inatomi *et al.*, 1996), also detected in this work. Mucin 5 was defined as a goblet cell characteristic marker (Gipson and Inatomi, 1997). In fact, altered expression of mucins by goblet cells have been associated to dry eye disease (Danjo, 1998) and mucin and interleukine secretion have been documented for IOBA-NHC cells to be altered under stress condition (Li *et al.*, 2009, Tau, 2012). Moreover, mucin 5AC, expressed in human conjunctiva (McKenzie *et al.*, 2000) and identified in this work, was found as a reduced protein in Sjögren's dry eye (Gipson *et al.*, 2004) documented together with mucin 16 to be related to atopic keratokonjunctivitis (Dogru *et al.*, 2008) and can therefore be studied in future projects by use of the developed LC ESI monitoring NHC-IOBA cells in different experimental setups. However it could not be identified with high frequency and purification attempts might be needed. Actually, most of the mucin studies focused on RNA or used immune histological approaches for mucin determination, whereby mucins maybe hampered by low ionization frequencies in MS workflows due to oligosaccharide side chains (Thomsson *et al.*, 1999). Besides, secretion of peptides and proteins, e.g. antimicrobial peptides (McDermott, 2009), glycoconjugate secretions encircling non-mucin glycoproteins (Dartt *et al.*, 1996) or mucin associated peptides like TFF-peptides (trefoil factors) (Langer *et al.*, 1999) have been reported for the human conjunctiva. In accordance, more secretory proteins could be detected, providing a detailed impression on ocular surface epithelia activity in this work. One of these secretory proteins is macrophage migration inhibitory factor, a cytokine, which could be detected with

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high score by the LC ESI method and was reported to be upregulated in conjunctival tissues in case of ocular cicatricial pemphigoid (OCP) (Razzaque *et al.*, 2004). Another identified secretory protein was serpin B5, an antimicrobial serine proteinase inhibitor, identified in ocular surface epithelia (Silverman *et al.*, 2001). Actually, serpin B5 could be detected with high score in tear fluid by the developed LC ESI method, which proves the origin of tear serpin B5 from conjunctival cells secreting the protein to the tear film. Moreover, the communicative activity through cytokines could be supported by the identification of a chemokine receptor, C-C chemokine receptor type 9. In accordance Gulati and coworkers proved the expression of chemokine receptor CCR5 on the surface of human conjunctival cells associated to dry eye (Gulati *et al.*, 2006). Interestingly, a GABAergic and also a dopaminergic neurotransmitter receptor could be revealed, which supports the finding of nerve innervation and nerve secretion control of goblet cells of the human conjunctiva (Diebold *et al.*, 2001). Beside secretory proteins, challenging recovery of membrane proteins has been realized. Thereby, vinculin, a focal adhesion marker (Sosne *et al.*, 2002), proposed to be involved in Sjögren's pathology (Hjelmervik *et al.*, 2009) could be identified by the developed LC ESI method. Numerous proteins, e.g. the nucleus protein, Ras GTPase-activating-like protein IQGAP1, which most likely plays a key role in corneal development associated to Ras (Burgess *et al.*, 2010) have not been identified for conjunctival cells before. In addition, the identification of oxidative stress proteins in conjunctival cells could clarify the origin of these proteins in tear film, e.g. superoxide dismutase [Mn], [Cu-Zn]; thioredoxin, thioredoxin reductase 1, thioredoxin domain containing protein 6, 17; glutaredoxin 3; glutathione synthetase, glutathione S-transferase P/ Ω -1; lactoglutathion lyase; microsomal glutathione S-transferase 1 and peroxiredoxin 2, 4, 5 and 6. Especially, peroxiredoxin 2 has been associated to ocular surface disorders with overexpression in pterygium conjunctiva tissue (Bautista-de Lucio *et al.*, 2013). Moreover, peroxiredoxin 1, 5 and glutathione S-transferase have been found to be overexpressed in tears of conjunctivochalasis (Acera *et al.*, 2011), which supports the strong pathological relationship between conjunctival cells and tear film. In this work, a deep insight view of the tear proteome could be achieved by MALDI and ESI MS with high literature congruency, while numerous proteins could be catalogued for tear fluid for the first time. For example, centaurin- γ 1 as a autism related brain protein (Wassink *et al.*, 2005), furthermore detected in cephalopod and vertebrate optic lobe neurons of camera eyes (Yoshida and Ogura, 2011). Also, cytosolic 5'-nucleotidase III-like protein, most likely derived from the lens (Cappiello *et al.*, 1992) and transmembrane channel-like protein 8, probably originated from serum (Zhang *et al.*, 2012) and related to Epidermodysplasia verruciformis (Kurima *et al.*, 2003), were documented for the first time in tears. Also forkhead-associated domain-containing protein 1, displaying a neurodegenerative relevant sequence motif (Guzik and Goldstein, 2004), could

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be detected in all experiments and was not MS documented for tear fluid before. Interestingly, certain retina proteins, e.g. retinal dehydrogenase 1 or selenium-binding protein 1 could be revealed from tear fluid. In confidence, retinol dehydrogenase has been identified in tear fluid of patients with diabetic retinopathy (Csoz *et al.*, 2012) and traffic between retina and tear film in the course of degenerative processes might be thinkable since intraocular lesions can effect the ocular surface (Lee and Hirst, 1995). Also, identified proteins like myozenin 2, a hypertrophic cardiomyopathy biomarker (Osio *et al.*, 2007) associated to skeletal muscles (Takada *et al.*, 2001) could be suggested to reach the tear film due to internal processes. In contrast, proteins like cadherin, desmoplakin, plektin, desmoglein are clearly derived from cell-cell interactions sites (Buxton and Magee, 1992, Gallicano *et al.*, 1998, Koch *et al.*, 1990, Schmelz and Franke, 1993, Seifert *et al.*, 1992, Wiche *et al.*, 1983). Also, calreticulin identified in tear fluid is associated to cell-cell contact and had been demonstrated to play a key role in integrin control (Coppolino *et al.*, 1997). Without a doubt, these proteins are useful to focus on in studies investigating on epithelia barrier functionality. Most of the tear proteins were found to be intracellular proteins with predominance in cytoplasmic organelle-bound proteins inferred from both workflows. This finding and the predominance in nucleus proteins was also reported by de Souza and colleagues using a comparable LC ESI Orbitrap workflow (de Souza *et al.*, 2006). Since a comparable protein distribution could be demonstrated for conjunctival cells, tear fluid provides a reflection of proteomic hierarchy in epithelia. Typical nucleus proteins, e.g. histones have been described for healthy human tear fluid by use of LC ESI (Zhou, 2006) and could confidently be detected in tears in this work. Numerous lipid-associated proteins, e.g. lipocalins, phospholipid transfer protein, apolipoprotein AI or transcobalamin-I could be detected in tears by use of the developed methods. Transcobalamins are lipid and vitamin B12 carriers (Carmel, 1972, Rosenfeld *et al.*, 2003) described for human tears (Csoz *et al.*, 2012). Also, lipocalin a prominent tear protein was intensively studied for its lipid binding features (Delaire *et al.*, 1992, Millar *et al.*, 2009, Redl, 2000). Interestingly, beside its affinity to lipids, it was documented to bind thioredoxin (Redl *et al.*, 1999), which was also detected in tear fluid in this study. Besides, thioredoxin reductase 3, peroxiredoxin (thioredoxin peroxidase 2) could be identified in tears. Most likely these proteins build a functional complex in tears counterstriking oxidative stress (Lechner *et al.*, 2001). Confidently, thioredoxin and associated proteins were detected in conjunctival cells concluding these proteins to be directly or indirectly released by the cells to the tear film. This is supported by a study of Sotozono and coworkers, which could prove thioredoxin in human tear fluid as well as in conjunctival and corneal epithelia proposing a protective role under oxidative stress condition in case of inflammation (Sotozono *et al.*, 2005). Probably, under oxidative stress conditions, the rate of lytic ocular surface epithelial cells increases with a

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proportional release rate of thioredoxin and related proteins to the tear film. Since peroxiredoxin has been reported to display elevated levels in diseased conjunctival tissue in pterygium and was proposed to protect cells against oxidative stress (Bautista-de Lucio *et al.*, 2013), it could be released to the tear film after lysis of overexpressing cells, thereby, restoring the oxidative status of the extracellular milieu. It is interesting, that also a complement associated protein; complement decay-accelerating factor, could be detected in the cells. Complement C3 was also detected in tears by use of the developed MALDI and ESI methods. In confidence, Ballow *et al.* had found evidence for complement factor production in conjunctival tissues (Ballow *et al.*, 1985), which underlines the key role of the conjunctiva for inflammatory processes manifesting in the tear film. In summary, by use of the developed BU workflows a sensitive image of the ocular surface proteome could be obtained showing high confidence with recent studies on the tear proteome. Thereby, numerous new proteins could be mapped for the tear proteome and moreover the origin of several proteins could be allocated to the conjunctiva. The portion of overlapping proteins regarding tear and conjunctival cells most likely suggests a more detailed biological role of the ocular surface epithelium traversing barrier function.

5.4 Effects of Taflotan[®] sine on the ocular surface proteome

At first instance, interestingly, most responding candidate proteins showed longitudinal diminishment. Functional analysis classified the majority of these candidates as cellular proteins deducting evidence for beneficial effects of the therapeutic switch to preservative-free Taflotan[®] sine appliance. Ocular surface regenerative processes are most likely reflected by the observed longitudinal cellular protein decline in in the POAG patients' tear film, which can be hypothesized to indicate a decreased leakage rate of epithelium cells. It is proposed, that most of cellular proteins in the tear film are originated from conjunctiva vessel leakage (Fukuda *et al.*, 1996, Janssen and van Bijsterveld, 1983, Tiffany, 2008, Tiffany *et al.*, 1989) and corneal cell leakage events due to cellular breakdown (Choy *et al.*, 2004, de Souza *et al.*, 2006), which can also be associated to atrophy of lacrimal glands (Stern *et al.*, 2004). Accordingly, increased epithelial breakdown in the cornea was reported to be correlated to dry eye (Holly and Lemp, 1977); (Ogawa *et al.*, 2001, Vroman *et al.*, 2005);(Brignole *et al.*, 2000). Also, Chang and colleagues clearly demonstrated that BAC included in latanoprost causes corneal cell epithelial membrane leakage (Chang *et al.*, 2000). As a matter of fact, BAC was able to extract intracellular lactat dehydrogenase from rabbit cornea due to its detergent qualities (Grant and Acosta, 1996), which supports the observation of a high number of enzymes and other intracellular proteins with high pre-therapeutic shift abundance in POAG tears in this work. Also, there is clear evidence for cell lytic processes encircling necrotic and apoptotic reactions of ocular surface epithelia in dose-dependant contact with

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BAC (Burstein, 1980, De Saint Jean *et al.*, 2000, Tonjum, 1975, Tripathi and Tripathi, 1989). Moreover, BAC was found to alter myosin modification impacting corneal actin cytoskeleton integrity (Guo *et al.*, 2007). Increased disruption of the perjunctional actomyosin ring corresponding to increased cell damage of endothelial corneal cells was reported after treatment of rabbit eyes with BAC eye drops (Chen *et al.*, 2011a). Accordingly, increased levels of myosin fragments most likely reflect tight junction and corneal endothelial barrier defects. Xu and coworkers proved SDS comparable detergent qualities of BAC able to disrupt corneal tight junctions (Nakamura *et al.*, 2010) and increased cornea permeability (Majumdar *et al.*, 2008). Actually, after the medication switch a linear decrease in myosin 10 was detected, most likely indicating corneal regeneration. Cadherin 5, another corneal integrity marker was detected to be linearly decreased after the switch. Cadherins are corneal junction proteins impaired by inflammation (Kimura *et al.*, 2009). This is why a decline in soluble cadherins is indicative for ocular surface restoring. Furthermore, in this study, the association to cellular apoptotic processes of several of the candidates fits well with the observation of an increase in cellular defect indicators. In accordance, Barki & Tahir reported high toxicity of BAC while observing apoptotic processes in corneal epithelia of guinea pigs treated with BAC eye drops (Barki and Tahir, 2007). De facto, numerous members of the annexin protein family are documented to be strongly correlated to cellular damage, apoptosis and inflammation (Aubry *et al.*, 1999, Gerke *et al.*, 2005, Gerke and Moss, 2002, Perretti and D'Acquisto, 2009, Reutelingsperger and van Heerde, 1997). Annexin A11 detected in tears in this study and significantly exhibiting a linear level decrease also could reflect ocular surface regeneration since overexpression of tear annexin had been associated to dry eye (Soria *et al.*, 2013). Interestingly, annexin 1 was found to be upregulated in tears resulting from parasympathetic defected lacrimal glands (Nguyen *et al.*, 2004). Besides, level increase of annexin 5 indicated apoptosis (Buehrlen *et al.*, 2007, Koopman *et al.*, 1994, Plasier *et al.*, 1999, van Engeland *et al.*, 1998, Vermes *et al.*, 1995) in cultured conjunctival cells exposed to BAC containing latanoprost, travoprost formulations and BAC alone in comparison to preservative-free travoprostZ 0.004% (Baudouin *et al.*, 2007). Since there is a strong relation between annexin family members and their role for physiological stress condition (Gerke and Moss, 2002), time-related decline of annexin 11 is most likely linked to a drop in apoptotic events and reflects ocular surface recovery. Beside its direct relation to annexin 1 as a dry eye marker (Soria *et al.*, 2013), annexin 11 is associated to calcyclin (S100A6) as a further dry eye marker, because of its calcyclin binding motif (Sudo and Hidaka, 1998, Tokumitsu *et al.*, 1993). Serotransferrin detected in tears with high abundance (Li *et al.*, 2005, Srinivasan *et al.*, 2012, Zhou *et al.*, 2009c) was documented to increase during inflammation (Hallquist and Klasing, 1994) and was found to be significantly linear reduced in this study. Interestingly, a conjunctival transferrin receptor had

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been identified (Baudouin *et al.*, 1992) and could also be detected in NHC-IOBA cells in this work. Possibly, increased serotransferrin as an acute phase protein (Hallquist and Klasing, 1994) could be sensed by conjunctival cells, leading to proteomic changes. Also, an increased iron supply of ocular surface epithelia under stress condition mediated by elevated tear serotransferrin levels can be assumed (Ismail and Brock, 1993, Kawakami *et al.*, 1990, Oria *et al.*, 1988). Alternatively, elevated serotransferrin levels in tear fluid indicate a counterstrategy to bind iron released from lytic cells (Jurado, 1997) due to preservative-induced damage. Nevertheless, other dry eye associated markers, which showed a clear level shift, are of speculative expressiveness to evaluate the ocular surface condition. For example, galectin 7, which had been documented for numerous ocular tissues including the cornea (Schlotzer-Schrehardt *et al.*, 2012), was found to be clearly reduced in the course of time in response to Taflotan[®] sine usage. In fact, gene expression of galectin 7 has been documented to be elevated in injured corneas and has been supposed to be involved in corneal wound healing (Cao *et al.*, 2002a). This implication fits well with the observed diminishing of the galectin 7 level, most likely reflecting a regenerating ocular surface. Contrary, galectin 7 was found to be decreased in tears of dry eye patients (Soria *et al.*, 2013). However, since galectins, especially galectin 7, are related to apoptosis (Kuwabara *et al.*, 2001, Kuwabara *et al.*, 2002) and wound healing (Cao *et al.*, 2002b), also documented for the cornea (Patron *et al.*, 2004), a time-related decrease of the galectin 7 level most likely corresponds to regenerative processes. Nevertheless it is thinkable that a deficiency of galectin 7 corresponds to the development of dry eye pathology under certain conditions and that preservatives induce a special dry eye phenotype due to multifactorial character of the disease. However, dry eye involvement of galectins (Argueso and Panjwani, 2011, Yabuta *et al.*, 2014), their abundance in epithelial wounds (Pepe *et al.*, 2014) and postulated role in apoptotic/necrotic cell clearance (Beer *et al.*, 2008, Mostowy, 2013, Sarter, 2010) makes the observed galectin 7 highly indicative for ocular surface recovery. The diminishment of leakage of cellular and tight junction proteins in the study course supports this assumption. Most likely, in first instance, ocular surface epithelia are affected by BAC followed by lacrimal and meibian alterations as a counter reaction to ocular surface damage which is in confidence of the proposed cornea and lacrimal feedback model of Mathers (Mathers, 2000). De facto, MS results were supported by MA findings since longitudinal diminished MS marker candidates encircling e.g. pyruvatekinase isozymes M1/M2, kinectin, annexin A11, serotransferrin or cadherin 5 displayed a significant convergence to the healthy stage in the course of Taflotan[®] sine appliance. Actually, the release of intracellular proteins has been documented to induce inflammation on target cells (Chase *et al.*, 2007, Harris and Andersson, 2004, Scaffidi *et al.*, 2002) and ocular surface inflammation, which has been indicated by elevated cytokine levels (Boehm *et al.*, 2011, de Paiva *et al.*, 2013, De Paiva *et*

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al., 2007, Luo *et al.*, 2004, Pflugfelder *et al.*, 2013). Interestingly, an increase in interleukine mRNA in ocular surface epithelia has been shown in response to topical use of prostaglandine analogues (Lopilly Park *et al.*, 2012), which fits well with MA results of this work. Accordingly, distinct approximation of the majority of analyzed cytokines to the healthy stage in the course of the study indicated a clear reduction of ocular surface inflammation in response to the therapeutic switch. In confidence, Baudouin *et al.* reported increased expression of inflammatory conjunctival cell membrane markers, e.g. cytokine receptors in glaucoma patients in the course of BAC related topical medication (Baudouin *et al.*, 1994). As already discussed, numerous successfully validated tear proteins are not typical dry eye markers, underlining the suggestion of a superordinate dry eye phenotype according to “medicative induced dry eye” (Fraunfelder *et al.*, 2012). Most likely, the observed proteomic changes indicate reshuffling conditions of the tear film reflected by a diminishment of epithelia cell leaking events and a correlating drop of inflammation represented by declined cytokine levels. Interestingly, Wong and colleagues could detect clear proteomic alterations comparing tear fluid of medicated glaucoma patients with healthy and dry eye patients, whereby the 14-3-3 ζ/δ protein, absent in dry eye tears, was found to be increased in medicated groups compared to healthy subjects (Wong *et al.*, 2011). This finding supports a medicative sicca phenotype. Confidently, 14-3-3 isoforms were also identified in POAG tears in this work, but displaying no response towards Taflotan[®] sine. However, the hypothesis is additionally supported by proteomic illumination of clinical dry eye phenotypes (Boehm *et al.*, 2013). In contrast, prominent enzymes showing an altered status towards preservative-free tafluprost utilization like pyruvate kinase isozymes M1/M2 or mitochondrial pantothenate kinase 2 have not been reported for dry eye. In this work, they certainly refer to cell leakage and have interestingly been documented for tear and lacrimal gland fluid (Vanhaeri.Nj and Glasius, 1974a, b). In general, lysosomal enzymes are derived from the lacrimal gland and levels of cytoplasmic proteins, especially enzymes should be low in tear film “under conditions where the epithelia cells of the conjunctiva remain intact”, whereby “the conjunctiva may act as a second source for lysosomal enzymes after mild trauma” (Van Haeringen, 1981). Recently, Nakamori and Koyama used tear film enzyme activity of lactat dehydrogenase to estimate ocular epithelial damage since the enzyme was proposed to reach the tear film through epithelial cell leakage (Koyama *et al.*, 1996, Nakamori *et al.*, 1993b), which was reported in accordance to early studies proposing cellular leakage of intracellular enzyme as direct consequence of cytotoxicity (Nakamura *et al.*, 1985). In confidence, enzymes like pyruvate kinase are known to be highly expressed in cornea keratocytes (Jester *et al.*, 2005) and have been reported to be released by ocular surface epithelia (Sariri and Ghafoori, 2008) to the tear film (Baeyens and Gurny, 1997). Actually, it had been shown that leakage rates correlate with expression levels of intracellular proteins

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(Barnes and Pielak, 2011), which is in confidence with the high abundant detection of pyruvate kinase isozymes M1/M2 in tear fluid in this work and its abundant detection in NHC-IOBA cells supporting its epithelial origin. In confidence, Menke and Jokusch used pyruvate kinase release to monitor effects of hyperosmotic shock in myotubes (Menke and Jokusch, 1995), supposing pyruvate kinase as a predominant releasing protein through membrane damage. Nevertheless, it remains unknown if level alterations in these enzymes have a superordinate functions for the ocular surface in case of stress condition. Interestingly pyruvate kinase had been reported to increase post-cautery and declines after 6-7 days during rat cornea angiogenesis (Thompson *et al.*, 2003), proposing a highlighted metabolic function, which fits well with the proposed key role of pyruvate kinase M2 in altered tumor metabolism (Mazurek, 2012). Furthermore, the supposed role of a specific pyruvate isozymes M1/M2 derived peptide as an autoantigen in human polymyositis (Kawachi *et al.*, 2001) may hint to inflammatory functions. Despite the thinkable functions of observed leakage enzymes, a disruption of the cellular barrier as well as cellular lysis is highly likely due to Xalatan® application >6 month considering the destructive BAC potential in extended therapy (Cha *et al.*, 2004), including tear film stability decline (Wilson *et al.*, 1975), tear production (Labbe *et al.*, 2006) and multifactorial ocular surface complication (Xiong *et al.*, 2008). Since BAC is also known to interact with protein membrane receptors, which influence signal transduction (Patarca and Fletcher, 1995), ocular surface processes resulting from long time application are likely to display high complexity. In summary, the study results show that switching to preservative-free formulation leads to ocular surface recovery, also supported by clinical parameters, TBUT and BST. The therapeutic switch provides a patient perspective for life quality enhancement.

5.5 Taurine effects on the tear and cell proteome

Regarding taurine eye drop application several tear proteins could be revealed responding with a distinct level decline. Interestingly, these effects were predominantly detected in the contact lens wearer group over the 5 week study period. However, in contrast, using a linear trend model no obvious taurine response reactions in the sicca group could be observed. An explanation could be pathological differences in contact lens ocular surface processes and related effectiveness of taurine as well as the selected application duration. For example, Kokke and colleagues studied effects of Ω -6 fatty acid utilization in a 6 month study and could document distinct improvements in the ocular surface health status after 3 month (Kokke *et al.*, 2008). However, since the substance has been applied orally, time dependent effects differ most likely from topical appliance in terms of bioavailability. Additionally, differences in the course of the disease or pathological dispositions may be reflected by the fact, that by use of linear regression analysis, decreased proteins could exclusively been revealed in the

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contact lens group. In contrast, the non-linear *Kruskal-Wallis* analysis detected candidate proteins displaying distinct level decreases in both taurine treatment groups (S_Taurine, CL_Taurine). This finding suggests that a strict linear regression model is not applicable to detect all responses in a 5 week study period considering tear protein variation according to time (Ng *et al.*, 2000a, Ng *et al.*, 2000b, Ng *et al.*, 2001) and hormone system influence (Sullivan *et al.*, 1984, Sullivan and Hann, 1989). For example, glycoproteins show a high degree of diurnal fluctuation (Sack *et al.*, 1997). Regarding the functional context, candidate proteins detected by use of both models, linear and non-linear, were predominantly associated to inflammation. Actually, they could be verified to be upregulated in contact lens and/or dry eye syndrome tears compared to a healthy non contact lens wearer reference. This fits well with the finding of elevated levels of inflammatory proteins in dry eye (Grus *et al.*, 2005b, Lam *et al.*, 2008) and contact lens tears (Grus *et al.*, 2005a). Secretoglobin 1D1, a small anti-inflammatory protein (Mukherjee *et al.*, 2008, Mukherjee *et al.*, 2007), also known as lipophilin A, which was previously found to be a decreased tear biomarker for contact lens wear (Kramann *et al.*, 2011) and dry eye (Nichols and Green-Church, 2009, Versura *et al.*, 2010) was contrary found to be elevated in tears of contact lens and sicca tears in the present study referring to a healthy non-contact lens wearer reference. The protein was reported to build a complex with mammaglobin B (Stoeckelhuber *et al.*, 2006). Interestingly, both proteins; secretoglobin 1D1 and mammaglobin B, were found to be elevated in tears of aqueous-deficient dry eye patients and patients with a combinatory deficit in the lipid and aqueous tear phase (Boehm *et al.*, 2013). In this work, secretoglobin 1D1 was demonstrated to be shifted in direction of the healthy stage in response to taurine utilization. α -2-macroglobin, another taurine responding candidate, was reported to be expressed in the cornea (Laurie, 2008, Twining *et al.*, 1994), but probably could also be originated from micro vascular bleeding (Lembach *et al.*, 2001) since it is a dominant blood protein (McMillan, 1976). It is interesting, that this high molecular weight proteinase can bind a wide range of targeted molecules, e.g. S100A9 (Eue *et al.*, 2002) or β -2-microglobulin (Gouin-Charnet *et al.*, 2000). Moreover, it is suggested to be involved in dry eye referring to a misbalance between proteinases and inhibitors in the disease pathology (Laurie, 2008), which was also proposed to play a role in contact lens wearing (Cejkova *et al.*, 1992). Also, a decline in complement C3 in response to taurine application was detected. Complement C3 activates the complement cascade and was reported to be the major player in closed eye tear inflammatory processes, that can be compared to inflammation associated to contact lens wearing (Sack *et al.*, 1992). Since hemolytic activity of complement C3 was proofed in tears (Mondino and Zaidman, 1983, Veerhuis and Kijlstra, 1982, Willcox *et al.*, 1997), a potential taurine depending decline could be associated with the observed level decrease of haptoglobin related protein and hemopexin, which are both anti-oxidant protective plasma

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proteins occurring in the course of inflammation as acute phase proteins (Ceciliani *et al.*, 2002, Kuhajda *et al.*, 1989). Haptoglobin and haptoglobin related protein can bind hemoglobin during red blood cell lysis (Chen *et al.*, 1998, Dobryzycka, 1997, Karring *et al.*, 2006, Nielsen *et al.*, 2006), whereby haptoglobin has been reported to be increased in tears of climatic droplet keratopathy patients (Zhou *et al.*, 2009d). Hemopexin, which binds free heme during inflammatory hemolysis was first detected by Pong *et al.* 2010 in tear fluid and moreover, it was demonstrated to be elevated in tears of patients suffering from vernal keratokunjunctivitis increasing with disease stages (Pong *et al.*, 2010, Pong *et al.*, 2011). The protein was also correlated to allergic processes of the ocular surface (Enriquez-de-Salamanca *et al.*, 2012). Hemopexin and α -2-macroglobin are biologically close related and have been reported to display elevated levels in cerebrospinal fluid and serum during inflammation (Saso *et al.*, 1999). Hemopexin was also found to be upregulated in inflammatory nerve injury in the eye (Camborieu *et al.*, 2000). Ig γ , also detected to respond to taurine application, has been proven to be elevated in tears of keratoconus patients (Chandler *et al.*, 1974, Pannebaker *et al.*, 2010) and has been detected in contact lens deposits (Green-Church and Nichols, 2008), probably reflecting its activity in contact lens tear inflammation. Regarding the classes of candidate proteins, it can be suggested, that the taurine impact is predominately focused on proteins occurring in the course of inflammation and hemolysis, whereby several of these proteins are known to play key roles in dry eye. Actually the response of inflammatory proteins like haptoglobin, α -2-macroglobin, hemopexin together with complement C3 as a prominent type 1 acute phase protein (Baumann and Schendel, 1991, Immenschuh *et al.*, 1995), as well as the response of immunoglobulines, clearly indicates the impact of taurine on the immune response of the ocular surface. It was shown that most of these acute phase proteins are coordinately regulated (Baumann and Mullereberhard, 1987), which fits well with the observation that the proteins responded simultaneously. The differential regulation of tear immunoglobuline components like Ig κ chain C region, Ig α -2 chain C region Ig J chain, Ig α -1 chain C region have been furthermore associated to inflammatory processes in terms of POAG (Pieragostino *et al.*, 2012). Ig κ C level alteration was also documented for keratoconus associated tear fluid (Lema *et al.*, 2010). However, several limitations of the study design have to be solved in future projects. Since tear proteins show a high degree of naturally variability, in future studies the monitoring time has to be extended to detect proteins that correspond later in the time course of taurine application. Additionally, taurine has to be tested in combination with other cleaning solution ingredients expected to have beneficial effects on contact lens-induced dry eye. In fact, only little is known about underlying mechanism how taurine may influence inflammatory response reflected by the observed proteomic changes. Taurine concentration of the supplemented eye drops was approximately 20fold higher than the natural taurine

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concentration in tears [195.5 μM] (Nakatsukasa *et al.*, 2011) and therefore it can be suggested, that this taurine overrun lead to a decline in the taurine uptake rate of ocular surface epithelia. Actually, Na^+ dependent uptake as well as a high affinity transporter system have been documented for taurine (Karring *et al.*, 2006, Oh *et al.*, 2006, Shioda *et al.*, 2002, Smith *et al.*, 1992). In confidence, by use of the developed LC ESI method numerous transmembrane amino acid transporters could be detected in conjunctival epithelia cells, e.g. large neutral amino acids transporter or sodium coupled neutral amino acid transporter 2, which could be involved in taurine uptake. Interestingly, probable cation-transporting ATPase 13A5 could be identified in the cells, which fits well with the finding of Garrett and colleagues, who proposed a carrier-mediated organic cation transport process mediating the absorption of organic amines, carnitine and drugs supported by the detection of carnitine/organic cation transporters OCTN1, 2 (Garrett *et al.*, 2008). Nevertheless, the question remains, how taurine may induce the observed effects regarding decreased inflammation in contact lens wearers and which cellular targets are affected. In accordance taurine was tested on NHC-IOBA conjunctival cells and a dose-dependent taurine effect could be determined. Whereas elevated concentrations >20 mM lead to stress reactions indicated by reduced viability, increased ROS production and upregulation of stress associated proteins, e.g. elevation of thioredoxin 1, peroxiredoxin 2 and glutaredoxin 1 levels, lower concentrations showed beneficial effects on cells, which is somehow contrary to the relative high effective level of taurine eye drops in the tear study. However, diffusion effects of the tear film are highly likely in comparison to cell culture experiments. Interestingly, taurine was highly effective to reduce ROS production in peroxide stressed cells, however, it was more effective to increase the viability in unstressed than in stressed cells without discrimination between peroxide or staurosporine stress condition. Cellular levels of numerous stress marker proteins (glutathion S-transferases, peroxiredoxins, thioredoxin, superoxide dismutase and cytochrome C) were clearly found to be decreased. Most of these proteins play a role in counterstriking oxidative stress. Peroxiredoxins had been intensively studied to be increased under oxidative stress condition in various tissues (Ishii and Yanagawa, 2007, Shibata *et al.*, 2003, Sideri *et al.*, 2010) and peroxiredoxin 4 had been demonstrated to successfully remove peroxide (Tavender and Bulleid, 2010). In addition, thioredoxin and glutaredoxin are two enzyme key players in the protection system against oxidative stress in case of inflammation and disease (Arodin *et al.*, 2013, Zhou *et al.*, 2010) and thioredoxin overexpression has been reported to extend life span in mice (Mitsui *et al.*, 2002). In contrast, Chen *et al.* reported glutathione and thioredoxin to be downregulated with hyperosmolarity in NHC-IOBA cells, but also reported upregulation of certain enzymes like pyrophosphorylase (Chen *et al.*, 2012). Nevertheless, thioredoxin expression has been reported to be increased by oxidative and hypoxic stresses documented for cancers (Karlenius *et al.*, 2012, Karlenius, 2010) and

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beneficial effects of overexpression in retinal pigment epithelial cells in response to oxidative stress could be documented (Sugano, 2013). Most likely, antioxidative effects of taurine also influence inflammatory and hemolytic processes caused by e.g. contact lens wear. Interestingly, also increased histone expression has been shown to be associated to life span extension in yeast (Feser *et al.*, 2010). It is thinkable, that a defined surplus in taurine availability relieves this anti-stress system, which is reflected by downregulation of these protein species and by ROS decline and increased cellular viability. Since thioredoxin was found to regenerate oxidative stress in endothelial cells (Fernando *et al.*, 1992), it can be suggested, that taurine acts like a substitute since it was documented to protect proteins from uncontrolled modification, e.g. nitration (Askwith *et al.*, 2012), glycation (Devamanocharan *et al.*, 1997), carbonylation (Son *et al.*, 2007) and denaturation (Arakawa and Timasheff, 1985) under stress condition. Interestingly, Kayuya and co-researchers detected a decline in natural cataract lens taurine levels correlating with glutathione decline (Kasuya *et al.*, 1992), which supports taurine-anti-oxidative enzyme interactions. Regarding the detected histone decline, it had been shown, that downregulation of histone H4 is associated to differentiation processes in single cells (Gerbaulet *et al.*, 1992) and histones have been found to be involved in inflammation (Gilthorpe, 2013, Nicodeme *et al.*, 2010, Xu *et al.*, 2011). Allam *et al.* intensively studied histones released from dying renal cells and found histones to increase TLR mediated inflammation and kill endothelial and epithelial cells (Allam *et al.*, 2012), which is interesting in case of conjunctival cells and ocular surface inflammation. Moreover, the effectivity of taurine regarding unstressed to peroxide stressed cells may reflect the potential of taurine to improve cell viability in natural to mild stress environment and to fail rescuing cells under harsh stress conditions. This is fitting well with the finding of Nonaka and colleagues, who reported taurine to prevent superoxide dismutase expression and secretion in case of homocysteine-induced stress (Nonaka *et al.*, 2001). It is thinkable, that under normal to mild stress condition taurine may exonerate the anti-oxidative enzyme system, but fails to prevent cells under strong stress. Furthermore, stress quality could play a role, since taurine showed good peroxide-counter action but low performance in case of staurosporine induced stress. Taken together, this might contribute to explain the differences in taurine tear proteomic response pattern observed between primary dry eye subjects and contact lens wearers, which might reflect different qualities of stress regarding molecular mechanism. It can be suggested from these findings, that taurine performs differentially depending on dry eye phenotype, stress degree and quality. However, taurine was supported in this work to influence numerous metabolic proteins in conjunctival cells, which emphasizes its modulative potential to improve cell survival under mild condition. De facto, the dramatically downregulation of calmodulin and the associated calmodulin/calcium apparatus in response to 0.05 mM taurine in unstressed cells is highly indicative for a cellular

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taurine response and illustrates the direct impact of taurine on epithelia cells of the ocular surface. Because calmodulin is a key mediator of taurine transport, a modified cellular taurine traffic (Falktoft and Lambert, 2004, Law, 1995) and a direct osmolytic effect on conjunctival cells can be assumed. Actually, for corneal cells a 4.5fold intracellular taurine concentration increase within 48 h had been demonstrated to elevate cell viability (Shioda *et al.*, 2002). Confidently, level decrease of galectin-1 and galectin 3-binding protein is interesting since elevated expression of galectin-1 was found to be associated to apoptosis in endothelial cells (Qiu *et al.*, 2008) and galectin 7 decline could also be documented as a potential ocular surface recovery response in the present Taflotan[®] sine study. Nevertheless, the discrepancy of beneficial effects obtained by topical application of high taurine concentrations in tear fluid and the dose-dependent effects observed in cell culture most likely reflect experimental design differences between in vitro and in vivo studies. The ocular surface as a highly complex functional unit can react to changes in the biochemical environment with a whole organ response, whereas cellular responses have limitations. However, in both study systems, tear film and conjunctival cells, as representing components of the ocular surface, taurine performed regeneratively. That is why taurine is highly interesting for future studies regarding therapeutic strategies of ocular surface pathologies. Thereby, more detailed studies have to be launched to illuminate the interaction between taurine and different sources of stress. Also, more investigations have to be tasked addressing the direct influence of taurine on tear and ocular surface related proteins, e.g. in terms of post-translational modifications (PTMs) and protein turnover.

5.6 Deviated strategies facing dry eye and ocular surface complications

A taurine counteraction on inflammation could be indicated by a decline in inflammatory tear proteins predominantly observed in contact lens tears. However, it can be assumed, that other dry eye forms, also, can be positively affected by taurine appliance, which might be revealed in prolonged studies. Up to date, taurine, beside other osmolytes, could be proposed as an effective agency against hyperosmolar stress and preventing of inflammation and damage of the ocular surface, especially in dry eye therapy (Baudouin *et al.*, 2013), which is in confidence with the beneficial effects of taurine on conjunctival cells in this work. Since taurine was demonstrated to benefit peroxide stressed and untreated NHC-IOBA cells, in this work a loss of mucin-producing goblet cells as well as associated inflammatory cascades in dry eye disease are likely to be prevented by the amino acid. Moreover, since neuronal control of goblet cell mucin secretion had been proposed (Diebold *et al.*, 2001) and taurine was demonstrated for its activity on ionotropic and metabotropic receptors of retina nerves (Rowan *et al.*, 2013), taurine may directly influence mucin secretion and emending tear film properties, which has to be examined intensively in future studies. However, care

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has to be taken regarding concentration settings of taurine since a dose dependent cellular agency could be substantiated in the present work. In case of preservative induced ocular surface disorders as shown for BAC containing Xalatan[®] utilization in terms of glaucoma therapy, taurine could be proposed for its ocular surface damage counteracting potential. De facto, osmolytes like taurine have been intensively studied for their protein stabilization features (Anjum *et al.*, 2000, Arakawa and Timasheff, 1985, Haque *et al.*, 2005, Street *et al.*, 2006) and e.g. trimethylamine *n*-oxide, a naturally occurring osmolyte had been demonstrated to stabilize proteins from urea influence (Zou *et al.*, 2002b). Interestingly, some deep-sea species use taurine derivatives as trimethylamine *n*-oxide substitutes, which emphasizes their analog role regarding osmolytic activity (Yin *et al.*, 1999, Yin *et al.*, 2000). The potential of taurine and other osmolytes for protein stabilization and rescuing proteins from misfolding lead to the definition of osmolytes to represent “chemical chaperones” (Rajan *et al.*, 2011, Yancey, 2001). Accordingly, stabilizing and refolding of proteins in a detergent environment could be valued as a promising key feature of osmolytes, especially a feature of taurine. In ocular surface complications referring to glaucoma therapy, the use of preservative-free Taflotan[®] sine can be recommended since a clear ocular surface regeneration could be inferred from the proteomic tear film results. It is thinkable, that a combination of the preservative-free glaucoma therapy with the use of taurine as an formulation “additive” (Hamada *et al.*, 2009) could represent a promising therapeutic strategy for glaucoma medication induced dry eye disease. Moreover, proteins responding to taurine could be studied more intensively for their value as therapeutic targets. For example, thioredoxin, glutathione or peroxiredoxin are trigger points of oxidative stress, which can be addressed for the appliance to counteract inflammatory processes. Actually, research on the therapeutic value of extracellular thioredoxin as an anti-inflammation tool has already been launched (Mahmood *et al.*, 2013, Matsuo and Yodoi, 2013). Also, tear enzymes, especially the pyruvate kinase system has to be studied more intensively, since promising effects have been postulated for pyruvate kinase in terms of cancer therapy (Mazurek, 2012, Tamada *et al.*, 2012). Combinatory effects of potential therapeutic proteins, taurine and osmolytes may show a new direction in dry eye therapy.

6 Conclusion

BU clearly exceeded TD performance in this work and a resulting LC MS BU platform encircling MALDI/ESI techniques could be developed, which enabled a deep insight view to the ocular surface containing tear film and conjunctival cells. Moreover, by use of the developed methods the effects of taurine and Taflotan[®] sine on the ocular surface could be studied intensively, illuminating new molecular proteomic players indicative for dry eye and “medication induced dry eye” providing a deeper view to molecular mechanism. Finally, the proteomic results could give a perspective to develop promising therapeutic strategies regarding dry eye management and encourage future studies.

7 Summary

In the present work a top down (TD) MALDI approach and two bottom up (BU) MALDI/ESI mass spectrometric (MS) based methods have been developed integrating HPLC to study the ocular surface encircling tear film and conjunctival cells. A detailed overview of development steps, benefits and limitations was given in the work. Whereas the TD approach was found to be applicable to examine crude cellular specimen, BU approaches resulted in high sensitivity and accuracy regarding the analysis of processed conjunctival cell and also tear film proteins. By use of the LC MALDI BU method more than 200 capillary tear proteins could be detected, LC ESI resulted in more than 1000 capillary tear proteins, which was also achieved for Schirmer strip tears and conjunctival cells. Regarding quality and quantity ESI and MALDI methods showed distinct differences discussed to be useful depending on the proteomic target. Moreover, by use of the developed LC MALDI/ESI BU platform, which clearly performed advantageous over the TD approach, the influence of taurine and Taflotan[®] sine application on the ocular surface could be studied. Taurine could be demonstrated to lead to anti-inflammatory effects on the ocular surface reflected by dynamic changes in tear film proteins, predominantly documented in contact lens induced dry eye. Beneficial effects could also be supported in conjunctival cell studies, however showing a dose-dependent taurine effect. Regarding the topical application of preservative-free Taflotan[®] sine, by use of LC ESI BU analysis, an ocular surface recovery could be confirmed in primary open angle glaucoma (POAG) patients with dry eye reflected by dynamic tear film changes after a therapeutic switch from preservative containing Xalatan[®]. This could also be validated by microarray (MA) experiments. In taurin studies as well as in the Taflotan[®] sine study, characteristic ocular surface protein candidates could be revealed, which can be used to objectively assess the ocular surface health status. A use of taurine for dry eye therapy and a combinatory strategy of Taflotan[®] sine and taurine have been proposed for future therapeutic strategies in terms of medication induced dry eye in glaucoma.

8 Abbreviations

- 1D/2D=1 Dimensional/2 Dimensional
- ACN= Acetonitrile
- Ambi= Ammoniumbicarbonate
- BAC= Benzalkonium chloride
- BST= Baal Schirmer tear
- BU= Bottom Up
- CBB= Coomassie Brilliant Blue
- CHCA= α -cyano-4-hydroxy.cinnamic acid
- CID= Collision induced decay
- DDM= Dodecyl- β -D-maltoside
- DMEM= Dulbecco's modified eagle's medium
- DPBS= Dulbecco's phosphate buffered saline
- EDTA= Ethylendiamintetraacetate
- ESI= Electro spray ionization
- FA= Formic acid
- FCS= Fetal calf serum
- GO= Gene ontology
- HPLC= High pressure liquid chromatography
- ID= Inner diameter
- IOP= Intra ocular pressure
- LP= Laser power
- MA= Micro array
- MALDI= Matrix assisted laser desorption ionization
- MeOH= Methanol
- MES=2-(*N*-morpholino)ethanesulfonic acid
- MOPS= 3-(*N*-morpholino)propanesulfonic acid
- MS= Mass spectrometry
- MWC= Molecular weight cut off
- OSDI= Ocular surface disease index
- PKM= Pyruvat kinase isozymes M1/M2
- POAG= Primary open angle glaucoma
- PSD= Post source decay
- RP= Reversed phase
- SDS PAGE= Sodium dodecyl sulfate poly acrylamide gel electrophorese

Abbreviations

- SDS= Sodium dodecyl sulfate
- SELDI= Surface enhanced laser desorption ionization
- SPE= Solid phase extraction
- TBUT= Tear break up time
- TD= Top Down
- TFA= trifluor acetic acid
- TIC= Total ion current
- TLR= Toll like receptor
- TOF= Time of flight

9 Instruments/Technical equipment

- 428™ array laser scanner (Affymetrix Inc.)
- A 316 0.5µm online precolumn filter (Upchurch Scientific, Washington, USA)
- Acrodisc 13 mm x 0.45 µm GHP filter (PALL Life Sciences, Ann Arbor, USA)
- BioBasic® C18 analytical column (150 x 0.5mm ID) (Thermo Fisher Scientific, Rockford, USA)
- BioBasic® C18 precolumn (30 x 0.5mm ID) (Thermo Fisher Scientific, Rockford, USA)
- BioBasic® Phenyl analytical column (100 x 0.5mm ID) (Thermo Fisher Scientific, Rockford, USA)
- Cellstar® V-bottom 96 well plates (Greiner Bio-One)
- Concentrator 5301 instrument (Eppendorf, Hamburg, Germany).
- DCP-9042 CDN benchtop scanner (Brother International GMBH)
- Defined capillary tube (20µl) (Eksigent, Dublin, Ireland)
- Heraeus™ Biofuge Primo R centrifuge (Thermo Fisher Scientific, Rockford, USA)
- Jupiter® 4 µ Proteo analytical column (150 x 0.5mm ID) (Phenomenex, Torrence, USA)
- KS250 Basic Intellimixer (Kika Labortechnik, Cologne, Germany)
- Labsonic® M (Sartorius)
- Low Flow Metal Needle for API 2 Probes (Thermo Fisher Scientific, Rockford, USA)
- LTQ Orbitrap XL mass spectrometer (Thermo Fisher Scientific, Rockford, USA)
- MTP 396 Polished/Matt Steel MALDI Target (Bruker Daltonics, Bremen, Germany)
- Multiscan Ascent scanner (Thermo Fisher Scientific, Rockford, USA)
- Nikon Eclipse TS 100 (Nikon)
- Oncyte Avid nitrocellulose slides (Grace Bio-Labs Inc., Oregon, USA)
- Onyx® column couplers (Phenomenex, Torrence, USA)
- P-470/ M-472 graduated micro-split valves (Upchurch Scientific, Washington, USA)
- PAL HTS robot (CTC Analytics, Zwingen, Switzerland)
- PEEK™ tubings/Fingertight fittings/ferrules (Upchurch Scientific, Washington, USA)
- Prospere P-HR (4µm, 150mm x 1mm ID) (Alltech Associates, Inc., Deerfield, Illinois, USA)
- Rheos Allegro HPLC Pump (Thermo Fisher Scientific, Rockford, USA)
- Ringcaps® Duran® disposable micro pipettes with ring mark (Hirschmann Laborgeräte GmbH & Co.Kg, Eberstadt, Germany).
- sciFLEXARRAYER microarray spotter (Sciension, Dortmund, Germany)
- Security Guard™ cartridge system/RP1 (4x2mmID) filter inlet (Phenomenex, Torrence, USA),
- Stainless steel NanoTight™ union (Upchurch Scientific, Washington, USA)

Instruments/Technical equipment

- Sun Collect MALDI spotter (Sunchrom, Friedrichsdorf, Switzerland)
- Tear Test ophthalmic strips (Optitech[®] Eyecare, Tarun Enterprises, Allahabad, India)
- Teflon-coated needle tip (Tecan, Männedorf, Switzerland)
- Ultraflex II mass spectrometer (Bruker Daltonics, Bremen, Germany)
- Unimax 1010 shaker
- Unimax 1010 shaker (Heidolph Instruments, Schwabach, Germany)
- XCell SureLock[™] Mini-Cell Electrophoresis System (Invitrogen, Carlsbad, USA)
- ZIPTIP[®] C18 pipette tips (EMD Millipore Corporation, Billerica, USA)
- Reaction tubes (0.5/1.5/2 ml) (Eppendorf AG, Hamburg, Germany)
- Pipette tips (Eppendorf AG, Hamburg, Germany)

10 Software

- XCalibur[®] (Thermo Fisher Scientific, Rockford, USA)
- Photoshop C53 (extended version) (Adobe Systems Inc.)
- Ingenuity[®] IPA[®] tool system (Ingenuity[®] Systems)
- P2M (Proteomics Pipeline Mainz)
- Cytoscape (version 2.8.3)
- MarathonControl (version 1.5.7) (Lasertechnik Berlin, Berlin, Germany)
- Warp-LC (version 1.1) (Bruker Daltonics, Bremen, Germany)
- Survey Viewer (version 1.1) (Bruker Daltonics, Bremen, Germany)
- Proteome Discoverer[®] (version 1.1) (Thermo Fisher Scientific, Rockford, USA)
- Flex Control (version 2.4) (Bruker Daltonics, Bremen, Germany)
- Flex Analysis (version 2.4) (Bruker Daltonics, Bremen, Germany)
- ImaGene[®] (version 5.5) (BioDiscovery, Hawthorne, USA)
- Statistica (versions 8/10) (StatSoft, Tulsa, USA)
- TramineR
- R-Project (version 2.8)

11 Chemicals

- ACN_{LC-MS} (Applichem GmbH, Illinois Tool Works Inc., Chicago, Illinois, USA)
- AMO Complete Moisture Plus[®] (Abbott Medical Optics Illinois, USA)
- CHCA (Fluka Analytical, Sigma-Aldrich, Co. LLC.)
- Crystall violet assay (Sigma-Aldrich Co. LLC)
- DCFH-DA ROS assays (Sigma-Aldrich Co, St. Louis, USA),
- DDM (Fluka Analytical, Sigma-Aldrich Co, St. Louis, USA)
- DMEM (Ham's F12 1:1) (Biochrome, Merck KGaA, Darmstadt, Germany)
- DPBS (Sigma-Aldrich Co, St. Louis, USA)
- EDTA (Sigma-Aldrich Co, St. Louis, USA)
- FCS (Biochrome, Merck KGaA, Darmstadt, Germany)
- FA_{LC-MS} (Applichem GmbH, Illinois Tool Works Inc., Chicago, Illinois, USA)
- L-alanin/L-glutamine (Biochrome, Merck KGaA, Darmstadt, Germany)
- MeOH_{LC-MS} (Applichem GmbH, Illinois Tool Works Inc., Chicago, Illinois, USA)
- Novex Colloidal Blue staining kit (Invitrogen, Carlsbad, USA)
- NuPage 12% Bis-Tris, 10-well prepared minigels (Invitrogen, Carlsbad, USA)
- NuPAGE LDS sample buffer [4x] (Invitrogen, Carlsbad, USA)
- Penicillin (Sigma-Aldrich Co, St. Louis, USA)
- Sequencing grade modified trypsin (Promega Co, Madison, USA)
- Streptomycin (Sigma-Aldrich Co, St. Louis, USA)
- Insulin (Sigma-Aldrich Co, St. Louis, USA)
- EGF (Invitrogen, Carlsbad, USA)
- Hydrocortison (Sigma-Aldrich Co, St. Louis, USA)
- Peptide Calibration Standard (Bruker Daltonics, Bremen, Germany)
- Peroxide (Carl Roth GMBH & Co.KG, Karlsruhe, Germany)
- Pierce[®] LTQ ESI positive ion calibration mix (Thermo Fisher Scientific, Rockford, USA)
- Protease inhibitor for use in tissue culture (Sigma-Aldrich Co, St. Louis, USA)
- Protein Calibration Standard I (Bruker Daltonics, Bremen, Germany)
- Reducing agent [10x] (Invitrogen, Carlsbad, USA)
- SeabluePlus 2 prestained protein standard (Invitrogen, Carlsbad, USA)
- Sequencing grade modified trypsin (Promega)
- Staurosporine (Calbiochem, Merck KGaA, Darmstadt, Germany)
- Streptomycin (Sigma-Aldrich Co, St. Louis, USA)
- Taurine (Carl Roth GmbH+Co.KG, Karlsruhe, Germany)
- TFA (Merck KGaA, Darmstadt, Germany)

Chemicals

- Water_{LC-MS} (Applichem GmbH, Illinois Tool Works Inc., Chicago, Illinois, USA)

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13 Appendices

Appendix 1 LC MALDI MS identified capillary tear proteins (Funke *et al.*, 2012)

1.	TRFL_HUMAN Mass: 78132 Score: 16010 Queries matched: 614 emPAI: 15.25 Lactotransferrin OS=Homo sapiens GN=LTF PE=1 SV=6
2.	LCN1_HUMAN Mass: 19238 Score: 15876 Queries matched: 489 emPAI: 4.60 Lipocalin-1 OS=Homo sapiens GN=LCN1 PE=1 SV=1
3.	PROL4_HUMAN Mass: 15116 Score: 14746 Queries matched: 371 emPAI: 3.28 Proline-rich protein 4 OS=Homo sapiens GN=PRR4 PE=1 SV=2
4.	LYSC_HUMAN Mass: 16526 Score: 4800 Queries matched: 208 emPAI: 6.34 Lysozyme C OS=Homo sapiens GN=LYZ PE=1 SV=1
5.	ALBU_HUMAN Mass: 69321 Score: 4415 Queries matched: 230 emPAI: 5.90 Serum albumin OS=Homo sapiens GN=ALB PE=1 SV=2
6.	PIGR_HUMAN Mass: 83232 Score: 4283 Queries matched: 211 emPAI: 1.09 Polymeric immunoglobulin receptor OS=Homo sapiens GN=PIGR PE=1 SV=4
7.	IGKC_HUMAN Mass: 11602 Score: 4135 Queries matched: 113 emPAI: 9.36 Ig kappa chain C region OS=Homo sapiens GN=IGKC PE=1 SV=1
8.	IGHA1_HUMAN Mass: 37631 Score: 4096 Queries matched: 126 emPAI: 1.43 Ig alpha-1 chain C region OS=Homo sapiens GN=IGHA1 PE=1 SV=2
9.	LC1L1_HUMAN Mass: 17907 Score: 3707 Queries matched: 156 emPAI: 0.36 Putative lipocalin 1-like protein 1 OS=Homo sapiens GN=LCN1L1 PE=5 SV=1
10.	IGHA2_HUMAN Mass: 36503 Score: 3488 Queries matched: 98 emPAI: 1.14 Ig alpha-2 chain C region OS=Homo sapiens GN=IGHA2 PE=1 SV=3
11.	CYTS_HUMAN Mass: 16204 Score: 3473 Queries matched: 116 emPAI: 4.45 Cystatin-S OS=Homo sapiens GN=CST4 PE=1 SV=3
12.	LACRT_HUMAN Mass: 14237 Score: 3135 Queries matched: 112 emPAI: 2.18 Extracellular glycoprotein lacritin OS=Homo sapiens GN=LACRT PE=1 SV=1
13.	PIP_HUMAN Mass: 16562 Score: 2998 Queries matched: 210 emPAI: 6.34 Prolactin-inducible protein OS=Homo sapiens GN=PIP PE=1 SV=1
14.	TRY1_HUMAN Mass: 26541 Score: 2917 Queries matched: 226 emPAI: 0.23 Trypsin-1 OS=Homo sapiens GN=PRSS1 PE=1 SV=1
15.	TRY2_HUMAN Mass: 26471 Score: 2917 Queries matched: 224 emPAI: 0.23 Trypsin-2 OS=Homo sapiens GN=PRSS2 PE=1 SV=1
16.	TRY6_HUMAN Mass: 26522 Score: 2917 Queries matched: 223 emPAI: 0.23 Putative trypsin-6 OS=Homo sapiens GN=TRY6 PE=5 SV=1
17.	ZA2G_HUMAN Mass: 34237 Score: 2514 Queries matched: 107 emPAI: 6.03 Zinc-alpha-2-glycoprotein OS=Homo sapiens GN=AZGP1 PE=1 SV=2
18.	SG2A1_HUMAN Mass: 10876 Score: 2373 Queries matched: 89 emPAI: 11.18 Mammaglobin-B OS=Homo sapiens GN=SCGB2A1 PE=1 SV=1
19.	CYTN_HUMAN Mass: 16351 Score: 1949 Queries matched: 75 emPAI: 2.85 Cystatin-SN OS=Homo sapiens GN=CST1 PE=1 SV=2
20.	DMBT1_HUMAN Mass: 260569 Score: 1305 Queries matched: 59 emPAI: 0.09 Deleted in malignant brain tumors 1 protein OS=Homo sapiens GN=DMBT1 PE=1 SV=2
21.	PROL1_HUMAN Mass: 27199 Score: 1081 Queries matched: 59 emPAI: 1.77 Proline-rich protein 1 OS=Homo sapiens GN=PROL1 PE=1 SV=2
22.	LAC2_HUMAN Mass: 11287 Score: 829 Queries matched: 46 emPAI: 3.23 Ig lambda-2 chain C regions OS=Homo sapiens GN=IGLC2 PE=1 SV=1
23.	LAC3_HUMAN Mass: 11231 Score: 829 Queries matched: 46 emPAI: 3.29 Ig lambda-3 chain C regions OS=Homo sapiens GN=IGLC3 PE=1 SV=1
24.	LAC7_HUMAN Mass: 11296 Score: 826 Queries matched: 41 emPAI: 1.62 Ig lambda-7 chain C region OS=Homo sapiens GN=IGLC7 PE=1 SV=2
25.	ZG16B_HUMAN Mass: 22725 Score: 806 Queries matched: 63 emPAI: 1.07 Zymogen granule protein 16 homolog B OS=Homo sapiens GN=ZG16B PE=1 SV=3
26.	HV305_HUMAN Mass: 13218 Score: 750 Queries matched: 32 emPAI: 0.51 Ig heavy chain V-III region BRO OS=Homo sapiens PE=1 SV=1
27.	HV316_HUMAN Mass: 12794 Score: 750 Queries matched: 32 emPAI: 0.53 Ig heavy chain V-III region TEI OS=Homo sapiens PE=1 SV=1
28.	SG1D1_HUMAN Mass: 9891 Score: 680 Queries matched: 50 emPAI: 2.00 Secretoglobin family 1D member 1 OS=Homo sapiens GN=SCGB1D1 PE=1 SV=1
29.	IGJ_HUMAN Mass: 18087 Score: 677 Queries matched: 43 emPAI: 0.84 Immunoglobulin J chain OS=Homo sapiens GN=IGJ PE=1 SV=4
30.	CLUS_HUMAN Mass: 52461 Score: 527 Queries matched: 34 emPAI: 0.53 Clusterin OS=Homo sapiens GN=CLU PE=1 SV=1

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31.	A2MG_HUMAN Mass: 163189 Score: 507 Queries matched: 45 emPAI: 0.46 Alpha-2-macroglobulin OS=Homo sapiens GN=A2M PE=1 SV=2
32.	IGHG1_HUMAN Mass: 36083 Score: 481 Queries matched: 23 emPAI: 1.94 Ig gamma-1 chain C region OS=Homo sapiens GN=IGHG1 PE=1 SV=1
33.	APOA1_HUMAN Mass: 30759 Score: 425 Queries matched: 43 emPAI: 1.95 Apolipoprotein A-I OS=Homo sapiens GN=APOA1 PE=1 SV=1
34.	CO3_HUMAN Mass: 187030 Score: 383 Queries matched: 74 emPAI: 0.39 Complement C3 OS=Homo sapiens GN=C3 PE=1 SV=2
35.	A1AT_HUMAN Mass: 46707 Score: 340 Queries matched: 24 emPAI: 1.59 Alpha-1-antitrypsin OS=Homo sapiens GN=SERPINA1 PE=1 SV=3
36.	ACTB_HUMAN Mass: 41710 Score: 331 Queries matched: 43 emPAI: 0.70 Actin, cytoplasmic 1 OS=Homo sapiens GN=ACTB PE=1 SV=1
37.	KV302_HUMAN Mass: 11768 Score: 331 Queries matched: 51 emPAI: 1.53 Ig kappa chain V-III region SIE OS=Homo sapiens PE=1 SV=1
38.	K2C1_HUMAN Mass: 65999 Score: 255 Queries matched: 81 emPAI: 0.81 Keratin, type II cytoskeletal 1 OS=Homo sapiens GN=KRT1 PE=1 SV=6
39.	TRFE_HUMAN Mass: 77000 Score: 249 Queries matched: 36 emPAI: 0.24 Serotransferrin OS=Homo sapiens GN=TF PE=1 SV=2
40.	KV301_HUMAN Mass: 11628 Score: 249 Queries matched: 33 emPAI: 0.60 Ig kappa chain V-III region B6 OS=Homo sapiens PE=1 SV=1
41.	POTEF_HUMAN Mass: 121367 Score: 242 Queries matched: 37 emPAI: 0.15 POTE ankyrin domain family member F OS=Homo sapiens GN=POTEF PE=1 SV=2
42.	POTEE_HUMAN Mass: 121286 Score: 242 Queries matched: 36 emPAI: 0.15 POTE ankyrin domain family member E OS=Homo sapiens GN=POTEE PE=1 SV=3
43.	ACTBM_HUMAN Mass: 41989 Score: 241 Queries matched: 25 emPAI: 0.30 Putative beta-actin-like protein 3 OS=Homo sapiens GN=POTEKP PE=5 SV=1
44.	TTHY_HUMAN Mass: 15877 Score: 241 Queries matched: 20 emPAI: 1.82 Transthyretin OS=Homo sapiens GN=TTR PE=1 SV=1
45.	HV318_HUMAN Mass: 12423 Score: 241 Queries matched: 19 emPAI: 0.55 Ig heavy chain V-III region TUR OS=Homo sapiens PE=1 SV=1
46.	HV315_HUMAN Mass: 13082 Score: 241 Queries matched: 19 emPAI: 0.52 Ig heavy chain V-III region WAS OS=Homo sapiens PE=1 SV=1
47.	HV304_HUMAN Mass: 12348 Score: 241 Queries matched: 19 emPAI: 0.56 Ig heavy chain V-III region TIL OS=Homo sapiens PE=1 SV=1
48.	HV313_HUMAN Mass: 12945 Score: 241 Queries matched: 19 emPAI: 0.52 Ig heavy chain V-III region POM OS=Homo sapiens PE=1 SV=1
49.	ACTBL_HUMAN Mass: 41976 Score: 200 Queries matched: 27 emPAI: 0.14 Beta-actin-like protein 2 OS=Homo sapiens GN=ACTBL2 PE=1 SV=2
50.	ZAGL1_HUMAN Mass: 22965 Score: 178 Queries matched: 20 emPAI: 0.62 Putative zinc-alpha-2-glycoprotein-like 1 OS=Homo sapiens PE=5 SV=2
51.	S10A8_HUMAN Mass: 10828 Score: 169 Queries matched: 13 emPAI: 1.72 Protein S100-A8 OS=Homo sapiens GN=S100A8 PE=1 SV=1
52.	EIF2A_HUMAN Mass: 64949 Score: 152 Queries matched: 126 emPAI: 0.09 Eukaryotic translation initiation factor 2A OS=Homo sapiens GN=EIF2A PE=1 SV=3
53.	AIFM2_HUMAN Mass: 40501 Score: 145 Queries matched: 15 emPAI: 0.15 Apoptosis-inducing factor 2 OS=Homo sapiens GN=AIFM2 PE=2 SV=1
54.	IGHM_HUMAN Mass: 49276 Score: 138 Queries matched: 22 emPAI: 0.12 Ig mu chain C region OS=Homo sapiens GN=IGHM PE=1 SV=3
55.	MUCB_HUMAN Mass: 43030 Score: 138 Queries matched: 21 emPAI: 0.14 Ig mu heavy chain disease protein OS=Homo sapiens PE=1 SV=1
56.	ELOA1_HUMAN Mass: 89853 Score: 129 Queries matched: 136 emPAI: 0.06 Transcription elongation factor B polypeptide 3 OS=Homo sapiens GN=TCEB3 PE=1 SV=2
57.	HPT_HUMAN Mass: 45177 Score: 125 Queries matched: 20 emPAI: 0.45 Haptoglobin OS=Homo sapiens GN=HP PE=1 SV=1
58.	HPTR_HUMAN Mass: 39005 Score: 125 Queries matched: 13 emPAI: 0.53 Haptoglobin-related protein OS=Homo sapiens GN=HPR PE=2 SV=2
59.	LAC1_HUMAN Mass: 11341 Score: 122 Queries matched: 19 emPAI: 3.23 Ig lambda-1 chain C regions OS=Homo sapiens GN=IGLC1 PE=1 SV=1
60.	IGHG3_HUMAN Mass: 41260 Score: 108 Queries matched: 16 emPAI: 0.31 Ig gamma-3 chain C region OS=Homo sapiens GN=IGHG3 PE=1 SV=2

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61.	KV312_HUMAN Mass: 14064 Score: 107 Queries matched: 26 emPAI: 0.47 Ig kappa chain V-III region HAH OS=Homo sapiens PE=2 SV=1
62.	IGHG2_HUMAN Mass: 35878 Score: 101 Queries matched: 10 emPAI: 0.17 Ig gamma-2 chain C region OS=Homo sapiens GN=IGHG2 PE=1 SV=2
63.	ABCA3_HUMAN Score: 97 Queries matched: 28 ATP-binding cassette sub-family A member 3 OS=Homo sapiens GN=ABCA3 PE=1 SV=2
64.	K22E_HUMAN Mass: 65393 Score: 94 Queries matched: 51 emPAI: 0.09 Keratin, type II cytoskeletal 2 epidermal OS=Homo sapiens GN=KRT2 PE=1 SV=2
65.	S10A9_HUMAN Mass: 13234 Score: 90 Queries matched: 8 emPAI: 0.51 Protein S100-A9 OS=Homo sapiens GN=S100A9 PE=1 SV=1
66.	HS90B_HUMAN Mass: 83212 Score: 84 Queries matched: 24 emPAI: 0.40 Heat shock protein HSP 90-beta OS=Homo sapiens GN=HSP90AB1 PE=1 SV=4
67.	HBA_HUMAN Mass: 15248 Score: 83 Queries matched: 8 emPAI: 1.06 Hemoglobin subunit alpha OS=Homo sapiens GN=HBA1 PE=1 SV=2
68.	K1C10_HUMAN Mass: 58792 Score: 79 Queries matched: 64 emPAI: 0.33 Keratin, type I cytoskeletal 10 OS=Homo sapiens GN=KRT10 PE=1 SV=6
69.	EF1A2_HUMAN Mass: 50438 Score: 70 Queries matched: 7 emPAI: 0.25 Elongation factor 1-alpha 2 OS=Homo sapiens GN=EEF1A2 PE=1 SV=1
70.	EF1A1_HUMAN Mass: 50109 Score: 70 Queries matched: 8 emPAI: 0.25 Elongation factor 1-alpha 1 OS=Homo sapiens GN=EEF1A1 PE=1 SV=1
71.	HEMO_HUMAN Mass: 51643 Score: 68 Queries matched: 10 emPAI: 0.11 Hemopexin OS=Homo sapiens GN=HPX PE=1 SV=2
72.	K1C9_HUMAN Mass: 62027 Score: 68 Queries matched: 23 emPAI: 0.20 Keratin, type I cytoskeletal 9 OS=Homo sapiens GN=KRT9 PE=1 SV=3
73.	KV105_HUMAN Mass: 11654 Score: 66 Queries matched: 2 emPAI: 0.60 Ig kappa chain V-I region DEE OS=Homo sapiens PE=1 SV=1
74.	H90B2_HUMAN Score: 66 Queries matched: 3 Putative heat shock protein HSP 90-beta 2 OS=Homo sapiens GN=HSP90AB2P PE=1 SV=2
75.	POTEI_HUMAN Mass: 121205 Score: 65 Queries matched: 21 emPAI: 0.10 POTE ankyrin domain family member I OS=Homo sapiens GN=POTEI PE=3 SV=1
76.	ENOG_HUMAN Mass: 47239 Score: 64 Queries matched: 16 emPAI: 0.13 Gamma-enolase OS=Homo sapiens GN=ENO2 PE=1 SV=3
77.	ENOA_HUMAN Mass: 47139 Score: 64 Queries matched: 13 emPAI: 0.13 Alpha-enolase OS=Homo sapiens GN=ENO1 PE=1 SV=2
78.	ENOB_HUMAN Mass: 46902 Score: 64 Queries matched: 9 emPAI: 0.13 Beta-enolase OS=Homo sapiens GN=ENO3 PE=1 SV=4
79.	POTEJ_HUMAN Mass: 117315 Score: 63 Queries matched: 23 emPAI: 0.05 POTE ankyrin domain family member J OS=Homo sapiens GN=POTEJ PE=3 SV=1
80.	NGAL_HUMAN Mass: 22574 Score: 62 Queries matched: 12 emPAI: 0.63 Neutrophil gelatinase-associated lipocalin OS=Homo sapiens GN=LCN2 PE=1 SV=2
81.	SMCA4_HUMAN Mass: 184530 Score: 59 Queries matched: 74 emPAI: 0.03 Transcription activator BRG1 OS=Homo sapiens GN=SMARCA4 PE=1 SV=2
82.	RRBP1_HUMAN Mass: 152381 Score: 59 Queries matched: 139 emPAI: 0.04 Ribosome-binding protein 1 OS=Homo sapiens GN=RRBP1 PE=1 SV=4
83.	HV310_HUMAN Mass: 13557 Score: 57 Queries matched: 11 emPAI: 0.50 Ig heavy chain V-III region HIL OS=Homo sapiens PE=1 SV=1
84.	A1AG1_HUMAN Mass: 23497 Score: 56 Queries matched: 3 emPAI: 0.26 Alpha-1-acid glycoprotein 1 OS=Homo sapiens GN=ORM1 PE=1 SV=1
85.	KV101_HUMAN Mass: 11985 Score: 54 Queries matched: 4 emPAI: 0.58 Ig kappa chain V-I region AG OS=Homo sapiens PE=1 SV=1
86.	KV118_HUMAN Mass: 11833 Score: 54 Queries matched: 3 emPAI: 0.58 Ig kappa chain V-I region WEA OS=Homo sapiens PE=1 SV=1
87.	NCKP5_HUMAN Mass: 208409 Score: 53 Queries matched: 27 emPAI: 0.03 Nck-associated protein 5 OS=Homo sapiens GN=NCKAP5 PE=1 SV=2
88.	LV403_HUMAN Mass: 11510 Score: 53 Queries matched: 4 emPAI: 0.60 Ig lambda chain V-IV region Hil OS=Homo sapiens PE=1 SV=1
89.	MECP2_HUMAN Score: 53 Queries matched: 24 Methyl-CpG-binding protein 2 OS=Homo sapiens GN=MECP2 PE=1 SV=1
90.	GSTCD_HUMAN Mass: 71033 Score: 52 Queries matched: 20 emPAI: 0.08 Glutathione S-transferase C-terminal domain-containing protein OS=Homo sapiens GN=GSTCD PE=1 SV=

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91.	IGHG4_HUMAN Mass: 35918 Score: 51 Queries matched: 9 emPAI: 0.36 Ig gamma-4 chain C region OS=Homo sapiens GN=IGHG4 PE=1 SV=1
92.	CO4A_HUMAN Mass: 192650 Score: 50 Queries matched: 20 emPAI: 0.03 Complement C4-A OS=Homo sapiens GN=C4A PE=1 SV=1
93.	K2C5_HUMAN Mass: 62340 Score: 50 Queries matched: 15 emPAI: 0.09 Keratin, type II cytoskeletal 5 OS=Homo sapiens GN=KRT5 PE=1 SV=3
94.	PATZ1_HUMAN Mass: 74013 Score: 49 Queries matched: 35 emPAI: 0.08 POZ-, AT hook-, and zinc finger-containing protein 1 OS=Homo sapiens GN=PATZ1 PE=1 SV=1
95.	CE164_HUMAN Mass: 164214 Score: 48 Queries matched: 26 emPAI: 0.07 Centrosomal protein of 164 kDa OS=Homo sapiens GN=CEP164 PE=1 SV=3
96.	PA2GA_HUMAN Mass: 16072 Score: 47 Queries matched: 10 emPAI: 0.41 Phospholipase A2, membrane associated OS=Homo sapiens GN=PLA2G2A PE=1 SV=2
97.	NOTC4_HUMAN Mass: 209480 Score: 46 Queries matched: 31 emPAI: 0.03 Neurogenic locus notch homolog protein 4 OS=Homo sapiens GN=NOTCH4 PE=1 SV=2
98.	KCNU1_HUMAN Score: 45 Queries matched: 27 Potassium channel subfamily U member 1 OS=Homo sapiens GN=KCNU1 PE=2 SV=2
99.	RXFP2_HUMAN Mass: 86396 Score: 43 Queries matched: 4 emPAI: 0.07 Relaxin receptor 2 OS=Homo sapiens GN=RXFP2 PE=1 SV=1
100.	A1AG2_HUMAN Mass: 23588 Score: 43 Queries matched: 4 emPAI: 0.26 Alpha-1-acid glycoprotein 2 OS=Homo sapiens GN=ORM2 PE=1 SV=2
101.	SSH2_HUMAN Mass: 158116 Score: 43 Queries matched: 20 emPAI: 0.04 Protein phosphatase Slingshot homolog 2 OS=Homo sapiens GN=SSH2 PE=1 SV=1
102.	CT194_HUMAN Mass: 132202 Score: 43 Queries matched: 30 emPAI: 0.04 Uncharacterized protein C20orf194 OS=Homo sapiens GN=C20orf194 PE=2 SV=1
103.	HV319_HUMAN Mass: 12555 Score: 43 Queries matched: 6 emPAI: 0.55 Ig heavy chain V-III region JON OS=Homo sapiens PE=1 SV=1
104.	SPIR2_HUMAN Mass: 79620 Score: 42 Queries matched: 25 emPAI: 0.07 Protein spire homolog 2 OS=Homo sapiens GN=SPIRE2 PE=1 SV=3
105.	LRC4C_HUMAN Score: 41 Queries matched: 13 Leucine-rich repeat-containing protein 4C OS=Homo sapiens GN=LRR4C PE=1 SV=1
106.	COBA2_HUMAN Mass: 171670 Score: 40 Queries matched: 44 emPAI: 0.03 Collagen alpha-2(XI) chain OS=Homo sapiens GN=COL11A2 PE=1 SV=4
107.	POP1_HUMAN Mass: 114636 Score: 40 Queries matched: 36 emPAI: 0.05 Ribonucleases P/MRP protein subunit POP1 OS=Homo sapiens GN=POP1 PE=1 SV=2
108.	DPOLN_HUMAN Mass: 100244 Score: 39 Queries matched: 20 emPAI: 0.06 DNA polymerase nu OS=Homo sapiens GN=POLN PE=1 SV=2
109.	AMBP_HUMAN Score: 39 Queries matched: 61 Protein AMBP OS=Homo sapiens GN=AMBP PE=1 SV=1
110.	CERU_HUMAN Mass: 122128 Score: 39 Queries matched: 23 emPAI: 0.10 Ceruloplasmin OS=Homo sapiens GN=CP PE=1 SV=1
111.	HV307_HUMAN Mass: 13659 Score: 38 Queries matched: 26 emPAI: 1.23 Ig heavy chain V-III region CAM OS=Homo sapiens PE=1 SV=1
112.	K2C4_HUMAN Mass: 57250 Score: 37 Queries matched: 26 emPAI: 0.22 Keratin, type II cytoskeletal 4 OS=Homo sapiens GN=KRT4 PE=1 SV=4
113.	FIBB_HUMAN Mass: 55892 Score: 37 Queries matched: 7 emPAI: 0.10 Fibrinogen beta chain OS=Homo sapiens GN=FGB PE=1 SV=2
114.	ANXA1_HUMAN Mass: 38690 Score: 37 Queries matched: 4 emPAI: 0.15 Annexin A1 OS=Homo sapiens GN=ANXA1 PE=1 SV=2
115.	TTL11_HUMAN Score: 37 Queries matched: 71 Tubulin polyglutamylase TTL11 OS=Homo sapiens GN=TTL11 PE=2 SV=1
116.	CENPE_HUMAN Mass: 316219 Score: 37 Queries matched: 73 emPAI: 0.02 Centromere-associated protein E OS=Homo sapiens GN=CENPE PE=1 SV=2
117.	RGPA2_HUMAN Mass: 210636 Score: 37 Queries matched: 120 emPAI: 0.03 Ral GTPase-activating protein subunit alpha-2 OS=Homo sapiens GN=RALGAP2 PE=1 SV=2
118.	E41LA_HUMAN Mass: 79010 Score: 36 Queries matched: 27 emPAI: 0.07 Band 4.1-like protein 4A OS=Homo sapiens GN=EPB41L4A PE=1 SV=2
119.	FGD6_HUMAN Mass: 160715 Score: 36 Queries matched: 39 emPAI: 0.04 FYVE, RhoGEF and PH domain-containing protein 6 OS=Homo sapiens GN=FGD6 PE=1 SV=2
120.	KLH34_HUMAN Mass: 70568 Score: 36 Queries matched: 32 emPAI: 0.08 Kelch-like protein 34 OS=Homo sapiens GN=KLHL34 PE=2 SV=1

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121.	SPYA_HUMAN Mass: 42982 Score: 36 Queries matched: 46 emPAI: 0.14 Serine--pyruvate aminotransferase OS=Homo sapiens GN=AGXT PE=1 SV=1
122.	TBA1A_HUMAN Mass: 50104 Score: 36 Queries matched: 14 emPAI: 0.25 Tubulin alpha-1A chain OS=Homo sapiens GN=TUBA1A PE=1 SV=1
123.	HMG2_HUMAN Mass: 9387 Score: 36 Queries matched: 1 emPAI: 0.77 Non-histone chromosomal protein HMG-17 OS=Homo sapiens GN=HMG2 PE=1 SV=3
124.	ZFAN1_HUMAN Score: 35 Queries matched: 17 AN1-type zinc finger protein 1 OS=Homo sapiens GN=ZFAND1 PE=1 SV=1
125.	BEST1_HUMAN Mass: 67640 Score: 35 Queries matched: 9 emPAI: 0.09 Bestrophin-1 OS=Homo sapiens GN=BEST1 PE=1 SV=1
126.	TLR4_HUMAN Mass: 95619 Score: 35 Queries matched: 23 emPAI: 0.06 Toll-like receptor 4 OS=Homo sapiens GN=TLR4 PE=1 SV=2
127.	NAV2_HUMAN Mass: 267969 Score: 35 Queries matched: 60 emPAI: 0.02 Neuron navigator 2 OS=Homo sapiens GN=NAV2 PE=1 SV=2
128.	SIRT6_HUMAN Mass: 39094 Score: 34 Queries matched: 39 emPAI: 0.15 NAD-dependent deacetylase sirtuin-6 OS=Homo sapiens GN=SIRT6 PE=1 SV=2
129.	MDGA1_HUMAN Mass: 105724 Score: 34 Queries matched: 37 emPAI: 0.05 MAM domain-containing glycosylphosphatidylinositol anchor protein 1 OS=Homo sapiens GN=MDGA1 PE=
130.	TUB_HUMAN Mass: 55617 Score: 34 Queries matched: 17 emPAI: 0.11 Tubby protein homolog OS=Homo sapiens GN=TUB PE=1 SV=1
131.	K2C73_HUMAN Mass: 58887 Score: 34 Queries matched: 11 emPAI: 0.10 Keratin, type II cytoskeletal 73 OS=Homo sapiens GN=KRT73 PE=1 SV=1
132.	H15_HUMAN Mass: 22566 Score: 34 Queries matched: 3 emPAI: 0.28 Histone H1.5 OS=Homo sapiens GN=HIST1H1B PE=1 SV=3
133.	DEF6_HUMAN Mass: 10968 Score: 34 Queries matched: 6 emPAI: 0.64 Defensin-6 OS=Homo sapiens GN=DEFA6 PE=1 SV=1
134.	URB2_HUMAN Mass: 170435 Score: 34 Queries matched: 49 emPAI: 0.03 Unhealthy ribosome biogenesis protein 2 homolog OS=Homo sapiens GN=URB2 PE=1 SV=2
135.	CP059_HUMAN Mass: 46373 Score: 34 Queries matched: 12 emPAI: 0.13 Uncharacterized protein C16orf59 OS=Homo sapiens GN=C16orf59 PE=2 SV=1
136.	SPA9_HUMAN Mass: 46527 Score: 34 Queries matched: 20 emPAI: 0.13 Serpins OS=Homo sapiens GN=SERPINA9 PE=1 SV=3
137.	FIBG_HUMAN Mass: 51479 Score: 34 Queries matched: 9 emPAI: 0.24 Fibrinogen gamma chain OS=Homo sapiens GN=FGG PE=1 SV=3
138.	CIC_HUMAN Mass: 163719 Score: 34 Queries matched: 15 emPAI: 0.03 Protein capicua homolog OS=Homo sapiens GN=CIC PE=1 SV=2
139.	CAPON_HUMAN Mass: 56115 Score: 34 Queries matched: 12 emPAI: 0.10 Carboxyl-terminal PDZ ligand of neuronal nitric oxide synthase protein OS=Homo sapiens GN=NOS1AP PE
140.	CLCA_HUMAN Mass: 27060 Score: 33 Queries matched: 27 emPAI: 0.23 Clathrin light chain A OS=Homo sapiens GN=CLTA PE=1 SV=1
141.	MCCB_HUMAN Mass: 61294 Score: 33 Queries matched: 57 emPAI: 0.10 Methylcrotonyl-CoA carboxylase beta chain, mitochondrial OS=Homo sapiens GN=MCCC2 PE=1 SV=1
142.	PDE12_HUMAN Score: 33 Queries matched: 14 2',5'-phosphodiesterase 12 OS=Homo sapiens GN=PDE12 PE=1 SV=2
143.	HSPB1_HUMAN Mass: 22768 Score: 33 Queries matched: 8 emPAI: 0.63 Heat shock protein beta-1 OS=Homo sapiens GN=HSPB1 PE=1 SV=2
144.	KIF26A_HUMAN Mass: 194468 Score: 33 Queries matched: 93 emPAI: 0.03 Kinesin-like protein KIF26A OS=Homo sapiens GN=KIF26A PE=1 SV=3
145.	CH60_HUMAN Mass: 61016 Score: 33 Queries matched: 15 emPAI: 0.10 60 kDa heat shock protein, mitochondrial OS=Homo sapiens GN=HSPD1 PE=1 SV=2
146.	PSME4_HUMAN Mass: 211199 Score: 33 Queries matched: 17 emPAI: 0.03 Proteasome activator complex subunit 4 OS=Homo sapiens GN=PSME4 PE=1 SV=2
147.	KIF17_HUMAN Mass: 115043 Score: 33 Queries matched: 19 emPAI: 0.05 Kinesin-like protein KIF17 OS=Homo sapiens GN=KIF17 PE=1 SV=2
148.	USP9Y_HUMAN Mass: 290891 Score: 33 Queries matched: 49 emPAI: 0.02 Probable ubiquitin carboxyl-terminal hydrolase FAF-Y OS=Homo sapiens GN=USP9Y PE=1 SV=2
149.	GPTC4_HUMAN Mass: 50351 Score: 33 Queries matched: 18 emPAI: 0.12 G patch domain-containing protein 4 OS=Homo sapiens GN=GPATCH4 PE=1 SV=2
150.	RSF1_HUMAN Mass: 163720 Score: 33 Queries matched: 33 emPAI: 0.03 Remodeling and spacing factor 1 OS=Homo sapiens GN=RSF1 PE=1 SV=2

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151.	USP9X_HUMAN Mass: 289356 Score: 33 Queries matched: 52 emPAI: 0.04 Probable ubiquitin carboxyl-terminal hydrolase FAF-X OS=Homo sapiens GN=USP9X PE=1 SV=2
152.	LPHN3_HUMAN Mass: 161710 Score: 32 Queries matched: 32 emPAI: 0.04 Latrophilin-3 OS=Homo sapiens GN=LPHN3 PE=1 SV=2
153.	HSP7C_HUMAN Mass: 70854 Score: 32 Queries matched: 14 emPAI: 0.08 Heat shock cognate 71 kDa protein OS=Homo sapiens GN=HSPA8 PE=1 SV=1
154.	HSP72_HUMAN Mass: 69978 Score: 32 Queries matched: 8 emPAI: 0.08 Heat shock-related 70 kDa protein 2 OS=Homo sapiens GN=HSPA2 PE=1 SV=1
155.	SNED1_HUMAN Mass: 152104 Score: 32 Queries matched: 36 emPAI: 0.04 Sushi, nidogen and EGF-like domain-containing protein 1 OS=Homo sapiens GN=SNED1 PE=1 SV=2
156.	DPYL1_HUMAN Score: 32 Queries matched: 5 Dihydropyrimidinase-related protein 1 OS=Homo sapiens GN=CRMP1 PE=1 SV=1
157.	TIFA_HUMAN Mass: 21431 Score: 32 Queries matched: 1 emPAI: 0.29 TRAF-interacting protein with FHA domain-containing protein A OS=Homo sapiens GN=TIFA PE=1 SV=1
158.	SCLT1_HUMAN Mass: 80861 Score: 32 Queries matched: 17 emPAI: 0.07 Sodium channel and clathrin linker 1 OS=Homo sapiens GN=SCLT1 PE=1 SV=2
159.	RRP5_HUMAN Mass: 208570 Score: 32 Queries matched: 46 emPAI: 0.06 Protein RRP5 homolog OS=Homo sapiens GN=PDCD11 PE=1 SV=3
160.	PGBM_HUMAN Mass: 468501 Score: 32 Queries matched: 58 emPAI: 0.05 Basement membrane-specific heparan sulfate proteoglycan core protein OS=Homo sapiens GN=HSPG2 P
161.	WDFY3_HUMAN Mass: 395006 Score: 32 Queries matched: 65 emPAI: 0.03 WD repeat and FYVE domain-containing protein 3 OS=Homo sapiens GN=WDFY3 PE=1 SV=2
162.	CSPP1_HUMAN Mass: 141736 Score: 31 Queries matched: 19 emPAI: 0.04 Centrosome and spindle pole-associated protein 1 OS=Homo sapiens GN=CSPP1 PE=1 SV=3
163.	RL31_HUMAN Mass: 14454 Score: 31 Queries matched: 31 emPAI: 0.46 60S ribosomal protein L31 OS=Homo sapiens GN=RPL31 PE=1 SV=1
164.	MCR_HUMAN Mass: 106999 Score: 31 Queries matched: 15 emPAI: 0.05 Mineralocorticoid receptor OS=Homo sapiens GN=NR3C2 PE=1 SV=1
165.	K1C28_HUMAN Mass: 50536 Score: 31 Queries matched: 24 emPAI: 0.12 Keratin, type I cytoskeletal 28 OS=Homo sapiens GN=KRT28 PE=1 SV=2
166.	FSIP2_HUMAN Mass: 780119 Score: 31 Queries matched: 134 emPAI: 0.01 Fibrous sheath-interacting protein 2 OS=Homo sapiens GN=FSIP2 PE=1 SV=4
167.	HS90A_HUMAN Mass: 84607 Score: 31 Queries matched: 14 emPAI: 0.14 Heat shock protein HSP 90-alpha OS=Homo sapiens GN=HSP90AA1 PE=1 SV=5
168.	UACA_HUMAN Mass: 162404 Score: 31 Queries matched: 44 emPAI: 0.04 Uveal autoantigen with coiled-coil domains and ankyrin repeats OS=Homo sapiens GN=UACA PE=1 SV=2
169.	GLDN_HUMAN Mass: 58920 Score: 31 Queries matched: 20 emPAI: 0.10 Gliomedin OS=Homo sapiens GN=GLDN PE=2 SV=1
170.	PRAML_HUMAN Mass: 58127 Score: 31 Queries matched: 6 emPAI: 0.10 Leucine-rich repeat-containing protein PRAME-like OS=Homo sapiens GN=PRAMEL PE=2 SV=2
171.	DGKK_HUMAN Mass: 141739 Score: 31 Queries matched: 47 emPAI: 0.04 Diacylglycerol kinase kappa OS=Homo sapiens GN=DGKK PE=1 SV=1
172.	CP057_HUMAN Score: 31 Queries matched: 5 UPF0406 protein C16orf57 OS=Homo sapiens GN=C16orf57 PE=1 SV=1
173.	WNK2_HUMAN Mass: 242525 Score: 31 Queries matched: 25 emPAI: 0.02 Serine/threonine-protein kinase WNK2 OS=Homo sapiens GN=WNK2 PE=1 SV=4
174.	K2C7_HUMAN Mass: 51373 Score: 31 Queries matched: 39 emPAI: 0.11 Keratin, type II cytoskeletal 7 OS=Homo sapiens GN=KRT7 PE=1 SV=4
175.	K2C8_HUMAN Mass: 53671 Score: 31 Queries matched: 17 emPAI: 0.11 Keratin, type II cytoskeletal 8 OS=Homo sapiens GN=KRT8 PE=1 SV=7
176.	GFAP_HUMAN Mass: 49850 Score: 31 Queries matched: 42 emPAI: 0.12 Glial fibrillary acidic protein OS=Homo sapiens GN=GFAP PE=1 SV=1
177.	K2C80_HUMAN Mass: 50494 Score: 31 Queries matched: 9 emPAI: 0.12 Keratin, type II cytoskeletal 80 OS=Homo sapiens GN=KRT80 PE=1 SV=2
178.	C114B_HUMAN Mass: 148003 Score: 31 Queries matched: 27 emPAI: 0.04 Transmembrane protein C9orf144B OS=Homo sapiens GN=C9orf144B PE=2 SV=4
179.	TRAK2_HUMAN Mass: 101356 Score: 30 Queries matched: 14 emPAI: 0.06 Trafficking kinesin-binding protein 2 OS=Homo sapiens GN=TRAK2 PE=1 SV=2
180.	COE4_HUMAN Mass: 64433 Score: 30 Queries matched: 5 emPAI: 0.09 Transcription factor COE4 OS=Homo sapiens GN=EBF4 PE=2 SV=2

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181.	SPTA2_HUMAN Mass: 284364 Score: 30 Queries matched: 36 emPAI: 0.02 Spectrin alpha chain, brain OS=Homo sapiens GN=SPTAN1 PE=1 SV=3
182.	GA2L2_HUMAN Mass: 96460 Score: 30 Queries matched: 48 emPAI: 0.06 GAS2-like protein 2 OS=Homo sapiens GN=GAS2L2 PE=2 SV=1
183.	FBX37_HUMAN Mass: 32977 Score: 30 Queries matched: 25 emPAI: 0.18 F-box only protein 37 OS=Homo sapiens GN=FBXL15 PE=2 SV=2
184.	CP4Z1_HUMAN Mass: 59048 Score: 30 Queries matched: 7 emPAI: 0.10 Cytochrome P450 4Z1 OS=Homo sapiens GN=CYP4Z1 PE=2 SV=1
185.	TR112_HUMAN Mass: 14190 Score: 30 Queries matched: 12 emPAI: 0.47 tRNA methyltransferase 112 homolog OS=Homo sapiens GN=TRMT112 PE=1 SV=1
186.	LAMA3_HUMAN Mass: 366414 Score: 30 Queries matched: 48 emPAI: 0.03 Laminin subunit alpha-3 OS=Homo sapiens GN=LAMA3 PE=1 SV=2
187.	DMXL2_HUMAN Score: 30 Queries matched: 40 DmX-like protein 2 OS=Homo sapiens GN=DMXL2 PE=1 SV=2
188.	PHS2_HUMAN Mass: 14356 Score: 30 Queries matched: 5 emPAI: 0.47 Pterin-4-alpha-carbinolamine dehydratase 2 OS=Homo sapiens GN=PCBD2 PE=1 SV=4
189.	ZN536_HUMAN Mass: 141328 Score: 30 Queries matched: 7 emPAI: 0.04 Zinc finger protein 536 OS=Homo sapiens GN=ZNF536 PE=1 SV=3
190.	ASPM_HUMAN Mass: 409540 Score: 30 Queries matched: 70 emPAI: 0.01 Abnormal spindle-like microcephaly-associated protein OS=Homo sapiens GN=ASPM PE=1 SV=2
191.	ZBTB6_HUMAN Score: 30 Queries matched: 10 Zinc finger and BTB domain-containing protein 6 OS=Homo sapiens GN=ZBTB6 PE=2 SV=1
192.	SEMF3_HUMAN Mass: 88325 Score: 30 Queries matched: 29 emPAI: 0.07 Semaphorin-3F OS=Homo sapiens GN=SEMA3F PE=2 SV=2
193.	IDH3A_HUMAN Mass: 39566 Score: 30 Queries matched: 9 emPAI: 0.15 Isocitrate dehydrogenase [NAD] subunit alpha, mitochondrial OS=Homo sapiens GN=IDH3A PE=1 SV=1
194.	LRC49_HUMAN Mass: 78845 Score: 30 Queries matched: 62 emPAI: 0.07 Leucine-rich repeat-containing protein 49 OS=Homo sapiens GN=LRRRC49 PE=2 SV=2
195.	M4K4_HUMAN Mass: 142013 Score: 30 Queries matched: 21 emPAI: 0.04 Mitogen-activated protein kinase kinase kinase 4 OS=Homo sapiens GN=MAP4K4 PE=1 SV=2
196.	KI67_HUMAN Mass: 358474 Score: 30 Queries matched: 78 emPAI: 0.02 Antigen KI-67 OS=Homo sapiens GN=MKI67 PE=1 SV=2
197.	YM012_HUMAN Mass: 59375 Score: 30 Queries matched: 55 emPAI: 0.10 Uncharacterized protein DKFZp434B061 OS=Homo sapiens PE=2 SV=2
198.	YV021_HUMAN Mass: 30934 Score: 30 Queries matched: 48 emPAI: 0.20 Uncharacterized protein LOC284861 OS=Homo sapiens PE=2 SV=1
199.	MRGRD_HUMAN Mass: 36094 Score: 30 Queries matched: 47 emPAI: 0.17 Mas-related G-protein coupled receptor member D OS=Homo sapiens GN=MRGPRD PE=2 SV=1
200.	HV320_HUMAN Mass: 12722 Score: 30 Queries matched: 5 emPAI: 0.53 Ig heavy chain V-III region GAL OS=Homo sapiens PE=1 SV=1
201.	KIF7_HUMAN Mass: 150495 Score: 30 Queries matched: 47 emPAI: 0.04 Kinesin-like protein KIF7 OS=Homo sapiens GN=KIF7 PE=1 SV=2
202.	MDN1_HUMAN Mass: 632420 Score: 30 Queries matched: 90 emPAI: 0.01 Midasin OS=Homo sapiens GN=MDN1 PE=1 SV=2
203.	RN150_HUMAN Mass: 48041 Score: 29 Queries matched: 5 emPAI: 0.12 RING finger protein 150 OS=Homo sapiens GN=RNF150 PE=2 SV=2
204.	CA195_HUMAN Score: 29 Queries matched: 22 Putative uncharacterized protein C1orf195 OS=Homo sapiens GN=C1orf195 PE=2 SV=1
205.	CCKN_HUMAN Mass: 12661 Score: 29 Queries matched: 3 emPAI: 0.54 Cholecystokinin OS=Homo sapiens GN=CCK PE=1 SV=1
206.	IRS1_HUMAN Mass: 131509 Score: 29 Queries matched: 28 emPAI: 0.04 Insulin receptor substrate 1 OS=Homo sapiens GN=IRS1 PE=1 SV=1
207.	CHD7_HUMAN Mass: 335717 Score: 29 Queries matched: 47 emPAI: 0.02 Chromodomain-helicase-DNA-binding protein 7 OS=Homo sapiens GN=CHD7 PE=1 SV=3
208.	SUCR1_HUMAN Mass: 38672 Score: 29 Queries matched: 27 emPAI: 0.15 Succinate receptor 1 OS=Homo sapiens GN=SUCNR1 PE=1 SV=2
209.	FLRT1_HUMAN Mass: 71313 Score: 29 Queries matched: 11 emPAI: 0.08 Leucine-rich repeat transmembrane protein FLRT1 OS=Homo sapiens GN=FLRT1 PE=1 SV=3
210.	PTC1_HUMAN Mass: 160442 Score: 29 Queries matched: 14 emPAI: 0.04 Protein patched homolog 1 OS=Homo sapiens GN=PTCH1 PE=1 SV=2

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211.	FBXW8_HUMAN Mass: 67352 Score: 29 Queries matched: 32 emPAI: 0.09 F-box/WD repeat-containing protein 8 OS=Homo sapiens GN=FBXW8 PE=1 SV=2
212.	AINX_HUMAN Score: 29 Queries matched: 7 Alpha-internexin OS=Homo sapiens GN=INA PE=1 SV=2
213.	TRIB1_HUMAN Mass: 40983 Score: 29 Queries matched: 24 emPAI: 0.15 Tribbles homolog 1 OS=Homo sapiens GN=TRIB1 PE=1 SV=2
214.	DIP2C_HUMAN Mass: 170658 Score: 29 Queries matched: 19 emPAI: 0.03 Disco-interacting protein 2 homolog C OS=Homo sapiens GN=DIP2C PE=2 SV=2
215.	WRIP1_HUMAN Mass: 72088 Score: 29 Queries matched: 13 emPAI: 0.08 ATPase WRNIP1 OS=Homo sapiens GN=WRNIP1 PE=1 SV=2
216.	GELS_HUMAN Mass: 85644 Score: 29 Queries matched: 13 emPAI: 0.07 Gelsolin OS=Homo sapiens GN=GSN PE=1 SV=1
217.	AL8A1_HUMAN Mass: 53367 Score: 29 Queries matched: 9 emPAI: 0.11 Aldehyde dehydrogenase family 8 member A1 OS=Homo sapiens GN=ALDH8A1 PE=1 SV=1
218.	SYCP3_HUMAN Mass: 27711 Score: 29 Queries matched: 9 emPAI: 0.22 Synaptonemal complex protein 3 OS=Homo sapiens GN=SYCP3 PE=2 SV=1
219.	ZC11A_HUMAN Mass: 89076 Score: 29 Queries matched: 17 emPAI: 0.06 Zinc finger CCCH domain-containing protein 11A OS=Homo sapiens GN=ZC3H11A PE=1 SV=3
220.	CT096_HUMAN Mass: 42832 Score: 29 Queries matched: 10 emPAI: 0.14 Uncharacterized protein C20orf96 OS=Homo sapiens GN=C20orf96 PE=2 SV=2
221.	ZF161_HUMAN Mass: 50924 Score: 28 Queries matched: 5 emPAI: 0.12 Zinc finger protein 161 homolog OS=Homo sapiens GN=ZFP161 PE=2 SV=2
222.	NPA1P_HUMAN Mass: 254227 Score: 28 Queries matched: 19 emPAI: 0.02 Nucleolar pre-ribosomal-associated protein 1 OS=Homo sapiens GN=URB1 PE=1 SV=4
223.	WBS22_HUMAN Score: 28 Queries matched: 8 Uncharacterized methyltransferase WBSCR22 OS=Homo sapiens GN=WBSCR22 PE=1 SV=2
224.	CHST3_HUMAN Mass: 54671 Score: 28 Queries matched: 15 emPAI: 0.11 Carbohydrate sulfotransferase 3 OS=Homo sapiens GN=CHST3 PE=1 SV=3
225.	K1C23_HUMAN Mass: 48101 Score: 28 Queries matched: 21 emPAI: 0.12 Keratin, type I cytoskeletal 23 OS=Homo sapiens GN=KRT23 PE=1 SV=2
226.	PF21B_HUMAN Mass: 57419 Score: 28 Queries matched: 8 emPAI: 0.10 PHD finger protein 21B OS=Homo sapiens GN=PHF21B PE=2 SV=1
227.	NCK2_HUMAN Mass: 42889 Score: 28 Queries matched: 29 emPAI: 0.14 Cytoplasmic protein NCK2 OS=Homo sapiens GN=NCK2 PE=1 SV=2
228.	FZD8_HUMAN Mass: 73252 Score: 28 Queries matched: 10 emPAI: 0.08 Frizzled-8 OS=Homo sapiens GN=FZD8 PE=1 SV=1
229.	FANCJ_HUMAN Mass: 140789 Score: 28 Queries matched: 62 emPAI: 0.04 Fanconi anemia group J protein OS=Homo sapiens GN=BRIP1 PE=1 SV=1
230.	ANT3_HUMAN Mass: 52569 Score: 28 Queries matched: 6 emPAI: 0.11 Antithrombin-III OS=Homo sapiens GN=SERPINC1 PE=1 SV=1
231.	VIP2_HUMAN Mass: 140318 Score: 28 Queries matched: 23 emPAI: 0.04 Inositol hexakisphosphate and diphosphoinositol-pentakisphosphate kinase 2 OS=Homo sapiens GN=PP1P
232.	SMC3_HUMAN Mass: 141454 Score: 28 Queries matched: 21 emPAI: 0.04 Structural maintenance of chromosomes protein 3 OS=Homo sapiens GN=SMC3 PE=1 SV=2
233.	CF132_HUMAN Mass: 124016 Score: 28 Queries matched: 119 emPAI: 0.05 Uncharacterized protein C6orf132 OS=Homo sapiens GN=C6orf132 PE=1 SV=3
234.	COOA1_HUMAN Mass: 175388 Score: 28 Queries matched: 54 emPAI: 0.03 Collagen alpha-1(XIV) chain OS=Homo sapiens GN=COL24A1 PE=2 SV=2
235.	CF168_HUMAN Mass: 46813 Score: 28 Queries matched: 6 emPAI: 0.13 Uncharacterized protein C6orf168 OS=Homo sapiens GN=C6orf168 PE=2 SV=2
236.	DTL_HUMAN Mass: 79417 Score: 28 Queries matched: 25 emPAI: 0.07 Denticleless protein homolog OS=Homo sapiens GN=DTL PE=1 SV=3
237.	P4R3A_HUMAN Mass: 95308 Score: 28 Queries matched: 19 emPAI: 0.06 Serine/threonine-protein phosphatase 4 regulatory subunit 3A OS=Homo sapiens GN=SMEK1 PE=1 SV=1
238.	STOX1_HUMAN Mass: 110893 Score: 28 Queries matched: 29 emPAI: 0.05 Storkhead-box protein 1 OS=Homo sapiens GN=STOX1 PE=1 SV=2
239.	MSLN_HUMAN Mass: 68942 Score: 28 Queries matched: 19 emPAI: 0.08 Mesothelin OS=Homo sapiens GN=MSLN PE=1 SV=2
240.	CABP1_HUMAN Mass: 39841 Score: 28 Queries matched: 14 emPAI: 0.15 Calcium-binding protein 1 OS=Homo sapiens GN=CABP1 PE=1 SV=4

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241.	APC_HUMAN Mass: 311455 Score: 27 Queries matched: 35 emPAI: 0.02 Adenomatous polyposis coli protein OS=Homo sapiens GN=APC PE=1 SV=2
242.	BAALC_HUMAN Mass: 19212 Score: 27 Queries matched: 30 emPAI: 0.33 Brain and acute leukemia cytoplasmic protein OS=Homo sapiens GN=BAALC PE=2 SV=3
243.	CSN6_HUMAN Score: 27 Queries matched: 4 COP9 signalosome complex subunit 6 OS=Homo sapiens GN=COPS6 PE=1 SV=1
244.	LUZP4_HUMAN Mass: 35916 Score: 27 Queries matched: 11 emPAI: 0.17 Leucine zipper protein 4 OS=Homo sapiens GN=LUZP4 PE=2 SV=1
245.	ANXA2_HUMAN Mass: 38580 Score: 27 Queries matched: 7 emPAI: 0.15 Annexin A2 OS=Homo sapiens GN=ANXA2 PE=1 SV=2
246.	HSP76_HUMAN Mass: 70984 Score: 27 Queries matched: 8 emPAI: 0.08 Heat shock 70 kDa protein 6 OS=Homo sapiens GN=HSPA6 PE=1 SV=2
247.	HS71L_HUMAN Mass: 70331 Score: 27 Queries matched: 7 emPAI: 0.08 Heat shock 70 kDa protein 1-like OS=Homo sapiens GN=HSPA1L PE=1 SV=2
248.	HSP71_HUMAN Mass: 70009 Score: 27 Queries matched: 6 emPAI: 0.08 Heat shock 70 kDa protein 1A/1B OS=Homo sapiens GN=HSPA1A PE=1 SV=5
249.	TARA_HUMAN Mass: 261217 Score: 27 Queries matched: 99 emPAI: 0.02 TRIO and F-actin-binding protein OS=Homo sapiens GN=TRIOBP PE=1 SV=3
250.	ZN831_HUMAN Mass: 177839 Score: 27 Queries matched: 43 emPAI: 0.03 Zinc finger protein 831 OS=Homo sapiens GN=ZNF831 PE=2 SV=4
251.	RIMKB_HUMAN Mass: 42436 Score: 27 Queries matched: 17 emPAI: 0.14 Ribosomal protein S6 modification-like protein B OS=Homo sapiens GN=RIMKLB PE=2 SV=2
252.	PAPOG_HUMAN Mass: 82751 Score: 27 Queries matched: 16 emPAI: 0.07 Poly(A) polymerase gamma OS=Homo sapiens GN=PAPOLG PE=1 SV=2
253.	HMGX3_HUMAN Mass: 168228 Score: 27 Queries matched: 30 emPAI: 0.03 HMG domain-containing protein 3 OS=Homo sapiens GN=HMGXB3 PE=2 SV=2
254.	ANKL2_HUMAN Mass: 104050 Score: 27 Queries matched: 54 emPAI: 0.06 Ankyrin repeat and LEM domain-containing protein 2 OS=Homo sapiens GN=ANKLE2 PE=1 SV=4
255.	SG269_HUMAN Mass: 192986 Score: 27 Queries matched: 49 emPAI: 0.03 Tyrosine-protein kinase SgK269 OS=Homo sapiens GN=SGK269 PE=1 SV=4
256.	LIME1_HUMAN Mass: 31268 Score: 27 Queries matched: 11 emPAI: 0.19 Lck-interacting transmembrane adapter 1 OS=Homo sapiens GN=LIME1 PE=1 SV=1
257.	TYW1B_HUMAN Mass: 76897 Score: 27 Queries matched: 37 emPAI: 0.08 tRNA wybutosine-synthesizing protein 1 homolog B OS=Homo sapiens GN=TYW1B PE=2 SV=2
258.	TYW1_HUMAN Mass: 83649 Score: 27 Queries matched: 33 emPAI: 0.07 tRNA wybutosine-synthesizing protein 1 homolog OS=Homo sapiens GN=TYW1 PE=1 SV=2
259.	PHLB2_HUMAN Mass: 142070 Score: 27 Queries matched: 12 emPAI: 0.04 Pleckstrin homology-like domain family B member 2 OS=Homo sapiens GN=PHLDB2 PE=1 SV=2
260.	HV306_HUMAN Mass: 12371 Score: 27 Queries matched: 5 emPAI: 1.43 Ig heavy chain V-III region BUT OS=Homo sapiens PE=1 SV=1
261.	AHNK_HUMAN Mass: 628699 Score: 27 Queries matched: 72 emPAI: 0.01 Neuroblast differentiation-associated protein AHNAK OS=Homo sapiens GN=AHNAK PE=1 SV=2
262.	RL7_HUMAN Mass: 29207 Score: 27 Queries matched: 10 emPAI: 0.21 60S ribosomal protein L7 OS=Homo sapiens GN=RPL7 PE=1 SV=1
263.	ZN284_HUMAN Mass: 68974 Score: 27 Queries matched: 6 emPAI: 0.08 Zinc finger protein 284 OS=Homo sapiens GN=ZNF284 PE=2 SV=1
264.	NU188_HUMAN Mass: 195917 Score: 26 Queries matched: 23 emPAI: 0.03 Nucleoporin NUP188 homolog OS=Homo sapiens GN=NUP188 PE=1 SV=1
265.	CCD38_HUMAN Mass: 65274 Score: 26 Queries matched: 31 emPAI: 0.09 Coiled-coil domain-containing protein 38 OS=Homo sapiens GN=CCDC38 PE=2 SV=1
266.	AXN2_HUMAN Mass: 93499 Score: 26 Queries matched: 35 emPAI: 0.06 Axin-2 OS=Homo sapiens GN=AXIN2 PE=1 SV=1
267.	CECR9_HUMAN Mass: 23587 Score: 26 Queries matched: 19 emPAI: 0.26 Putative cat eye syndrome critical region protein 9 OS=Homo sapiens GN=CECR9 PE=5 SV=1

Appendix 2: LC ESI MS identified capillary tear proteins

Accession	# Peptides	# AAs	MW [kDa]	calc. pI	Score	Description
P02788	1821	710	78,1	8,12	1981	Lactotransferrin OS=Homo sapiens GN=LTF PE=1 SV=6 - [TRFL_HUMAN]
P01024	179	1663	187,0	6,4	1680	Complement C3 precursor [Contains: Complement C3 beta chain; Complement C
P98160	177	4391	468,5	6,51	1669	Basement membrane-specific heparan sulfate proteoglycan core protein OS=Hor
P02768	306	609	69,3	6,28	1448	Serum albumin precursor - Homo sapiens (Human) - [ALBU_HUMAN]
P04264	199	644	66,0	8,12	1389	Keratin, type II cytoskeletal 1 OS=Homo sapiens GN=KRT1 PE=1 SV=6 - [K2C1_HUM
Q8WZ42	271	34350	3813,8	6,34	1300	Titin OS=Homo sapiens GN=TTN PE=1 SV=2 - [TITIN_HUMAN]
P25311	540	298	34,2	6,05	1090	Zinc-alpha-2-glycoprotein OS=Homo sapiens GN=AZGP1 PE=1 SV=2 - [ZA2G_HUM
Q13421	102	630	68,9	6,38	845	Mesothelin OS=Homo sapiens GN=MSLN PE=1 SV=2 - [MSLN_HUMAN]
P35527	83	623	62,0	5,24	742	Keratin, type I cytoskeletal 9 OS=Homo sapiens GN=KRT9 PE=1 SV=3 - [K1C9_HUM
P01833	286	764	83,2	5,74	741	Polymeric immunoglobulin receptor OS=Homo sapiens GN=PIGR PE=1 SV=4 - [PIC
Q9UGM3	229	2413	260,6	5,44	718	Deleted in malignant brain tumors 1 protein OS=Homo sapiens GN=DMBT1 PE=1 S
P31025	706	176	19,2	5,58	704	Lipocalin-1 OS=Homo sapiens GN=LCN1 PE=1 SV=1 - [LCN1_HUMAN]
P00450	82	1065	122,1	5,72	697	Ceruloplasmin OS=Homo sapiens GN=CP PE=1 SV=1 - [CERU_HUMAN]
P10909	145	449	52,5	6,27	679	Clusterin OS=Homo sapiens GN=CLU PE=1 SV=1 - [CLUS_HUMAN]
P01876	710	353	37,6	6,51	615	Ig alpha-1 chain C region OS=Homo sapiens GN=IGHA1 PE=1 SV=2 - [IGHA1_HUMA
P60709	97	375	41,7	5,48	609	Actin, cytoplasmic 1 (Beta-actin) - Homo sapiens (Human) - [ACTB_HUMAN]
P01871	83	452	49,3	6,77	563	Ig mu chain C region OS=Homo sapiens GN=IGHM PE=1 SV=3 - [IGHM_HUMAN]
P00738	97	406	45,2	6,58	550	Haptoglobin OS=Homo sapiens GN=HP PE=1 SV=1 - [HPT_HUMAN]
P01877	580	340	36,5	6,1	550	Ig alpha-2 chain C region OS=Homo sapiens GN=IGHA2 PE=1 SV=3 - [IGHA2_HUMA
P62737	79	377	42,0	5,39	514	Actin, aortic smooth muscle (Alpha-actin-2) - Mus musculus (Mouse) - [ACTA_MC
Q96DA0	213	208	22,7	7,39	474	Zymogen granule protein 16 homolog B OS=Homo sapiens GN=ZG16B PE=1 SV=3 -
P01009	23	418	46,7	5,59	460	Alpha-1-antitrypsin OS=Homo sapiens GN=SERPINA1 PE=1 SV=3 - [A1AT_HUMAN]
P61626	959	148	16,5	9,16	460	Lysozyme C precursor (EC 3.2.1.17) (1,4-beta-N-acetylmuramidase C) - Homo sapi
Q08380	54	585	65,3	5,27	447	Galectin-3-binding protein OS=Homo sapiens GN=LGALS3BP PE=1 SV=1 - [LG3BP_I
P13645	68	584	58,8	5,21	443	Keratin, type I cytoskeletal 10 OS=Homo sapiens GN=KRT10 PE=1 SV=6 - [K1C10_H
P04083	25	346	38,7	7,02	428	Annexin A1 OS=Homo sapiens GN=ANXA1 PE=1 SV=2 - [ANXA1_HUMAN]
P12273	288	146	16,6	8,05	422	Prolactin-inducible protein OS=Homo sapiens GN=PIP PE=1 SV=1 - [PIP_HUMAN]
P20061	83	433	48,2	5,03	416	Transcobalamin-1 OS=Homo sapiens GN=TCN1 PE=1 SV=2 - [TCO1_HUMAN]
Q9GZ28	498	138	14,2	5,5	415	Extracellular glycoprotein lacritin OS=Homo sapiens GN=LACRT PE=1 SV=1 - [LACR
P55058	37	493	54,7	7,01	412	Phospholipid transfer protein OS=Homo sapiens GN=PLTP PE=1 SV=1 - [PLTP_HUM
P35908	49	639	65,4	8	408	Keratin, type II cytoskeletal 2 epidermal OS=Homo sapiens GN=KRT2 PE=1 SV=2 -
P02647	36	267	30,8	5,76	398	Apolipoprotein A-I precursor (Apo-AI) (ApoA-I) [Contains: Apolipoprotein A-I(-I-
P01623	96	109	11,7	8,91	376	Ig kappa chain V-III region WOL OS=Homo sapiens PE=1 SV=1 - [KV305_HUMAN]
Q8NF91	53	8797	1010,4	5,53	363	Nesprin-1 OS=Homo sapiens GN=SYNE1 PE=1 SV=3 - [SYNE1_HUMAN]
P01036	189	141	16,2	5,02	354	Cystatin-S precursor (Cystatin-4) (Salivary acidic protein 1) (Cystatin-SA-III) - Hom
Q8I2T6	111	3477	409,5	10,45	344	Abnormal spindle-like microcephaly-associated protein OS=Homo sapiens GN=A
Q03001	43	7570	860,1	5,25	337	Dystonin OS=Homo sapiens GN=DST PE=1 SV=4 - [DYST_HUMAN]
P80188	62	198	22,6	8,91	334	Neutrophil gelatinase-associated lipocalin OS=Homo sapiens GN=LCN2 PE=1 SV=.
P06733	42	434	47,1	7,39	334	Alpha-enolase (EC 4.2.1.11) (2-phospho-D-glycerate hydro-lyase) (Non-neural en
P80303	29	420	50,2	5,12	324	Nucleobindin-2 OS=Homo sapiens GN=NUCB2 PE=1 SV=2 - [NUCB2_HUMAN]
POCG05	192	106	11,3	7,24	317	Ig lambda-2 chain C regions OS=Homo sapiens GN=IGLC2 PE=1 SV=1 - [LAC2_HUM
O75556	326	95	10,9	5,78	306	Mammaglobin-B OS=Homo sapiens GN=SCGB2A1 PE=1 SV=1 - [SG2A1_HUMAN]
Q658J3	37	1075	121,3	6,2	304	POTE ankyrin domain family member E OS=Homo sapiens GN=POTEE PE=1 SV=3 -
P04259	25	564	60,0	8	302	Keratin, type II cytoskeletal 6B OS=Homo sapiens GN=KRT6B PE=1 SV=5 - [K2C6B_
ASA3E0	33	1075	121,4	6,2	301	POTE ankyrin domain family member F OS=Homo sapiens GN=POTEF PE=1 SV=2 -
Q15149	66	4684	531,5	5,96	293	Plectin OS=Homo sapiens GN=PLEC PE=1 SV=3 - [PLEC_HUMAN]
O15078	92	2479	290,2	5,95	289	Centrosomal protein of 290 kDa OS=Homo sapiens GN=CEP290 PE=1 SV=2 - [CE290
P01011	27	423	47,6	5,52	283	Alpha-1-antichymotrypsin OS=Homo sapiens GN=SERPINA3 PE=1 SV=2 - [AACT_H
P20929	52	6669	772,4	9,07	278	Nebulin OS=Homo sapiens GN=NEB PE=1 SV=4 - [NEBU_HUMAN]
Q99935	149	248	27,2	10,42	274	Proline-rich protein 1 OS=Homo sapiens GN=PROL1 PE=1 SV=2 - [PROL1_HUMAN]
Q16378	472	134	15,1	7,06	272	Proline-rich protein 4 OS=Homo sapiens GN=PRR4 PE=1 SV=3 - [PROL4_HUMAN]
Q9UQ35	36	2752	299,4	12,06	272	Serine/arginine repetitive matrix protein 2 OS=Homo sapiens GN=SRRM2 PE=1 SV
Q8IVF4	76	4471	514,5	5,88	266	Dynein heavy chain 10, axonemal OS=Homo sapiens GN=DNAH10 PE=1 SV=4 - [DY
Q06830	14	199	22,1	8,13	264	Peroxiredoxin-1 (EC 1.11.1.15) (Thioredoxin peroxidase 2) (Thioredoxin-depende
P15924	53	2871	331,6	6,81	263	Desmoplakin OS=Homo sapiens GN=DSP PE=1 SV=3 - [DESP_HUMAN]
Q562R1	55	376	42,0	5,59	257	Beta-actin-like protein 2 OS=Homo sapiens GN=ACTBL2 PE=1 SV=2 - [ACTBL_HUM
POCG38	56	1075	121,2	6,21	256	POTE ankyrin domain family member I OS=Homo sapiens GN=POTEI PE=3 SV=1 - [
Q5VSP4	218	162	17,9	5	251	Putative lipocalin 1-like protein 1 OS=Homo sapiens GN=LCN1L1 PE=5 SV=1 - [LC1
Q8WXH0	32	6885	795,9	5,36	250	Nesprin-2 OS=Homo sapiens GN=SYNE2 PE=1 SV=3 - [SYNE2_HUMAN]
P49792	67	3224	358,0	6,2	246	E3 SUMO-protein ligase RanBP2 OS=Homo sapiens GN=RANBP2 PE=1 SV=2 - [RBP2
P04406	21	335	36,0	8,46	245	Glyceraldehyde-3-phosphate dehydrogenase OS=Homo sapiens GN=GAPDH PE=1
Q8WXI7	48	22152	2351,2	6	245	Mucin-16 OS=Homo sapiens GN=MUC16 PE=1 SV=2 - [MUC16_HUMAN]
P01619	78	108	11,6	9,25	239	Ig kappa chain V-III region B6 OS=Homo sapiens PE=1 SV=1 - [KV301_HUMAN]
A0M8Q6	104	106	11,3	8,28	236	Ig lambda-7 chain C region OS=Homo sapiens GN=IGLC7 PE=1 SV=2 - [LAC7_HUMA

Appendices

Q12955	64	4377	480,1	6,49	234	Ankyrin-3 OS=Homo sapiens GN=ANK3 PE=1 SV=3 - [ANK3_HUMAN]
P01625	40	114	12,6	7,93	225	Ig kappa chain V-IV region Len OS=Homo sapiens PE=1 SV=2 - [KV402_HUMAN]
P13647	15	590	62,3	7,74	225	Keratin, type II cytoskeletal 5 OS=Homo sapiens GN=KRT5 PE=1 SV=3 - [K2C5_HUN
Q8NCM8	160	4307	492,3	6,54	223	Cytoplasmic dynein 2 heavy chain 1 OS=Homo sapiens GN=DYNC2H1 PE=1 SV=4 - I
P25054	112	2843	311,5	7,8	221	Adenomatous polyposis coli protein OS=Homo sapiens GN=APC PE=1 SV=2 - [APC
Q9Y4A5	32	3859	437,3	8,19	220	Transformation/transcription domain-associated protein (350/400 kDa PCAF-assc
Q02224	22	2701	316,2	5,64	218	Centromere-associated protein E OS=Homo sapiens GN=CENPE PE=1 SV=2 - [CENI
P26447	31	101	11,7	6,11	214	Protein S100-A4 OS=Homo sapiens GN=S100A4 PE=1 SV=1 - [S10A4_HUMAN]
O95968	171	90	9,9	9,25	214	Secretoglobin family 1D member 1 OS=Homo sapiens GN=SCGB1D1 PE=1 SV=1 - [S
P46100	28	2492	282,4	6,58	212	Transcriptional regulator ATRX OS=Homo sapiens GN=ATRX PE=1 SV=5 - [ATRX_HU
P06702	39	114	13,2	6,13	211	Protein S100-A9 OS=Homo sapiens GN=S100A9 PE=1 SV=1 - [S10A9_HUMAN]
O75592	21	4640	509,8	7,03	209	Probable E3 ubiquitin-protein ligase MYCBP2 OS=Homo sapiens GN=MYCBP2 PE=
Q9P225	107	4427	507,4	6,37	209	Dynein heavy chain 2, axonemal OS=Homo sapiens GN=DNAH2 PE=1 SV=3 - [DYH2
P01834	224	106	11,6	5,87	207	Ig kappa chain C region - Homo sapiens (Human) - [KAC_HUMAN]
Q9Y6V0	44	5183	566,3	6,54	202	Protein piccolo OS=Homo sapiens GN=PCLO PE=1 SV=3 - [PCLO_HUMAN]
P01598	25	108	11,8	8,44	199	Ig kappa chain V-I region EU OS=Homo sapiens PE=1 SV=1 - [KV106_HUMAN]
Q01484	45	3924	430,0	5,12	199	Ankyrin-2 OS=Homo sapiens GN=ANK2 PE=1 SV=3 - [ANK2_HUMAN]
POCG04	76	106	11,3	7,87	197	Ig lambda-1 chain C regions OS=Homo sapiens GN=IGLC1 PE=1 SV=1 - [LAC1_HUM
POC6X7	57	7073	789,7	6,62	195	Replicase polyprotein 1ab OS=Human SARS coronavirus GN=rep PE=1 SV=1 - [RIA
O95613	20	3336	377,8	5,55	194	Precentrin OS=Homo sapiens GN=PCNT PE=1 SV=4 - [PCNT_HUMAN]
P01781	55	116	12,7	8,48	191	Ig heavy chain V-III region GAL OS=Homo sapiens PE=1 SV=1 - [HV320_HUMAN]
P21817	49	5038	564,8	5,3	191	Ryanodine receptor 1 OS=Homo sapiens GN=RYR1 PE=1 SV=3 - [RYR1_HUMAN]
P01593	48	108	12,0	5,99	189	Ig kappa chain V-I region AG - Homo sapiens (Human) - [KV101_HUMAN]
P01591	87	159	18,1	5,24	188	Immunoglobulin J chain OS=Homo sapiens GN=IGJ PE=1 SV=4 - [IGJ_HUMAN]
Q5TZA2	25	2017	228,4	5,5	187	Rootletin OS=Homo sapiens GN=CROCC PE=1 SV=1 - [CROCC_HUMAN]
Q92736	47	4967	564,2	6,07	184	Ryanodine receptor 2 OS=Homo sapiens GN=RYR2 PE=1 SV=3 - [RYR2_HUMAN]
Q12789	22	2109	238,7	7,3	184	General transcription factor 3C polypeptide 1 OS=Homo sapiens GN=GTF3C1 PE=1
Q2LD37	25	5005	555,1	6,58	183	Uncharacterized protein KIAA1109 OS=Homo sapiens GN=KIAA1109 PE=1 SV=2 - [I
P09211	11	210	23,3	5,64	182	Glutathione S-transferase P OS=Homo sapiens GN=GSTP1 PE=1 SV=2 - [GSTP1_HU
P01857	16	330	36,1	8,19	182	Ig gamma-1 chain C region - Homo sapiens (Human) - [IGHG1_HUMAN]
A6NKT7	58	1766	198,3	6,39	181	RanBP2-like and GRIP domain-containing protein 3 OS=Homo sapiens GN=RGPD3
P08758	11	320	35,9	5,05	181	Annexin A5 OS=Homo sapiens GN=ANXA5 PE=1 SV=2 - [ANXA5_HUMAN]
Q5VST9	42	7968	867,9	5,99	181	Obscurin OS=Homo sapiens GN=OBSCN PE=1 SV=3 - [OBSCN_HUMAN]
P06313	35	133	14,6	6,58	181	Ig kappa chain V-IV region JI OS=Homo sapiens PE=4 SV=1 - [KV403_HUMAN]
Q09666	35	5890	628,7	6,15	180	Neuroblast differentiation-associated protein AHNK OS=Homo sapiens GN=AHN
Q9BYX7	25	375	42,0	6,33	177	Putative beta-actin-like protein 3 OS=Homo sapiens GN=POTEKP PE=5 SV=1 - [AC
P06703	43	90	10,2	5,48	177	Protein S100-A6 OS=Homo sapiens GN=S100A6 PE=1 SV=1 - [S10A6_HUMAN]
P00338	11	332	36,7	8,27	177	L-lactate dehydrogenase A chain OS=Homo sapiens GN=LDHA PE=1 SV=2 - [LDHA_
P63104	18	245	27,7	4,79	174	14-3-3 protein zeta/delta OS=Homo sapiens GN=YWHAZ PE=1 SV=1 - [1433Z_HUM
Q8TCU4	23	4167	460,7	6,28	172	Alstrom syndrome protein 1 OS=Homo sapiens GN=ALMS1 PE=1 SV=3 - [ALMS1_H
O95678	9	551	59,5	7,74	170	Keratin, type II cytoskeletal 75 OS=Homo sapiens GN=KRT75 PE=1 SV=2 - [K2C75_
P80748	30	111	11,9	5,08	169	Ig lambda chain V-III region LOI OS=Homo sapiens PE=1 SV=1 - [LV302_HUMAN]
Q96HP0	26	2047	229,4	6,74	169	Dedicator of cytokinesis protein 6 OS=Homo sapiens GN=DOCK6 PE=1 SV=3 - [DO
P02533	21	472	51,5	5,16	168	Keratin, type I cytoskeletal 14 OS=Homo sapiens GN=KRT14 PE=1 SV=4 - [K1C14_H
Q5XKE5	27	535	57,8	7,2	168	Keratin, type II cytoskeletal 79 OS=Homo sapiens GN=KRT79 PE=1 SV=2 - [K2C79_
P05109	40	93	10,8	7,03	167	Protein S100-A8 OS=Homo sapiens GN=S100A8 PE=1 SV=1 - [S10A8_HUMAN]
Q8IVL1	29	2488	268,0	9,03	166	Neuron navigator 2 OS=Homo sapiens GN=NAV2 PE=1 SV=3 - [NAV2_HUMAN]
Q4G0P3	67	5120	575,5	6,07	165	Hydrocephalus-inducing protein homolog OS=Homo sapiens GN=HYDIN PE=1 SV=
Q9UFH2	76	4485	511,5	5,77	165	Dynein heavy chain 17, axonemal OS=Homo sapiens GN=DNAH17 PE=2 SV=2 - [DY
Q14515	12	664	75,2	4,81	164	SPARC-like protein 1 OS=Homo sapiens GN=SPARCL1 PE=1 SV=2 - [SPRL1_HUMAN
Q5CZC0	34	6907	780,1	6,71	163	Fibrous sheath-interacting protein 2 OS=Homo sapiens GN=FSIP2 PE=1 SV=4 - [FSI
Q15782	17	390	43,5	7,53	162	Chitinase-3-like protein 2 OS=Homo sapiens GN=CHI3L2 PE=1 SV=1 - [CH3L2_HUM
O75122	74	1294	141,0	8,47	161	CLIP-associating protein 2 OS=Homo sapiens GN=CLASP2 PE=1 SV=2 - [CLAP2_HUN
Q5T4S7	28	5183	573,5	6,04	160	E3 ubiquitin-protein ligase UBR4 OS=Homo sapiens GN=UBR4 PE=1 SV=1 - [UBR4_
Q14643	26	2758	313,7	6,04	159	Inositol 1,4,5-trisphosphate receptor type 1 OS=Homo sapiens GN=ITPR1 PE=1 SV
Q96T23	21	1441	163,7	5,01	159	Remodeling and spacing factor 1 OS=Homo sapiens GN=RSF1 PE=1 SV=2 - [RSF1_H
A6NE01	34	2351	262,6	8,6	159	Protein FAM186A OS=Homo sapiens GN=FAM186A PE=2 SV=3 - [F186A_HUMAN]
P07339	35	412	44,5	6,54	158	Cathepsin D OS=Homo sapiens GN=CTSD PE=1 SV=1 - [CATD_HUMAN]
Q8N3K9	25	4069	448,9	4,78	157	Cardiomyopathy-associated protein 5 OS=Homo sapiens GN=CMYA5 PE=1 SV=3 - I
P01766	68	120	13,2	6,57	157	Ig heavy chain V-III region BRO OS=Homo sapiens PE=1 SV=1 - [HV305_HUMAN]
P61769	64	119	13,7	6,52	156	Beta-2-microglobulin OS=Homo sapiens GN=B2M PE=1 SV=1 - [B2MG_HUMAN]
Q5T200	21	1668	196,5	9,42	155	Zinc finger CCH domain-containing protein 13 OS=Homo sapiens GN=ZC3H13 PE=
P49454	13	3210	367,5	5,07	155	Centromere protein F OS=Homo sapiens GN=CENPF PE=1 SV=2 - [CENPF_HUMAN
Q9UMN6	34	2715	293,3	8,22	154	Histone-lysine N-methyltransferase MLL4 OS=Homo sapiens GN=WBP7 PE=1 SV=:
B1AJZ9	324	1412	161,8	6,95	154	Forkhead-associated domain-containing protein 1 OS=Homo sapiens GN=FHAD1

Appendices

P35580	17	1976	228,9	5,54	153	Myosin-10 OS=Homo sapiens GN=MYH10 PE=1 SV=3 - [MYH10_HUMAN]
O14578	59	2027	231,3	6,57	153	Citron Rho-interacting kinase OS=Homo sapiens GN=CITR PE=1 SV=2 - [CTRO_HUM]
Q9UIF8	52	2168	240,3	6,54	153	Bromodomain adjacent to zinc finger domain protein 2B OS=Homo sapiens GN=B
O95153	32	1857	199,9	5,11	151	Peripheral-type benzodiazepine receptor-associated protein 1 OS=Homo sapiens
Q9BXX2	18	1477	167,6	6,16	151	Ankyrin repeat domain-containing protein 30B OS=Homo sapiens GN=ANKRD30B
Q07864	36	2286	261,4	6,39	150	DNA polymerase epsilon catalytic subunit A OS=Homo sapiens GN=POLE PE=1 SV:
Q92614	66	2054	233,0	6,3	149	Myosin-XVIIIa OS=Homo sapiens GN=MYO18A PE=1 SV=3 - [MY18A_HUMAN]
P01594	33	108	11,9	5,33	149	Ig kappa chain V-I region AU OS=Homo sapiens PE=1 SV=1 - [KV102_HUMAN]
Q13029	23	1718	188,8	7,31	149	PR domain zinc finger protein 2 OS=Homo sapiens GN=PRDM2 PE=1 SV=3 - [PRDM
Q14152	44	1382	166,5	6,79	149	Eukaryotic translation initiation factor 3 subunit A OS=Homo sapiens GN=EIF3A PI
P01606	51	108	11,8	9,91	148	Ig kappa chain V-I region OU OS=Homo sapiens PE=1 SV=1 - [KV114_HUMAN]
Q14669	20	1992	220,3	8,48	147	Probable E3 ubiquitin-protein ligase TRIP12 OS=Homo sapiens GN=TRIP12 PE=1 S'
Q86VF7	30	1730	197,0	9,2	147	Nebulin-related-anchoring protein OS=Homo sapiens GN=NRAP PE=2 SV=2 - [NR/
P01596	24	107	11,7	9,41	147	Ig kappa chain V-I region CAR OS=Homo sapiens PE=1 SV=1 - [KV104_HUMAN]
Q13428	51	1488	152,0	9,04	145	Treacle protein OS=Homo sapiens GN=TCOF1 PE=1 SV=3 - [TCOF_HUMAN]
Q92817	26	2033	231,5	6,96	144	Envoplakin OS=Homo sapiens GN=EVPL PE=1 SV=3 - [EVPL_HUMAN]
Q9UIG0	60	1483	170,8	8,48	142	Tyrosine-protein kinase BAZ1B OS=Homo sapiens GN=BAZ1B PE=1 SV=2 - [BAZ1B_
Q8TE73	23	4624	528,7	6,1	142	Dynein heavy chain 5, axonemal OS=Homo sapiens GN=DNAH5 PE=1 SV=3 - [DYHE
P22079	11	712	80,2	8,62	140	Lactoperoxidase OS=Homo sapiens GN=LPO PE=1 SV=2 - [PERL_HUMAN]
P01768	60	122	13,7	9,63	140	Ig heavy chain V-III region CAM OS=Homo sapiens PE=1 SV=1 - [HV307_HUMAN]
A7E2Y1	21	1941	221,3	6,01	140	Myosin-7B OS=Homo sapiens GN=MYH7B PE=2 SV=3 - [MYH7B_HUMAN]
P80362	25	108	11,7	5,31	139	Ig kappa chain V-I region WAT OS=Homo sapiens PE=1 SV=1 - [KV125_HUMAN]
P24043	17	3122	343,7	6,4	138	Laminin subunit alpha-2 OS=Homo sapiens GN=LAMA2 PE=1 SV=4 - [LAMA2_HUM
O60673	17	3130	352,6	8,47	138	DNA polymerase zeta catalytic subunit OS=Homo sapiens GN=REV3L PE=1 SV=2 -]
P07858	10	339	37,8	6,3	137	Cathepsin B OS=Homo sapiens GN=CTSB PE=1 SV=3 - [CATB_HUMAN]
A2RUB6	47	948	109,3	8,25	137	Coiled-coil domain-containing protein 66 OS=Homo sapiens GN=CCDC66 PE=1 SV:
Q96Q89	20	1820	210,5	5,67	137	Kinesin-like protein KIF20B OS=Homo sapiens GN=KIF20B PE=1 SV=3 - [KIF20B_HUI
Q8WXG9	20	6307	692,2	4,64	137	G-protein coupled receptor 98 OS=Homo sapiens GN=GPR98 PE=1 SV=1 - [GPR98_
Q68DN6	48	1748	196,5	6,1	137	RANBP2-like and GRIP domain-containing protein 1/2 OS=Homo sapiens GN=RGP
Q725J4	46	1906	203,2	8,79	137	Retinoic acid-induced protein 1 OS=Homo sapiens GN=RAI1 PE=1 SV=2 - [RAI1_HL
Q5VT06	16	3117	350,7	6,33	136	Centrosome-associated protein 350 OS=Homo sapiens GN=CEP350 PE=1 SV=1 - [C
Q12923	18	2485	276,7	6,42	135	Tyrosine-protein phosphatase non-receptor type 13 OS=Homo sapiens GN=PTPN
P01607	26	108	11,9	6,51	135	Ig kappa chain V-I region Rei OS=Homo sapiens PE=1 SV=1 - [KV115_HUMAN]
Q8TD26	14	2715	305,2	6,27	135	Chromodomain-helicase-DNA-binding protein 6 OS=Homo sapiens GN=CHD6 PE=
Q14980	22	2115	238,1	5,78	135	Nuclear mitotic apparatus protein 1 OS=Homo sapiens GN=NUMA1 PE=1 SV=2 - [N
P01765	49	115	12,3	9,13	134	Ig heavy chain V-III region TIL - Homo sapiens (Human) - [HV304_HUMAN]
Q8IZD9	14	2030	233,0	6,98	134	Dedicator of cytokinesis protein 3 OS=Homo sapiens GN=DOCK3 PE=1 SV=1 - [DO
Q04695	12	432	48,1	5,02	134	Keratin, type I cytoskeletal 17 OS=Homo sapiens GN=KRT17 PE=1 SV=2 - [K1C17_H
O60293	33	1989	226,2	8,13	134	Zinc finger C3H1 domain-containing protein OS=Homo sapiens GN=ZFC3H1 PE=1 S
Q96SN8	74	1893	214,9	5,58	134	CDK5 regulatory subunit-associated protein 2 OS=Homo sapiens GN=CDK5RAP2 P
Q96PE2	28	2063	221,5	6,29	134	Rho guanine nucleotide exchange factor 17 OS=Homo sapiens GN=ARHGEF17 PE=
Q726E9	17	1792	201,4	9,64	133	E3 ubiquitin-protein ligase RBBP6 OS=Homo sapiens GN=RBBP6 PE=1 SV=1 - [RBB
Q9Y4F4	167	1720	189,2	8,5	132	Protein FAM179B OS=Homo sapiens GN=FAM179B PE=1 SV=4 - [F179B_HUMAN]
Q9Y4D8	64	3996	439,1	6,19	132	Probable E3 ubiquitin-protein ligase C12orf51 OS=Homo sapiens GN=C12orf51 PE
P07711	11	333	37,5	5,45	132	Cathepsin L1 OS=Homo sapiens GN=CTSL1 PE=1 SV=2 - [CATL1_HUMAN]
Q9BV73	31	2442	281,0	5,02	131	Centrosome-associated protein CEP250 OS=Homo sapiens GN=CEP250 PE=1 SV=2
P01624	32	109	11,9	8,94	131	Ig kappa chain V-III region POM OS=Homo sapiens PE=1 SV=1 - [KV306_HUMAN]
Q8N474	15	314	35,4	8,85	131	Secreted frizzled-related protein 1 OS=Homo sapiens GN=SFRP1 PE=1 SV=1 - [SFR
Q5VU65	13	1888	210,5	7,5	131	Nuclear pore membrane glycoprotein 210-like OS=Homo sapiens GN=NUP210L PI
P07602	11	524	58,1	5,17	130	Proactivator polypeptide OS=Homo sapiens GN=PSAP PE=1 SV=2 - [SAP_HUMAN]
Q7Z460	28	1538	169,3	9,03	130	CLIP-associating protein 1 OS=Homo sapiens GN=CLASP1 PE=1 SV=1 - [CLAP1_HU
P07355	12	339	38,6	7,75	129	Annexin A2 OS=Homo sapiens GN=ANXA2 PE=1 SV=2 - [ANXA2_HUMAN]
Q7Z406	11	1995	227,9	5,92	129	Myosin-14 OS=Homo sapiens GN=MYH14 PE=1 SV=1 - [MYH14_HUMAN]
Q8WZ64	25	1704	193,3	7,39	129	Arf-GAP with Rho-GAP domain, ANK repeat and PH domain-containing protein 2
P30414	76	1462	165,6	9,99	129	NK-tumor recognition protein OS=Homo sapiens GN=NKTR PE=1 SV=2 - [NKTR_HL
Q9H2D6	23	2365	261,2	8,48	128	TRIO and F-actin-binding protein OS=Homo sapiens GN=TRIOBP PE=1 SV=3 - [TAR
Q7Z7A1	19	2325	268,7	5,55	128	Centriolin OS=Homo sapiens GN=CEP110 PE=1 SV=2 - [CE110_HUMAN]
Q14966	47	1978	220,5	6,38	128	Zinc finger protein 638 OS=Homo sapiens GN=ZNF638 PE=1 SV=2 - [ZN638_HUMA
P11717	24	2491	274,2	5,94	127	Cation-independent mannose-6-phosphate receptor OS=Homo sapiens GN=IGF2
Q2KHM9	16	967	109,3	7,68	127	Uncharacterized protein KIAA0753 OS=Homo sapiens GN=KIAA0753 PE=2 SV=3 - [I
Q86TE4	81	346	38,9	8,73	127	Leucine zipper protein 2 OS=Homo sapiens GN=LUZP2 PE=2 SV=2 - [LUZP2_HUMA
Q14004	18	1512	164,8	9,69	127	Cyclin-dependent kinase 13 OS=Homo sapiens GN=CDK13 PE=1 SV=2 - [CDK13_HL
Q63HN8	15	3280	373,7	6,92	126	RING finger protein 213 OS=Homo sapiens GN=RNF213 PE=1 SV=2 - [RN213_HUM
Q9Y2K3	11	1946	224,5	5,85	126	Myosin-15 OS=Homo sapiens GN=MYH15 PE=1 SV=5 - [MYH15_HUMAN]
Q9NR99	50	2828	312,0	8,32	126	Matrix-remodeling-associated protein 5 OS=Homo sapiens GN=MXRA5 PE=2 SV=3

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Q14571	29	2701	307,9	6,43	125	Inositol 1,4,5-trisphosphate receptor type 2 OS=Homo sapiens GN=ITPR2 PE=1 SV=1
Q72794	31	578	61,9	5,99	125	Keratin, type II cytoskeletal 1b OS=Homo sapiens GN=KRT77 PE=1 SV=3 - [K2C1B_HUMAN]
P01597	54	108	11,7	9,36	125	Ig kappa chain V-I region DEE OS=Homo sapiens PE=1 SV=1 - [KV105_HUMAN]
Q62Q06	12	2873	333,0	7,28	124	WD repeat-containing protein 87 OS=Homo sapiens GN=WDR87 PE=2 SV=3 - [WDR87_HUMAN]
Q86UR5	22	1692	189,0	9,66	124	Regulating synaptic membrane exocytosis protein 1 OS=Homo sapiens GN=RIMS1
P12270	23	2363	267,1	5,02	124	Nucleoprotein TPR OS=Homo sapiens GN=TPR PE=1 SV=3 - [TPR_HUMAN]
Q9NS87	19	1388	160,1	6	122	Kinesin-like protein KIF15 OS=Homo sapiens GN=KIF15 PE=1 SV=1 - [KIF15_HUMAN]
Q96AY4	12	2481	270,7	6,89	122	Tetratricopeptide repeat protein 28 OS=Homo sapiens GN=TTC28 PE=1 SV=4 - [TTC28_HUMAN]
P01767	48	115	12,4	9,25	122	Ig heavy chain V-III region BUT OS=Homo sapiens PE=1 SV=1 - [HV306_HUMAN]
O00192	35	962	104,6	6,81	122	Armadillo repeat protein deleted in velo-cardio-facial syndrome OS=Homo sapiens GN=VCF1
P06309	25	117	12,7	7,12	122	Ig kappa chain V-II region GM607 (Fragment) OS=Homo sapiens PE=4 SV=1 - [KV2C07_HUMAN]
Q9Y2F5	30	2266	247,7	5,48	121	Uncharacterized protein KIAA0947 OS=Homo sapiens GN=KIAA0947 PE=1 SV=5 - [KIAA0947_HUMAN]
P20930	20	4061	434,9	9,25	121	Filaggrin OS=Homo sapiens GN=FLG PE=1 SV=3 - [FLA_HUMAN]
O00584	9	256	29,5	7,08	121	Ribonuclease T2 OS=Homo sapiens GN=RNASET2 PE=1 SV=2 - [RNT2_HUMAN]
Q13402	23	2215	254,2	8,56	120	Myosin-VIIa OS=Homo sapiens GN=MYO7A PE=1 SV=1 - [MYO7A_HUMAN]
P01617	25	113	12,3	6	120	Ig kappa chain V-II region TEW OS=Homo sapiens PE=1 SV=1 - [KV204_HUMAN]
Q9UKJ3	26	1502	164,1	8,66	120	G patch domain-containing protein 8 OS=Homo sapiens GN=GPATCH8 PE=1 SV=2 - [GPATCH8_HUMAN]
Q13129	72	1914	217,8	6,77	120	Zinc finger protein Rlf OS=Homo sapiens GN=RLF PE=1 SV=2 - [RLF_HUMAN]
Q9H583	11	2144	242,2	6,54	119	HEAT repeat-containing protein 1 OS=Homo sapiens GN=HEATR1 PE=1 SV=3 - [HEATR1_HUMAN]
Q9P2P6	31	4614	506,4	6,46	118	STAR-related lipid transfer protein 9 OS=Homo sapiens GN=STAR9 PE=2 SV=2 - [STAR9_HUMAN]
P35498	35	2009	228,8	5,81	118	Sodium channel protein type 1 subunit alpha OS=Homo sapiens GN=SCN1A PE=1 SV=1 - [SCN1A_HUMAN]
Q96JB1	22	4490	514,3	6,32	118	Dynein heavy chain 8, axonemal OS=Homo sapiens GN=DNAH8 PE=1 SV=2 - [DNAH8_HUMAN]
Q8WXG6	123	1647	183,2	6,04	118	MAP kinase-activating death domain protein OS=Homo sapiens GN=MADD PE=1 SV=1 - [MADD_HUMAN]
O60244	24	1454	160,5	8,73	117	Mediator of RNA polymerase II transcription subunit 14 OS=Homo sapiens GN=MTF14
Q5THJ4	40	4387	491,5	6,58	117	Vacuolar protein sorting-associated protein 13D OS=Homo sapiens GN=VPS13D
Q8TF72	21	1996	216,7	7,8	116	Protein Shroom3 OS=Homo sapiens GN=SHROOM3 PE=1 SV=2 - [SHRM3_HUMAN]
O95359	37	2948	309,2	4,79	116	Transforming acidic coiled-coil-containing protein 2 OS=Homo sapiens GN=TACC2
Q9UQE7	11	1217	141,5	7,18	116	Structural maintenance of chromosomes protein 3 OS=Homo sapiens GN=SMC3
O14490	56	977	108,8	7,08	116	Disks large-associated protein 1 OS=Homo sapiens GN=DLGAP1 PE=1 SV=1 - [DLGAP1_HUMAN]
Q9P2D3	19	2071	224,2	7,17	116	HEAT repeat-containing protein 5B OS=Homo sapiens GN=HEATR5B PE=1 SV=2 - [HEATR5B_HUMAN]
P33992	15	734	82,2	8,37	115	DNA replication licensing factor MCM5 OS=Homo sapiens GN=MCM5 PE=1 SV=5 - [MCM5_HUMAN]
Q9HAU5	11	1272	147,7	5,69	115	Regulator of nonsense transcripts 2 OS=Homo sapiens GN=UPF2 PE=1 SV=1 - [UPF2_HUMAN]
P16591	29	822	94,6	7,14	113	Proto-oncogene tyrosine-protein kinase FER (EC 2.7.10.2) (p94-FER) (c-FER) (Tyro:FER) OS=Homo sapiens GN=FER PE=1 SV=1 - [SHR2_HUMAN]
Q13796	16	1616	176,3	7,09	113	Protein Shroom2 OS=Homo sapiens GN=SHROOM2 PE=1 SV=1 - [SHRM2_HUMAN]
O15417	13	2968	314,3	8,7	112	Trinucleotide repeat-containing gene 18 protein OS=Homo sapiens GN=TNRC18
Q8NCU4	10	936	110,5	9,6	112	Coiled-coil domain-containing protein KIAA1407 OS=Homo sapiens GN=KIAA1407
Q9Y496	19	699	80,0	6,54	111	Kinesin-like protein KIF3A OS=Homo sapiens GN=KIF3A PE=1 SV=4 - [KIF3A_HUMAN]
Q6WRI0	22	2623	290,7	9,13	111	Immunoglobulin superfamily member 10 OS=Homo sapiens GN=IGSF10 PE=1 SV=1 - [IGSF10_HUMAN]
Q96L93	36	1317	151,9	6,16	111	Kinesin-like protein KIF16B OS=Homo sapiens GN=KIF16B PE=1 SV=2 - [KIF16B_HUMAN]
P12036	53	1026	112,4	6,18	111	Neurofilament heavy polypeptide OS=Homo sapiens GN=NEFH PE=1 SV=4 - [NEFH_HUMAN]
Q8NB90	24	893	97,8	5,66	110	Spermatogenesis-associated protein 5 OS=Homo sapiens GN=SPATA5 PE=1 SV=3 - [SPATA5_HUMAN]
O75747	116	1445	165,6	6,93	110	Phosphatidylinositol-4-phosphate 3-kinase C2 domain-containing subunit gamma OS=Homo sapiens GN=PIP3K2C2
Q9Y2W1	20	955	108,6	10,15	109	Thyroid hormone receptor-associated protein 3 OS=Homo sapiens GN=THRAP3
Q9H254	20	2564	288,8	6,01	109	Spectrin beta chain, brain 3 OS=Homo sapiens GN=SPTBN4 PE=1 SV=2 - [SPTN4_HUMAN]
Q8IWI2	12	1684	195,8	5,14	109	GRIP and coiled-coil domain-containing protein 2 OS=Homo sapiens GN=GCC2 PE=1 SV=1 - [GCC2_HUMAN]
Q13576	30	1575	180,5	5,64	108	Ras GTPase-activating-like protein IQGAP2 OS=Homo sapiens GN=IQGAP2 PE=1 SV=1 - [IQGAP2_HUMAN]
P01859	9	326	35,9	7,59	108	Ig gamma-2 chain C region OS=Homo sapiens GN=IGHG2 PE=1 SV=2 - [IGHG2_HUMAN]
Q15772	21	3267	354,1	8,51	108	Striated muscle preferentially expressed protein kinase OS=Homo sapiens GN=SMEL1
Q726G8	24	1249	138,1	6,37	108	Ankyrin repeat and sterile alpha motif domain-containing protein 1B OS=Homo sapiens GN=ANKRD1B
Q6AWC2	18	1192	133,8	5,53	108	Protein WWC2 OS=Homo sapiens GN=WWC2 PE=1 SV=2 - [WWC2_HUMAN]
Q07954	15	4544	504,3	5,39	107	Pro-low-density lipoprotein receptor-related protein 1 OS=Homo sapiens GN=LRF1
Q72736	28	793	85,3	7,83	107	Pleckstrin homology domain-containing family H member 3 OS=Homo sapiens GN=PHLNC3
Q92616	18	2671	292,6	7,47	106	Translational activator GCN1 OS=Homo sapiens GN=GCN1L1 PE=1 SV=6 - [GCN1L1_HUMAN]
O60281	22	2723	304,6	7,39	106	Zinc finger protein 292 OS=Homo sapiens GN=ZNF292 PE=1 SV=3 - [ZNF292_HUMAN]
Q8NFU7	26	2136	235,2	8,24	106	Methylcytosine dioxygenase TET1 OS=Homo sapiens GN=TET1 PE=1 SV=2 - [TET1_HUMAN]
Q02779	57	954	103,6	7,01	106	Mitogen-activated protein kinase kinase kinase 10 OS=Homo sapiens GN=MAP3K10
Q5VYK3	116	1845	204,2	7,12	106	Proteasome-associated protein ECM29 homolog OS=Homo sapiens GN=ECM29
Q6TFL3	23	1326	152,7	6,81	106	Uncharacterized protein C9orf93 OS=Homo sapiens GN=C9orf93 PE=2 SV=1 - [C9orf93_HUMAN]
Q9COD4	23	1074	119,5	9,45	105	Zinc finger protein 518B OS=Homo sapiens GN=ZNF518B PE=2 SV=2 - [ZNF518B_HUMAN]
P42695	17	1498	168,8	7,5	105	Condensin-2 complex subunit D3 OS=Homo sapiens GN=NCAPD3 PE=1 SV=2 - [NCAPD3_HUMAN]
Q92878	18	1312	153,8	6,89	104	DNA repair protein RAD50 OS=Homo sapiens GN=RAD50 PE=1 SV=1 - [RAD50_HUMAN]
P01764	46	117	12,6	8,28	104	Ig heavy chain V-III region VH26 OS=Homo sapiens PE=1 SV=1 - [HV303_HUMAN]
Q15147	57	1175	134,4	6,9	104	1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase beta-4 OS=Homo sapiens GN=PLD4
Q727B0	24	1213	138,0	8,32	104	Filamin-A-interacting protein 1 OS=Homo sapiens GN=FLILP1 PE=1 SV=1 - [FLILP1_HUMAN]
Q8N8E3	32	955	112,7	6,68	103	Coiled-coil domain-containing protein 46 OS=Homo sapiens GN=CCDC46 PE=2 SV=1 - [CCDC46_HUMAN]

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Q494V2	18	611	71,1	7,11	103	Coiled-coil domain-containing protein 37 OS=Homo sapiens GN=CCDC37 PE=1 SV=
Q16651	8	343	36,4	5,85	103	Prostasin OS=Homo sapiens GN=PRSS8 PE=1 SV=1 - [PRSS8_HUMAN]
P31949	20	105	11,7	7,12	103	Protein S100-A11 (S100 calcium-binding protein A11) (Protein S100C) (Calgizzarin
Q8N9B5	19	988	111,4	6,18	103	Junction-mediating and -regulatory protein OS=Homo sapiens GN=JMY PE=1 SV=2
Q96JM4	18	1722	199,2	6,19	103	Leucine-rich repeat and IQ domain-containing protein 1 OS=Homo sapiens GN=Lf
Q9NPG3	20	1134	121,4	9,33	102	Ubiquitin-1 OS=Homo sapiens GN=UBN1 PE=1 SV=2 - [UBN1_HUMAN]
O75165	27	2243	254,3	6,74	102	DnaJ homolog subfamily C member 13 OS=Homo sapiens GN=DNAJC13 PE=1 SV=5
Q9NSD9	11	589	66,1	6,84	102	Phenylalanyl-tRNA synthetase beta chain OS=Homo sapiens GN=FARSB PE=1 SV=
Q9H6R4	24	1146	127,5	7,64	102	Nucleolar protein 6 OS=Homo sapiens GN=NOL6 PE=1 SV=2 - [NOL6_HUMAN]
P04792	10	205	22,8	6,4	102	Heat-shock protein beta-1 (HspB1) (Heat shock 27 kDa protein) (HSP 27) (Stress-r
Q8IY37	14	1157	129,5	8,1	102	Probable ATP-dependent RNA helicase DHX37 OS=Homo sapiens GN=DHX37 PE=
P12035	23	628	64,4	6,48	102	Keratin, type II cytoskeletal 3 OS=Homo sapiens GN=KRT3 PE=1 SV=3 - [K2C3_HUN
O14727	46	1248	141,7	6,4	102	Apoptotic protease-activating factor 1 OS=Homo sapiens GN=APAF1 PE=1 SV=2 - [
Q14687	16	1217	136,1	7,74	101	Genetic suppressor element 1 OS=Homo sapiens GN=GSE1 PE=1 SV=3 - [GSE1_HU
Q29RF7	14	1337	150,7	7,91	101	Sister chromatid cohesion protein PDS5 homolog A OS=Homo sapiens GN=PDS5A
Q9NPF5	29	467	53,0	9,5	101	DNA methyltransferase 1-associated protein 1 OS=Homo sapiens GN=DMAP1 PE=
Q4L180	10	1135	130,3	6,57	101	Filamin A-interacting protein 1-like OS=Homo sapiens GN=FLIP1L PE=1 SV=2 - [FI
Q5T2S8	32	1044	115,6	7,77	101	Armadillo repeat-containing protein 4 OS=Homo sapiens GN=ARMC4 PE=2 SV=1 -
Q8ND23	70	1372	150,1	7,44	101	Leucine-rich repeat-containing protein 16B OS=Homo sapiens GN=LRRCL16B PE=2
Q5TCY1	84	1321	142,6	5,6	100	Tau-tubulin kinase 1 OS=Homo sapiens GN=TTBK1 PE=1 SV=2 - [TTBK1_HUMAN]
O95477	9	2261	254,1	6,86	100	ATP-binding cassette sub-family A member 1 OS=Homo sapiens GN=ABCA1 PE=1
Q96JP5	35	570	63,4	7,36	100	E3 ubiquitin-protein ligase ZFP91 OS=Homo sapiens GN=ZFP91 PE=1 SV=1 - [ZFP9
Q96DT5	34	4523	520,7	6,44	100	Dynein heavy chain 11, axonemal OS=Homo sapiens GN=DNAH11 PE=1 SV=3 - [DY
P35579	17	1960	226,4	5,6	100	Myosin-9 OS=Homo sapiens GN=MYH9 PE=1 SV=4 - [MYH9_HUMAN]
Q14566	156	821	92,8	5,41	100	DNA replication licensing factor MCM6 OS=Homo sapiens GN=MCM6 PE=1 SV=1 -
Q8IWK6	43	1321	146,1	8,48	99	Probable G-protein coupled receptor 125 OS=Homo sapiens GN=GPR125 PE=1 SV=
P05787	40	483	53,7	5,59	98	Keratin, type II cytoskeletal 8 OS=Homo sapiens GN=KRT8 PE=1 SV=7 - [K2C8_HUN
Q9NZM1	21	2061	234,6	6,18	98	Myoferlin OS=Homo sapiens GN=MYOF PE=1 SV=1 - [MYOF_HUMAN]
P78344	24	907	102,3	7,14	98	Eukaryotic translation initiation factor 4 gamma 2 OS=Homo sapiens GN=EIF4G2 P
P01616	16	112	12,0	9,29	98	Ig kappa chain V-II region MIL OS=Homo sapiens PE=1 SV=1 - [KV203_HUMAN]
Q9BRK5	9	362	41,8	4,86	98	45 kDa calcium-binding protein OS=Homo sapiens GN=SDF4 PE=1 SV=1 - [CAB45_
Q8WVW8	30	1544	169,7	7,08	97	Arf-GAP with Rho-GAP domain, ANK repeat and PH domain-containing protein 3
Q01546	9	638	65,8	8,12	97	Keratin, type II cytoskeletal 2 oral OS=Homo sapiens GN=KRT76 PE=1 SV=2 - [K22C
Q9Y4D1	10	1078	123,4	7,23	97	Dishevelled-associated activator of morphogenesis 1 OS=Homo sapiens GN=DAAI
Q7L7X3	11	1001	116,0	7,65	97	Serine/threonine-protein kinase TAO1 OS=Homo sapiens GN=TAO1 PE=1 SV=1 -
Q8TBY8	15	1022	119,0	6,29	97	Polyamine-modulated factor 1-binding protein 1 OS=Homo sapiens GN=PMFBP1
P01621	49	100	10,7	6,52	97	Ig kappa chain V-III region NG9 (Fragment) OS=Homo sapiens PE=1 SV=1 - [KV303
P01037	62	141	16,4	7,21	97	Cystatin-SN OS=Homo sapiens GN=CST1 PE=1 SV=3 - [CYTN_HUMAN]
O75151	17	1096	120,7	9,17	97	PHD finger protein 2 OS=Homo sapiens GN=PHF2 PE=1 SV=4 - [PHF2_HUMAN]
Q6VAB6	42	950	107,6	8,69	96	Kinase suppressor of Ras 2 OS=Homo sapiens GN=KSR2 PE=1 SV=2 - [KSR2_HUMAN]
Q8N9T8	36	709	83,2	5,17	96	Protein KRI1 homolog OS=Homo sapiens GN=KRI1 PE=1 SV=2 - [KRI1_HUMAN]
Q9NTI5	12	1447	164,6	8,47	96	Sister chromatid cohesion protein PDS5 homolog B OS=Homo sapiens GN=PDS5B
Q8WVW7	26	1642	186,4	6,95	96	ATP-binding cassette sub-family A member 5 OS=Homo sapiens GN=ABCA5 PE=2
Q96RV3	7	2341	258,5	7,21	96	Pecanex-like protein 1 OS=Homo sapiens GN=PCNX PE=2 SV=2 - [PCX1_HUMAN]
Q9UHB6	17	759	85,2	6,84	96	LIM domain and actin-binding protein 1 OS=Homo sapiens GN=LIMA1 PE=1 SV=1 -
Q92797	80	1274	141,1	6,13	95	Symplekin OS=Homo sapiens GN=SYMPK PE=1 SV=2 - [SYMPK_HUMAN]
Q8IYB3	25	904	102,3	11,84	95	Serine/arginine repetitive matrix protein 1 OS=Homo sapiens GN=SRRM1 PE=1 SV=
Q14997	13	1843	211,2	6,9	95	Proteasome activator complex subunit 4 OS=Homo sapiens GN=PSME4 PE=1 SV=2
Q96SB8	10	1091	126,2	6,99	95	Structural maintenance of chromosomes protein 6 OS=Homo sapiens GN=SMC6 P
Q7Z3E2	14	898	103,6	6,27	95	Uncharacterized protein C10orf118 OS=Homo sapiens GN=C10orf118 PE=2 SV=2 - [
P09104	5	434	47,2	5,03	95	Gamma-enolase (EC 4.2.1.11) (2-phospho-D-glycerate hydro-lyase) (Neural enol
P23468	21	1912	214,6	6,57	95	Receptor-type tyrosine-protein phosphatase delta OS=Homo sapiens GN=PTPRD
Q6ZV29	11	1317	145,6	7,74	95	Patatin-like phospholipase domain-containing protein 7 OS=Homo sapiens GN=P
Q5TH69	37	2177	240,5	5,82	94	Brefeldin A-inhibited guanine nucleotide-exchange protein 3 OS=Homo sapiens
Q9UPU5	26	2620	294,2	6,14	94	Ubiquitin carboxyl-terminal hydrolase 24 OS=Homo sapiens GN=USP24 PE=1 SV=3
Q9BYW2	31	2564	287,4	6,14	94	Histone-lysine N-methyltransferase SETD2 OS=Homo sapiens GN=SETD2 PE=1 SV=
Q9HBT8	12	521	60,1	8,32	94	Zinc finger protein 286A OS=Homo sapiens GN=ZNF286A PE=2 SV=1 - [Z286A_HUM
O60437	23	1756	204,6	5,6	94	Periplakin OS=Homo sapiens GN=PPL PE=1 SV=4 - [PEPL_HUMAN]
Q8IWX3	30	2542	269,3	5,73	94	Ankyrin repeat and KH domain-containing protein 1 OS=Homo sapiens GN=ANKH
P14555	49	144	16,1	9,23	94	Phospholipase A2, membrane associated OS=Homo sapiens GN=PLA2G2A PE=1 SV=
P06396	6	782	85,6	6,28	94	Gelsolin OS=Homo sapiens GN=GSN PE=1 SV=1 - [GELS_HUMAN]
Q9BQS8	10	1478	166,9	4,92	94	FYVE and coiled-coil domain-containing protein 1 OS=Homo sapiens GN=FYCO1 P
P31946	12	246	28,1	4,83	94	14-3-3 protein beta/alpha OS=Homo sapiens GN=YWHAB PE=1 SV=3 - [1433B_HUN
Q5RHP9	16	1530	168,4	4,88	93	Uncharacterized protein C1orf173 OS=Homo sapiens GN=C1orf173 PE=2 SV=1 - [C/
O14802	14	1390	155,5	8,48	93	DNA-directed RNA polymerase III subunit RPC1 OS=Homo sapiens GN=POLR3A P

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Q8NA31	30	727	83,0	7,46	93	Coiled-coil domain-containing protein 79 OS=Homo sapiens GN=CCDC79 PE=2 SV=2
Q9HCE1	12	1003	113,6	8,82	93	Putative helicase MOV-10 OS=Homo sapiens GN=MOV10 PE=1 SV=2 - [MOV10_HL
Q9Y5S2	46	1711	194,2	6,37	93	Serine/threonine-protein kinase MRCK beta OS=Homo sapiens GN=CDC42BPB PE
O75923	12	2080	237,1	5,64	92	Dysferlin OS=Homo sapiens GN=DYSF PE=1 SV=1 - [DYSF_HUMAN]
O75417	36	1762	197,5	7,01	92	DNA polymerase theta OS=Homo sapiens GN=POLQ PE=1 SV=1 - [DPOLQ_HUMAN
Q6N069	73	864	101,4	7,87	92	N-alpha-acetyltransferase 16, NatA auxiliary subunit OS=Homo sapiens GN=NAA:
P53367	23	373	41,7	6,7	92	Arfaptin-1 OS=Homo sapiens GN=ARFIP1 PE=1 SV=2 - [ARFP1_HUMAN]
Q99490	59	1192	124,6	9,89	92	Centaurin-gamma 1 (ARF-GAP with GTP-binding protein-like, ankyrin repeat and
P02787	73	698	77,0	7,12	92	Serotransferrin OS=Homo sapiens GN=TF PE=1 SV=3 - [TRFE_HUMAN]
Q5VT97	11	1116	124,9	8,15	92	Rho GTPase-activating protein SYDE2 OS=Homo sapiens GN=SYDE2 PE=1 SV=1 - [S
O75691	11	2785	318,2	7,39	92	Small subunit processome component 20 homolog OS=Homo sapiens GN=UTP20
Q8TDY2	15	1594	183,0	5,41	92	RB1-inducible coiled-coil protein 1 OS=Homo sapiens GN=RB1CC1 PE=1 SV=3 - [RE
Q9NRA8	15	985	108,1	8,32	92	Eukaryotic translation initiation factor 4E transporter OS=Homo sapiens GN=EIF4E
Q13393	12	1074	124,1	8,78	92	Phospholipase D1 OS=Homo sapiens GN=PLD1 PE=1 SV=1 - [PLD1_HUMAN]
O43166	10	1804	199,9	8,19	92	Signal-induced proliferation-associated 1-like protein 1 OS=Homo sapiens GN=SI
Q8IYE0	13	955	112,7	8,48	91	Coiled-coil domain-containing protein 146 OS=Homo sapiens GN=CCDC146 PE=2 S
P11137	34	1827	199,4	4,91	91	Microtubule-associated protein 2 OS=Homo sapiens GN=MAP2 PE=1 SV=4 - [MAP
Q2M296	20	383	42,1	8,73	91	Methenyltetrahydrofolate synthase domain-containing protein OS=Homo sapier
Q2TAC6	10	998	111,3	8,69	91	Kinesin-like protein KIF19 OS=Homo sapiens GN=KIF19 PE=2 SV=2 - [KIF19_HUMA
Q9NYF0	45	836	90,1	8,69	91	Dapper homolog 1 OS=Homo sapiens GN=DACT1 PE=2 SV=2 - [DACT1_HUMAN]
Q5VSL9	20	837	95,5	6,29	90	Protein FAM40A OS=Homo sapiens GN=FAM40A PE=1 SV=1 - [FA40A_HUMAN]
Q86WJ1	39	897	100,9	6,9	90	Chromodomain-helicase-DNA-binding protein 1-like OS=Homo sapiens GN=CHD:
Q9Y4I1	79	1855	215,3	8,48	90	Myosin-Va OS=Homo sapiens GN=MYOSA PE=1 SV=2 - [MYOSA_HUMAN]
P68104	7	462	50,1	9,01	89	Elongation factor 1-alpha 1 OS=Homo sapiens GN=EEF1A1 PE=1 SV=1 - [EF1A1_HU
P08069	25	1367	154,7	5,8	89	Insulin-like growth factor 1 receptor OS=Homo sapiens GN=IGF1R PE=1 SV=1 - [IG
Q9Y2K7	11	1162	132,7	7,58	89	Lysine-specific demethylase 2A OS=Homo sapiens GN=KDM2A PE=1 SV=3 - [KDM:
P31146	25	461	51,0	6,68	89	Coronin-1A OS=Homo sapiens GN=CORO1A PE=1 SV=4 - [COR1A_HUMAN]
O60229	28	2985	340,0	6,07	88	Kalirin OS=Homo sapiens GN=KALRN PE=1 SV=2 - [KALRN_HUMAN]
Q96JG9	12	3925	409,9	7,72	88	Zinc finger protein 469 OS=Homo sapiens GN=ZNF469 PE=1 SV=3 - [ZN469_HUMAN]
Q5VUA4	13	2279	251,0	7,2	88	Zinc finger protein 318 OS=Homo sapiens GN=ZNF318 PE=1 SV=2 - [ZN318_HUMAN]
P13533	12	1939	223,6	5,73	87	Myosin-6 OS=Homo sapiens GN=MYH6 PE=1 SV=5 - [MYH6_HUMAN]
P08727	12	400	44,1	5,14	87	Keratin, type I cytoskeletal 19 OS=Homo sapiens GN=KRT19 PE=1 SV=4 - [K1C19_H
Q9Y2G9	50	1366	150,2	6,52	87	Protein strawberry notch homolog 2 OS=Homo sapiens GN=SBNO2 PE=2 SV=3 - [S
Q96MR6	10	1250	144,9	5,8	87	WD repeat-containing protein 65 OS=Homo sapiens GN=WDR65 PE=2 SV=3 - [WDR
Q6KC79	34	2804	315,9	7,91	87	Nipped-B-like protein OS=Homo sapiens GN=NIPBL PE=1 SV=2 - [NIPBL_HUMAN]
Q13618	20	768	88,9	8,48	87	Cullin-3 OS=Homo sapiens GN=CUL3 PE=1 SV=2 - [CUL3_HUMAN]
Q96QB1	46	1528	170,5	6,4	86	Rho GTPase-activating protein 7 OS=Homo sapiens GN=DLC1 PE=1 SV=4 - [RHG07_
Q13233	10	1512	164,4	7,74	86	Mitogen-activated protein kinase kinase kinase 1 OS=Homo sapiens GN=MAP3K1
Q8TDN4	26	633	67,6	9,17	85	CDK5 and ABL1 enzyme substrate 1 OS=Homo sapiens GN=CABLES1 PE=1 SV=2 - [C
Q9Y3M8	17	1113	124,9	7,02	85	STAR-related lipid transfer protein 13 OS=Homo sapiens GN=STARD13 PE=1 SV=2 -
Q9NXH9	11	659	72,2	7,64	85	N(2),N(2)-dimethylguanosine tRNA methyltransferase OS=Homo sapiens GN=TRI
Q8NG31	8	2342	265,2	5,47	85	Protein CASC5 OS=Homo sapiens GN=CASC5 PE=1 SV=3 - [CASC5_HUMAN]
Q9ULV0	79	1848	213,5	7,2	85	Myosin-Vb OS=Homo sapiens GN=MYO5B PE=1 SV=3 - [MYO5B_HUMAN]
Q9ULJ8	12	1098	123,3	5,1	85	Neurabin-1 OS=Homo sapiens GN=PPP1R9A PE=1 SV=2 - [NEB1_HUMAN]
Q9H582	98	1327	149,5	8,16	85	Zinc finger protein 644 OS=Homo sapiens GN=ZNF644 PE=1 SV=2 - [ZN644_HUMAN]
Q68D51	11	928	106,8	8,48	85	DENN domain-containing protein 2C OS=Homo sapiens GN=DENND2C PE=2 SV=2 -
Q7Z2Y8	20	2422	278,9	6,55	85	Interferon-induced very large GTPase 1 OS=Homo sapiens GN=GVIN1 PE=2 SV=2 -
Q149N8	11	1683	193,0	7,47	85	E3 ubiquitin-protein ligase SHPRH OS=Homo sapiens GN=SHPRH PE=1 SV=2 - [SHP
O14777	7	642	73,9	5,6	85	Kinetochores protein NDC80 homolog OS=Homo sapiens GN=NDC80 PE=1 SV=1 - [I
Q9UPZ9	100	632	71,4	9,77	84	Serine/threonine-protein kinase ICK OS=Homo sapiens GN=ICK PE=1 SV=1 - [ICK_
Q9NYC9	45	4486	511,6	5,91	84	Dynein heavy chain 9, axonemal OS=Homo sapiens GN=DNAH9 PE=1 SV=3 - [DYHE
Q9P2L0	34	1181	133,5	6,38	84	WD repeat-containing protein 35 OS=Homo sapiens GN=WDR35 PE=2 SV=3 - [WDR
O15021	79	2626	284,2	8,62	84	Microtubule-associated serine/threonine-protein kinase 4 OS=Homo sapiens GN
Q7Z3Y7	9	464	50,5	5,47	84	Keratin, type I cytoskeletal 28 OS=Homo sapiens GN=KRT28 PE=1 SV=2 - [K1C28_H
P12883	10	1935	223,0	5,8	84	Myosin-7 OS=Homo sapiens GN=MYH7 PE=1 SV=5 - [MYH7_HUMAN]
Q9ULD4	12	1205	135,7	6,58	84	Bromodomain and PHD finger-containing protein 3 OS=Homo sapiens GN=BRPF3
P01700	21	112	11,9	8,91	84	Ig lambda chain V-I region HA OS=Homo sapiens PE=1 SV=1 - [LV102_HUMAN]
A6NKB5	52	2137	237,1	6,76	84	Pecanex-like protein 2 OS=Homo sapiens GN=PCNXL2 PE=1 SV=3 - [PCX2_HUMAN
P08582	72	738	80,2	6,01	84	Melanotransferrin OS=Homo sapiens GN=MF12 PE=1 SV=1 - [TRFM_HUMAN]
P01611	8	108	11,6	7,28	84	Ig kappa chain V-I region Wes OS=Homo sapiens PE=1 SV=1 - [KV119_HUMAN]
Q9P212	20	2302	258,6	6,48	83	1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase epsilon-1 OS=Homc
Q14722	14	419	46,5	8,94	83	Voltage-gated potassium channel subunit beta-1 OS=Homo sapiens GN=KCNAB1
A6NCI4	13	1184	133,9	8,46	83	von Willebrand factor A domain-containing protein 3A OS=Homo sapiens GN=VW
Q7Z4L5	8	1316	150,8	6,96	83	Tetratricopeptide repeat protein 21B OS=Homo sapiens GN=TTCT21B PE=2 SV=2 - [
Q9H251	28	3354	369,3	4,67	83	Cadherin-23 OS=Homo sapiens GN=CDH23 PE=1 SV=1 - [CAD23_HUMAN]

Appendices

Q9C0H6	9	718	80,2	6,86	83	Kelch-like protein 4 OS=Homo sapiens GN=KLHL4 PE=2 SV=2 - [KLHL4_HUMAN]
Q9NRZ9	78	838	97,0	7,93	83	Lymphoid-specific helicase OS=Homo sapiens GN=HELLS PE=1 SV=1 - [HELLS_HUM
Q9NQ76	44	525	58,4	8,57	83	Matrix extracellular phosphoglycoprotein OS=Homo sapiens GN=MEPE PE=1 SV=1
Q6PCB5	16	846	94,8	8,78	83	Round spermatid basic protein 1-like protein OS=Homo sapiens GN=RSNB11 PE=1
Q9H611	17	641	69,8	9,72	83	ATP-dependent DNA helicase PIF1 OS=Homo sapiens GN=PIF1 PE=1 SV=2 - [PIF1_
P52209	5	483	53,1	7,23	83	6-phosphogluconate dehydrogenase, decarboxylating OS=Homo sapiens GN=PGI
Q9NQX4	10	1742	202,7	7,71	82	Myosin-Vc OS=Homo sapiens GN=MYO5C PE=1 SV=2 - [MYO5C_HUMAN]
Q8NEC7	43	633	71,0	7,78	82	Glutathione S-transferase C-terminal domain-containing protein OS=Homo sapie
Q03111	19	559	62,0	8,59	82	Protein ENL OS=Homo sapiens GN=MLLT1 PE=1 SV=2 - [ENL_HUMAN]
Q9Y6A5	34	838	90,3	5,05	82	Transforming acidic coiled-coil-containing protein 3 OS=Homo sapiens GN=TACC3
Q9Y566	16	2161	224,8	8,15	82	SH3 and multiple ankyrin repeat domains protein 1 OS=Homo sapiens GN=SHANK
Q8N3T6	16	1108	121,7	6,35	82	Transmembrane protein 132C OS=Homo sapiens GN=TMEM132C PE=2 SV=3 - [T13
Q9C0F0	49	2248	241,8	6,14	82	Putative Polycomb group protein ASXL3 OS=Homo sapiens GN=ASXL3 PE=2 SV=3 -
Q8WV44	10	630	71,6	5,06	82	E3 ubiquitin-protein ligase TRIM41 OS=Homo sapiens GN=TRIM41 PE=1 SV=3 - [TR
Q09161	10	790	91,8	6,43	82	Nuclear cap-binding protein subunit 1 OS=Homo sapiens GN=NCBP1 PE=1 SV=1 -
P49815	17	1807	200,5	7,31	82	Tuberin OS=Homo sapiens GN=TSC2 PE=1 SV=2 - [TSC2_HUMAN]
Q9NW08	19	1133	127,7	8,5	82	DNA-directed RNA polymerase III subunit RPC2 OS=Homo sapiens GN=POLR3B PE
Q66K14	16	1250	140,4	5,25	82	TBC1 domain family member 9B OS=Homo sapiens GN=TBC1D9B PE=1 SV=3 - [TBC
Q9BXX3	15	1397	158,7	6,48	82	Ankyrin repeat domain-containing protein 30A OS=Homo sapiens GN=ANKRD30A
Q9H5P4	6	517	55,6	8,92	81	PDZ domain-containing protein 7 OS=Homo sapiens GN=PDZD7 PE=1 SV=1 - [PDZI
P14136	14	432	49,8	5,52	81	Glial fibrillary acidic protein OS=Homo sapiens GN=GFAP PE=1 SV=1 - [GFAP_HUM
Q14684	12	758	84,4	9,76	81	Ribosomal RNA processing protein 1 homolog B OS=Homo sapiens GN=RRP1B PE=
Q9Y4C0	24	1643	180,5	5,54	81	Neurexin-3-alpha OS=Homo sapiens GN=NRXN3 PE=2 SV=4 - [NRX3A_HUMAN]
Q9BWS9	16	393	44,9	8,63	81	Chitinase domain-containing protein 1 OS=Homo sapiens GN=CHID1 PE=1 SV=1 -
Q05D60	49	604	70,9	6,21	81	Coiled-coil domain-containing protein 67 OS=Homo sapiens GN=CCDC67 PE=2 SV=
Q9HAU0	18	1116	127,4	7,53	81	Pleckstrin homology domain-containing family A member 5 OS=Homo sapiens GI
P20807	21	821	94,2	6,18	81	Calpain-3 OS=Homo sapiens GN=CAPN3 PE=1 SV=2 - [CAN3_HUMAN]
Q9C093	32	1822	209,7	5,54	81	Sperm flagellar protein 2 OS=Homo sapiens GN=SPEF2 PE=1 SV=2 - [SPEF2_HUMA
Q6ZU35	14	1233	136,7	5,6	80	Uncharacterized protein KIAA1211 OS=Homo sapiens GN=KIAA1211 PE=1 SV=3 - [I
Q9P227	25	1491	162,1	9,19	80	Rho GTPase-activating protein 23 OS=Homo sapiens GN=ARGAP23 PE=1 SV=2 - [I
Q9HCM1	15	1747	194,7	8,78	80	Uncharacterized protein C12orf35 OS=Homo sapiens GN=C12orf35 PE=1 SV=3 - [C1
Q75037	25	1637	182,5	7,08	80	Kinesin-like protein KIF21B OS=Homo sapiens GN=KIF21B PE=1 SV=2 - [KI21B_HU
Q8NEG5	13	633	72,7	8,62	80	E3 ubiquitin-protein ligase ZSWIM2 OS=Homo sapiens GN=ZSWIM2 PE=1 SV=2 - [z
Q9ULI0	12	1458	164,8	6,8	80	ATPase family AAA domain-containing protein 2B OS=Homo sapiens GN=ATAD2B
P14618	3	531	57,9	7,84	80	Pyruvate kinase isozymes M1/M2 (EC 2.7.1.40) (Pyruvate kinase muscle isozyme)
Q9BY89	12	1806	196,6	8,47	80	Uncharacterized protein KIAA1671 OS=Homo sapiens GN=KIAA1671 PE=1 SV=2 - [I
P42345	8	2549	288,7	7,17	80	Serine/threonine-protein kinase mTOR OS=Homo sapiens GN=MTOR PE=1 SV=1 -
Q70CQ2	25	3546	404,0	5,82	80	Ubiquitin carboxyl-terminal hydrolase 34 OS=Homo sapiens GN=USP34 PE=1 SV=2
Q14667	20	2235	253,5	7,14	80	UPF0378 protein KIAA0100 OS=Homo sapiens GN=KIAA0100 PE=1 SV=3 - [K0100_H
Q5VZK9	28	1371	151,5	7,85	80	Leucine-rich repeat-containing protein 16A OS=Homo sapiens GN=LRRC16A PE=1
Q9NQ38	30	1064	120,6	8,06	80	Serine protease inhibitor Kazal-type 5 OS=Homo sapiens GN=SPINK5 PE=1 SV=2 -
P01702	22	111	11,4	4,89	80	Ig lambda chain V-1 region NIG-64 OS=Homo sapiens PE=1 SV=1 - [LV104_HUMAN]
P29728	13	719	82,4	8,25	79	2'-5'-oligoadenylate synthase 2 OS=Homo sapiens GN=OAS2 PE=1 SV=3 - [OAS2_H
P02549	7	2419	279,8	5,05	79	Spectrin alpha chain, erythrocyte OS=Homo sapiens GN=SPTA1 PE=1 SV=5 - [SPTA
Q96028	7	1365	152,2	8,69	79	Probable histone-lysine N-methyltransferase NSD2 OS=Homo sapiens GN=WHSC
P04433	14	115	12,6	4,96	79	Ig kappa chain V-III region VG (Fragment) OS=Homo sapiens PE=1 SV=1 - [KV309_f
Q9NSB2	75	600	64,8	7,56	79	Keratin, type II cuticular Hb4 OS=Homo sapiens GN=KRT84 PE=1 SV=2 - [KRT84_HU
Q2M2Z5	11	673	75,1	5,62	79	Centrosomal protein kizuna OS=Homo sapiens GN=PLK1S1 PE=1 SV=2 - [KIZ_HUM
Q15678	11	1187	135,2	8,31	79	Tyrosine-protein phosphatase non-receptor type 14 OS=Homo sapiens GN=PTPN
Q9H1D0	11	725	83,2	7,62	79	Transient receptor potential cation channel subfamily V member 6 (TrpV6) (Epitf
Q9BZJ0	18	848	100,4	8	79	Crooked neck-like protein 1 OS=Homo sapiens GN=CRNKL1 PE=1 SV=4 - [CRNL1_H
Q8TCU6	11	1659	186,1	6,44	79	Phosphatidylinositol 3,4,5-trisphosphate-dependent Rac exchanger 1 protein OS
P19013	7	534	57,2	6,61	78	Keratin, type II cytoskeletal 4 OS=Homo sapiens GN=KRT4 PE=1 SV=4 - [K2C4_HUN
P78539	14	464	51,5	8,66	78	Sushi repeat-containing protein SRPX OS=Homo sapiens GN=SRPX PE=2 SV=1 - [Sf
Q12774	8	1597	176,7	5,53	78	Rho guanine nucleotide exchange factor 5 OS=Homo sapiens GN=ARHGEF5 PE=1 S
Q8NFT6	24	615	67,2	8,34	78	Protein DBF4 homolog B OS=Homo sapiens GN=DBF4B PE=1 SV=1 - [DBF4B_HUMA
Q6YHU6	8	1953	219,5	6,06	78	Thyroid adenoma-associated protein OS=Homo sapiens GN=THADA PE=1 SV=1 - [
Q05519	24	484	53,5	10,52	78	Serine/arginine-rich splicing factor 11 OS=Homo sapiens GN=SRSF11 PE=1 SV=1 -
Q5TCS8	15	1911	221,3	5,01	78	Adenylate kinase domain-containing protein 1 OS=Homo sapiens GN=AKD1 PE=1
P16157	11	1881	206,1	6,01	77	Ankyrin-1 OS=Homo sapiens GN=ANK1 PE=1 SV=3 - [ANK1_HUMAN]
Q9Y666	22	1083	119,0	6,71	77	Solute carrier family 12 member 7 OS=Homo sapiens GN=SLC12A7 PE=1 SV=3 - [S1
P01034	3	146	15,8	8,75	77	Cystatin-C OS=Homo sapiens GN=CST3 PE=1 SV=1 - [CYTC_HUMAN]
Q9UC06	28	446	50,8	8,31	77	Zinc finger protein 70 OS=Homo sapiens GN=ZNF70 PE=2 SV=2 - [ZNF70_HUMAN]
P25705	14	553	59,7	9,13	77	ATP synthase subunit alpha, mitochondrial OS=Homo sapiens GN=ATP5A1 PE=1 S
Q8IZJ3	17	1885	206,6	6,42	77	C3 and PZP-like alpha-2-macroglobulin domain-containing protein 8 OS=Homo sa

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Q8WXR4	6	1341	151,7	8,15	77	Myosin-IIb OS=Homo sapiens GN=MYO3B PE=2 SV=4 - [MYO3B_HUMAN]
P12107	9	1806	181,0	5,17	77	Collagen alpha-1(XI) chain OS=Homo sapiens GN=COL11A1 PE=1 SV=4 - [COBA1_HUMAN]
P13929	16	434	46,9	7,71	77	Beta-enolase OS=Homo sapiens GN=ENO3 PE=1 SV=4 - [ENOB_HUMAN]
O95803	17	873	100,8	8,06	77	Bifunctional heparan sulfate N-deacetylase/N-sulfotransferase 3 OS=Homo sapiens GN=HSPG3 PE=1 SV=3 - [HSPG3_HUMAN]
P23634	11	1241	137,8	6,6	77	Plasma membrane calcium-transporting ATPase 4 OS=Homo sapiens GN=ATP2B4 PE=1 SV=1 - [ATP2B4_HUMAN]
Q5CZ79	12	823	93,9	8,32	77	Ankyrin repeat domain-containing protein 20B OS=Homo sapiens GN=ANKRD20B PE=1 SV=1 - [ANKRD20B_HUMAN]
Q8NAP3	7	1195	134,2	8,03	77	Zinc finger and BTB domain-containing protein 38 OS=Homo sapiens GN=ZBTB38 PE=1 SV=1 - [ZBTB38_HUMAN]
Q9Y6D9	8	718	83,0	5,92	77	Mitotic spindle assembly checkpoint protein MAD1 OS=Homo sapiens GN=MAD1 PE=1 SV=1 - [MAD1_HUMAN]
Q8N157	20	1196	137,0	7,12	77	Joubertin OS=Homo sapiens GN=AH11 PE=1 SV=1 - [AH11_HUMAN]
Q86V15	103	1759	189,9	7,03	77	Zinc finger protein castor homolog 1 OS=Homo sapiens GN=CASZ1 PE=2 SV=4 - [CASZ1_HUMAN]
Q8TBF8	21	365	42,0	9,04	76	Protein FAM81A OS=Homo sapiens GN=FAM81A PE=2 SV=1 - [FAM81A_HUMAN]
Q6ZRQ5	21	1243	142,2	7,12	76	Protein MMS22-like OS=Homo sapiens GN=MMS22L PE=1 SV=3 - [MMS22_HUMAN]
Q13395	49	1621	181,6	7,05	76	Probable methyltransferase TARBP1 OS=Homo sapiens GN=TARBP1 PE=1 SV=1 - [TARBP1_HUMAN]
Q8NCA5	48	519	55,4	9,03	76	Protein FAM98A OS=Homo sapiens GN=FAM98A PE=1 SV=1 - [FAM98A_HUMAN]
Q9HC62	25	589	67,8	9,48	76	Sentrin-specific protease 2 OS=Homo sapiens GN=SEN2 PE=1 SV=3 - [SEN2_HUMAN]
Q86SQ7	24	713	82,6	5,81	75	Serologically defined colon cancer antigen 8 OS=Homo sapiens GN=SDCCAG8 PE=1 SV=1 - [SDCCAG8_HUMAN]
O95185	22	931	103,1	6,1	75	Netrin receptor UNC5C OS=Homo sapiens GN=UNC5C PE=1 SV=2 - [UNC5C_HUMAN]
Q86VH2	10	1401	160,2	7,28	75	Kinesin-like protein KIF27 OS=Homo sapiens GN=KIF27 PE=2 SV=1 - [KIF27_HUMAN]
Q8WW11	7	1683	192,6	8,09	75	LIM domain only protein 7 OS=Homo sapiens GN=LMO7 PE=1 SV=3 - [LMO7_HUMAN]
Q9BQI9	89	281	31,3	8,4	75	Nuclear receptor-interacting protein 2 OS=Homo sapiens GN=NRIP2 PE=2 SV=3 - [NRIP2_HUMAN]
O00257	52	560	61,3	9,36	75	E3 SUMO-protein ligase CBX4 OS=Homo sapiens GN=CBX4 PE=1 SV=3 - [CBX4_HUMAN]
Q9Y6X9	44	1032	117,7	8,38	75	MORC family CW-type zinc finger protein 2 OS=Homo sapiens GN=MORC2 PE=1 SV=1 - [MORC2_HUMAN]
Q5JU85	6	1478	161,6	8,56	75	IQ motif and SEC7 domain-containing protein 2 OS=Homo sapiens GN=IQSEC2 PE=1 SV=1 - [IQSEC2_HUMAN]
O43548	10	720	80,7	6,44	75	Protein-glutamine gamma-glutamyltransferase 5 OS=Homo sapiens GN=TGM5 PE=1 SV=1 - [TGM5_HUMAN]
Q9UPP5	21	1409	155,6	6,19	75	Uncharacterized protein KIAA1107 OS=Homo sapiens GN=KIAA1107 PE=1 SV=2 - [KIAA1107_HUMAN]
Q86VQ6	116	682	74,8	8,22	75	Thioredoxin reductase 3 (Fragment) OS=Homo sapiens GN=TXNRD3 PE=1 SV=3 - [TXNRD3_HUMAN]
O43272	9	516	59,2	6,89	75	Proline dehydrogenase, mitochondrial OS=Homo sapiens GN=PRODH PE=1 SV=2 - [PRODH_HUMAN]
Q0P6D6	15	951	110,4	6,47	75	Coiled-coil domain-containing protein 15 OS=Homo sapiens GN=CCDC15 PE=2 SV=1 - [CCDC15_HUMAN]
P07197	5	916	102,4	4,91	74	Neurofilament medium polypeptide OS=Homo sapiens GN=NEFM PE=1 SV=3 - [NEFM_HUMAN]
Q5KSL6	107	1271	141,7	5,53	74	Diacylglycerol kinase kappa OS=Homo sapiens GN=DGKK PE=1 SV=1 - [DGKK_HUMAN]
Q9H4I2	11	956	104,6	6,07	74	Zinc fingers and homeoboxes protein 3 OS=Homo sapiens GN=ZHX3 PE=1 SV=3 - [ZHX3_HUMAN]
O75164	12	1064	120,6	5,85	74	Lysine-specific demethylase 4A OS=Homo sapiens GN=KDM4A PE=1 SV=2 - [KDM4A_HUMAN]
Q9GRS0	10	853	96,6	4,94	74	Trimethylguanosine synthase OS=Homo sapiens GN=TGS1 PE=1 SV=3 - [TGS1_HUMAN]
P17040	11	1043	117,5	6,43	74	Zinc finger and SCAN domain-containing protein 20 OS=Homo sapiens GN=ZSCAN20 PE=1 SV=1 - [ZSCAN20_HUMAN]
Q86Y22	6	540	51,9	7,25	74	Collagen alpha-1(XVIII) chain OS=Homo sapiens GN=COL23A1 PE=1 SV=1 - [COL23A1_HUMAN]
Q9H0R1	17	490	54,7	6,61	74	MHD domain-containing death-inducing protein OS=Homo sapiens GN=MUDENG PE=1 SV=1 - [MUDENG_HUMAN]
Q1MSJ5	11	1256	145,4	6,8	74	Centrosome and spindle pole-associated protein 1 OS=Homo sapiens GN=CSPP1 PE=1 SV=1 - [CSPP1_HUMAN]
Q14678	9	1352	147,2	5,3	73	KN motif and ankyrin repeat domain-containing protein 1 OS=Homo sapiens GN=ANKRD1 PE=1 SV=1 - [ANKRD1_HUMAN]
Q92901	17	407	46,3	10,45	73	60S ribosomal protein L3-like OS=Homo sapiens GN=RPL3L PE=1 SV=3 - [RPL3L_HUMAN]
Q7L2E3	14	1194	133,9	8,78	73	Putative ATP-dependent RNA helicase DHX30 OS=Homo sapiens GN=DHX30 PE=1 SV=1 - [DHX30_HUMAN]
Q17R98	29	1081	119,1	6,89	73	Zinc finger protein 827 OS=Homo sapiens GN=ZNF827 PE=2 SV=1 - [ZNF827_HUMAN]
Q05397	35	1052	119,2	6,62	73	Focal adhesion kinase 1 OS=Homo sapiens GN=PTK2 PE=1 SV=2 - [FAK1_HUMAN]
Q9H3R0	11	1056	119,9	6,46	73	Lysine-specific demethylase 4C OS=Homo sapiens GN=KDM4C PE=1 SV=2 - [KDM4C_HUMAN]
O95453	6	639	73,4	6,2	73	Poly(A)-specific ribonuclease PARN OS=Homo sapiens GN=PARN PE=1 SV=1 - [PARN_HUMAN]
Q6UWX4	29	724	80,7	9,01	73	HHIP-like protein 2 OS=Homo sapiens GN=HHIP2 PE=2 SV=1 - [HHIP2_HUMAN]
O95071	33	2799	309,2	5,85	73	E3 ubiquitin-protein ligase UBR5 OS=Homo sapiens GN=UBR5 PE=1 SV=2 - [UBR5_HUMAN]
Q9Y285	9	508	57,5	7,8	73	Phenylalanyl-tRNA synthetase alpha chain OS=Homo sapiens GN=FARSA PE=1 SV=1 - [FARSA_HUMAN]
Q9Y6K9	93	419	48,2	5,71	73	NF-kappa-B essential modulator (NEMO) (NF-kappa-B essential modifier) (Inhibitor of I-kappaB kinase) OS=Homo sapiens GN=IKBKG PE=1 SV=1 - [IKBKG_HUMAN]
Q9HAV4	11	1204	136,2	5,8	73	Exportin-5 OS=Homo sapiens GN=XPO5 PE=1 SV=1 - [XPO5_HUMAN]
Q96T17	24	732	81,9	8,84	73	MAP7 domain-containing protein 2 OS=Homo sapiens GN=MAP7D2 PE=1 SV=2 - [MAP7D2_HUMAN]
Q86Z14	12	1044	119,7	9,22	73	Beta-klotho OS=Homo sapiens GN=KLB PE=1 SV=1 - [KLOTB_HUMAN]
Q96A65	39	974	110,4	6,49	73	Exocyst complex component 4 OS=Homo sapiens GN=EXOC4 PE=1 SV=1 - [EXOC4_HUMAN]
Q72478	24	1369	155,1	8,09	72	ATP-dependent RNA helicase DHX29 OS=Homo sapiens GN=DHX29 PE=1 SV=2 - [DHX29_HUMAN]
Q5VV67	18	1664	177,4	6,51	72	Peroxisome proliferator-activated receptor gamma coactivator-related protein 1 OS=Homo sapiens GN=PPARGC1B PE=1 SV=1 - [PPARGC1B_HUMAN]
Q502W7	33	563	65,3	8,75	72	Coiled-coil domain-containing protein 38 OS=Homo sapiens GN=CCDC38 PE=2 SV=1 - [CCDC38_HUMAN]
Q8NB25	15	1140	132,9	5,83	72	Protein FAM184A OS=Homo sapiens GN=FAM184A PE=2 SV=3 - [FAM184A_HUMAN]
Q9BXL6	20	1004	113,2	5,92	72	Caspase recruitment domain-containing protein 14 OS=Homo sapiens GN=CARD14 PE=1 SV=1 - [CARD14_HUMAN]
Q9P2K1	11	1620	186,1	6,74	72	Coiled-coil and C2 domain-containing protein 2A OS=Homo sapiens GN=CC2D2A PE=1 SV=1 - [CC2D2A_HUMAN]
Q9BPU6	15	564	61,4	7,2	72	Dihydropyrimidinase-related protein 5 OS=Homo sapiens GN=DPYSL5 PE=1 SV=1 - [DPYSL5_HUMAN]
P23458	49	1154	133,2	7,55	72	Tyrosine-protein kinase JAK1 OS=Homo sapiens GN=JAK1 PE=1 SV=2 - [JAK1_HUMAN]
P08922	15	2347	263,7	6,11	72	Proto-oncogene tyrosine-protein kinase ROS OS=Homo sapiens GN=ROS1 PE=2 SV=1 - [ROS1_HUMAN]
A4D1F6	29	860	98,0	7,46	72	Leucine-rich repeat and death domain-containing protein LOC401387 OS=Homo sapiens GN=LOC401387 PE=1 SV=1 - [LOC401387_HUMAN]
Q727G8	16	4022	448,4	6,46	72	Vacuolar protein sorting-associated protein 13B OS=Homo sapiens GN=VPS13B PE=1 SV=1 - [VPS13B_HUMAN]
Q9P2E9	13	1410	152,4	8,6	72	Ribosome-binding protein 1 OS=Homo sapiens GN=RRBP1 PE=1 SV=4 - [RRBP1_HUMAN]
Q9P2E3	17	1918	220,1	7,3	72	NFX1-type zinc finger-containing protein 1 OS=Homo sapiens GN=ZNF1 PE=1 SV=1 - [ZNF1_HUMAN]

Appendices

O95425	11	2214	247,6	6,98	72	Supervillin OS=Homo sapiens GN=SVIL PE=1 SV=2 - [SVIL_HUMAN]
Q07283	4	1943	253,8	5,78	71	Trichohyalin OS=Homo sapiens GN=TCHH PE=1 SV=2 - [TRHY_HUMAN]
P51825	12	1210	131,3	9,2	71	AF4/FMR2 family member 1 OS=Homo sapiens GN=AFF1 PE=1 SV=1 - [AFF1_HUM]
Q4W5G0	93	525	59,6	9	71	Tigger transposable element-derived protein 2 OS=Homo sapiens GN=TIGD2 PE=
Q9HCK8	16	2581	290,3	6,47	71	Chromodomain-helicase-DNA-binding protein 8 OS=Homo sapiens GN=CHD8 PE=
Q9C0D6	10	1143	124,7	9,03	71	FH2 domain-containing protein 1 OS=Homo sapiens GN=FHDC1 PE=2 SV=2 - [FHDC
Q86UK5	15	1308	147,9	6,96	71	Limbin OS=Homo sapiens GN=EVC2 PE=1 SV=1 - [LBN_HUMAN]
Q9NYB0	15	399	44,2	4,73	71	Telomeric repeat-binding factor 2-interacting protein 1 OS=Homo sapiens GN=TE
Q86WI1	16	4243	465,4	6,11	71	Fibrocystin-L OS=Homo sapiens GN=PKHD1L1 PE=2 SV=2 - [PKHL1_HUMAN]
A4FU69	7	1503	173,3	5,85	71	EF-hand calcium-binding domain-containing protein 5 OS=Homo sapiens GN=EFC
Q7Z5Q5	21	900	100,2	8,29	71	DNA polymerase nu OS=Homo sapiens GN=POLN PE=1 SV=2 - [DPOLN_HUMAN]
Q5VVJ2	110	828	95,0	5,53	71	Histone H2A deubiquitinase MYSM1 OS=Homo sapiens GN=MYSM1 PE=1 SV=1 - [M
Q08AN1	17	781	90,2	9,57	71	Zinc finger protein 616 OS=Homo sapiens GN=ZNF616 PE=2 SV=2 - [ZNF616_HUMA
Q81WN7	15	2480	261,0	4,41	71	Retinitis pigmentosa 1-like 1 protein OS=Homo sapiens GN=RP1L1 PE=1 SV=4 - [R]
Q53ET0	25	693	73,3	7,11	71	CREB-regulated transcription coactivator 2 OS=Homo sapiens GN=CRTC2 PE=1 SV=
P17480	6	764	89,4	5,81	71	Nucleolar transcription factor 1 OS=Homo sapiens GN=UBTF PE=1 SV=1 - [UBF1_H
Q9UPX8	14	1470	158,7	6,9	70	SH3 and multiple ankyrin repeat domains protein 2 OS=Homo sapiens GN=SHANP
Q9NRC6	9	3674	416,6	6,67	70	Spectrin beta chain, brain 4 OS=Homo sapiens GN=SPTBN5 PE=1 SV=1 - [SPTN5_H
Q58F21	13	947	107,9	8,95	70	Bromodomain testis-specific protein OS=Homo sapiens GN=BRDT1 PE=1 SV=4 - [B
O43306	6	1168	130,5	8,22	70	Adenylate cyclase type 6 OS=Homo sapiens GN=ADCY6 PE=1 SV=2 - [ADCY6_HUM
Q96JG6	12	964	111,1	6,2	70	Coiled-coil domain-containing protein 132 OS=Homo sapiens GN=CCDC132 PE=1 S
O15234	11	703	76,2	6,48	70	Protein CASC3 OS=Homo sapiens GN=CASC3 PE=1 SV=2 - [CASC3_HUMAN]
Q96CP6	68	724	80,6	6,74	70	GRAM domain-containing protein 1A OS=Homo sapiens GN=GRAMD1A PE=1 SV=2
Q8NFA0	7	1604	181,5	6,44	70	Ubiquitin carboxyl-terminal hydrolase 32 OS=Homo sapiens GN=USP32 PE=1 SV=1
Q9BZF1	70	889	101,1	6,96	70	Oxysterol-binding protein-related protein 8 OS=Homo sapiens GN=OSBPL8 PE=1 :
Q96J17	15	2443	278,7	5,97	70	Spatacsin OS=Homo sapiens GN=SPG11 PE=1 SV=3 - [SPTCS_HUMAN]
Q502W6	22	1294	145,7	7,33	70	von Willebrand factor A domain-containing protein 3B OS=Homo sapiens GN=VW
Q13217	4	504	57,5	6,15	70	DnaJ homolog subfamily C member 3 OS=Homo sapiens GN=DNAJC3 PE=1 SV=1 -
Q7Z3Z0	8	450	49,3	5,08	70	Keratin, type I cytoskeletal 25 OS=Homo sapiens GN=KRT25 PE=1 SV=1 - [K1C25_H
Q13535	41	2644	301,2	7,43	69	Serine/threonine-protein kinase ATR OS=Homo sapiens GN=ATR PE=1 SV=3 - [AT
P01763	39	114	12,2	8,5	69	Ig heavy chain V-III region WEA OS=Homo sapiens PE=1 SV=1 - [HV302_HUMAN]
Q96BP3	8	646	73,5	7,15	69	Peptidylprolyl isomerase domain and WD repeat-containing protein 1 (EC 5.2.1.8
Q9H9Y6	80	1135	128,1	7,83	69	DNA-directed RNA polymerase I subunit RPA2 OS=Homo sapiens GN=POLR1B PE=
P01033	4	207	23,2	8,1	69	Metalloproteinase inhibitor 1 OS=Homo sapiens GN=TIMP1 PE=1 SV=1 - [TIMP1_H
Q81WC1	12	876	98,4	9,32	69	MAP7 domain-containing protein 3 OS=Homo sapiens GN=MAP7D3 PE=1 SV=2 - [P
O94762	12	991	108,8	8,56	69	ATP-dependent DNA helicase Q5 OS=Homo sapiens GN=RECQL5 PE=1 SV=2 - [REC
Q9ULH0	51	1771	196,4	6,62	69	Kinase D-interacting substrate of 220 kDa OS=Homo sapiens GN=KIDINS220 PE=1 :
Q96M83	10	486	55,7	7,78	69	Coiled-coil domain-containing protein 7 OS=Homo sapiens GN=CCDC7 PE=2 SV=2
P01613	19	112	12,2	5,36	69	Ig kappa chain V-I region Ni - Homo sapiens (Human) - [KV121_HUMAN]
Q9H9B1	12	1267	138,2	5,94	69	Histone-lysine N-methyltransferase, H3 lysine-9 specific 5 OS=Homo sapiens GN=
P20591	9	662	75,5	5,83	69	Interferon-induced GTP-binding protein Mx1 OS=Homo sapiens GN=MX1 PE=1 SV
Q55RES	13	1749	195,9	6,73	69	Nucleoporin NUP188 homolog OS=Homo sapiens GN=NUP188 PE=1 SV=1 - [NUP18
Q6DJT9	10	500	55,9	8,56	69	Zinc finger protein PLAG1 OS=Homo sapiens GN=PLAG1 PE=1 SV=1 - [PLAG1_HUM
Q14C86	19	1478	164,9	5,22	68	GTPase-activating protein and VPS9 domain-containing protein 1 OS=Homo sapie
Q3ZCX4	50	644	74,3	8,22	68	Zinc finger protein 568 OS=Homo sapiens GN=ZNF568 PE=2 SV=2 - [ZNF568_HUMA
Q7Z3V4	10	1068	123,0	8,19	68	Ubiquitin-protein ligase E3B OS=Homo sapiens GN=UBE3B PE=1 SV=3 - [UBE3B_H
Q9UPW8	11	1703	192,9	5,35	68	Protein unc-13 homolog A OS=Homo sapiens GN=UNC13A PE=2 SV=3 - [UN13A_H
P13639	13	858	95,3	6,83	68	Elongation factor 2 OS=Homo sapiens GN=EEF2 PE=1 SV=4 - [EF2_HUMAN]
P14616	7	1297	143,6	6,47	68	Insulin receptor-related protein OS=Homo sapiens GN=INSRR PE=2 SV=2 - [INSRR
Q14444	130	709	78,3	5,25	68	Caprin-1 OS=Homo sapiens GN=CAPRIN1 PE=1 SV=2 - [CAPR1_HUMAN]
Q8NA03	15	581	66,1	5,16	68	Fibrous sheath-interacting protein 1 OS=Homo sapiens GN=FSIP1 PE=2 SV=1 - [FSI
A8MUU9	7	505	55,3	13,3	68	Putative uncharacterized protein ENSP00000383309 OS=Homo sapiens PE=5 SV=3
Q06210	36	699	78,8	7,11	68	Glucosamine--fructose-6-phosphate aminotransferase [isomerizing] 1 OS=Homo
Q9Y696	58	253	28,8	5,59	68	Chloride intracellular channel protein 4 OS=Homo sapiens GN=CLIC4 PE=1 SV=4 -
O00571	9	662	73,2	7,18	67	ATP-dependent RNA helicase DDX3X OS=Homo sapiens GN=DDX3X PE=1 SV=3 - [D
A7MCY6	12	615	67,7	5,86	67	TANK-binding kinase 1-binding protein 1 OS=Homo sapiens GN=TBKBP1 PE=1 SV=
Q14416	8	872	95,5	8,12	67	Metabotropic glutamate receptor 2 OS=Homo sapiens GN=GRM2 PE=2 SV=2 - [GRI
Q9UFD9	12	1545	169,7	6,54	67	RIMS-binding protein 3A OS=Homo sapiens GN=RIMBP3 PE=1 SV=3 - [RIM3A_HUN
Q9P2M7	6	1197	136,3	5,54	67	Cingulin OS=Homo sapiens GN=CGN PE=1 SV=2 - [CING_HUMAN]
O94906	19	941	106,9	8,25	67	Pre-mRNA-processing factor 6 OS=Homo sapiens GN=PRPF6 PE=1 SV=1 - [PRPF6_H
Q9NRW7	9	570	65,0	8,24	67	Vacuolar protein sorting-associated protein 45 OS=Homo sapiens GN=VPS45 PE=1
Q92750	117	862	91,0	9,54	67	Transcription initiation factor TFIID subunit 4B OS=Homo sapiens GN=TAF4B PE=1
P42694	11	1942	218,8	7,42	67	Probable helicase with zinc finger domain OS=Homo sapiens GN=HELZ PE=1 SV=2
P09884	12	1462	165,8	5,85	67	DNA polymerase alpha catalytic subunit OS=Homo sapiens GN=POLA1 PE=1 SV=2
P52333	9	1124	125,0	7,18	67	Tyrosine-protein kinase JAK3 OS=Homo sapiens GN=JAK3 PE=1 SV=2 - [JAK3_HUM

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P04208	23	109	11,7	6,54	67	Ig lambda chain V-l region WAH OS=Homo sapiens PE=1 SV=1 - [LV106_HUMAN]
Q8N283	11	1001	109,9	6,02	67	Ankyrin repeat domain-containing protein 35 OS=Homo sapiens GN=ANKRD35 PE=
Q13206	28	875	100,8	8,63	67	Probable ATP-dependent RNA helicase DDX10 OS=Homo sapiens GN=DDX10 PE=
Q6P2H3	9	762	85,6	6	66	Coiled-coil domain-containing protein 21 OS=Homo sapiens GN=CCDC21 PE=1 SV=
Q9NS91	26	495	56,2	7,58	66	E3 ubiquitin-protein ligase RAD18 OS=Homo sapiens GN=RAD18 PE=1 SV=2 - [RAD
Q08378	32	1498	167,3	5,44	66	Golgin subfamily A member 3 OS=Homo sapiens GN=GOLGA3 PE=1 SV=2 - [GOGA:
Q14118	8	895	97,4	8,56	66	Dysglycolan OS=Homo sapiens GN=DAG1 PE=1 SV=2 - [DAG1_HUMAN]
P31040	9	664	72,6	7,39	66	Succinate dehydrogenase [ubiquinone] flavoprotein subunit, mitochondrial OS=
O94822	6	1766	200,4	6,25	66	E3 ubiquitin-protein ligase listerin OS=Homo sapiens GN=LTN1 PE=1 SV=6 - [LTN1
O60504	19	671	75,3	9,45	66	Vinexin OS=Homo sapiens GN=SORBS3 PE=1 SV=2 - [VINEX_HUMAN]
Q5VYS8	13	1495	171,1	6,83	66	Terminal uridylyltransferase 7 OS=Homo sapiens GN=ZCCHC6 PE=1 SV=1 - [TUT7_I
Q9H8K7	23	445	49,2	6,25	66	Uncharacterized protein C10orf88 OS=Homo sapiens GN=C10orf88 PE=2 SV=2 - [CJ
P48553	53	1259	142,1	6,04	66	Trafficking protein particle complex subunit 10 OS=Homo sapiens GN=TRAPPC10
Q8IXR9	14	625	71,3	9,16	66	Uncharacterized protein C12orf56 OS=Homo sapiens GN=C12orf56 PE=2 SV=2 - [CJ
Q92608	94	1830	211,8	6,87	66	Dedicator of cytokinesis protein 2 OS=Homo sapiens GN=DOCK2 PE=1 SV=2 - [DO
O15068	22	1137	128,0	6,43	66	Guanine nucleotide exchange factor DBS OS=Homo sapiens GN=MCF2L PE=1 SV=2
Q727J5	17	298	33,8	9,2	66	Developmental pluripotency-associated protein 2 (Pluripotent embryonic stem c
Q9Y388	10	322	37,3	9,83	66	RNA-binding motif protein, X-linked 2 OS=Homo sapiens GN=RBMX2 PE=1 SV=2 -
Q5TAX3	16	1644	185,0	7,97	66	Terminal uridylyltransferase 4 OS=Homo sapiens GN=ZCCHC11 PE=1 SV=3 - [TUT4
Q9BQI3	46	630	71,1	5,99	66	Eukaryotic translation initiation factor 2-alpha kinase 1 OS=Homo sapiens GN=EIF
P35749	11	1972	227,2	5,5	65	Myosin-11 OS=Homo sapiens GN=MYH11 PE=1 SV=3 - [MYH11_HUMAN]
Q9GT21	48	854	95,4	8,12	65	Selenocysteine insertion sequence-binding protein 2 OS=Homo sapiens GN=SECI
Q55R76	14	1530	173,4	8,06	65	TPR repeat-containing protein C10orf93 OS=Homo sapiens GN=C10orf93 PE=2 SV=
Q68CP9	11	1835	197,3	7,42	65	AT-rich interactive domain-containing protein 2 OS=Homo sapiens GN=ARID2 PE=
P98179	3	157	17,2	8,91	65	Putative RNA-binding protein 3 OS=Homo sapiens GN=RBM3 PE=1 SV=1 - [RBM3
Q14674	6	2120	233,0	7,55	65	Separin OS=Homo sapiens GN=ESPL1 PE=1 SV=3 - [ESPL1_HUMAN]
O75410	36	805	87,7	4,88	65	Transforming acidic coiled-coil-containing protein 1 OS=Homo sapiens GN=TACC:
Q8TDD5	14	553	64,2	6,18	65	Mucolipin-3 OS=Homo sapiens GN=MCOLN3 PE=2 SV=1 - [MCLN3_HUMAN]
A6PVC2	12	814	90,7	8,13	65	Protein monoglycylase TTL8 OS=Homo sapiens GN=TTL8 PE=1 SV=3 - [TTL8_HUI
Q8IU68	40	726	81,6	9,36	65	Transmembrane channel-like protein 8 OS=Homo sapiens GN=TMCC8 PE=1 SV=1 -
P57768	17	344	39,1	4,65	65	Sorting nexin-16 OS=Homo sapiens GN=SNX16 PE=1 SV=2 - [SNX16_HUMAN]
Q9GZR2	22	422	46,6	9,77	65	RNA exonuclease 4 OS=Homo sapiens GN=REXO4 PE=1 SV=2 - [REXO4_HUMAN]
Q9Y3Z3	7	626	72,2	7,14	65	SAM domain and HD domain-containing protein 1 OS=Homo sapiens GN=SAMHD:
Q9H3S7	5	1636	178,9	6,92	65	Tyrosine-protein phosphatase non-receptor type 23 OS=Homo sapiens GN=PTPN
O15550	8	1401	154,1	7,44	65	Lysine-specific demethylase 6A OS=Homo sapiens GN=KDM6A PE=1 SV=2 - [KDM6
Q9P2D1	18	2997	335,7	6,34	65	Chromodomain-helicase-DNA-binding protein 7 OS=Homo sapiens GN=CHD7 PE=
Q9Y239	25	953	107,6	7,11	65	Nucleotide-binding oligomerization domain-containing protein 1 OS=Homo sapi
Q9NYQ6	7	3014	329,3	5,92	65	Cadherin EGF LAG seven-pass G-type receptor 1 OS=Homo sapiens GN=CELSR1 PE
Q13523	6	1007	116,9	10,26	65	Serine/threonine-protein kinase PRP4 homolog OS=Homo sapiens GN=PRPF4B P
Q9HDC5	9	661	71,6	9,32	65	Junctophilin-1 OS=Homo sapiens GN=JPH1 PE=1 SV=2 - [JPH1_HUMAN]
Q6ZR08	11	3092	356,7	6,19	65	Dynein heavy chain 12, axonemal OS=Homo sapiens GN=DNAH12 PE=1 SV=2 - [DY
Q0ZGT2	5	675	80,6	5,33	65	Nexilin OS=Homo sapiens GN=NEXN PE=1 SV=1 - [NEXN_HUMAN]
Q8TEW0	9	1356	151,3	7,68	64	Partitioning defective 3 homolog OS=Homo sapiens GN=PARD3 PE=1 SV=2 - [PARI
Q8NDI1	11	1231	139,9	5,35	64	EH domain-binding protein 1 OS=Homo sapiens GN=EHBP1 PE=1 SV=3 - [EHBP1_H
Q9UDT6	9	1046	115,8	6,73	64	CAP-Gly domain-containing linker protein 2 OS=Homo sapiens GN=CLIP2 PE=1 SV
Q14241	15	798	89,9	9,57	64	Transcription elongation factor B polypeptide 3 OS=Homo sapiens GN=TCEB3 PE=
Q8IW93	13	802	89,1	7,53	64	Rho guanine nucleotide exchange factor 19 OS=Homo sapiens GN=ARHGEF19 PE=
O94911	6	1581	179,1	7,18	64	ATP-binding cassette sub-family A member 8 OS=Homo sapiens GN=ABCA8 PE=1
O75145	25	1194	133,4	5,68	64	Liprin-alpha-3 OS=Homo sapiens GN=PPFIA3 PE=1 SV=3 - [LIPA3_HUMAN]
Q9H4L7	40	1026	117,3	5,55	64	SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subf
Q13098	86	491	55,5	6,74	64	COP9 signalosome complex subunit 1 OS=Homo sapiens GN=GPS1 PE=1 SV=4 - [C
P32780	40	548	62,0	8,66	64	General transcription factor IIH subunit 1 OS=Homo sapiens GN=GTF2H1 PE=1 SV=
Q9UQ03	33	480	54,9	8,27	64	Coronin-2B OS=Homo sapiens GN=CORO2B PE=2 SV=4 - [COR2B_HUMAN]
Q93100	11	1093	124,8	6,95	64	Phosphorylase b kinase regulatory subunit beta OS=Homo sapiens GN=PHKB PE=
B1AK53	32	854	91,7	6,93	64	Espin OS=Homo sapiens GN=ESPN PE=1 SV=1 - [ESPN_HUMAN]
Q9HCL2	6	828	93,7	7,74	64	Glycerol-3-phosphate acyltransferase 1, mitochondrial OS=Homo sapiens GN=GP
Q55QS7	43	431	51,2	6,23	64	SH2 domain-containing protein 4B OS=Homo sapiens GN=SH2D4B PE=2 SV=1 - [SH
Q6UXY8	7	1006	114,7	8,21	63	Transmembrane channel-like protein 5 OS=Homo sapiens GN=TMCC5 PE=2 SV=3 -
P17987	9	556	60,3	6,11	63	T-complex protein 1 subunit alpha OS=Homo sapiens GN=TCP1 PE=1 SV=1 - [TCPA
P58107	18	5090	555,3	5,6	63	Epiplakin OS=Homo sapiens GN=EPPK1 PE=1 SV=2 - [EPIPL_HUMAN]
Q8WVM7	6	1258	144,3	5,59	63	Cohesin subunit SA-1 OS=Homo sapiens GN=STAG1 PE=1 SV=3 - [STAG1_HUMAN]
Q9BTW9	8	1192	132,5	6,19	63	Tubulin-specific chaperone D OS=Homo sapiens GN=TBCD PE=1 SV=2 - [TBCD_HUI
O75460	15	977	109,7	6,42	63	Serine/threonine-protein kinase/endoribonuclease IRE1 OS=Homo sapiens GN=I
Q96RL7	9	3174	360,0	6,33	63	Vacuolar protein sorting-associated protein 13A OS=Homo sapiens GN=VPS13A P
Q6DN14	17	999	111,6	8,15	63	Multiple C2 and transmembrane domain-containing protein 1 OS=Homo sapiens

Appendices

Q460N3	38	656	72,5	8,82	63	Poly [ADP-ribose] polymerase 15 OS=Homo sapiens GN=PARP15 PE=1 SV=1 - [PAF
O94885	10	1247	136,6	6,09	63	SAM and SH3 domain-containing protein 1 OS=Homo sapiens GN=SASH1 PE=1 SV=
Q7Z6J4	94	655	74,8	6,93	63	FYVE, RhoGEF and PH domain-containing protein 2 OS=Homo sapiens GN=FGD2 P
Q9UK32	22	745	83,8	6,34	63	Ribosomal protein S6 kinase alpha-6 OS=Homo sapiens GN=RPS6KA6 PE=1 SV=1 -
Q6PRD1	6	2367	257,2	5,71	63	Probable G-protein coupled receptor 179 OS=Homo sapiens GN=GPR179 PE=2 SV=
P58317	31	390	44,7	8,05	63	Zinc finger protein 121 OS=Homo sapiens GN=ZNF121 PE=2 SV=2 - [ZNF121_HUMA
Q6ZMW3	49	1958	217,8	7,44	63	Echinoderm microtubule-associated protein-like 6 OS=Homo sapiens GN=EML6 P
P51523	33	738	85,4	8,78	63	Zinc finger protein 84 (Zinc finger protein HPF2) - Homo sapiens (Human) - [ZNF8
Q96RU3	7	617	71,3	5,72	63	Formin-binding protein 1 OS=Homo sapiens GN=FBNP1 PE=1 SV=2 - [FBNP1_HUM
Q9H078	10	707	78,7	9,01	63	Caseolytic peptidase B protein homolog OS=Homo sapiens GN=CLPB PE=1 SV=1
Q99743	9	824	91,7	6,81	63	Neuronal PAS domain-containing protein 2 OS=Homo sapiens GN=NPAS2 PE=2 SV
P56715	5	2156	240,5	5,8	63	Oxygen-regulated protein 1 OS=Homo sapiens GN=RP1 PE=1 SV=1 - [RP1_HUMAN
Q8IZY2	7	2146	234,2	7,24	63	ATP-binding cassette sub-family A member 7 OS=Homo sapiens GN=ABCA7 PE=1
Q96BY6	7	2183	249,2	6,93	62	Dedicator of cytokinesis protein 10 OS=Homo sapiens GN=DOCK10 PE=1 SV=2 - [D
Q16363	20	1823	202,4	6,28	62	Laminin subunit alpha-4 OS=Homo sapiens GN=LAMA4 PE=1 SV=4 - [LAMA4_HUM
Q9V6I0	6	2220	246,2	6,02	62	Calcineurin-binding protein cabin-1 OS=Homo sapiens GN=CABIN1 PE=1 SV=1 - [C
Q9H1Z4	12	485	53,7	9,14	62	WD repeat-containing protein 13 OS=Homo sapiens GN=WDR13 PE=1 SV=2 - [WDR
P33261	30	490	55,9	7,39	62	Cytochrome P450 2C19 OS=Homo sapiens GN=CYP2C19 PE=1 SV=3 - [CP2CJ_HUMA
Q9BYB0	15	1741	186,2	9,03	62	SH3 and multiple ankyrin repeat domains protein 3 OS=Homo sapiens GN=SHAN
Q8IXH8	15	852	95,2	5,94	62	Cadherin-like protein 26 OS=Homo sapiens GN=CDH26 PE=2 SV=3 - [CAD26_HUM
A4D1E1	84	1349	152,5	8,54	62	Zinc finger protein 804B OS=Homo sapiens GN=ZNF804B PE=1 SV=2 - [Z804B_HUM
O43830	15	326	38,2	9,17	62	Zinc finger protein 73 OS=Homo sapiens GN=ZNF73 PE=3 SV=1 - [ZNF73_HUMAN]
Q9Y2W6	13	606	67,0	5,08	62	Tudor and KH domain-containing protein OS=Homo sapiens GN=TDRKH PE=1 SV=
O75096	17	1905	211,9	5,27	62	Low-density lipoprotein receptor-related protein 4 OS=Homo sapiens GN=LRP4 P
Q13813	5	2472	284,4	5,35	62	Spectrin alpha chain, brain OS=Homo sapiens GN=SPTAN1 PE=1 SV=3 - [SPTA2_HL
Q96A00	10	147	16,7	9,38	62	Protein phosphatase 1 regulatory subunit 14A OS=Homo sapiens GN=PPP1R14A P
Q680I6	7	523	58,6	9,16	62	Lysine-specific demethylase 4D OS=Homo sapiens GN=KDM4D PE=1 SV=3 - [KDM
Q6ZNF79	7	300	34,7	9,2	62	Zinc finger protein 705A OS=Homo sapiens GN=ZNF705A PE=2 SV=1 - [Z705A_HUM
P10632	37	490	55,8	8,5	62	Cytochrome P450 2C8 OS=Homo sapiens GN=CYP2C8 PE=1 SV=2 - [CP2C8_HUMAN
O43852	8	315	37,1	4,64	62	Calumenin OS=Homo sapiens GN=CALU PE=1 SV=2 - [CALU_HUMAN]
Q9HCC0	50	563	61,3	7,68	62	Methylcrotonoyl-CoA carboxylase beta chain, mitochondrial OS=Homo sapiens G
Q9UKK3	34	1724	192,5	5,66	62	Poly [ADP-ribose] polymerase 4 OS=Homo sapiens GN=PARP4 PE=1 SV=3 - [PARP
Q96RR4	5	588	64,7	6,68	62	Calcium/calmodulin-dependent protein kinase kinase 2 OS=Homo sapiens GN=C
O94953	12	1096	121,8	7,09	61	Lysine-specific demethylase 4B OS=Homo sapiens GN=KDM4B PE=1 SV=4 - [KDM
Q8NAN2	23	632	71,0	5,63	61	Protein FAM73A OS=Homo sapiens GN=FAM73A PE=2 SV=1 - [FA73A_HUMAN]
P13942	8	1736	171,7	6,21	61	Collagen alpha-2(XI) chain OS=Homo sapiens GN=COL11A2 PE=1 SV=4 - [COBA2_H
Q6F5E8	27	1435	154,6	6,76	61	Leucine-rich repeat-containing protein 16C OS=Homo sapiens GN=RLTPR PE=1 SV
O75179	7	2603	274,1	6,52	61	Ankyrin repeat domain-containing protein 17 OS=Homo sapiens GN=ANKRD17 PE
Q9NP60	4	686	78,6	6,46	61	X-linked interleukin-1 receptor accessory protein-like 2 OS=Homo sapiens GN=IL
Q96L91	26	3159	343,3	9,19	61	E1A-binding protein p400 OS=Homo sapiens GN=EP400 PE=1 SV=4 - [EP400_HUMA
O60290	9	1169	131,6	7,23	61	Zinc finger protein 862 OS=Homo sapiens GN=ZNF862 PE=2 SV=2 - [ZN862_HUMA
Q9UKV0	4	1011	111,2	6,89	61	Histone deacetylase 9 OS=Homo sapiens GN=HDAC9 PE=1 SV=2 - [HDAC9_HUMAN
Q9Y2R2	8	807	91,6	7,59	61	Tyrosine-protein phosphatase non-receptor type 22 OS=Homo sapiens GN=PTPN
Q8WVB6	14	975	107,3	7,21	61	Chromosome transmission fidelity protein 18 homolog OS=Homo sapiens GN=CH
P51654	6	580	65,5	6,37	61	Glypican-3 OS=Homo sapiens GN=GPC3 PE=1 SV=1 - [GPC3_HUMAN]
Q8NEE6	8	735	83,9	8,72	61	F-box/LRR-repeat protein 13 OS=Homo sapiens GN=FBXL13 PE=2 SV=3 - [FXL13_H
P04745	6	511	57,7	6,93	61	Alpha-amylase 1 OS=Homo sapiens GN=AMY1A PE=1 SV=2 - [AMY1_HUMAN]
P55040	12	296	33,9	8,51	61	GTP-binding protein GEM OS=Homo sapiens GN=GEM PE=1 SV=1 - [GEM_HUMAN]
P06753	19	284	32,8	4,72	61	Tropomyosin alpha-3 chain (Tropomyosin-3) (Tropomyosin gamma) (hTMS) - Hon
P10155	8	538	60,6	8,03	61	60 kDa SS-A/Ro ribonucleoprotein OS=Homo sapiens GN=TROVE2 PE=1 SV=2 - [RC
Q15596	33	1464	159,1	6,64	61	Nuclear receptor coactivator 2 OS=Homo sapiens GN=NCOA2 PE=1 SV=2 - [NCOA2
Q8NAV1	21	312	37,5	9,96	61	Pre-mRNA-splicing factor 38A OS=Homo sapiens GN=PRPF38A PE=1 SV=1 - [PR38
Q9NZ08	4	941	107,2	6,46	61	Endoplasmic reticulum aminopeptidase 1 OS=Homo sapiens GN=ERAP1 PE=1 SV=
Q9Y232	39	598	66,4	9,45	61	Chromodomain Y-like protein OS=Homo sapiens GN=CDYL PE=1 SV=2 - [CDYL1_HL
Q96SB3	20	815	89,1	4,97	60	Neurabin-2 OS=Homo sapiens GN=PPP1R9B PE=1 SV=2 - [NEB2_HUMAN]
Q8TC71	29	538	61,1	8,63	60	Spermatogenesis-associated protein 18 OS=Homo sapiens GN=SPATA18 PE=1 SV=
Q8N9H9	23	656	69,7	5,43	60	Uncharacterized protein C1orf127 OS=Homo sapiens GN=C1orf127 PE=2 SV=2 - [C
Q8N9M1	11	422	44,7	10,11	60	Uncharacterized protein C19orf47 OS=Homo sapiens GN=C19orf47 PE=1 SV=1 - [C
O75376	13	2440	270,0	7,11	60	Nuclear receptor corepressor 1 OS=Homo sapiens GN=NCOR1 PE=1 SV=2 - [NCOR
Q9UL54	7	1235	138,2	7,27	60	Serine/threonine-protein kinase TAO2 (EC 2.7.11.1) (Thousand and one amino ac
Q7Z591	6	1439	155,0	6,32	60	AT-hook-containing transcription factor OS=Homo sapiens GN=AKNA PE=1 SV=2 -
Q2M2I5	8	525	55,1	4,96	60	Keratin, type I cytoskeletal 24 OS=Homo sapiens GN=KRT24 PE=1 SV=1 - [K1C24_H
Q8N961	17	839	93,2	5,9	60	Ankyrin repeat and BTB/POZ domain-containing protein 2 OS=Homo sapiens GN=
O15360	5	1455	162,7	6,6	60	Fanconi anemia group A protein OS=Homo sapiens GN=FANCA PE=1 SV=2 - [FANC
Q8N239	101	644	70,6	5,6	60	Kelch-like protein 34 OS=Homo sapiens GN=KLHL34 PE=2 SV=1 - [KLH34_HUMAN]

Appendices

Q9Y4E6	32	1490	163,7	6,92	60	WD repeat-containing protein 7 OS=Homo sapiens GN=WDR7 PE=2 SV=2 - [WDR7
Q5T4T6	12	812	93,5	5,8	60	Synaptonemal complex protein 2-like OS=Homo sapiens GN=SYCP2L PE=1 SV=2 -
Q13424	8	505	53,9	6,8	60	Alpha-1-syntrophin OS=Homo sapiens GN=SNTA1 PE=1 SV=1 - [SNTA1_HUMAN]
P21580	24	790	89,6	8,22	60	Tumor necrosis factor alpha-induced protein 3 OS=Homo sapiens GN=TNFAIP3 PE
Q8N1T3	19	1032	119,0	9,09	60	Myosin-Ih OS=Homo sapiens GN=MYO1H PE=2 SV=2 - [MYO1H_HUMAN]
Q92973	41	898	102,3	4,98	60	Transportin-1 OS=Homo sapiens GN=TNPO1 PE=1 SV=2 - [TNPO1_HUMAN]
Q96N67	37	2140	242,4	6,8	60	Dedicator of cytokinesis protein 7 OS=Homo sapiens GN=DOCK7 PE=1 SV=4 - [DO
Q08462	16	1091	123,5	8,09	60	Adenylate cyclase type 2 OS=Homo sapiens GN=ADCY2 PE=1 SV=5 - [ADCY2_HUM
O15523	7	660	73,1	7,55	60	ATP-dependent RNA helicase DDX3Y OS=Homo sapiens GN=DDX3Y PE=1 SV=2 - [C
Q8NEM0	27	835	92,8	8,25	60	Microcephalin OS=Homo sapiens GN=MCPH1 PE=1 SV=3 - [MCPH1_HUMAN]
Q13075	6	1403	159,5	5,99	60	Baculoviral IAP repeat-containing protein 1 OS=Homo sapiens GN=NAIP PE=1 SV=
Q6PKG0	72	1096	123,4	8,82	60	La-related protein 1 OS=Homo sapiens GN=LARP1 PE=1 SV=2 - [LARP1_HUMAN]
Q81ZQ1	7	3526	395,0	6,76	60	WD repeat and FYVE domain-containing protein 3 OS=Homo sapiens GN=WDFY3 I
Q15413	22	4870	551,6	5,66	60	Ryanodine receptor 3 OS=Homo sapiens GN=RYP3 PE=1 SV=2 - [RYP3_HUMAN]
Q5TID7	10	509	60,1	5,91	59	Uncharacterized protein C1orf114 OS=Homo sapiens GN=C1orf114 PE=2 SV=1 - [C/
Q96S53	92	571	63,6	7,06	59	Dual specificity testis-specific protein kinase 2 OS=Homo sapiens GN=TESK2 PE=2
O94779	25	1100	120,6	6,34	59	Contactin-5 OS=Homo sapiens GN=CNTN5 PE=1 SV=2 - [CNTN5_HUMAN]
A3KN83	5	1393	154,2	7,88	59	Protein strawberry notch homolog 1 OS=Homo sapiens GN=SBNO1 PE=1 SV=1 - [S
P49788	5	294	33,3	8,51	59	Retinoic acid receptor responder protein 1 OS=Homo sapiens GN=RARRES1 PE=2
Q9NQW6	26	1124	124,1	8,07	59	Actin-binding protein anillin OS=Homo sapiens GN=ANLN PE=1 SV=2 - [ANLN_HU
Q53F19	12	620	70,5	5,73	59	Uncharacterized protein C17orf85 OS=Homo sapiens GN=C17orf85 PE=1 SV=2 - [C/
Q4G0X9	5	1142	130,0	5,29	59	Coiled-coil domain-containing protein 40 OS=Homo sapiens GN=CCDC40 PE=2 SV=
Q9Y6N6	13	1575	171,1	6,58	59	Laminin subunit gamma-3 OS=Homo sapiens GN=LAMC3 PE=2 SV=3 - [LAMC3_HUI
Q95197	12	1032	112,5	4,96	59	Reticulon-3 OS=Homo sapiens GN=RTN3 PE=1 SV=2 - [RTN3_HUMAN]
Q9NX58	4	379	43,6	9,54	59	Cell growth-regulating nucleolar protein OS=Homo sapiens GN=LYAR PE=1 SV=2 -
A0AVI2	8	2093	241,8	7,96	59	Fer-1-like protein 5 OS=Homo sapiens GN=FER1L5 PE=2 SV=2 - [FER1L5_HUMAN]
Q9NXD2	12	777	88,2	8,53	59	Myotubularin-related protein 10 OS=Homo sapiens GN=MTMR10 PE=1 SV=3 - [MT
Q13057	11	564	62,3	6,99	59	Bifunctional coenzyme A synthase OS=Homo sapiens GN=COASY PE=1 SV=4 - [CO
Q8NEP3	12	725	80,0	4,67	59	Leucine-rich repeat-containing protein 50 OS=Homo sapiens GN=LRRCS0 PE=1 SV=
Q9NQ87	96	328	35,1	10,68	59	Hairy/enhancer-of-split related with YRPW motif-like protein OS=Homo sapiens
Q05BV3	8	1969	219,3	7,74	59	Echinoderm microtubule-associated protein-like 5 OS=Homo sapiens GN=EML5 P
O94913	5	1555	172,9	8,48	59	Pre-mRNA cleavage complex 2 protein Pcf11 OS=Homo sapiens GN=PFC11 PE=1 S
Q52LW3	5	1261	142,0	6,74	59	Rho GTPase-activating protein 29 OS=Homo sapiens GN=ARHGAP29 PE=1 SV=2 - [I
Q9Y485	6	3027	337,6	6,34	59	Dmx-like protein 1 OS=Homo sapiens GN=DMXL1 PE=1 SV=3 - [DMXL1_HUMAN]
Q9BRD0	4	619	70,5	9,86	59	BUD13 homolog OS=Homo sapiens GN=BUD13 PE=1 SV=1 - [BUD13_HUMAN]
P26599	9	531	57,2	9,17	58	Polypyrimidine tract-binding protein 1 OS=Homo sapiens GN=PTBP1 PE=1 SV=1 -
Q4FZB7	11	885	99,1	8,78	58	Histone-lysine N-methyltransferase SUV420H1 OS=Homo sapiens GN=SUV420H1
O95263	22	885	98,9	6,83	58	High affinity cAMP-specific and IBMX-insensitive 3',5'-cyclic phosphodiesterase 8
Q6ZRV2	9	1179	127,0	6,98	58	Protein FAM83H OS=Homo sapiens GN=FAM83H PE=1 SV=3 - [FAM83H_HUMAN]
O60524	4	1076	122,9	6,35	58	Serologically defined colon cancer antigen 1 OS=Homo sapiens GN=SDCCAG1 PE=
Q9UDY4	20	337	37,8	8,5	58	DnaJ homolog subfamily B member 4 OS=Homo sapiens GN=DNAJB4 PE=1 SV=1 -
Q9UN79	16	622	69,2	6,71	58	Transcription factor SOX-13 OS=Homo sapiens GN=SOX13 PE=1 SV=3 - [SOX13_HU
Q5T6S3	11	580	65,5	8,84	58	PHD finger protein 19 OS=Homo sapiens GN=PHF19 PE=1 SV=1 - [PHF19_HUMAN]
Q9HCS4	16	588	62,6	8,94	58	Transcription factor 7-like 1 OS=Homo sapiens GN=TCF7L1 PE=2 SV=1 - [TF7L1_HU
Q96NH3	21	1257	144,7	6,74	58	Protein broad-minded OS=Homo sapiens GN=BROMI PE=2 SV=4 - [BROMI_HUMA
Q6NV74	5	962	102,1	7,96	58	Uncharacterized protein C2orf55 OS=Homo sapiens GN=C2orf55 PE=1 SV=3 - [CB0
Q2VWP7	31	1150	127,0	7,59	58	Proteogenin OS=Homo sapiens GN=PRTG PE=2 SV=1 - [PRTG_HUMAN]
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Q9HCE3	7	1301	141,6	8,65	58	Zinc finger protein 532 OS=Homo sapiens GN=ZNF532 PE=1 SV=2 - [ZNF532_HUMA
Q8NFP4	50	955	105,7	8,34	58	MAM domain-containing glycosylphosphatidylinositol anchor protein 1 OS=Homo
Q8NB66	7	2214	250,8	5,92	58	Protein unc-13 homolog C OS=Homo sapiens GN=UNC13C PE=1 SV=3 - [UNC13_HU
Q9UPW6	30	733	82,5	6,9	58	DNA-binding protein SATB2 OS=Homo sapiens GN=SATB2 PE=1 SV=2 - [SATB2_HU
Q9BZV3	6	1241	138,5	4,61	58	Interphotoreceptor matrix proteoglycan 2 OS=Homo sapiens GN=IMP2 PE=1 SV=
O43424	23	1007	113,3	6,07	58	Glutamate receptor delta-2 subunit OS=Homo sapiens GN=GRID2 PE=2 SV=2 - [GR
Q9NNX1	9	390	44,2	6	58	Tuftelin OS=Homo sapiens GN=TUFT1 PE=2 SV=1 - [TUFT1_HUMAN]
P05177	4	515	58,3	9,06	58	Cytochrome P450 1A2 OS=Homo sapiens GN=CYP1A2 PE=1 SV=3 - [CP1A2_HUMAN
Q9NQ90	18	1003	113,9	6,55	58	Anoctamin-2 OS=Homo sapiens GN=ANO2 PE=1 SV=2 - [ANO2_HUMAN]
O14964	11	777	86,1	6,16	57	Hepatocyte growth factor-regulated tyrosine kinase substrate OS=Homo sapiens
Q16352	3	499	55,4	5,4	57	Alpha-internexin OS=Homo sapiens GN=INA PE=1 SV=2 - [AINX_HUMAN]
Q9Y597	15	815	88,9	7,03	57	BTB/POZ domain-containing protein KCTD3 OS=Homo sapiens GN=KCTD3 PE=1 SV
Q92613	6	823	93,7	7,18	57	Protein Jade-3 OS=Homo sapiens GN=PHF16 PE=1 SV=1 - [JADE3_HUMAN]
Q5T481	5	1227	134,3	5,69	57	Probable RNA-binding protein 20 OS=Homo sapiens GN=RBM20 PE=1 SV=3 - [RBM
Q9BY77	71	421	46,1	9,99	57	Polymerase delta-interacting protein 3 (46 kDa DNA polymerase delta interactor
Q8TDG4	12	1101	124,1	6,58	57	Helicase POLQ-like OS=Homo sapiens GN=HELQ PE=1 SV=2 - [HELQ_HUMAN]
Q9NS56	10	1045	119,1	9,51	57	E3 ubiquitin-protein ligase Topors OS=Homo sapiens GN=TOPORS PE=1 SV=1 - [TC

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O60522	14	2096	236,4	5,25	57	Tudor domain-containing protein 6 OS=Homo sapiens GN=TDRD6 PE=2 SV=2 - [TD
P52848	5	882	100,8	7,97	57	Bifunctional heparan sulfate N-deacetylase/N-sulfotransferase 1 OS=Homo sapie
Q6NSJ5	9	796	90,2	6,96	57	Leucine-rich repeat-containing protein 8E OS=Homo sapiens GN=LRR8E PE=2 SV
P04430	2	108	11,8	8,44	57	Ig kappa chain V-I region BAN OS=Homo sapiens PE=1 SV=1 - [KV122_HUMAN]
P35711	21	763	84,0	6,6	57	Transcription factor SOX-5 OS=Homo sapiens GN=SOX5 PE=2 SV=3 - [SOX5_HUMA
P17213	27	487	53,9	9,38	57	Bactericidal permeability-increasing protein OS=Homo sapiens GN=BPI PE=1 SV=
Q9BQA5	28	517	59,6	6,19	57	Histone H4 transcription factor OS=Homo sapiens GN=HINFP PE=1 SV=2 - [HINFP_
P17252	17	672	76,7	7,05	57	Protein kinase C alpha type OS=Homo sapiens GN=PRKCA PE=1 SV=4 - [PKCA_HU
Q9NYU2	4	1555	177,1	5,63	57	UDP-glucose:glycoprotein glucosyltransferase 1 OS=Homo sapiens GN=UGGT1 PE
Q6ZNA1	49	936	107,6	9,22	57	Zinc finger protein 836 OS=Homo sapiens GN=ZNF836 PE=2 SV=2 - [ZN836_HUMA
P15822	26	2718	296,7	7,84	56	Zinc finger protein 40 OS=Homo sapiens GN=HIVEP1 PE=1 SV=3 - [ZEP1_HUMAN]
A5D8W1	15	941	105,8	7,17	56	Uncharacterized protein C7orf63 OS=Homo sapiens GN=C7orf63 PE=2 SV=3 - [CGO
Q9NXF1	16	929	105,6	9,36	56	Testis-expressed sequence 10 protein OS=Homo sapiens GN=TEX10 PE=1 SV=2 - [
Q8IY92	10	1834	199,9	6,06	56	Structure-specific endonuclease subunit SLX4 OS=Homo sapiens GN=SLX4 PE=1 S
Q9NRX1	8	252	27,9	9,73	56	RNA-binding protein PNO1 OS=Homo sapiens GN=PNO1 PE=1 SV=1 - [PNO1_HUM
O75676	15	772	85,6	8,28	56	Ribosomal protein S6 kinase alpha-4 OS=Homo sapiens GN=RPS6KA4 PE=1 SV=1 -
Q9ULR0	6	285	33,0	5,17	56	Pre-mRNA-splicing factor ISY1 homolog OS=Homo sapiens GN=ISY1 PE=1 SV=3 - [I
P98196	5	1134	129,7	6,6	56	Probable phospholipid-transporting ATPase 1H OS=Homo sapiens GN=ATP11A PE
Q9NQ75	9	786	87,1	7,08	56	Cas scaffolding protein family member 4 OS=Homo sapiens GN=CASS4 PE=1 SV=2
Q00341	3	1268	141,4	6,87	56	Vigilin OS=Homo sapiens GN=HDLBP PE=1 SV=2 - [VIGLN_HUMAN]
Q9BWW8	13	343	38,1	8,35	56	Apolipoprotein L6 OS=Homo sapiens GN=APOL6 PE=2 SV=1 - [APOL6_HUMAN]
Q9UPT5	9	735	83,3	6,79	56	Exocyst complex component 7 OS=Homo sapiens GN=EXOC7 PE=1 SV=3 - [EXOC7_
Q5T447	4	861	97,1	5,64	56	E3 ubiquitin-protein ligase HECTD3 OS=Homo sapiens GN=HECTD3 PE=1 SV=1 - [H
P35251	9	1148	128,2	9,36	56	Replication factor C subunit 1 OS=Homo sapiens GN=RFC1 PE=1 SV=4 - [RFC1_HUN
Q96BN8	32	352	40,2	5,47	56	Protein FAM105B OS=Homo sapiens GN=FAM105B PE=1 SV=3 - [F105B_HUMAN]
Q9UQL6	5	1122	121,9	6,24	56	Histone deacetylase 5 OS=Homo sapiens GN=HDAC5 PE=1 SV=2 - [HDAC5_HUMAN
O94808	35	682	76,9	7,37	56	Glucosamine--fructose-6-phosphate aminotransferase [isomerizing] 2 OS=Homo
Q5JTW2	5	689	76,3	8,18	56	Centrosomal protein of 78 kDa OS=Homo sapiens GN=CEP78 PE=1 SV=1 - [CEP78_
Q6ZV70	23	420	46,3	7,09	56	LanC-like protein 3 OS=Homo sapiens GN=LANCL3 PE=2 SV=2 - [LANC3_HUMAN]
Q8TB72	9	1066	114,1	7,08	56	Pumilio homolog 2 OS=Homo sapiens GN=PUM2 PE=1 SV=2 - [PUM2_HUMAN]
Q96C24	46	671	76,0	8,98	56	Synaptotagmin-like protein 4 OS=Homo sapiens GN=SYTL4 PE=1 SV=2 - [SYTL4_HU
Q6P9B6	7	456	51,0	6,24	55	TLD domain-containing protein KIAA1609 OS=Homo sapiens GN=KIAA1609 PE=1 S
P05165	66	728	80,0	7,52	55	Propionyl-CoA carboxylase alpha chain, mitochondrial OS=Homo sapiens GN=PC
Q14D04	21	833	94,7	6,71	55	Ventricular zone-expressed PH domain-containing protein homolog 1 OS=Homo
Q5JPE7	12	1267	139,4	5,76	55	Nodal modulator 2 OS=Homo sapiens GN=NOMO2 PE=1 SV=1 - [NOMO2_HUMAN]
P01714	2	108	11,4	6,52	55	Ig lambda chain V-III region SH OS=Homo sapiens PE=1 SV=1 - [LV301_HUMAN]
Q9C0B2	3	762	86,9	5,52	55	Uncharacterized protein KIAA1751 OS=Homo sapiens GN=KIAA1751 PE=2 SV=2 - [I
Q9UKG1	6	709	79,6	5,41	55	DCC-interacting protein 13-alpha OS=Homo sapiens GN=APPL1 PE=1 SV=1 - [DP13
Q9HCE0	8	2579	292,3	6,43	55	UPF0493 protein KIAA1632 OS=Homo sapiens GN=KIAA1632 PE=2 SV=2 - [K1632_H
Q8GXH1	12	822	95,3	9,47	55	IQ and AAA domain-containing protein 1 OS=Homo sapiens GN=IQCA1 PE=2 SV=1
Q8TDI7	6	906	102,5	9,47	55	Transmembrane channel-like protein 2 OS=Homo sapiens GN=TMC2 PE=2 SV=3 -]
P29597	10	1187	133,6	7,15	55	Non-receptor tyrosine-protein kinase TYK2 OS=Homo sapiens GN=TYK2 PE=1 SV=
Q99250	11	2005	227,8	5,73	55	Sodium channel protein type 2 subunit alpha OS=Homo sapiens GN=SCN2A PE=1
Q7Z403	15	805	90,0	8,66	55	Transmembrane channel-like protein 6 OS=Homo sapiens GN=TMC6 PE=1 SV=2 -]
Q96CW5	8	907	103,5	8,12	55	Gamma-tubulin complex component 3 OS=Homo sapiens GN=TUBGCP3 PE=1 SV=
A1L390	16	1219	134,3	6,55	55	Pleckstrin homology domain-containing family G member 3 OS=Homo sapiens Gf
P01780	37	115	12,6	9,29	55	Ig heavy chain V-III region JON - Homo sapiens (Human) - [HV319_HUMAN]
O75381	10	377	41,2	4,94	55	Peroxisomal membrane protein PEX14 OS=Homo sapiens GN=PEX14 PE=1 SV=1 -]
Q59EK9	22	446	49,7	5,27	55	RUN domain-containing protein 3A OS=Homo sapiens GN=RUNDC3A PE=2 SV=2 -
Q5T1M5	8	1219	133,5	5,2	55	FK506-binding protein 15 OS=Homo sapiens GN=FKBP15 PE=1 SV=2 - [FKB15_HUM
Q9ULM3	15	1422	150,7	8,98	55	YEATS domain-containing protein 2 OS=Homo sapiens GN=YEATS2 PE=1 SV=2 - [YE
Q96H22	54	339	39,5	9,13	55	Centromere protein N OS=Homo sapiens GN=CENPN PE=1 SV=2 - [CENPN_HUMAI
Q5U651	3	963	103,4	7,96	55	Ras-interacting protein 1 OS=Homo sapiens GN=RASIP1 PE=1 SV=1 - [RAIN_HUMA
P19174	47	1290	148,4	6,05	55	1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase gamma-1 OS=Homc
Q8N884	101	522	58,8	9,48	55	Uncharacterized protein C6orf150 OS=Homo sapiens GN=C6orf150 PE=1 SV=2 - [Cf
Q8IXA5	19	215	23,4	7,94	54	Sperm acrosome membrane-associated protein 3 OS=Homo sapiens GN=SPACA3
Q9UP95	14	1085	120,6	6,44	54	Solute carrier family 12 member 4 OS=Homo sapiens GN=SLC12A4 PE=1 SV=2 - [S1
Q8N6Q8	6	603	68,2	7,08	54	Uncharacterized protein C12orf26 OS=Homo sapiens GN=C12orf26 PE=2 SV=2 - [Cf
Q96FB5	21	475	52,9	7,97	54	UPF0431 protein C1orf66 OS=Homo sapiens GN=C1orf66 PE=2 SV=2 - [CA066_HUM
Q96977	11	292	33,5	6,68	54	Cytosolic 5'-nucleotidase III-like protein OS=Homo sapiens GN=NT5C3L PE=1 SV=
Q68D86	11	513	60,4	5,95	54	Coiled-coil domain-containing protein 102B OS=Homo sapiens GN=CCDC102B PE=
Q63HK5	13	1081	118,5	7,25	54	Teashirt homolog 3 OS=Homo sapiens GN=TSHZ3 PE=1 SV=2 - [TSH3_HUMAN]
P27815	6	886	98,1	5,21	54	cAMP-specific 3',5'-cyclic phosphodiesterase 4A OS=Homo sapiens GN=PDE4A PE
Q6ZUT9	60	1274	144,9	6,73	54	DENN domain-containing protein 5B OS=Homo sapiens GN=DENND5B PE=1 SV=2
Q8NCM2	14	988	111,8	7,58	54	Potassium voltage-gated channel subfamily H member 5 OS=Homo sapiens GN=k

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P48995	19	793	91,2	8,05	54	Short transient receptor potential channel 1 OS=Homo sapiens GN=TRPC1 PE=1 SV=1
Q6EMK4	18	673	71,7	7,39	54	Vasorin OS=Homo sapiens GN=VASN PE=1 SV=1 - [VASN_HUMAN]
Q9UHL9	69	959	106,0	6,87	54	General transcription factor II-I repeat domain-containing protein 1 OS=Homo sapiens GN=TFIIID1 PE=1 SV=1
Q3SY89	5	546	59,7	9,85	54	RNA polymerase II transcription factor SIII subunit A3-like-1 OS=Homo sapiens GN=POLR2B PE=1 SV=1
Q9H3U1	18	944	103,0	6,07	54	Protein unc-45 homolog A OS=Homo sapiens GN=UNC45A PE=1 SV=1 - [UNC45A_HUMAN]
O75152	22	810	89,1	8,37	54	Zinc finger CCH domain-containing protein 11A OS=Homo sapiens GN=ZC3H11A
Q7Z3T8	13	1539	168,8	4,82	54	Zinc finger FYVE domain-containing protein 16 OS=Homo sapiens GN=ZFYVE16 PE=1 SV=1
P19971	2	482	49,9	5,53	54	Thymidine phosphorylase OS=Homo sapiens GN=TYMP PE=1 SV=2 - [TYMP_HUMAN]
O43150	7	1006	111,6	6,68	54	Arf-GAP with SH3 domain, ANK repeat and PH domain-containing protein 2 OS=Homo sapiens GN=ARFGAP2 PE=1 SV=1
Q96G42	19	594	63,3	9	54	Kelch domain-containing protein 7B OS=Homo sapiens GN=KLHDC7B PE=2 SV=2 - [KLHDC7B_HUMAN]
Q86XE3	4	530	60,7	8,21	53	EF-hand domain-containing family member A2 OS=Homo sapiens GN=EFHA2 PE=1 SV=1
Q92771	3	950	105,9	8,24	53	Probable ATP-dependent RNA helicase DDX12 OS=Homo sapiens GN=DDX12 PE=1 SV=1
Q8TED9	6	768	86,4	6,8	53	Actin filament-associated protein 1-like 1 OS=Homo sapiens GN=AFAP1L1 PE=1 SV=1
A8MX4	46	1036	120,0	9,31	53	Zinc finger protein 99 OS=Homo sapiens GN=ZNF99 PE=2 SV=2 - [ZNF99_HUMAN]
Q9BVG8	32	833	92,7	7,69	53	Kinesin-like protein KIFC3 OS=Homo sapiens GN=KIFC3 PE=1 SV=3 - [KIFC3_HUMAN]
Q8NEN9	42	1154	128,5	6,09	53	PDZ domain-containing protein 8 OS=Homo sapiens GN=PDZD8 PE=1 SV=1 - [PDZD8_HUMAN]
Q9H1J1	36	476	54,7	9,03	53	Regulator of nonsense transcripts 3A OS=Homo sapiens GN=UPF3A PE=1 SV=1 - [UPF3A_HUMAN]
Q8NC67	8	525	59,4	6,81	53	Neuropilin and tolloid-like protein 2 precursor (Brain-specific transmembrane protein 2) OS=Homo sapiens GN=NPPL2 PE=1 SV=1
Q5VU57	15	503	58,2	8,41	53	Cytosolic carboxypeptidase 6 OS=Homo sapiens GN=AGBL4 PE=2 SV=3 - [CBPC6_HUMAN]
A6NI56	6	674	76,0	8,38	53	Coiled-coil domain-containing protein 154 OS=Homo sapiens GN=CCDC154 PE=2 SV=1
Q6P179	4	960	110,4	6,71	53	Endoplasmic reticulum aminopeptidase 2 OS=Homo sapiens GN=ERAP2 PE=1 SV=1
Q9BTV7	8	478	52,2	9,82	53	CDK5 and ABL1 enzyme substrate 2 OS=Homo sapiens GN=CABLES2 PE=1 SV=3 - [CABLES2_HUMAN]
Q15005	19	226	25,0	8,47	53	Signal peptidase complex subunit 2 OS=Homo sapiens GN=SPCS2 PE=1 SV=3 - [SPCS2_HUMAN]
PO2763	4	201	23,5	5,02	53	Alpha-1-acid glycoprotein 1 precursor (AGP 1) (Orosomucoid-1) (OMD 1) - Homo sapiens GN=AGP1 PE=1 SV=1
Q6ZNG1	10	722	83,1	9,25	53	Zinc finger protein 600 OS=Homo sapiens GN=ZNF600 PE=1 SV=2 - [ZNF600_HUMAN]
Q96A61	16	297	34,6	4,25	53	Tripartite motif-containing protein 52 OS=Homo sapiens GN=TRIM52 PE=2 SV=1 - [TRIM52_HUMAN]
Q8WY91	13	577	62,9	9,28	53	THAP domain-containing protein 4 OS=Homo sapiens GN=THAP4 PE=1 SV=2 - [THAP4_HUMAN]
Q587J8	8	217	24,3	9,48	53	ES cell-associated transcript 1 protein OS=Homo sapiens GN=ECAT1 PE=2 SV=1 - [ECAT1_HUMAN]
Q9ULQ0	16	834	95,3	5,91	53	Protein FAM40B OS=Homo sapiens GN=FAM40B PE=2 SV=2 - [FAM40B_HUMAN]
Q6NUK1	27	477	53,3	6,33	53	Calcium-binding mitochondrial carrier protein ScaMC-1 OS=Homo sapiens GN=SLC25A10 PE=1 SV=1
PO1703	16	103	10,9	9,29	53	Ig lambda chain V-1 region NEWM OS=Homo sapiens GN=LV105 PE=1 SV=1 - [LV105_HUMAN]
Q01844	9	656	68,4	9,33	53	RNA-binding protein EWS OS=Homo sapiens GN=EWSR1 PE=1 SV=1 - [EWS_HUMAN]
Q9Y679	15	476	53,0	8,09	53	Ancient ubiquitously expressed protein 1 OS=Homo sapiens GN=AUP1 PE=1 SV=1 - [AUP1_HUMAN]
Q8N3Y1	4	598	67,4	5,67	53	F-box/WD repeat-containing protein 8 OS=Homo sapiens GN=FBXW8 PE=1 SV=2 - [FBXW8_HUMAN]
Q9UBT7	102	734	81,8	6,64	53	Alpha-catulin OS=Homo sapiens GN=CTNNA1 PE=1 SV=2 - [CTNNA1_HUMAN]
P08237	16	780	85,1	7,99	53	6-phosphofructokinase, muscle type OS=Homo sapiens GN=PFKM PE=1 SV=2 - [PFKM_HUMAN]
O76074	14	875	99,9	6,09	53	cGMP-specific 3',5'-cyclic phosphodiesterase OS=Homo sapiens GN=PDE5A PE=1 SV=1
Q8NEH6	5	495	60,5	7,12	52	Meiosis-specific nuclear structural protein 1 OS=Homo sapiens GN=MNS1 PE=2 SV=1
Q9Y263	12	795	87,1	6,37	52	Phospholipase A-2-activating protein OS=Homo sapiens GN=PLAA PE=1 SV=2 - [PLAA_HUMAN]
Q8N8U2	34	506	56,5	8,79	52	Chromodomain Y-like protein 2 OS=Homo sapiens GN=CDYL2 PE=2 SV=2 - [CDYL2_HUMAN]
Q15057	7	778	88,0	6,8	52	Arf-GAP with coiled-coil, ANK repeat and PH domain-containing protein 2 OS=Homo sapiens GN=ARFGAP2 PE=1 SV=1
Q9UPS6	7	1923	208,6	4,96	52	Histone-lysine N-methyltransferase SETD1B OS=Homo sapiens GN=SETD1B PE=1 SV=1
Q8IXK2	35	581	66,9	6,8	52	Polypeptide N-acetylgalactosaminyltransferase 12 OS=Homo sapiens GN=GALNT12 PE=1 SV=1
Q9ULJ1	7	636	73,7	6,48	52	Outer dense fiber protein 2-like OS=Homo sapiens GN=ODF2L PE=2 SV=2 - [ODF2L_HUMAN]
Q07343	6	736	83,3	5,25	52	cAMP-specific 3',5'-cyclic phosphodiesterase 4B OS=Homo sapiens GN=PDE4B PE=1 SV=1
Q13619	10	759	87,6	8,13	52	Cullin-4A OS=Homo sapiens GN=CUL4A PE=1 SV=3 - [CUL4A_HUMAN]
Q8WZ75	4	1007	107,4	6,64	52	Roundabout homolog 4 precursor (Magic roundabout) - Homo sapiens (Human) - GN=ROBO4 PE=1 SV=1
Q16236	19	605	67,8	4,78	52	Nuclear factor erythroid 2-related factor 2 OS=Homo sapiens GN=NFE2L2 PE=1 SV=1
Q13332	17	1948	216,9	6,46	52	Receptor-type tyrosine-protein phosphatase S OS=Homo sapiens GN=PTPRS PE=1 SV=1
Q8N4A0	35	578	66,6	7,75	52	Polypeptide N-acetylgalactosaminyltransferase 4 OS=Homo sapiens GN=GALNT4 PE=1 SV=1
Q96AD5	19	504	55,3	7,08	52	Patatin-like phospholipase domain-containing protein 2 OS=Homo sapiens GN=PLD2 PE=1 SV=1
P61247	6	264	29,9	9,73	52	40S ribosomal protein S3a OS=Homo sapiens GN=RPS3A PE=1 SV=2 - [RPS3A_HUMAN]
Q8IUF8	8	465	52,8	6,7	52	MYC-induced nuclear antigen OS=Homo sapiens GN=MINA PE=1 SV=1 - [MINA_HUMAN]
O75558	4	287	33,2	6,55	52	Syntaxin-11 OS=Homo sapiens GN=STX11 PE=1 SV=1 - [STX11_HUMAN]
Q9NVE4	134	849	96,3	8,59	52	Coiled-coil domain-containing protein 87 OS=Homo sapiens GN=CCDC87 PE=2 SV=1
Q14679	34	1199	133,3	8,85	52	Tubulin polyglutamylase TLL4 OS=Homo sapiens GN=TLL4 PE=1 SV=2 - [TLL4_HUMAN]
P50995	44	505	54,4	7,65	52	Annexin A11 OS=Homo sapiens GN=ANXA11 PE=1 SV=1 - [ANXA11_HUMAN]
O75093	45	1534	167,8	6,57	52	Slit homolog 1 protein OS=Homo sapiens GN=SLIT1 PE=2 SV=4 - [SLIT1_HUMAN]
Q96JF6	8	807	93,8	8,7	52	Zinc finger protein 594 OS=Homo sapiens GN=ZNF594 PE=2 SV=3 - [ZNF594_HUMAN]
Q15468	10	1287	142,9	6,46	52	SCL-interrupting locus protein OS=Homo sapiens GN=STIL PE=1 SV=2 - [STIL_HUMAN]
Q9ULT8	5	2610	289,2	5,35	52	E3 ubiquitin-protein ligase HECTD1 OS=Homo sapiens GN=HECTD1 PE=1 SV=3 - [HECTD1_HUMAN]
O60870	11	393	45,3	8,95	52	DNA/RNA-binding protein KIN17 OS=Homo sapiens GN=KIN17 PE=1 SV=2 - [KIN17_HUMAN]
Q96MX3	10	618	67,8	9,36	52	Zinc finger protein 48 OS=Homo sapiens GN=ZNF48 PE=2 SV=2 - [ZNF48_HUMAN]
Q9Y267	9	594	66,6	8,38	51	Solute carrier family 22 member 14 OS=Homo sapiens GN=SLC22A14 PE=2 SV=4 - [SLC22A14_HUMAN]
Q99728	12	777	86,6	8,72	51	BRCA1-associated RING domain protein 1 OS=Homo sapiens GN=BARD1 PE=1 SV=1

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Q5T601	39	910	101,3	8,62	51	Probable G-protein coupled receptor 110 OS=Homo sapiens GN=GPR110 PE=2 SV=
P17302	15	382	43,0	8,76	51	Gap junction alpha-1 protein OS=Homo sapiens GN=GJA1 PE=1 SV=2 - [CXA1_HUM
P41219	9	470	53,6	5,47	51	Peripherin OS=Homo sapiens GN=PRPH PE=1 SV=2 - [PERI_HUMAN]
P06133	7	528	60,5	8,51	51	UDP-glucuronosyltransferase 2B4 OS=Homo sapiens GN=UGT2B4 PE=1 SV=2 - [UD
Q92888	5	912	102,4	5,66	51	Rho guanine nucleotide exchange factor 1 (p115-RhoGEF) (p115RhoGEF) (115 kDa
Q68CJ6	31	796	91,1	8,63	51	GTPase SLIP-GC OS=Homo sapiens GN=C8orf80 PE=1 SV=3 - [SLIP_HUMAN]
Q8NET8	5	790	90,6	6,6	51	Transient receptor potential cation channel subfamily V member 3 OS=Homo sap
O14531	9	572	61,8	7,09	51	Dihydropyrimidinase-related protein 4 OS=Homo sapiens GN=DPYSL4 PE=1 SV=2
Q9BT78	52	406	46,2	5,83	51	COP9 signalosome complex subunit 4 OS=Homo sapiens GN=COPS4 PE=1 SV=1 - [
Q9HCZ1	11	680	79,6	9,16	51	Zinc finger protein 334 - Homo sapiens (Human) - [ZN334_HUMAN]
Q9GIL0	15	193	22,9	9,91	51	UPF0671 protein C14orf153 OS=Homo sapiens GN=C14orf153 PE=1 SV=2 - [CN153
O60568	6	738	84,7	6,05	51	Procollagen-lysine,2-oxoglutarate 5-dioxygenase 3 OS=Homo sapiens GN=PLOD3
A4D150	12	409	42,8	5,87	51	Killer cell lectin-like receptor subfamily G member 2 OS=Homo sapiens GN=KLRG
O75808	19	1086	117,2	6,71	51	Calpain-15 OS=Homo sapiens GN=SOLH PE=1 SV=1 - [CAN15_HUMAN]
Q9NRD9	16	1551	177,1	7,9	51	Dual oxidase 1 OS=Homo sapiens GN=DUOX1 PE=1 SV=1 - [DUOX1_HUMAN]
P13716	20	330	36,3	6,79	51	Delta-aminolevulinic acid dehydratase OS=Homo sapiens GN=ALAD PE=1 SV=1 - [
O00555	3	2505	282,2	8,84	51	Voltage-dependent P/Q-type calcium channel subunit alpha-1A OS=Homo sapier
Q9Y211	11	1504	166,5	5,14	51	Nischarin OS=Homo sapiens GN=NISCH PE=1 SV=3 - [NISCH_HUMAN]
Q8TAA5	20	225	25,4	7,72	51	GrpE protein homolog 2, mitochondrial precursor (Mt-GrpE#2) - Homo sapiens (H
Q3B820	31	660	76,7	8,03	51	Protein FAM161A OS=Homo sapiens GN=FAM161A PE=2 SV=2 - [F161A_HUMAN]
Q5W5X9	6	447	50,0	8,38	51	Tetratricopeptide repeat protein 23 OS=Homo sapiens GN=TTC23 PE=2 SV=1 - [TT
Q96JM3	7	812	89,0	8,44	51	Zinc finger protein 828 OS=Homo sapiens GN=ZNF828 PE=1 SV=2 - [ZN828_HUMAN]
P09758	14	323	35,7	8,87	51	Tumor-associated calcium signal transducer 2 OS=Homo sapiens GN=TACSTD2 PE=
Q9H1X1	23	276	31,3	5,43	51	Radial spoke head protein 9 homolog OS=Homo sapiens GN=RSPH9 PE=1 SV=1 - [f
Q96JQ0	4	3298	346,0	4,94	50	Protocadherin-16 OS=Homo sapiens GN=DCHS1 PE=2 SV=1 - [PCD16_HUMAN]
Q7Z494	23	1330	150,8	6,76	50	Nephrocystin-3 OS=Homo sapiens GN=NPHP3 PE=1 SV=1 - [NPHP3_HUMAN]
Q49MI3	68	558	62,6	8,27	50	Ceramide kinase-like protein OS=Homo sapiens GN=CERKL PE=1 SV=1 - [CERKL_H
P16150	7	400	40,3	5,1	50	Leukosialin OS=Homo sapiens GN=SPN PE=1 SV=1 - [LEUK_HUMAN]
Q9Y5H2	11	935	101,5	4,96	50	Protocadherin gamma-A11 OS=Homo sapiens GN=PCDHGA11 PE=1 SV=1 - [PCDGB
Q9P2B2	15	879	98,5	6,61	50	Prostaglandin F2 receptor negative regulator OS=Homo sapiens GN=PTGFRN PE=
Q9UPN9	11	1127	122,5	6,67	50	E3 ubiquitin-protein ligase TRIM33 OS=Homo sapiens GN=TRIM33 PE=1 SV=3 - [TR
P52895	11	323	36,7	7,49	50	Aldo-keto reductase family 1 member C2 OS=Homo sapiens GN=AKR1C2 PE=1 SV=
Q86YH6	29	399	44,1	8,24	50	Decaprenyl-diphosphate synthase subunit 2 OS=Homo sapiens GN=PDSS2 PE=1 S'
P00558	4	417	44,6	8,1	50	Phosphoglycerate kinase 1 (EC 2.7.2.3) (Primer recognition protein 2) (PRP 2) - Hc
Q96L96	6	1907	201,1	7,58	50	Alpha-protein kinase 3 OS=Homo sapiens GN=ALPK3 PE=2 SV=2 - [ALPK3_HUMAN
Q9H6Q4	5	476	53,0	7,23	50	Cytosolic Fe-S cluster assembly factor NARFL OS=Homo sapiens GN=NARFL PE=1 S
Q12767	6	1356	151,1	6,44	50	Uncharacterized protein KIAA0195 OS=Homo sapiens GN=KIAA0195 PE=1 SV=1 - [I
Q7Z401	5	1863	209,1	7,31	50	C-myc promoter-binding protein OS=Homo sapiens GN=DENND4A PE=1 SV=2 - [M
P35240	8	595	69,6	6,47	50	Merlin OS=Homo sapiens GN=NF2 PE=1 SV=1 - [MERL_HUMAN]
Q4G0T1	4	1027	108,5	6,15	50	Putative scavenger receptor cysteine-rich domain-containing protein LOC619207
Q8NDM7	4	1665	191,9	5,99	50	WD repeat-containing protein C10orf79 OS=Homo sapiens GN=C10orf79 PE=2 SV=
Q9BQK8	4	851	93,6	5,52	50	Phosphatidate phosphatase LPIN3 OS=Homo sapiens GN=LPIN3 PE=1 SV=3 - [LPIN
Q9H825	16	291	33,4	6,95	50	Methyltransferase-like protein 8 OS=Homo sapiens GN=METTL8 PE=2 SV=2 - [MET
Q9H992	6	704	78,0	6,77	50	E3 ubiquitin-protein ligase MARCH7 OS=Homo sapiens GN=MARCH7 PE=1 SV=1 - [
P52746	14	1687	187,8	7,91	50	Zinc finger protein 142 OS=Homo sapiens GN=ZNF142 PE=1 SV=4 - [ZN142_HUMAN]
Q8TD22	6	1067	117,8	6,4	50	NEDD9-interacting protein with calponin homology and LIM domains OS=Homo s
Q9NP72	6	206	23,0	5,24	50	Ras-related protein Rab-18 OS=Homo sapiens GN=RAB18 PE=1 SV=1 - [RAB18_HUI
Q6Z5B9	4	765	85,0	6,98	50	Zinc finger and BTB domain-containing protein 49 OS=Homo sapiens GN=ZBTB49 I
Q9UHK0	5	495	56,3	9,13	50	Nuclear fragile X mental retardation-interacting protein 1 OS=Homo sapiens GN=
Q6PJ61	6	646	69,0	7,99	50	F-box only protein 46 OS=Homo sapiens GN=FBXO46 PE=1 SV=2 - [FBX46_HUMAN
Q6QEF8	18	472	52,7	5,96	50	Coronin-6 OS=Homo sapiens GN=CORO6 PE=1 SV=2 - [CORO6_HUMAN]
Q8IY17	7	1366	149,9	7,81	50	Neuropathy target esterase OS=Homo sapiens GN=PNPLA6 PE=1 SV=2 - [PLPL6_H
Q9BWW1	16	1114	121,0	7,01	49	Brother of CDO OS=Homo sapiens GN=BOC PE=1 SV=1 - [BOC_HUMAN]
Q9UBL3	13	628	68,7	5,69	49	Set1/Ash2 histone methyltransferase complex subunit ASH2 (ASH2-like protein)
Q14289	7	1009	115,8	6,25	49	Protein-tyrosine kinase 2-beta OS=Homo sapiens GN=PTK2B PE=1 SV=2 - [FAK2_H
Q9UIK4	6	370	42,9	6,92	49	Death-associated protein kinase 2 OS=Homo sapiens GN=DAPK2 PE=1 SV=1 - [DAI
O95684	48	399	43,0	4,81	49	FGFR1 oncogene partner OS=Homo sapiens GN=FGFR1OP PE=1 SV=1 - [FR1OP_HU
Q55YE7	14	1610	170,6	6,96	49	NHS-like protein 1 OS=Homo sapiens GN=NHSL1 PE=1 SV=2 - [NHSL1_HUMAN]
Q8NOX2	10	631	70,8	6,33	49	Sperm-associated antigen 16 protein OS=Homo sapiens GN=SPAG16 PE=2 SV=2 - [
Q8TDR2	11	534	58,0	9,74	49	Serine/threonine-protein kinase 35 OS=Homo sapiens GN=STK35 PE=1 SV=2 - [STI
Q5TEA3	12	1177	132,2	6,57	49	Uncharacterized protein C20orf194 OS=Homo sapiens GN=C20orf194 PE=2 SV=1 - [
Q8NBIG	168	393	43,8	8,13	49	Uncharacterized protein C3orf21 OS=Homo sapiens GN=C3orf21 PE=2 SV=1 - [CCO
Q6P3X8	14	592	68,0	8,54	49	PiggyBac transposable element-derived protein 2 OS=Homo sapiens GN=PGBD2 I
Q70UQ0	15	350	39,3	9,17	49	Inhibitor of nuclear factor kappa-B kinase-interacting protein OS=Homo sapiens C
P42680	8	631	73,5	8,44	49	Tyrosine-protein kinase Tec OS=Homo sapiens GN=TEC PE=1 SV=2 - [TEC_HUMAN

Appendices

Q92621	9	2012	227,8	6,19	49	Nuclear pore complex protein Nup205 OS=Homo sapiens GN=NUP205 PE=1 SV=3
Q9UBJ2	11	740	83,2	8,92	49	ATP-binding cassette sub-family D member 2 OS=Homo sapiens GN=ABCD2 PE=1
P61164	10	376	42,6	6,64	49	Alpha-centractin OS=Mus musculus GN=Actr1a PE=2 SV=1 - [ACTZ_MOUSE]
Q9NRJ5	15	636	71,6	6,44	49	Poly(A) polymerase beta OS=Homo sapiens GN=PAPOLB PE=2 SV=1 - [PAPOB_HU]
Q86UX7	11	667	75,9	6,98	49	Fermitin family homolog 3 OS=Homo sapiens GN=FERMT3 PE=1 SV=1 - [URP2_HUI]
P55011	6	1212	131,4	6,4	49	Solute carrier family 12 member 2 OS=Homo sapiens GN=SLC12A2 PE=1 SV=1 - [S1]
Q9HC36	31	420	47,0	8,73	49	RNA methyltransferase-like protein 1 OS=Homo sapiens GN=RNMTL1 PE=1 SV=2 -
Q96PQ7	6	755	84,4	6,64	49	Kelch-like protein 5 OS=Homo sapiens GN=KLHL5 PE=2 SV=3 - [KLHL5_HUMAN]
Q15326	5	562	66,2	8,22	49	Zinc finger MYND domain-containing protein 11 OS=Homo sapiens GN=ZMYND11
C9J069	27	976	106,6	9,03	49	Uncharacterized protein C9orf172 OS=Homo sapiens GN=C9orf172 PE=3 SV=1 - [C]
Q15424	8	915	102,6	5,47	49	Scaffold attachment factor B1 OS=Homo sapiens GN=SAFB PE=1 SV=4 - [SAFB1_HL]
O60812	13	293	32,1	5,06	49	Heterogeneous nuclear ribonucleoprotein C-like 1 OS=Homo sapiens GN=HNRNF
Q494X1	5	144	16,6	8,88	49	Protein FAM153C OS=Homo sapiens GN=FAM153C PE=2 SV=1 - [F153C_HUMAN]
Q15561	10	434	48,3	7,33	49	Transcriptional enhancer factor TEF-3 OS=Homo sapiens GN=TEAD4 PE=1 SV=3 - [T]
O43566	144	566	61,4	8,19	48	Regulator of G-protein signaling 14 OS=Homo sapiens GN=RG514 PE=1 SV=4 - [RG'
P51795	16	746	83,1	6,81	48	H(+)/Cl(-) exchange transporter 5 OS=Homo sapiens GN=CLCN5 PE=1 SV=1 - [CLCN
O15335	36	359	40,5	9,39	48	Chondroadherin OS=Homo sapiens GN=CHAD PE=2 SV=2 - [CHAD_HUMAN]
Q8IXQ6	27	854	96,3	7,91	48	Poly [ADP-ribose] polymerase 9 OS=Homo sapiens GN=PARP9 PE=1 SV=2 - [PARP'
Q96ST8	4	783	89,5	6,8	48	Coiled-coil domain-containing protein 123, mitochondrial OS=Homo sapiens GN=
Q9UMS0	6	254	28,4	5,07	48	NFU1 iron-sulfur cluster scaffold homolog, mitochondrial OS=Homo sapiens GN=
Q15025	19	636	71,8	6,62	48	TNFAIP3-interacting protein 1 OS=Homo sapiens GN=TNIP1 PE=1 SV=2 - [TNIP1_H
P11498	6	1178	129,6	6,84	48	Pyruvate carboxylase, mitochondrial OS=Homo sapiens GN=PC PE=1 SV=2 - [PYC_
Q8IUC6	16	712	76,4	5,41	48	TIR domain-containing adapter molecule 1 OS=Homo sapiens GN=TICAM1 PE=1 S'
Q8NDL9	3	886	97,5	9,17	48	Cytosolic carboxypeptidase-like protein 5 OS=Homo sapiens GN=AGBL5 PE=2 SV=
Q16589	24	344	38,8	5,48	48	Cyclin-G2 OS=Homo sapiens GN=CCNG2 PE=1 SV=1 - [CCNG2_HUMAN]
Q02446	5	784	81,9	7,05	48	Transcription factor Sp4 OS=Homo sapiens GN=SP4 PE=1 SV=2 - [SP4_HUMAN]
Q96JN2	5	1154	134,0	4,65	48	Coiled-coil domain-containing protein 136 OS=Homo sapiens GN=CCDC136 PE=1 S
P35221	4	906	100,0	6,29	48	Catenin alpha-1 (Cadherin-associated protein) (Alpha E-catenin) (NY-REN-13 anti
Q9H0M0	6	922	105,1	5,9	48	NEDD4-like E3 ubiquitin-protein ligase WWP1 OS=Homo sapiens GN=WWP1 PE=1
Q96JX3	9	654	74,1	7,68	48	Protein SERAC1 OS=Homo sapiens GN=SERAC1 PE=2 SV=1 - [SRAC1_HUMAN]
Q4G148	14	440	50,5	8,65	48	Glucoside xylosyltransferase 1 OS=Homo sapiens GN=GXYLT1 PE=1 SV=2 - [GXYLT1
Q9H2S9	3	585	64,1	6,86	48	Zinc finger protein Eos OS=Homo sapiens GN=IKZF4 PE=1 SV=2 - [IKZF4_HUMAN]
Q8N1V2	10	620	68,3	6,95	48	WD repeat-containing protein 16 OS=Homo sapiens GN=WDR16 PE=1 SV=3 - [WDR
Q9NP99	7	234	26,4	8,84	48	Triggering receptor expressed on myeloid cells 1 OS=Homo sapiens GN=TREM1 P
P31939	10	592	64,6	6,71	48	Bifunctional purine biosynthesis protein PURH OS=Homo sapiens GN=ATIC PE=1 S
Q00839	8	825	90,5	6	48	Heterogeneous nuclear ribonucleoprotein U OS=Homo sapiens GN=HNRNPU PE=
Q96MT3	17	831	94,2	6,24	48	Prickle-like protein 1 OS=Homo sapiens GN=PRICKLE1 PE=1 SV=2 - [PRIC1_HUMAN
Q0VDD8	54	3507	399,6	6,93	48	Dynein heavy chain 14, axonemal OS=Homo sapiens GN=DNAH14 PE=2 SV=3 - [DY
P17024	11	532	61,5	8,62	48	Zinc finger protein 20 OS=Homo sapiens GN=ZNF20 PE=2 SV=2 - [ZNF20_HUMAN]
Q9BSK4	21	669	73,6	6,07	48	Protein fem-1 homolog A OS=Homo sapiens GN=FEM1A PE=1 SV=1 - [FEM1A_HU]
Q9NVH2	7	962	106,8	8,02	48	Integrator complex subunit 7 OS=Homo sapiens GN=INTS7 PE=1 SV=1 - [INT7_HU]
O14793	3	375	42,7	6,76	48	Growth/differentiation factor 8 OS=Homo sapiens GN=MSTN PE=1 SV=1 - [GDF8_
O60733	17	806	89,8	7,27	48	85 kDa calcium-independent phospholipase A2 OS=Homo sapiens GN=PLA2G6 PE
Q9Y651	16	276	28,6	9,73	47	Transcription factor SOX-21 OS=Homo sapiens GN=SOX21 PE=2 SV=1 - [SOX21_HU
Q9BWF2	10	469	53,3	8,48	47	TRAF-interacting protein OS=Homo sapiens GN=TRAIIP PE=1 SV=1 - [TRAIIP_HUMA
Q9Y6R7	6	5405	571,6	5,34	47	IgGfC-binding protein OS=Homo sapiens GN=FCGBP PE=1 SV=3 - [FCGBP_HUMAN]
O35593	11	310	34,6	6,52	47	26S proteasome non-ATPase regulatory subunit 14 OS=Mus musculus GN=Psmd1
Q9NZ56	17	1722	180,0	5,47	47	Formin-2 OS=Homo sapiens GN=FMN2 PE=1 SV=4 - [FMN2_HUMAN]
Q2M329	40	555	62,7	4,94	47	Coiled-coil domain-containing protein 96 OS=Homo sapiens GN=CCDC96 PE=2 SV
Q96RK0	10	1608	163,7	8,56	47	Protein capicua homolog OS=Homo sapiens GN=CIC PE=1 SV=2 - [CIC_HUMAN]
Q9P2E2	6	1029	115,0	5,5	47	Kinesin-like protein KIF17 OS=Homo sapiens GN=KIF17 PE=1 SV=3 - [KIF17_HUMA
Q9Y620	13	910	102,9	8,12	47	DNA repair and recombination protein RAD54B OS=Homo sapiens GN=RAD54B PE
O60307	12	1309	143,0	8,06	47	Microtubule-associated serine/threonine-protein kinase 3 OS=Homo sapiens GN
P14625	11	803	92,4	4,84	47	Endoplasmic reticulum protein OS=Homo sapiens GN=HSP90B1 PE=1 SV=1 - [ENPL_HUMAN]
Q9GZU2	12	1588	180,7	5,48	47	Paternally-expressed gene 3 protein OS=Homo sapiens GN=PEG3 PE=2 SV=1 - [PE
Q86V48	5	1076	120,2	8,5	47	Leucine zipper protein 1 OS=Homo sapiens GN=LUZP1 PE=1 SV=2 - [LUZP1_HUMA]
Q658Y4	27	838	93,8	6,39	47	Protein FAM91A1 OS=Homo sapiens GN=FAM91A1 PE=1 SV=3 - [F91A1_HUMAN]
Q9BTE3	20	642	72,9	5,87	47	UPF0557 protein C10orf119 OS=Homo sapiens GN=C10orf119 PE=1 SV=2 - [C119_
Q8NEF3	12	446	53,5	9,48	47	Coiled-coil domain-containing protein 112 OS=Homo sapiens GN=CCDC112 PE=2 S
Q5QJ66	90	756	84,4	6,16	47	Deoxynucleotidyltransferase terminal-interacting protein 2 OS=Homo sapiens G
Q8TBPO	55	767	86,3	5,82	47	TBC1 domain family member 16 OS=Homo sapiens GN=TBC1D16 PE=1 SV=1 - [TBC
Q9HG6Y	151	334	35,2	10,81	47	PDZ domain-containing protein MAGIX OS=Homo sapiens GN=MAGIX PE=1 SV=3 -
P23141	9	567	62,5	6,6	47	Liver carboxylesterase 1 OS=Homo sapiens GN=CES1 PE=1 SV=2 - [EST1_HUMAN]
A6NNM8	7	815	93,6	8,91	47	Tubulin polyglutamylase TLL13 OS=Homo sapiens GN=TLL13 PE=1 SV=2 - [TLL13
Q9NQX6	16	463	53,7	8,91	47	Zinc finger protein 331 (Zinc finger protein 463) (C2H2-like zinc finger protein rea

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P23470	19	1445	161,9	6,42	47	Receptor-type tyrosine-protein phosphatase gamma OS=Homo sapiens GN=PTPF
Q03252	4	600	67,6	5,35	47	Lamin-B2 OS=Homo sapiens GN=LMNB2 PE=1 SV=3 - [LMNB2_HUMAN]
P32927	9	897	97,3	5,52	47	Cytokine receptor common subunit beta OS=Homo sapiens GN=CSF2RB PE=1 SV=
P00480	50	354	39,9	8,63	47	Ornithine carbamoyltransferase, mitochondrial precursor (EC 2.1.3.3) (OTCase) (C
Q9H0I9	19	626	67,8	6,33	47	Transketolase-like protein 2 OS=Homo sapiens GN=TKTL2 PE=1 SV=1 - [TKTL2_HUI
Q62WJ8	10	1503	159,8	5,74	47	Kielin/chordin-like protein OS=Homo sapiens GN=KCP PE=2 SV=2 - [KCP_HUMAN]
Q8TBC3	16	707	76,3	8,28	47	SH3KBP1-binding protein 1 OS=Homo sapiens GN=SHKBP1 PE=1 SV=2 - [SHKB1_HU
Q13825	10	339	35,6	9,48	46	Methylglutaconyl-CoA hydratase, mitochondrial OS=Homo sapiens GN=AUH PE=1
P09327	6	827	92,6	6,39	46	Villin-1 OS=Homo sapiens GN=VIL1 PE=1 SV=4 - [VILI_HUMAN]
O15075	13	740	82,2	8,66	46	Serine/threonine-protein kinase DCLK1 OS=Homo sapiens GN=DCLK1 PE=1 SV=2 -
Q8WUM0	31	1156	128,9	5,1	46	Nuclear pore complex protein Nup133 OS=Homo sapiens GN=NUP133 PE=1 SV=2 -
Q15825	24	494	56,9	6,62	46	Neuronal acetylcholine receptor subunit alpha-6 OS=Homo sapiens GN=CHRNA6
Q9NY12	6	217	22,3	10,92	46	H/ACA ribonucleoprotein complex subunit 1 OS=Homo sapiens GN=GAR1 PE=1 SV
P19634	3	815	90,7	7,21	46	Sodium/hydrogen exchanger 1 OS=Homo sapiens GN=SLC9A1 PE=1 SV=2 - [SL9A1
O75683	10	361	41,4	10,64	46	Surfeit locus protein 6 OS=Homo sapiens GN=SURF6 PE=1 SV=3 - [SURF6_HUMAN]
P15311	96	586	69,4	6,27	46	Ezrin OS=Homo sapiens GN=EZR PE=1 SV=4 - [EZRI_HUMAN]
Q76M96	9	950	108,1	9,72	46	Coiled-coil domain-containing protein 80 OS=Homo sapiens GN=CCDC80 PE=1 SV=
Q5T0Z8	15	1188	124,0	9,42	46	Uncharacterized protein C6orf132 OS=Homo sapiens GN=C6orf132 PE=1 SV=3 - [C6
Q9COB5	26	715	77,5	9,01	46	Probable palmitoyltransferase ZDHHC5 OS=Homo sapiens GN=ZDHHC5 PE=1 SV=2
P56730	15	875	97,0	8,03	46	Neurotrypsin OS=Homo sapiens GN=PRSS12 PE=2 SV=2 - [NETR_HUMAN]
Q15746	11	1914	210,6	6,15	46	Myosin light chain kinase, smooth muscle OS=Homo sapiens GN=MYLK PE=1 SV=4
Q5M8T2	14	416	44,2	7,27	46	Solute carrier family 35 member D3 OS=Homo sapiens GN=SLC35D3 PE=2 SV=1 - [S
Q96C03	4	454	49,2	5,12	46	Smith-Magenis syndrome chromosomal region candidate gene 7 protein OS=Hon
P05181	3	493	56,8	8,1	46	Cytochrome P450 2E1 OS=Homo sapiens GN=CYP2E1 PE=1 SV=1 - [CP2E1_HUMAN]
Q9UI12	4	483	55,8	6,48	46	Vacuolar ATP synthase subunit H (EC 3.6.3.14) (V-ATPase H subunit) (Vacuolar pro
Q86TC9	6	1320	145,2	6,77	46	Myopalladin OS=Homo sapiens GN=MYPN PE=1 SV=2 - [MYPN_HUMAN]
Q9UJV3	5	735	83,2	7,39	46	Probable E3 ubiquitin-protein ligase MID2 OS=Homo sapiens GN=MID2 PE=1 SV=3
Q16222	13	522	58,7	6,33	46	UDP-N-acetylhexosamine pyrophosphorylase OS=Homo sapiens GN=UAP1 PE=1 SV
Q9BSV6	58	310	33,6	8,43	46	tRNA-splicing endonuclease subunit Sen34 OS=Homo sapiens GN=TSEN34 PE=1 SV
Q5VTE6	3	544	62,3	7,81	46	Protein angel homolog 2 OS=Homo sapiens GN=ANGEL2 PE=2 SV=1 - [ANGE2_HUM
Q12912	109	555	62,1	5,85	46	Lymphoid-restricted membrane protein OS=Homo sapiens GN=LRMP PE=1 SV=3 -
Q9NZN5	16	1544	173,1	5,74	46	Rho guanine nucleotide exchange factor 12 OS=Homo sapiens GN=ARHGEF12 PE=
Q01581	50	520	57,3	5,41	46	Hydroxymethylglutaryl-CoA synthase, cytoplasmic (EC 2.3.3.10) (HMG-CoA synth
Q3MIX3	4	580	65,9	8,97	45	Uncharacterized narF domain-containing protein kinase 5 OS=Homo sapiens GN=
O14744	6	637	72,6	6,29	45	Protein arginine N-methyltransferase 5 OS=Homo sapiens GN=PRMT5 PE=1 SV=4
P10074	5	688	77,0	8,18	45	Zinc finger and BTB domain-containing protein 48 OS=Homo sapiens GN=ZBTB48 I
P81408	14	668	71,3	8,46	45	Protein FAM189B OS=Homo sapiens GN=FAM189B PE=1 SV=2 - [F189B_HUMAN]
Q9HD64	8	160	17,7	9,67	45	G antigen family D member 2 OS=Homo sapiens GN=XAG6A1 PE=2 SV=2 - [GAGD2
P06331	11	146	16,2	8,28	45	Ig heavy chain V-II region ARH-77 OS=Homo sapiens PE=4 SV=1 - [HV209_HUMAN]
P19525	3	551	62,1	8,4	45	Interferon-induced, double-stranded RNA-activated protein kinase OS=Homo sa
Q726M2	10	555	62,6	7,36	45	F-box only protein 33 OS=Homo sapiens GN=FBXO33 PE=2 SV=1 - [FBX33_HUMAN]
Q9NP61	9	516	56,9	7,36	45	ADP-ribosylation factor GTPase-activating protein 3 OS=Homo sapiens GN=ARFG/
P17676	6	345	36,1	8,31	45	CCAAT/enhancer-binding protein beta OS=Homo sapiens GN=CEBPB PE=1 SV=2 -
O95759	25	1140	130,8	5,52	45	TBC1 domain family member 8 OS=Homo sapiens GN=TBC1D8 PE=1 SV=3 - [TBCD8
Q0IIM8	34	1120	128,6	5,95	45	TBC1 domain family member 8B OS=Homo sapiens GN=TBC1D8B PE=2 SV=2 - [TBC
Q99575	9	1024	114,6	9,22	45	Ribonucleases P/MRP protein subunit POP1 OS=Homo sapiens GN=POP1 PE=1 SV
O15446	8	510	55,0	8,51	45	DNA-directed RNA polymerase I subunit RPA34 OS=Homo sapiens GN=CD3EAP PE
P01612	7	109	11,9	6,57	45	Ig kappa chain V-I region Mev - Homo sapiens (Human) - [KV120_HUMAN]
Q86UE8	44	772	87,6	8,41	45	Serine/threonine-protein kinase tousled-like 2 OS=Homo sapiens GN=TLK2 PE=1
O95948	21	504	54,3	9,61	45	One cut domain family member 2 OS=Homo sapiens GN=ONECUT2 PE=2 SV=2 - [C
Q96S11	9	283	31,9	7,44	45	BTB/POZ domain-containing protein KCTD15 OS=Homo sapiens GN=KCTD15 PE=1
O60662	4	606	68,0	5,29	45	Kelch repeat and BTB domain-containing protein 10 OS=Homo sapiens GN=KBTBL
Q6NT55	4	531	61,9	8,73	45	Cytochrome P450 4F22 OS=Homo sapiens GN=CYP4F22 PE=2 SV=1 - [CP4FN_HUM
Q8WXK1	15	588	65,8	5,83	45	Ankyrin repeat and SOCS box protein 15 OS=Homo sapiens GN=ASB15 PE=2 SV=3 -
O94827	5	1062	117,4	6,3	45	Pleckstrin homology domain-containing family G member 5 OS=Homo sapiens GN
Q7KYR7	6	527	59,6	6,48	45	Butyrophilin subfamily 2 member A1 OS=Homo sapiens GN=BTN2A1 PE=1 SV=3 -]
P06401	4	933	98,9	6,49	45	Progesterone receptor OS=Homo sapiens GN=PGR PE=1 SV=4 - [PRGR_HUMAN]
Q86XX4	8	4007	442,6	5,59	45	Extracellular matrix protein FRAS1 OS=Homo sapiens GN=FRAS1 PE=2 SV=1 - [FRA
O75818	8	363	41,8	6,67	45	Ribonuclease P protein subunit p40 OS=Homo sapiens GN=RPP40 PE=1 SV=3 - [RP
P78318	22	339	39,2	5,38	45	Immunoglobulin-binding protein 1 OS=Homo sapiens GN=IGBP1 PE=1 SV=1 - [IGB
Q6P1J9	2	531	60,5	9,61	45	Parafibromin OS=Homo sapiens GN=CDC73 PE=1 SV=1 - [CDC73_HUMAN]
P51451	31	505	57,7	7,87	45	Tyrosine-protein kinase Blk OS=Homo sapiens GN=BLK PE=1 SV=3 - [BLK_HUMAN]
Q96CX6	7	371	40,6	6,84	45	Leucine-rich repeat-containing protein 58 OS=Homo sapiens GN=LRRC58 PE=1 SV
Q8N1G4	24	583	63,4	8,28	45	Leucine-rich repeat-containing protein 47 OS=Homo sapiens GN=LRRC47 PE=1 SV
Q9UKD2	33	239	27,5	8,29	44	mRNA turnover protein 4 homolog OS=Homo sapiens GN=MRTO4 PE=1 SV=2 - [MI

Appendices

Q9BXW9	7	1471	166,4	6,24	44	Fanconi anemia group D2 protein OS=Homo sapiens GN=FANCD2 PE=1 SV=1 - [FA
P08174	10	381	41,4	7,59	44	Complement decay-accelerating factor OS=Homo sapiens GN=CD55 PE=1 SV=4 - [
Q8WZ94	3	311	34,3	8,31	44	Olfactory receptor 5P3 OS=Homo sapiens GN=OR5P3 PE=2 SV=1 - [OR5P3_HUMAN
P49754	12	854	98,5	5,85	44	Vacuolar protein sorting-associated protein 41 homolog OS=Homo sapiens GN=V
O43639	11	380	42,9	6,95	44	Cytoplasmic protein NCK2 OS=Homo sapiens GN=NCK2 PE=1 SV=2 - [NCK2_HUMA
Q6ZUL3	4	223	24,7	9,45	44	Uncharacterized protein C8orf86 OS=Homo sapiens GN=C8orf86 PE=2 SV=1 - [CHO
O75762	14	1119	127,4	7,12	44	Transient receptor potential cation channel subfamily A member 1 OS=Homo sap
Q16854	14	277	32,0	8,66	44	Deoxyguanosine kinase, mitochondrial OS=Homo sapiens GN=DGUOK PE=1 SV=2
P78333	17	572	63,7	6,81	44	Glypican-5 OS=Homo sapiens GN=GPC5 PE=1 SV=1 - [GPC5_HUMAN]
P36955	2	418	46,3	6,38	44	Pigment epithelium-derived factor OS=Homo sapiens GN=SERPINF1 PE=1 SV=4 - [
Q9BXX0	6	1053	115,6	6,46	44	EMILIN-2 OS=Homo sapiens GN=EMILIN2 PE=1 SV=3 - [EMIL2_HUMAN]
Q6PIW4	11	674	74,0	7,85	44	Fidgetin-like protein 1 OS=Homo sapiens GN=FIGL1 PE=1 SV=2 - [FIGL1_HUMAN
Q9BXT2	16	260	28,1	9,17	44	Voltage-dependent calcium channel gamma-6 subunit OS=Homo sapiens GN=CA
P30740	7	379	42,7	6,28	44	Leukocyte elastase inhibitor OS=Homo sapiens GN=SERPINB1 PE=1 SV=1 - [ILEU_
Q8TDY3	12	377	41,7	5,47	44	Actin-related protein T2 OS=Homo sapiens GN=ACTRT2 PE=2 SV=2 - [ACTT2_HUM
P37802	2	199	22,4	8,25	44	Transgelin-2 (SM22-alpha homolog) - Homo sapiens (Human) - [TAGL2_HUMAN]
B4DS77	80	424	46,9	8,66	44	Protein shisa-9 OS=Homo sapiens GN=SHISA9 PE=2 SV=3 - [SHSA9_HUMAN]
Q9NX95	3	663	72,3	6,25	44	Syntabulin OS=Homo sapiens GN=SYBU PE=1 SV=2 - [SYBU_HUMAN]
Q15633	10	366	39,0	6,54	44	RISC-loading complex subunit TARBP2 OS=Homo sapiens GN=TARBP2 PE=1 SV=3 -
O60258	32	216	24,9	10,42	44	Fibroblast growth factor 17 OS=Homo sapiens GN=FGF17 PE=2 SV=1 - [FGF17_HUN
A6NES4	18	1706	193,1	6,68	44	HEAT repeat-containing protein 7B1 OS=Homo sapiens GN=HEATR7B1 PE=4 SV=3 -
Q96PH1	12	765	86,4	8,59	44	NADPH oxidase 5 OS=Homo sapiens GN=NOX5 PE=1 SV=1 - [NOX5_HUMAN]
Q9NVP4	3	579	62,9	7,59	44	Ankyrin repeat-containing protein C20orf12 OS=Homo sapiens GN=C20orf12 PE=2
P01920	14	261	30,0	7,25	44	HLA class II histocompatibility antigen, DQ beta 1 chain OS=Homo sapiens GN=HL
Q9P2D8	9	2635	295,1	6,32	43	Protein unc-79 homolog OS=Homo sapiens GN=KIAA1409 PE=2 SV=3 - [UNC79_HU
Q7L622	15	706	80,5	7,65	43	G2/M phase-specific E3 ubiquitin-protein ligase OS=Homo sapiens GN=G2E3 PE=1
Q577W7	12	516	58,2	7,2	43	Thiosulfate sulfurtransferase/rhodanese-like domain-containing protein 2 OS=H
Q8N137	4	903	101,2	5,54	43	Centrobilin OS=Homo sapiens GN=CENTROB PE=1 SV=1 - [CNTRB_HUMAN]
Q9Y5N6	10	252	28,1	8,66	43	Origin recognition complex subunit 6 OS=Homo sapiens GN=ORC6 PE=1 SV=1 - [O
O15321	9	606	68,8	7,17	43	Transmembrane 9 superfamily member 1 OS=Homo sapiens GN=TM9SF1 PE=2 SV
Q8WY54	45	764	84,9	5,03	43	Protein phosphatase 1E OS=Homo sapiens GN=PPM1E PE=1 SV=2 - [PPM1E_HUM
Q13733	9	1029	114,1	6,64	43	Sodium/potassium-transporting ATPase subunit alpha-4 OS=Homo sapiens GN=A
P01715	18	106	11,3	5,24	43	Ig lambda chain V-IV region Bau OS=Homo sapiens PE=1 SV=1 - [LV401_HUMAN]
P06889	18	106	11,3	4,42	43	Ig lambda chain V-IV region MOL OS=Homo sapiens PE=1 SV=1 - [LV405_HUMAN]
Q7RTX1	7	841	93,0	7,97	43	Taste receptor type 1 member 1 OS=Homo sapiens GN=TAS1R1 PE=2 SV=1 - [TS1R
Q17RN3	3	349	37,3	7,18	43	Protein FAM98C OS=Homo sapiens GN=FAM98C PE=2 SV=1 - [FA98C_HUMAN]
Q8TD55	16	490	53,3	5,43	43	Pleckstrin homology domain-containing family O member 2 OS=Homo sapiens Gi
O75888	3	250	27,4	9,63	43	Tumor necrosis factor ligand superfamily member 13 OS=Homo sapiens GN=TNFS
Q9Y6C2	5	1016	106,6	5,15	43	EMILIN-1 OS=Homo sapiens GN=EMILIN1 PE=1 SV=2 - [EMIL1_HUMAN]
P48357	4	1165	132,4	6,52	43	Leptin receptor OS=Homo sapiens GN=LEPR PE=1 SV=2 - [LEPR_HUMAN]
Q14146	21	1524	170,4	7,31	43	Unhealthy ribosome biogenesis protein 2 homolog OS=Homo sapiens GN=URB2 F
Q9BV36	3	600	65,9	6,07	43	Melanophilin OS=Homo sapiens GN=MLPH PE=1 SV=1 - [MELPH_HUMAN]
Q86YP4	9	633	68,0	9,94	43	Transcriptional repressor p66-alpha OS=Homo sapiens GN=GATAD2A PE=1 SV=1 -
O43148	34	476	54,8	6,61	43	mRNA cap guanine-N7 methyltransferase OS=Homo sapiens GN=RNMT PE=1 SV=
Q9HCB6	12	807	90,9	6,11	43	Spondin-1 OS=Homo sapiens GN=SPON1 PE=1 SV=2 - [SPON1_HUMAN]
O43615	8	452	51,3	8,32	43	Mitochondrial import inner membrane translocase subunit TIM44 OS=Homo sapi
Q92556	3	727	83,8	6,28	43	Engulfment and cell motility protein 1 OS=Homo sapiens GN=ELMO1 PE=1 SV=2 -
Q96NB3	6	372	42,0	5,31	43	Zinc finger protein 830 OS=Homo sapiens GN=ZNF830 PE=1 SV=2 - [ZN830_HUMA
Q96JP2	3	1530	167,0	8,41	43	Putative myosin-XVB OS=Homo sapiens GN=MYO15B PE=5 SV=2 - [MY15B_HUMA
Q68J44	16	220	25,3	6,04	43	Dual specificity phosphatase DUPD1 OS=Homo sapiens GN=DUPD1 PE=2 SV=1 - [D
Q9NXN4	55	497	56,2	5,74	43	Ganglioside-induced differentiation-associated protein 2 OS=Homo sapiens GN=
P14780	10	707	78,4	6,06	42	Matrix metalloproteinase-9 OS=Homo sapiens GN=MMP9 PE=1 SV=3 - [MMP9_HU
Q8IY50	14	662	76,0	7,24	42	GRAM domain-containing protein 1C OS=Homo sapiens GN=GRAMD1C PE=2 SV=2
P01699	3	111	11,5	5,29	42	Ig lambda chain V-I region VOR OS=Homo sapiens PE=1 SV=1 - [LV101_HUMAN]
Q9NWN3	5	711	78,7	8	42	F-box only protein 34 OS=Homo sapiens GN=FBXO34 PE=1 SV=2 - [FBX34_HUMAN
Q8NDY6	10	225	23,7	9,22	42	Class E basic helix-loop-helix protein 23 OS=Homo sapiens GN=BHLHE23 PE=2 SV=
Q9H4K7	14	406	43,9	9,45	42	GTP-binding protein 5 OS=Homo sapiens GN=GTPBP5 PE=2 SV=1 - [GTPB5_HUMA
Q9P283	8	1151	125,8	7,65	42	Semaphorin-5B OS=Homo sapiens GN=SEMA5B PE=2 SV=4 - [SEM5B_HUMAN]
P43490	34	491	55,5	7,15	42	Nicotinamide phosphoribosyltransferase OS=Homo sapiens GN=NAMPT PE=1 SV:
Q9BY31	13	904	105,2	8,57	42	Zinc finger protein 717 OS=Homo sapiens GN=ZNF717 PE=1 SV=2 - [ZN717_HUMA
Q15842	7	424	47,9	9,26	42	ATP-sensitive inward rectifier potassium channel 8 OS=Homo sapiens GN=KCNJ8
O43299	3	807	88,5	7,01	42	Uncharacterized protein KIAA0415 OS=Homo sapiens GN=KIAA0415 PE=2 SV=2 - [I
A6NM62	6	1247	140,7	7,91	42	Leucine-rich repeat-containing protein 53 OS=Homo sapiens GN=LRRC53 PE=4 SV:
Q14671	6	1186	126,4	6,84	42	Pumilio homolog 1 OS=Homo sapiens GN=PUM1 PE=1 SV=3 - [PUM1_HUMAN]
P07814	14	1512	170,5	7,33	42	Bifunctional aminoacyl-tRNA synthetase OS=Homo sapiens GN=EPRS PE=1 SV=5 -

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Q8WVL7	12	239	27,3	5,15	42	Ankyrin repeat domain-containing protein 49 OS=Homo sapiens GN=ANKRD49 PE
Q6ZNI0	6	430	49,3	8,68	42	Beta-1,3-galactosyl-O-glycosyl-glycoprotein beta-1,6-N-acetylglucosaminyltransf
Q11203	19	375	42,1	8,98	42	CMP-N-acetylneuraminate-beta-1,4-galactoside alpha-2,3-sialyltransferase OS=H
O00338	10	296	34,9	7,52	42	Sulfotransferase 1C2 OS=Homo sapiens GN=SULT1C2 PE=1 SV=1 - [ST1C2_HUMAN]
A6NMZ2	4	147	16,5	8,27	42	Sentatan OS=Homo sapiens GN=SNTN PE=2 SV=1 - [SNTAN_HUMAN]
Q9BV20	16	369	39,1	6,3	42	Methylthioribose-1-phosphate isomerase OS=Homo sapiens GN=MRI1 PE=1 SV=1
A6NEY8	3	169	18,6	6,25	42	PrdX deacylase domain-containing protein 1 OS=Homo sapiens GN=PRDXDD1P PE
Q14CX7	24	972	112,2	6,64	42	N-alpha-acetyltransferase 25, NatB auxiliary subunit OS=Homo sapiens GN=NAA:
P29122	7	969	106,4	7,68	42	Protein convertase subtilisin/kexin type 6 OS=Homo sapiens GN=PCSK6 PE=1
O43674	8	189	21,7	9,63	42	NADH dehydrogenase [ubiquinone] 1 beta subcomplex subunit 5, mitochondrial
Q9H5I5	10	2752	317,9	6,15	42	Protein PIEZO2 OS=Homo sapiens GN=FAM38B PE=2 SV=2 - [PIEZ2_HUMAN]
O14544	12	535	59,5	7,25	42	Suppressor of cytokine signaling 6 OS=Homo sapiens GN=SOCS6 PE=1 SV=2 - [SOC
Q9H4Z2	8	1342	144,8	6,42	42	Zinc finger protein 335 OS=Homo sapiens GN=ZNF335 PE=1 SV=1 - [ZNF335_HUMA
P01743	2	117	12,9	8,92	42	Ig heavy chain V-I region HG3 OS=Homo sapiens PE=4 SV=1 - [HV102_HUMAN]
Q6ZRR9	8	648	72,7	8,85	42	Doublecortin domain-containing protein 5 OS=Homo sapiens GN=DCDC5 PE=2 SV
Q9Y2C2	12	406	47,6	8,69	42	Uronyl 2-sulfotransferase OS=Homo sapiens GN=UST PE=2 SV=1 - [UST_HUMAN]
P42681	6	527	61,2	7,97	42	Tyrosine-protein kinase TXK OS=Homo sapiens GN=TXK PE=1 SV=3 - [TXK_HUMAN
P83105	3	476	50,9	8,02	42	Probable serine protease HTRA4 OS=Homo sapiens GN=HTRA4 PE=2 SV=1 - [HTRA
P01604	19	108	12,1	8,81	42	Ig kappa chain V-I region Kue OS=Homo sapiens PE=1 SV=1 - [KV112_HUMAN]
Q86YQ8	21	564	63,1	5,96	41	Copine-8 OS=Homo sapiens GN=CPNE8 PE=1 SV=2 - [CPNE8_HUMAN]
Q9UPP1	9	1060	117,8	8,72	41	Histone lysine demethylase PHF8 OS=Homo sapiens GN=PHF8 PE=1 SV=3 - [PHF8
Q6PII3	15	467	53,9	6,33	41	Uncharacterized protein C3orf19 OS=Homo sapiens GN=C3orf19 PE=2 SV=3 - [CCO
P52740	5	706	80,6	8,4	41	Zinc finger protein 132 OS=Homo sapiens GN=ZNF132 PE=2 SV=2 - [ZNF132_HUMA
Q15293	3	331	38,9	5	41	Reticulocalbin-1 OS=Homo sapiens GN=RCN1 PE=1 SV=1 - [RCN1_HUMAN]
P01713	16	110	11,4	5,96	41	Ig lambda chain V-II region NIG-58 OS=Homo sapiens PE=1 SV=1 - [LV210_HUMAN
P46952	36	286	32,5	5,88	41	3-hydroxyanthranilate 3,4-dioxygenase OS=Homo sapiens GN=HAO PE=1 SV=2 -
Q2M3W8	16	571	65,8	8,9	41	Zinc finger protein 181 OS=Homo sapiens GN=ZNF181 PE=2 SV=1 - [ZNF181_HUMA
Q9Y575	11	518	57,7	6,27	41	Ankyrin repeat and SOCS box protein 3 OS=Homo sapiens GN=ASB3 PE=1 SV=1 - [
O60884	7	412	45,7	6,48	41	DnaJ homolog subfamily A member 2 OS=Homo sapiens GN=DNAJA2 PE=1 SV=1 -
Q92896	16	1179	134,5	6,9	41	Golgi apparatus protein 1 OS=Homo sapiens GN=GLG1 PE=1 SV=2 - [GSLG1_HUMA
P32248	5	378	42,8	8,48	41	C-C chemokine receptor type 7 OS=Homo sapiens GN=CCR7 PE=1 SV=2 - [CCR7_H
Q8TBZ3	10	569	62,9	8	41	WD repeat-containing protein 20 OS=Homo sapiens GN=WDR20 PE=1 SV=2 - [WDI
Q14134	9	588	65,8	7,15	41	Tripartite motif-containing protein 29 OS=Homo sapiens GN=TRIM29 PE=1 SV=2 -
Q8IXM7	10	274	31,0	9,54	41	Outer dense fiber protein 3-like protein 1 OS=Homo sapiens GN=ODF3L1 PE=2 SV
P11586	86	935	101,5	7,3	41	C-1-tetrahydrofolate synthase, cytoplasmic OS=Homo sapiens GN=MTHFD1 PE=1
Q99594	4	435	48,6	8,4	41	Transcriptional enhancer factor TEF-5 (TEA domain family member 3) (TEAD-3) (D
Q6ZU11	4	926	104,4	8,88	41	Uncharacterized protein FLJ44066 OS=Homo sapiens PE=1 SV=1 - [YD002_HUMAN]
P07864	40	332	36,3	7,46	41	L-lactate dehydrogenase C chain OS=Homo sapiens GN=LDHC PE=2 SV=4 - [LDHC_
Q8TC20	11	777	90,2	5,29	41	Cancer-associated gene 1 protein OS=Homo sapiens GN=CAGE1 PE=2 SV=2 - [CAG
Q96A46	12	364	39,2	8,59	41	Mitoferrin-2 OS=Homo sapiens GN=SLC25A28 PE=2 SV=1 - [MFRN2_HUMAN]
O76070	24	127	13,3	4,86	41	Gamma-synuclein OS=Homo sapiens GN=SNCG PE=1 SV=2 - [SYNG_HUMAN]
P01603	15	108	11,9	8,85	41	Ig kappa chain V-I region Ka OS=Homo sapiens PE=1 SV=1 - [KV111_HUMAN]
A6NJ16	7	123	13,6	9,6	41	Putative V-set and immunoglobulin domain-containing protein 6 OS=Homo sapie
Q86WR0	118	208	24,5	6,8	41	Coiled-coil domain-containing protein 25 OS=Homo sapiens GN=CCDC25 PE=1 SV:
Q9BQ52	45	826	92,2	7,9	41	Zinc phosphodiesterase ELAC protein 2 OS=Homo sapiens GN=ELAC2 PE=1 SV=2 -
Q16820	14	701	79,5	5,74	41	Mepripin A subunit beta OS=Homo sapiens GN=MEP1B PE=1 SV=3 - [MEP1B_HUMA
Q6NUM6	23	668	76,9	6,27	41	tRNA wybutosine-synthesizing protein 1 homolog B OS=Homo sapiens GN=TYW1
Q96PE7	18	176	18,7	9,09	41	Methylmalonyl-CoA epimerase, mitochondrial OS=Homo sapiens GN=MCEE PE=1
O15403	16	523	57,4	7,81	41	Monocarboxylate transporter 7 OS=Homo sapiens GN=SLC16A6 PE=1 SV=2 - [MOT
P0C2Y1	5	421	48,1	4,79	40	Putative neuroblastoma breakpoint family member 7 OS=Homo sapiens GN=NBP
Q96T68	13	719	81,8	7,5	40	Histone-lysine N-methyltransferase SETDB2 OS=Homo sapiens GN=SETDB2 PE=1 :
P26196	6	483	54,4	8,66	40	Probable ATP-dependent RNA helicase DDX6 (EC 3.6.1.-) (DEAD box protein 6) (A
Q13907	10	227	26,3	6,34	40	Isopentenyl-diphosphate Delta-isomerase 1 OS=Homo sapiens GN=IDI1 PE=1 SV=
Q9NUS5	10	200	22,5	6,87	40	Uncharacterized protein C20orf29 OS=Homo sapiens GN=C20orf29 PE=2 SV=1 - [C
Q5NE16	3	218	25,0	7,15	40	Putative cathepsin L-like protein 3 OS=Homo sapiens GN=CTSL3 PE=5 SV=1 - [CAT
Q6UXR4	10	307	34,8	6,4	40	Putative serpin A13 OS=Homo sapiens GN=SERPINA13 PE=5 SV=1 - [SPA13_HUMA
Q9NSE7	6	274	30,8	9,51	40	Putative ATP-binding cassette sub-family C member 13 OS=Homo sapiens GN=AE
A4D263	4	438	49,6	8,48	40	Uncharacterized protein C7orf72 OS=Homo sapiens GN=C7orf72 PE=4 SV=2 - [CG0
Q9Y6K1	43	912	101,8	6,57	40	DNA (cytosine-5)-methyltransferase 3A OS=Homo sapiens GN=DNMT3A PE=1 SV=
Q9BQI5	13	828	89,1	8,22	40	SH3-containing GRB2-like protein 3-interacting protein 1 OS=Homo sapiens GN=S
P35475	6	653	72,6	9,14	40	Alpha-L-iduronidase OS=Homo sapiens GN=IDUA PE=1 SV=2 - [IDUA_HUMAN]
Q16621	18	373	41,4	4,94	40	Transcription factor NF-E2 45 kDa subunit OS=Homo sapiens GN=NFE2 PE=1 SV=1
O75953	34	348	39,1	9,04	40	DnaJ homolog subfamily B member 5 OS=Homo sapiens GN=DNAJB5 PE=2 SV=1 -
Q9HCH5	2	934	104,9	8	40	Synaptotagmin-like protein 2 OS=Homo sapiens GN=SYTL2 PE=1 SV=3 - [SYTL2_HU
Q6ZT98	12	887	102,9	9,29	40	Tubulin polyglutamylase TLL7 OS=Homo sapiens GN=TLL7 PE=2 SV=2 - [TLL7_H

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O00299	1	241	26,9	5,17	40	Chloride intracellular channel protein 1 OS=Homo sapiens GN=CLIC1 PE=1 SV=4 -
Q14576	35	367	39,5	9,28	40	ELAV-like protein 3 OS=Homo sapiens GN=ELAVL3 PE=1 SV=3 - [ELAV3_HUMAN]
Q35XY7	13	634	69,9	5,69	40	Leucine-rich repeat, immunoglobulin-like domain and transmembrane domain-c
Q9H6E5	9	874	93,8	6,16	40	Speckle targeted PIP5K1A-regulated poly(A) polymerase OS=Homo sapiens GN=1
Q8N475	5	847	95,7	5,95	40	Follistatin-related protein 5 OS=Homo sapiens GN=FSTL5 PE=2 SV=2 - [FSTL5_HUN
Q5T699	3	396	45,7	5,62	40	Putative uncharacterized protein C6orf183 OS=Homo sapiens GN=C6orf183 PE=5
P53004	6	296	33,4	6,44	40	Biliverdin reductase A OS=Homo sapiens GN=BLVRA PE=1 SV=2 - [BIEA_HUMAN]
Q9BYX2	27	928	105,3	6,58	40	TBC1 domain family member 2A OS=Homo sapiens GN=TBC1D2 PE=1 SV=3 - [TBD
O75185	29	946	103,1	5,66	40	Calcium-transporting ATPase type 2C member 2 OS=Homo sapiens GN=ATP2C2 P
O00339	2	956	106,8	6,19	40	Matrilin-2 OS=Homo sapiens GN=MATN2 PE=1 SV=3 - [MATN2_HUMAN]
Q8N2K0	5	398	45,1	8,65	40	Monoacylglycerol lipase ABHD12 OS=Homo sapiens GN=ABHD12 PE=2 SV=2 - [AB
Q5T160	5	578	65,5	8,21	39	Probable arginyl-tRNA synthetase, mitochondrial precursor (EC 6.1.1.19) (Arginin
P61576	8	105	11,7	9,92	39	HERV-K_5q13.3 provirus Rec protein (HERV-K104 Rec protein) - Homo sapiens (Hu
Q8N3F9	7	429	47,0	7,42	39	Integral membrane protein GPR137C OS=Homo sapiens GN=GPR137C PE=2 SV=2 -
O60888	5	179	19,1	5,5	39	Protein CutA OS=Homo sapiens GN=CUTA PE=1 SV=2 - [CUTA_HUMAN]
Q96J88	2	318	36,8	9,89	39	Epithelial-stromal interaction protein 1 OS=Homo sapiens GN=EPST11 PE=2 SV=2 -
Q8N393	5	782	89,8	9,13	39	Zinc finger protein 786 OS=Homo sapiens GN=ZNF786 PE=2 SV=2 - [ZNF786_HUMA
P17174	3	413	46,2	7,01	39	Aspartate aminotransferase, cytoplasmic OS=Homo sapiens GN=GOT1 PE=1 SV=3
Q96BY7	33	2078	232,6	5,76	39	Autophagy-related protein 2 homolog B OS=Homo sapiens GN=ATG2B PE=1 SV=5
O43593	8	1189	127,4	7,08	39	Protein hairless OS=Homo sapiens GN=HR PE=1 SV=5 - [HAIR_HUMAN]
P62257	9	183	20,6	4,67	39	Ubiquitin-conjugating enzyme E2 H OS=Mus musculus GN=Ube2h PE=2 SV=1 - [UE
Q8N554	11	614	67,2	8,51	39	Zinc finger protein 276 OS=Homo sapiens GN=ZNF276 PE=1 SV=4 - [ZNF276_HUMA
P42261	8	906	101,4	7,71	39	Glutamate receptor 1 OS=Homo sapiens GN=GRIA1 PE=1 SV=2 - [GRIA1_HUMAN]
P98198	8	1209	137,4	7,01	39	Probable phospholipid-transporting ATPase ID OS=Homo sapiens GN=ATP8B2 PE
Q99941	73	703	76,7	6,27	39	Cyclic AMP-dependent transcription factor ATF-6 beta OS=Homo sapiens GN=ATF
Q9C0B9	4	1178	125,9	7,01	39	Zinc finger CCHC domain-containing protein 2 OS=Homo sapiens GN=ZCCHC2 PE=
Q9NV06	10	445	51,4	9,19	39	DDB1- and CUL4-associated factor 13 OS=Homo sapiens GN=DCAF13 PE=1 SV=2 - [
A6NKL6	3	621	63,9	10,08	39	Transmembrane protein 200C OS=Homo sapiens GN=TMEM200C PE=2 SV=2 - [T20
P40121	12	348	38,5	6,19	39	Macrophage-capping protein OS=Homo sapiens GN=CAPG PE=1 SV=2 - [CAPG_HU
Q9UNN5	11	650	73,9	4,88	39	FAS-associated factor 1 OS=Homo sapiens GN=FAF1 PE=1 SV=2 - [FAF1_HUMAN]
Q9BSC4	3	688	80,3	8,46	39	Nucleolar protein 10 OS=Homo sapiens GN=NOL10 PE=1 SV=1 - [NOL10_HUMAN]
Q9NSI2	3	230	25,4	11,08	39	Uncharacterized protein C21orf70 OS=Homo sapiens GN=C21orf70 PE=1 SV=2 - [C
Q6W3E5	43	623	71,9	9,03	39	Glycerophosphodiester phosphodiesterase domain-containing protein 4 OS=Hor
Q08211	2	1270	140,9	6,84	39	ATP-dependent RNA helicase A OS=Homo sapiens GN=DHX9 PE=1 SV=4 - [DHX9_
Q9Y6L7	9	1015	113,5	5,94	39	Toll-like protein 2 OS=Homo sapiens GN=TLL2 PE=1 SV=1 - [TLL2_HUMAN]
Q9UJU3	3	913	105,8	8,27	39	Zinc finger protein 112 homolog OS=Homo sapiens GN=ZFP112 PE=2 SV=2 - [ZF11
Q9NZI7	34	540	60,5	6,35	39	Upstream-binding protein 1 OS=Homo sapiens GN=UBP1 PE=2 SV=1 - [UBIP1_HUN
Q8IZU1	12	332	37,3	4,83	39	Protein FAM9A OS=Homo sapiens GN=FAM9A PE=2 SV=1 - [FAM9A_HUMAN]
Q8N9Z2	8	235	26,2	11,71	39	Uncharacterized protein FLJ36031 OS=Homo sapiens PE=2 SV=2 - [YG034_HUMAN]
Q96L42	7	1107	123,7	6,92	39	Potassium voltage-gated channel subfamily H member 8 OS=Homo sapiens GN=k
P04438	7	147	16,3	7,09	39	Ig heavy chain V-II region SESS OS=Homo sapiens PE=2 SV=1 - [HV208_HUMAN]
Q9BRG2	7	576	63,1	7,42	39	SH2 domain-containing protein 3A OS=Homo sapiens GN=SH2D3A PE=1 SV=1 - [S
Q9Y4B4	6	1467	162,7	6,09	39	Helicase ARIP4 OS=Homo sapiens GN=RAD54L2 PE=1 SV=4 - [ARIP4_HUMAN]
Q15173	4	497	57,4	6,71	38	Serine/threonine-protein phosphatase 2A 56 kDa regulatory subunit beta isoform
Q8WXD2	3	468	53,0	5,03	38	Secretogranin-3 precursor (Secretogranin III) (SgIII) - Homo sapiens (Human) - [S
Q92974	15	986	111,5	7,27	38	Rho guanine nucleotide exchange factor 2 OS=Homo sapiens GN=ARHGEF2 PE=1
Q9UK59	3	544	61,5	5,47	38	Lariat debranching enzyme OS=Homo sapiens GN=DBR1 PE=1 SV=2 - [DBR1_HUM
POC1H6	6	257	28,2	9,82	38	Histone H2B type F-M OS=Homo sapiens GN=H2BFM PE=2 SV=1 - [H2BFM_HUMAN
Q92539	5	896	99,3	5,33	38	Lipin-2 - Homo sapiens (Human) - [LPIN2_HUMAN]
Q9Y426	14	696	75,5	6,92	38	C2 domain-containing protein 2 OS=Homo sapiens GN=C2CD2 PE=1 SV=2 - [CU025
Q15858	9	1988	226,2	6,99	38	Sodium channel protein type 9 subunit alpha OS=Homo sapiens GN=SCN9A PE=1
Q9H0H3	3	589	65,9	6,57	38	Ectoderm-neural cortex protein 2 OS=Homo sapiens GN=KLHL25 PE=2 SV=1 - [ENC
P09210	2	222	25,6	8,59	38	Glutathione S-transferase A2 OS=Homo sapiens GN=GSTA2 PE=1 SV=4 - [GSTA2_H
P18077	6	110	12,5	11,06	38	60S ribosomal protein L35a - Homo sapiens (Human) - [RL35A_HUMAN]
P61009	43	180	20,3	8,62	38	Signal peptidase complex subunit 3 OS=Homo sapiens GN=SPCS3 PE=1 SV=1 - [SP
P28336	25	390	43,4	8,66	38	Neuromedin-B receptor OS=Homo sapiens GN=NMBR PE=2 SV=2 - [NMBR_HUMA
Q86XW9	1	330	36,8	4,89	38	Thioredoxin domain-containing protein 6 OS=Homo sapiens GN=TXNDC6 PE=1 SV
P98082	54	770	82,4	5,53	38	Disabled homolog 2 OS=Homo sapiens GN=DAB2 PE=1 SV=3 - [DAB2_HUMAN]
P56545	2	445	48,9	6,95	38	C-terminal-binding protein 2 OS=Homo sapiens GN=CTBP2 PE=1 SV=1 - [CTBP2_H
O95461	29	756	88,0	7,84	38	Glycosyltransferase-like protein LARGE1 OS=Homo sapiens GN=LARGE PE=1 SV=1
Q9Y2F9	10	522	58,4	7,46	38	BTB/POZ domain-containing protein 3 OS=Homo sapiens GN=BTBD3 PE=2 SV=1 - [
Q15404	52	277	31,5	8,65	38	Ras suppressor protein 1 OS=Homo sapiens GN=RSU1 PE=1 SV=3 - [RSU1_HUMAN]
Q9HCF6	101	1732	197,4	7,15	38	Transient receptor potential cation channel subfamily M member 3 OS=Homo sa
Q5HYI8	35	236	26,4	7,11	38	Rab-like protein 3 OS=Homo sapiens GN=RABL3 PE=1 SV=1 - [RABL3_HUMAN]
Q14435	34	633	72,6	7,99	38	Polypeptide N-acetylgalactosaminyltransferase 3 OS=Homo sapiens GN=GALNT3

Appendices

Q7L5D6	116	327	36,5	5,41	38	Golgi to ER traffic protein 4 homolog OS=Homo sapiens GN=GET4 PE=1 SV=1 - [GE
P17544	21	494	52,9	8,65	38	Cyclic AMP-dependent transcription factor ATF-7 OS=Homo sapiens GN=ATF7 PE=
Q9UHR6	4	403	42,9	5,99	38	Zinc finger HIT domain-containing protein 2 OS=Homo sapiens GN=ZNHIT2 PE=1 S
P48058	4	902	100,8	8,1	38	Glutamate receptor 4 OS=Homo sapiens GN=GRIA4 PE=2 SV=2 - [GRIA4_HUMAN]
Q5VSD8	43	79	8,2	9,74	38	Putative uncharacterized protein LOC401522 OS=Homo sapiens PE=4 SV=2 - [YIO2
Q8ND61	8	904	101,2	9,13	38	Uncharacterized protein C3orf20 OS=Homo sapiens GN=C3orf20 PE=1 SV=2 - [CCO
Q14005	19	1332	141,7	8,06	38	Pro-interleukin-16 OS=Homo sapiens GN=IL16 PE=1 SV=4 - [IL16_HUMAN]
P01708	3	109	11,5	7,87	38	Ig lambda chain V-II region BUR OS=Homo sapiens PE=1 SV=1 - [LV205_HUMAN]
Q96RY5	2	1269	134,6	7,87	38	Protein cramped-like OS=Homo sapiens GN=CRAMP1L PE=1 SV=3 - [CRML_HUMAI
Q96I23	3	114	12,6	9,33	37	Protein preY, mitochondrial OS=Homo sapiens GN=PREY PE=1 SV=1 - [PREY_HUM/
P51160	23	858	99,1	5,72	37	Cone cGMP-specific 3',5'-cyclic phosphodiesterase subunit alpha' OS=Homo sapie
P49917	9	911	103,9	7,96	37	DNA ligase 4 OS=Homo sapiens GN=LIG4 PE=1 SV=2 - [DNLI4_HUMAN]
O76014	2	449	49,7	4,96	37	Keratin, type I cuticular Ha7 OS=Homo sapiens GN=KRT37 PE=2 SV=3 - [KRT37_HUI
Q9Y689	13	179	20,7	6,79	37	ADP-ribosylation factor-like protein 5A OS=Homo sapiens GN=ARL5A PE=1 SV=1 -
Q9HCD5	10	579	65,5	9,6	37	Nuclear receptor coactivator 5 OS=Homo sapiens GN=NCOA5 PE=1 SV=2 - [NCOA5
Q95785	10	1651	178,6	6,86	37	Protein Wiz OS=Homo sapiens GN=WIZ PE=1 SV=2 - [WIZ_HUMAN]
Q9H0M4	7	648	72,0	7,53	37	Zinc finger CW-type PWWP domain protein 1 OS=Homo sapiens GN=ZCWPW1 PE=
Q15014	13	288	32,3	9,72	37	Mortality factor 4-like protein 2 OS=Homo sapiens GN=MORF4L2 PE=1 SV=1 - [MO
Q9H4B6	8	383	44,6	9,09	37	Protein salvador homolog 1 OS=Homo sapiens GN=SAV1 PE=1 SV=2 - [SAV1_HUM
Q6M2M9	14	219	22,7	4,87	37	Uncharacterized protein C4orf40 OS=Homo sapiens GN=C4orf40 PE=1 SV=1 - [CDO
Q8N201	14	2190	244,1	6,13	37	Integrator complex subunit 1 OS=Homo sapiens GN=INTS1 PE=1 SV=2 - [INT1_HUN
Q99814	23	870	96,4	6,28	37	Endothelial PAS domain-containing protein 1 OS=Homo sapiens GN=EPAS1 PE=1 '
Q9UBL0	3	812	89,1	6,95	37	cAMP-regulated phosphoprotein 21 OS=Homo sapiens GN=ARPP21 PE=1 SV=2 - [/
P40818	1	1118	127,4	8,51	37	Ubiquitin carboxyl-terminal hydrolase 8 OS=Homo sapiens GN=USP8 PE=1 SV=1 -
P10828	3	461	52,8	7,11	37	Thyroid hormone receptor beta OS=Homo sapiens GN=THRβ PE=1 SV=2 - [THB_HL
Q8N1Q8	27	247	27,7	7,72	37	Thioesterase superfamily member 5 OS=Homo sapiens GN=THEM5 PE=2 SV=2 - [T
Q9P0W5	10	487	53,4	5,12	37	Schwannomin-interacting protein 1 OS=Homo sapiens GN=SHIP1 PE=1 SV=1 - [S
Q96I99	6	432	46,5	6,39	37	Succinyl-CoA ligase [GDP-forming] subunit beta, mitochondrial OS=Homo sapien
Q9Y662	12	390	43,3	9,63	37	Heparan sulfate glucosamine 3-O-sulfotransferase 3B1 OS=Homo sapiens GN=HS
Q9UJZ1	2	356	38,5	7,39	37	Stomatin-like protein 2 OS=Homo sapiens GN=STOML2 PE=1 SV=1 - [STML2_HUM/
Q8NCU1	2	140	15,8	6,54	37	Uncharacterized protein C14orf48 OS=Homo sapiens GN=C14orf48 PE=2 SV=1 - [C
Q16665	23	826	92,6	5,33	37	Hypoxia-inducible factor 1-alpha OS=Homo sapiens GN=HIF1A PE=1 SV=1 - [HIF1A
Q00872	4	1141	128,2	6,04	37	Myosin-binding protein C, slow-type OS=Homo sapiens GN=MYBPC1 PE=1 SV=2 -
Q62N19	8	808	93,1	9,33	37	Zinc finger protein 841 OS=Homo sapiens GN=ZNF841 PE=2 SV=1 - [ZN841_HUMAN]
Q5HYM0	2	825	93,0	7,8	37	Probable ribonuclease ZC3H12B OS=Homo sapiens GN=ZC3H12B PE=2 SV=2 - [ZC1
Q70Z35	6	1606	182,5	7,44	37	Phosphatidylinositol 3,4,5-trisphosphate-dependent Rac exchanger 2 protein OS
P33260	23	490	55,7	7,2	37	Cytochrome P450 2C18 (EC 1.14.14.1) (CYPIIC18) (P450-6B/29C) - Homo sapiens (H
Q5TAH2	15	1124	129,0	6,92	37	Sodium/hydrogen exchanger 11 OS=Homo sapiens GN=SLC9A11 PE=2 SV=1 - [S9A
O60911	3	334	37,3	8,76	37	Cathepsin L2 OS=Homo sapiens GN=CTSL2 PE=1 SV=2 - [CATL2_HUMAN]
P16234	3	1089	122,6	5,17	37	Alpha-type platelet-derived growth factor receptor OS=Homo sapiens GN=PDGFR
Q8TED0	4	518	58,4	9,11	37	U3 small nucleolar RNA-associated protein 15 homolog OS=Homo sapiens GN=UT
Q8IV19	2	506	57,6	8,97	37	Nostrin OS=Homo sapiens GN=NOSTRIN PE=1 SV=2 - [NOSTN_HUMAN]
Q9H6S0	4	1430	160,1	8,4	37	Probable ATP-dependent RNA helicase YTHDC2 OS=Homo sapiens GN=YTHDC2 PE
Q86Y26	22	1132	120,2	5,45	37	Protein NUT OS=Homo sapiens GN=NUT PE=1 SV=2 - [NUT_HUMAN]
Q9NXB0	4	559	64,5	6,46	37	Meckel syndrome type 1 protein OS=Homo sapiens GN=MKS1 PE=1 SV=2 - [MKS1
O94933	15	977	108,9	7,37	37	SLIT and NTRK-like protein 3 OS=Homo sapiens GN=SLITRK3 PE=2 SV=2 - [SLIK3_H
P38435	3	758	87,5	8,02	37	Vitamin K-dependent gamma-carboxylase OS=Homo sapiens GN=GGCX PE=1 SV=
O60330	12	932	100,9	5,05	36	Protocadherin gamma-A12 OS=Homo sapiens GN=PCDHGA12 PE=2 SV=1 - [PCDGC
P51948	11	309	35,8	6,09	36	CDK-activating kinase assembly factor MAT1 OS=Homo sapiens GN=MNAT1 PE=1 :
Q9Y6L6	7	691	76,4	8,57	36	Solute carrier organic anion transporter family member 1B1 OS=Homo sapiens Gf
Q8TEY5	15	395	43,4	5,43	36	Cyclic AMP-responsive element-binding protein 3-like protein 4 OS=Homo sapie
P42566	9	896	98,6	4,64	36	Epidermal growth factor receptor substrate 15 OS=Homo sapiens GN=EPS15 PE=1
Q9P2M4	4	693	78,1	7,01	36	TBC1 domain family member 14 OS=Homo sapiens GN=TBC1D14 PE=1 SV=3 - [TBC
O14672	5	748	84,1	7,77	36	Disintegrin and metalloproteinase domain-containing protein 10 OS=Homo sapie
Q9NWX6	6	298	34,8	8	36	Probable tRNA(His) guanylyltransferase OS=Homo sapiens GN=THG1L PE=1 SV=2.
O43542	107	346	37,8	8,48	36	DNA-repair protein XRCC3 (X-ray repair cross-complementing protein 3) - Homo :
Q96PB1	8	797	91,6	8,82	36	CAS1 domain-containing protein 1 OS=Homo sapiens GN=CASD1 PE=2 SV=1 - [CAS
Q86XG9	4	351	40,5	5,05	36	Neuroblastoma breakpoint family member 5 OS=Homo sapiens GN=NBPF5 PE=2 :
P39023	7	403	46,1	10,18	36	60S ribosomal protein L3 OS=Homo sapiens GN=RPL3 PE=1 SV=2 - [RL3_HUMAN]
Q92674	33	756	86,7	8,76	36	Centromere protein I OS=Homo sapiens GN=CENPI PE=1 SV=2 - [CENPI_HUMAN]
P01825	5	117	12,8	7,08	36	Ig heavy chain V-II region NEWM OS=Homo sapiens PE=1 SV=1 - [HV207_HUMAN]
O75420	11	1035	114,5	5,39	36	PERQ amino acid-rich with GYF domain-containing protein 1 OS=Homo sapiens Gf
Q15650	10	581	66,1	7,85	36	Activating signal cointegrator 1 OS=Homo sapiens GN=TRIP4 PE=1 SV=4 - [TRIP4_H
O60347	4	775	85,6	5,74	36	TBC1 domain family member 12 OS=Homo sapiens GN=TBC1D12 PE=1 SV=3 - [TBC
Q5T7N3	3	995	107,3	5,27	36	KN motif and ankyrin repeat domain-containing protein 4 OS=Homo sapiens GN=

Appendices

Q96GW9	9	593	66,5	8,09	36	Methionyl-tRNA synthetase, mitochondrial OS=Homo sapiens GN=MARS2 PE=1 S
O95163	33	1332	150,2	5,94	36	Elongator complex protein 1 OS=Homo sapiens GN=IKBKAP PE=1 SV=3 - [ELP1_HU
A6NNW6	2	628	68,8	6	36	Enolase-like protein ENO4 OS=Homo sapiens GN=ENO4 PE=2 SV=2 - [ENO4_HUM
Q14457	3	450	51,9	4,89	36	Bec1-1 OS=Homo sapiens GN=BECN1 PE=1 SV=2 - [BECN1_HUMAN]
Q8WW83	17	523	58,8	9,26	36	Solute carrier family 35 member F5 OS=Homo sapiens GN=SLC35F5 PE=2 SV=1 - [S
Q62MV7	4	388	45,1	6,68	36	Leucine-, glutamate- and lysine-rich protein 1 OS=Homo sapiens GN=LEKR1 PE=2
Q99965	4	735	82,4	6,15	36	Disintegrin and metalloproteinase domain-containing protein 2 OS=Homo sapier
Q99543	8	621	72,0	8,7	36	DnaJ homolog subfamily C member 2 OS=Homo sapiens GN=DNAJC2 PE=1 SV=4 -
P78412	16	446	48,2	6,02	36	Iroquois-class homeodomain protein IRX-6 OS=Homo sapiens GN=IRX6 PE=2 SV=3
O95755	6	333	36,3	7,85	36	Ras-related protein Rab-36 OS=Homo sapiens GN=RAB36 PE=1 SV=2 - [RAB36_HUI
P51003	9	745	82,8	7,37	36	Poly(A) polymerase alpha OS=Homo sapiens GN=PAPOLA PE=1 SV=4 - [PAPOA_HI
Q727A4	3	578	64,9	9,35	36	PX domain-containing protein kinase-like protein OS=Homo sapiens GN=PXX PE=
Q8N5H7	20	860	94,4	8,07	36	SH2 domain-containing protein 3C OS=Homo sapiens GN=SH2D3C PE=1 SV=1 - [SH
P05783	2	430	48,0	5,45	36	Keratin, type I cytoskeletal 18 OS=Homo sapiens GN=KRT18 PE=1 SV=2 - [K1C18_H
P11177	28	359	39,2	6,65	36	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial OS=Homo s:
A6NNL0	18	878	93,9	7,84	36	Protein FAM22B OS=Homo sapiens GN=FAM22B PE=2 SV=2 - [FA22B_HUMAN]
Q8TEK3	8	1739	184,7	9,33	36	Histone-lysine N-methyltransferase, H3 lysine-79 specific OS=Homo sapiens GN=
Q96S52	6	555	61,6	6,49	36	GPI transamidase component PIG-S OS=Homo sapiens GN=PIGS PE=1 SV=3 - [PIGS
Q15345	3	812	88,6	8,38	36	Leucine-rich repeat-containing protein 41 OS=Homo sapiens GN=LRRC41 PE=1 SV:
O75600	2	419	45,3	8,05	36	2-amino-3-ketobutyrate coenzyme A ligase, mitochondrial OS=Homo sapiens GN
Q2M3V2	7	549	57,4	10,18	36	Ankyrin repeat domain-containing protein 43 OS=Homo sapiens GN=ANKRD43 PE
Q5T1N1	16	836	92,8	6,81	36	Protein AKNAD1 OS=Homo sapiens GN=AKNAD1 PE=2 SV=3 - [AKND1_HUMAN]
A8MX80	17	341	37,6	10,45	36	Putative UPF0607 protein ENSP00000383144 OS=Homo sapiens PE=3 SV=2 - [YM01
A6PVS8	12	624	73,6	9,72	36	Leucine-rich repeat and IQ domain-containing protein 3 OS=Homo sapiens GN=LF
Q9BR76	15	489	54,2	5,88	36	Coronin-1B OS=Homo sapiens GN=CORO1B PE=1 SV=1 - [COR1B_HUMAN]
O95672	4	775	87,7	7,03	36	Endothelin-converting enzyme-like 1 OS=Homo sapiens GN=ECEL1 PE=2 SV=3 - [E
P51956	11	506	57,7	7,17	35	Serine/threonine-protein kinase Nek3 OS=Homo sapiens GN=NEK3 PE=1 SV=2 - [I
P04808	8	185	21,1	8,69	35	Prorelaxin H1 precursor [Contains: Relaxin B chain; Relaxin A chain] - Homo sapie
Q9H2A3	4	272	28,6	7,75	35	Neurogenin-2 OS=Homo sapiens GN=NEUROG2 PE=1 SV=2 - [NGN2_HUMAN]
P84996	2	626	67,9	11,55	35	Protein ALEX OS=Homo sapiens GN=GNAS PE=1 SV=1 - [ALEX_HUMAN]
Q5VYM1	2	1079	117,6	7,36	35	Uncharacterized protein C9orf131 OS=Homo sapiens GN=C9orf131 PE=2 SV=3 - [C
P22914	14	178	21,0	6,89	35	Beta-crystallin S OS=Homo sapiens GN=CRYGS PE=1 SV=4 - [CRBS_HUMAN]
Q96S38	2	1066	118,6	4,87	35	Ribosomal protein S6 kinase delta-1 OS=Homo sapiens GN=RPS6K1 PE=1 SV=2 - [
Q8WU58	41	562	59,6	9,23	35	Uncharacterized protein C17orf63 OS=Homo sapiens GN=C17orf63 PE=1 SV=1 - [C
Q8TB36	16	358	41,3	8,34	35	Ganglioside-induced differentiation-associated protein 1 OS=Homo sapiens GN=
Q5JPI3	69	329	37,5	6,47	35	Uncharacterized protein C3orf38 OS=Homo sapiens GN=C3orf38 PE=2 SV=1 - [CC0
O15079	3	494	53,5	5,6	35	Syntaphilin OS=Homo sapiens GN=SNPH PE=1 SV=2 - [SNPH_HUMAN]
O95049	3	933	102,7	6,54	35	Tight junction protein ZO-3 OS=Homo sapiens GN=TJP3 PE=1 SV=2 - [ZO3_HUMAN
Q13905	14	1077	120,5	5,92	35	Rap guanine nucleotide exchange factor 1 OS=Homo sapiens GN=RAPGEF1 PE=1 S
A6NED2	28	376	40,1	5,27	35	RCC1 domain-containing protein 1 OS=Homo sapiens GN=RCCD1 PE=1 SV=1 - [RCC
O15165	5	306	33,9	6,28	35	Uncharacterized protein C18orf1 OS=Homo sapiens GN=C18orf1 PE=2 SV=1 - [CR0
Q7RTS3	5	328	34,9	5,25	35	Pancreas transcription factor 1 subunit alpha OS=Homo sapiens GN=PTF1A PE=1 S
Q6ZP65	4	573	64,8	5	35	Bicaudal D-related protein 1 OS=Homo sapiens GN=CCDC64 PE=2 SV=2 - [BICR1_H
Q92764	3	455	50,3	4,91	35	Keratin, type I cuticular Ha5 OS=Homo sapiens GN=KRT35 PE=1 SV=5 - [KRT35_HUI
Q14774	3	488	50,8	8,5	35	H2.0-like homeobox protein OS=Homo sapiens GN=HLX PE=1 SV=3 - [HLX_HUMAN
Q9NTZ6	3	932	97,3	8,63	35	RNA-binding protein 12 OS=Homo sapiens GN=RBM12 PE=1 SV=1 - [RBM12_HUM
Q13425	2	540	57,9	8,82	35	Beta-2-syntrophin (59 kDa dystrophin-associated protein A1 basic component 2)
A2RUR9	10	1427	165,0	5,36	35	Coiled-coil domain-containing protein 144A OS=Homo sapiens GN=CCDC144A PE:
Q8N0U7	6	546	62,0	8,65	35	Uncharacterized protein C1orf87 OS=Homo sapiens GN=C1orf87 PE=2 SV=1 - [CA0
Q9NZL6	11	768	86,6	6,09	35	Ral guanine nucleotide dissociation stimulator-like 1 OS=Homo sapiens GN=RGL1
Q5JRM2	7	361	39,9	9,54	35	Uncharacterized protein CXorf66 OS=Homo sapiens GN=CXorf66 PE=2 SV=1 - [CX0
O14746	4	1132	126,9	10,52	35	Telomerase reverse transcriptase OS=Homo sapiens GN=TERT PE=1 SV=1 - [TERT_
Q9H6R7	23	721	79,1	6,7	35	WD repeat-containing protein C2orf44 OS=Homo sapiens GN=C2orf44 PE=1 SV=1 -
O75629	4	220	24,1	7,59	35	Protein CREG1 OS=Homo sapiens GN=CREG1 PE=1 SV=1 - [CREG1_HUMAN]
Q9UP65	4	541	60,9	6,93	35	Cytosolic phospholipase A2 gamma OS=Homo sapiens GN=PLA2G4C PE=1 SV=2 - [
Q04609	6	750	84,3	6,98	35	Glutamate carboxypeptidase 2 OS=Homo sapiens GN=FOLH1 PE=1 SV=1 - [FOLH1_
Q9Y4K1	10	1723	188,6	5,86	35	Absent in melanoma 1 protein OS=Homo sapiens GN=AIM1 PE=1 SV=3 - [AIM1_HL
Q9P032	4	175	20,3	8,82	35	UPF0240 protein C6orf66 - Homo sapiens (Human) - [CF066_HUMAN]
Q7Z6J6	3	570	65,0	8,35	35	FERM domain-containing protein 5 OS=Homo sapiens GN=FRMD5 PE=2 SV=1 - [FR
P35236	5	360	40,5	6,79	35	Tyrosine-protein phosphatase non-receptor type 7 OS=Homo sapiens GN=PTPN7
Q0ZLH3	9	352	39,9	9,01	35	Pejvakin OS=Homo sapiens GN=PJVK PE=1 SV=1 - [PJVK_HUMAN]
Q99608	5	321	36,1	8,78	35	Necdin OS=Homo sapiens GN=NDN PE=1 SV=1 - [NECD_HUMAN]
Q96AP4	6	578	65,9	6,49	35	Zinc finger with UFM1-specific peptidase domain protein OS=Homo sapiens GN=

Appendix 3: LC ESI MS identified Schirmer strip tear proteins (Funke et al. 2014)

Accession	# Peptides	# AAs	MW [kDa]	calc. pI	Score	Description
P02788	7530	710	78,1	8,12	2196	Lactotransferrin GN=LTF PE=1 SV=6 [TRFL]
P02768	1668	609	69,3	6,28	1721	Serum albumin precursor [ALBU]
P04264	324	644	66,0	8,12	1525	Keratin, type II cytoskeletal 1 GN=KRT1 PE=1 SV=6 [K2C1]
P13645	228	584	58,8	5,21	1365	Keratin, type I cytoskeletal 10 GN=KRT10 PE=1 SV=6 [K1C10]
P06733	470	434	47,1	7,39	1134	Alphaenolase (EC 4.2.1.11) (2phosphoglycerate hydrolyase) (Nonneural enolase) (
P25311	1276	298	34,2	6,05	1075	Zinc alpha 2 glycoprotein GN=AZGP1 PE=1 SV=2 [ZA2G]
P98160	81	4391	468,5	6,51	1073	Basement membranespecific heparan sulfate proteoglycan core protein GN=HSPG:
P00352	174	501	54,8	6,73	1029	Retinal dehydrogenase 1 GN=ALDH1A1 PE=1 SV=2 [AL1A1]
P01009	167	418	46,7	5,59	886	Alpha1antitrypsin GN=SERPINA1 PE=1 SV=3 [A1AT]
P60709	434	375	41,7	5,48	878	Actin, cytoplasmic 1 (Betaactin) [ACTB]
P01024	62	1663	187,0	6,40	776	Complement C3 precursor [CO3]
Q8WZ42	265	34350	3813,7	6,35	765	Titin GN=TTN PE=1 SV=4 [TITIN]
P04083	128	346	38,7	7,02	747	Annexin A1 GN=ANXA1 PE=1 SV=2 [ANXA1]
P30740	108	379	42,7	6,28	747	Leukocyte elastase inhibitor GN=SERPINB1 PE=1 SV=1 [ILEU]
P68032	291	377	42,0	5,39	727	Actin, alpha cardiac muscle 1 (Alphacardiac actin) [ACTC]
P14618	62	531	57,9	7,84	666	Pyruvate kinase isozymes M1/M2 (EC 2.7.1.40) (Pyruvate kinase muscle isozyme) (P
Q06830	111	199	22,1	8,13	654	Peroxiredoxin1 (EC 1.11.1.15) (Thioredoxin peroxidase 2) (Thioredoxindependent p
P04792	183	205	22,8	6,40	646	Heatshock protein beta1 (HspB1) (Heat shock 27 kDa protein) (HSP 27) (Stressrespor
P01833	446	764	83,2	5,74	641	Polymeric immunoglobulin receptor GN=PIGR PE=1 SV=4 [PIGR]
Q9UGM3	206	2413	260,6	5,44	640	Deleted in malignant brain tumors 1 protein GN=DMBT1 PE=1 SV=2 [DMBT1]
P31025	3987	176	19,2	5,58	603	Lipocalin1 GN=LCN1 PE=1 SV=1 [LCN1]
P01876	1313	353	37,6	6,51	588	Ig alpha1 chain C region GN=IGHA1 PE=1 SV=2 [IGHA1]
Q658J3	245	1075	121,3	6,20	553	POTE ankyrin domain family member E GN=POTEE PE=1 SV=3 [POTEE]
P00738	67	406	45,2	6,58	550	Haptoglobin GN=HP PE=1 SV=1 [HPT]
P30838	56	453	50,4	6,54	540	Aldehyde dehydrogenase, dimeric NADPpreferring GN=ALDH3A1 PE=1 SV=3 [AL3A
A5A3E0	235	1075	121,4	6,20	535	POTE ankyrin domain family member F GN=POTEF PE=1 SV=2 [POTEF]
P01877	1100	340	36,5	6,10	525	Ig alpha2 chain C region GN=IGHA2 PE=1 SV=3 [IGHA2]
P07355	35	339	38,6	7,75	519	Annexin A2 GN=ANXA2 PE=1 SV=2 [ANXA2]
P09211	161	210	23,3	5,64	516	Glutathione Stransferase P GN=GSTP1 PE=1 SV=2 [GSTP1]
P02538	60	564	60,0	8,00	488	Keratin, type II cytoskeletal 6A GN=KRT6A PE=1 SV=3 [K2C6A]
P12273	1635	146	16,6	8,05	487	Prolactininducible protein GN=PIP PE=1 SV=1 [PIP]
P08758	31	320	35,9	5,05	473	Annexin A5 GN=ANXA5 PE=1 SV=2 [ANXA5]
P00338	80	332	36,7	8,27	454	Lactate dehydrogenase A chain GN=LDHA PE=1 SV=2 [LDHA]
P0CG38	206	1075	121,2	6,21	449	POTE ankyrin domain family member I GN=POTEI PE=3 SV=1 [POTEI]
P02787	400	698	77,0	7,12	444	Serotransferrin GN=TF PE=1 SV=3 [TRFE]
P01857	256	330	36,1	8,19	434	Ig gamma1 chain C region [IGHG1]
P06702	271	114	13,2	6,13	432	Protein S100A9 GN=S100A9 PE=1 SV=1 [S10A9]
P61626	3675	148	16,5	9,16	431	Lysozyme C precursor (EC 3.2.1.17) (1,4betaNacetylmuramidase C) [LYSC]
Q9GZZ8	912	138	14,2	5,50	415	Extracellular glycoprotein lacritin GN=LACRT PE=1 SV=1 [LACRT]
P10909	83	449	52,5	6,27	412	Clusterin GN=CLU PE=1 SV=1 [CLUS]
P00450	61	1065	122,1	5,72	403	Ceruloplasmin GN=CP PE=1 SV=1 [CERU]
P01623	67	109	11,7	8,91	381	Ig kappa chain VIII region WOL PE=1 SV=1 [KV305]
P31947	103	248	27,8	4,74	373	1433 protein sigma GN=SFN PE=1 SV=1 [1433S]
P35908	73	639	65,4	8,00	372	Keratin, type II cytoskeletal 2 epidermal GN=KRT2 PE=1 SV=2 [K22E]
P60174	34	286	30,8	5,92	367	Triosephosphate isomerase GN=TP1 PE=1 SV=3 [TPIS]
P20061	96	433	48,2	5,03	362	Transcobalamin1 GN=TCN1 PE=1 SV=2 [TCO1]
P00558	52	417	44,6	8,10	357	Phosphoglycerate kinase 1 (EC 2.7.2.3) (Primer recognition protein 2) (PRP 2) [PG
P14550	32	325	36,5	6,79	343	Alcohol dehydrogenase [NADP+] GN=AKR1A1 PE=1 SV=3 [AK1A1]
Q562R1	219	376	42,0	5,59	341	Betaactinlike protein 2 GN=ACTBL2 PE=1 SV=2 [ACTBL]
P05109	234	93	10,8	7,03	337	Protein S100A8 GN=S100A8 PE=1 SV=1 [S10A8]
Q13421	39	630	68,9	6,38	332	Mesothelin GN=MSLN PE=1 SV=2 [MSLN]
P01036	254	141	16,2	5,02	329	CystatinS precursor (Cystatin4) (Salivary acidic protein 1) (CystatinSAIII) [CYTS]
P63104	59	245	27,7	4,79	320	1433 protein zeta/delta GN=YWHAZ PE=1 SV=1 [1433Z]
P0CG05	460	106	11,3	7,24	311	Ig lambda2 chain C regions GN=IGLC2 PE=1 SV=1 [LAC2]
P02533	63	472	51,5	5,16	308	Keratin, type I cytoskeletal 14 GN=KRT14 PE=1 SV=4 [K1C14]
O75556	1021	95	10,9	5,78	301	MammaglobinB GN=SCGB2A1 PE=1 SV=1 [SG2A1]
P01622	55	109	11,8	8,50	299	Ig kappa chain VIII region Ti PE=1 SV=1 [KV304]
P40925	39	334	36,4	7,36	297	Malate dehydrogenase, cytoplasmic GN=MDH1 PE=1 SV=4 [MDHC]
P30041	47	224	25,0	6,38	294	Peroxiredoxin6 GN=PRDX6 PE=1 SV=3 [PRDX6]
P02763	54	201	23,5	5,02	284	Alpha1acid glycoprotein 1 precursor (AGP 1) (Orosomuroid1) (OMD 1) [A1AG1]
P52209	30	483	53,1	7,23	282	6phosphogluconate dehydrogenase, decarboxylating GN=PGD PE=1 SV=3 [6PGD]
P06396	64	782	85,6	6,28	281	Gelsolin GN=GSN PE=1 SV=1 [GELS]
P15924	4	2871	331,6	6,81	273	Desmoplakin GN=DSP PE=1 SV=3 [DSP]
P68372	26	445	49,8	4,89	273	Tubulin beta2C chain [TBB2C]

Appendices

P68104	31	462	50,1	9,01	271	Elongation factor 1alpha 1 GN=EEF1A1 PE=1 SV=1 [EF1A1]
P01859	129	326	35,9	7,59	268	Ig gamma2 chain C region GN=IGHG2 PE=1 SV=2 [IGHG2]
Q99935	242	248	27,2	10,42	262	Prolinerich protein 1 GN=PROL1 PE=1 SV=2 [PROL1]
P62259	32	255	29,2	4,74	260	1433 protein epsilon (1433E) [1433E]
P26447	106	101	11,7	6,11	256	Protein S100A4 GN=S100A4 PE=1 SV=1 [S10A4]
Q96KP4	24	475	52,8	5,97	254	Cytosolic nonspecific dipeptidase (Glutamate carboxypeptidase-like protein 1) (CNC
O95968	550	90	9,9	9,25	247	Secretoglobin family 1D member 1 GN=SCGB1D1 PE=1 SV=1 [SG1D1]
P09104	99	434	47,2	5,03	247	Gammaenolase (EC 4.2.1.11) (2phosphoDglycerate hydrolyase) (Neural enolase) (N
P40394	30	386	41,5	7,85	242	Alcohol dehydrogenase class 4 mu/sigma chain GN=ADH7 PE=1 SV=2 [ADH7]
B9A064	216	214	23,0	8,84	233	Immunoglobulin lambda dalike polypeptide 5 GN=IGLL5 PE=2 SV=2 [IGLL5]
P02647	19	267	30,8	5,76	233	Apolipoprotein AI precursor (ApoAI) (ApoAI) [Contains: Apolipoprotein AI(1242)]
P62987	21	128	14,7	9,83	232	Ubiquitin60S ribosomal protein L40 GN=UBA52 PE=1 SV=2 [RL40]
Q15149	3	4684	531,5	5,96	231	Plectin GN=PLEC PE=1 SV=3 [PLEC]
Q8NF91	37	8797	1010,4	5,53	230	Nesprin1 GN=SYNE1 PE=1 SV=3 [SYNE1]
P61982	39	247	28,3	4,89	229	1433 protein gamma [1433G]
O00299	29	241	26,9	5,17	216	Chloride intracellular channel protein 1 GN=CLIC1 PE=1 SV=4 [CLIC1]
P35527	13	623	62,0	5,24	212	Keratin, type I cytoskeletal 9 GN=KRT9 PE=1 SV=3 [K1C9]
Q04695	57	432	48,1	5,02	211	Keratin, type I cytoskeletal 17 GN=KRT17 PE=1 SV=2 [K1C17]
P09228	58	141	16,4	4,93	210	CystatinSA GN=CST2 PE=1 SV=1 [CYTT]
P01834	476	106	11,6	5,87	193	Ig kappa chain C region [KAC]
P07737	32	140	15,0	8,27	192	Profilin1 GN=PFN1 PE=1 SV=2 [PROF1]
P01037	122	141	16,4	7,21	188	CystatinSN GN=CST1 PE=1 SV=3 [CYTN]
Q81VF4	129	4471	514,5	5,88	186	Dynein heavy chain 10, axonemal GN=DNAH10 PE=1 SV=4 [DYH10]
O15078	39	2479	290,2	5,95	183	Centrosomal protein of 290 kDa GN=CEP290 PE=1 SV=2 [CE290]
P12429	12	323	36,4	5,92	182	Annexin A3 GN=ANXA3 PE=1 SV=3 [ANXA3]
P06703	30	90	10,2	5,48	178	Protein S100A6 GN=S100A6 PE=1 SV=1 [S10A6]
P01766	87	120	13,2	6,57	178	Ig heavy chain VIII region BRO PE=1 SV=1 [HV305]
Q5XKE5	25	535	57,8	7,20	176	Keratin, type II cytoskeletal 79 GN=KRT79 PE=1 SV=2 [K2C79]
A7E2Y1	1	1941	221,3	6,01	174	Myosin7B GN=MYH7B PE=2 SV=3 [MYH7B]
P02790	10	462	51,6	7,02	174	Hemopexin precursor (Beta1Bglycoprotein) [HEMO]
P20929	1	6669	772,4	9,07	174	Nebulin GN=NEB PE=1 SV=4 [NEBU]
P04745	10	511	57,7	6,93	173	Alphaamylase 1 GN=AMY1A PE=1 SV=2 [AMY1]
P04080	15	98	11,1	7,56	173	CystatinB GN=CSTB PE=1 SV=2 [CYTB]
P49454	2	3210	367,5	5,07	171	Centromere protein F GN=CENPF PE=1 SV=2 [CENPF]
Q16378	1671	134	15,1	7,06	167	Prolinerich protein 4 GN=PRR4 PE=1 SV=3 [PROL4]
P27348	84	245	27,7	4,78	164	1433 protein theta GN=YWHAQ PE=1 SV=1 [1433T]
P46100	54	2492	282,4	6,58	163	Transcriptional regulator ATRX GN=ATRX PE=1 SV=5 [ATRX]
P01591	91	159	18,1	5,24	163	Immunoglobulin J chain GN=IGJ PE=1 SV=4 [IGJ]
Q99996	35	3911	453,4	4,98	162	Akinase anchor protein 9 (Protein kinase Anchoring protein 9) (PRKA9) (Akinase ar
P01781	62	116	12,7	8,48	161	Ig heavy chain VIII region GAL PE=1 SV=1 [HV320]
Q9UQE7	190	1217	141,5	7,18	159	Structural maintenance of chromosomes protein 3 GN=SMC3 PE=1 SV=2 [SMC3]
Q8TD57	2	4116	470,5	6,43	159	Dynein heavy chain 3, axonemal GN=DNAH3 PE=2 SV=1 [DYH3]
O95239	115	1232	139,8	6,27	158	Chromosomeassociated kinesin KIF4A GN=KIF4A PE=1 SV=3 [KIF4A]
Q14643	83	2758	313,7	6,04	157	Inositol 1,4,5trisphosphate receptor type 1 GN=ITPR1 PE=1 SV=2 [ITPR1]
Q9Y524	6	205	22,9	4,63	156	Hemebinding protein 2 GN=HEBP2 PE=1 SV=1 [HEBP2]
Q92736	101	4967	564,2	6,07	156	Ryanodine receptor 2 GN=RYSR2 PE=1 SV=3 [RYSR2]
Q14789	4	3259	375,8	5,00	154	Golgin subfamily B member 1 GN=GOLGB1 PE=1 SV=2 [GOGB1]
Q5VST9	9	7968	867,9	5,99	153	Obscurin GN=OBSCN PE=1 SV=3 [OBSCN]
P19652	16	201	23,6	5,11	152	Alpha1acid glycoprotein 2 GN=ORM2 PE=1 SV=2 [A1AG2]
Q02224	8	2701	316,2	5,64	151	Centromereassociated protein E GN=CENPE PE=1 SV=2 [CENPE]
P04746	10	511	57,7	7,05	147	Pancreatic alphaamylase GN=AMY2A PE=1 SV=2 [AMYP]
P01765	63	115	12,3	9,13	146	Ig heavy chain VIII region TIL [HV304]
Q96T58	51	3664	402,0	7,64	145	Msx2interacting protein GN=SPEN PE=1 SV=1 [MINT]
Q9UPN3	26	7388	837,8	5,39	145	Microtubuleactin crosslinking factor 1, isoforms 1/2/3/5 GN=MACF1 PE=1 SV=4 [MA
Q8TDY2	20	1594	183,0	5,41	144	RB1inducible coiledcoil protein 1 GN=RB1CC1 PE=1 SV=3 [RBCC1]
P19961	10	511	57,7	7,09	143	Alphaamylase 2B GN=AMY2B PE=1 SV=1 [AMY2B]
Q5CZC0	1	6907	780,1	6,71	142	Fibrous sheathinteracting protein 2 GN=FSIP2 PE=1 SV=4 [FSIP2]
Q14980	20	2115	238,1	5,78	141	Nuclear mitotic apparatus protein 1 GN=NUMA1 PE=1 SV=2 [NUMA1]
Q9NQX4	31	1742	202,7	7,71	141	MyosinVc GN=MYO5C PE=1 SV=2 [MYO5C]
P01597	38	108	11,7	9,36	140	Ig kappa chain VI region DEE PE=1 SV=1 [KV105]
Q2M2I5	45	525	55,1	4,96	138	Keratin, type I cytoskeletal 24 GN=KRT24 PE=1 SV=1 [K1C24]
Q9BV73	49	2442	281,0	5,02	138	Centrosomeassociated protein CEP250 GN=CEP250 PE=1 SV=2 [CP250]
P18669	6	254	28,8	7,18	131	Phosphoglycerate mutase 1 GN=PGAM1 PE=1 SV=2 [PGAM1]
O75037	49	1637	182,5	7,08	131	Kinesinlike protein KIF21B GN=KIF21B PE=1 SV=2 [KI21B]
P47989	27	1333	146,3	7,66	130	Xanthine dehydrogenase/oxidase GN=XDH PE=1 SV=4 [XDH]

Appendices

P35580	1	1976	228,9	5,54	129	Myosin10 GN=MYH10 PE=1 SV=3 [MYH10]
P25054	12	2843	311,5	7,80	129	Adenomatous polyposis coli protein GN=APC PE=1 SV=2 [APC]
P80748	9	111	11,9	5,08	128	Ig lambda chain VIII region LOI PE=1 SV=1 [LV302]
P52895	8	323	36,7	7,49	127	Aldoketo reductase family 1 member C2 GN=AKR1C2 PE=1 SV=3 [AK1C2]
Q72320	26	450	49,3	5,08	125	Keratin, type I cytoskeletal 25 GN=KRT25 PE=1 SV=1 [K1C25]
P06309	30	117	12,7	7,12	124	Ig kappa chain VII region GM607 (Fragment) PE=4 SV=1 [KV205]
P04207	29	129	14,3	8,51	124	Ig kappa chain VIII region CLL PE=1 SV=2 [KV308]
P46013	42	3256	358,5	9,45	123	Antigen KI67 GN=MKI67 PE=1 SV=2 [KI67]
P04075	13	364	39,4	8,09	123	Fructosebiphosphate aldolase A GN=ALDOA PE=1 SV=2 [ALDOA]
Q72794	26	578	61,9	5,99	122	Keratin, type II cytoskeletal 1b GN=KRT77 PE=1 SV=3 [K2C1B]
Q04828	5	323	36,8	7,88	122	Aldoketo reductase family 1 member C1 GN=AKR1C1 PE=1 SV=1 [AK1C1]
Q62R08	1	3092	356,7	6,19	120	Dynein heavy chain 12, axonemal GN=DNAH12 PE=1 SV=2 [DYH12]
Q8TE73	29	4624	528,7	6,10	120	Dynein heavy chain 5, axonemal GN=DNAH5 PE=1 SV=3 [DYH5]
Q15772	18	3267	354,1	8,51	119	Striated muscle preferentially expressed protein kinase GN=SPEG PE=1 SV=4 [SPEG]
Q15643	1	1979	227,4	5,26	118	Thyroid receptorinteracting protein 11 GN=TRIP11 PE=1 SV=3 [TRIPB]
P15311	121	586	69,4	6,27	118	Ezrin GN=EZR PE=1 SV=4 [EZRI]
Q8NEN9	292	1154	128,5	6,09	118	PDZ domaincontaining protein 8 GN=PDZD8 PE=1 SV=1 [PDZD8]
O75417	14	2590	289,4	7,36	116	DNA polymerase theta GN=POLQ PE=1 SV=2 [DPOLO]
P13716	22	330	36,3	6,79	116	Deltaaminolevulinic acid dehydratase GN=ALAD PE=1 SV=1 [HEM2]
P27797	7	417	48,1	4,44	115	Calreticulin GN=CALR PE=1 SV=1 [CALR]
Q14008	54	2032	225,4	7,80	115	Cytoskeletonassociated protein 5 GN=CKAP5 PE=1 SV=3 [CKAP5]
P47929	4	136	15,1	7,62	114	Galectin7 GN=LGALS7 PE=1 SV=2 [LEG7]
Q01484	87	3957	433,4	5,14	114	Ankyrin2 GN=ANK2 PE=1 SV=4 [ANK2]
A4UGR9	23	3374	382,1	6,38	114	Xin actinbinding repeatcontaining protein 2 GN=XIRP2 PE=1 SV=2 [XIRP2]
Q92608	386	1830	211,8	6,87	114	Dedicator of cytokinesis protein 2 GN=DOCK2 PE=1 SV=2 [DOCK2]
Q72460	31	1538	169,3	9,03	113	CLIPassociating protein 1 GN=CLASP1 PE=1 SV=1 [CLAP1]
Q12955	17	4377	480,1	6,49	113	Ankyrin3 GN=ANK3 PE=1 SV=3 [ANK3]
Q14683	32	1233	143,1	7,64	113	Structural maintenance of chromosomes protein 1A GN=SMC1A PE=1 SV=2 [SMC1A]
Q9UIG0	4	1483	170,8	8,48	113	Tyrosineprotein kinase BAZ1B GN=BAZ1B PE=1 SV=2 [BAZ1B]
Q13439	19	2230	261,0	5,39	112	Golgin subfamily A member 4 GN=GOLGA4 PE=1 SV=1 [GOGA4]
P21817	98	5038	564,8	5,30	112	Ryanodine receptor 1 GN=RYR1 PE=1 SV=3 [RYR1]
Q2M1P5	18	1343	150,5	6,79	111	Kinesinlike protein KIF7 GN=KIF7 PE=1 SV=2 [KIF7]
POC6X7	154	7073	789,7	6,62	110	Replicase polyprotein 1ab SARS coronavirus GN=rep PE=1 SV=1 [R1ABCVHSA]
P11717	55	2491	274,2	5,94	110	Cationindependent mannose6phosphate receptor GN=IGF2R PE=1 SV=3 [MPRI]
Q8NCM8	426	4307	492,3	6,54	109	Cytoplasmic dynein 2 heavy chain 1 GN=DYNC2H1 PE=1 SV=4 [DYHC2]
Q14004	16	1512	164,8	9,69	109	Cyclindependent kinase 13 GN=CDK13 PE=1 SV=2 [CDK13]
Q03001	18	7570	860,1	5,25	109	Dystonin GN=DST PE=1 SV=4 [DYST]
Q93008	28	2570	292,1	5,80	109	Probable ubiquitin carboxylterminal hydrolase FAFX GN=USP9X PE=1 SV=3 [USP9X]
Q6N069	1	864	101,4	7,87	108	Nalpaacetyltransferase 16, NatA auxiliary subunit GN=NAA16 PE=1 SV=2 [NAA16]
Q9P273	105	2699	300,8	6,42	107	Teneurin3 GN=ODZ3 PE=2 SV=3 [TEN3]
Q8WXH0	18	6885	795,9	5,36	107	Nesprin2 GN=SYNE2 PE=1 SV=3 [SYNE2]
Q9Y2W1	28	955	108,6	10,15	107	Thyroid hormone receptorassociated protein 3 GN=THRAP3 PE=1 SV=2 [TR150]
Q9Y552	156	1711	194,2	6,37	106	Serine/threonineprotein kinase MRCK beta GN=CDC42BPB PE=1 SV=2 [MRCKB]
P01764	57	117	12,6	8,28	105	Ig heavy chain VIII region VH26 PE=1 SV=1 [HV303]
P01768	58	122	13,7	9,63	105	Ig heavy chain VIII region CAM PE=1 SV=1 [HV307]
P78527	25	4128	468,8	7,12	105	DNAdependent protein kinase catalytic subunit GN=PRKDC PE=1 SV=3 [PRKDC]
P06744	15	558	63,1	8,32	105	Glucose6phosphate isomerase GN=GPI PE=1 SV=4 [G6PI]
Q8IVL1	10	2488	268,0	9,03	104	Neuron navigator 2 GN=NAV2 PE=1 SV=3 [NAV2]
Q8WXG6	6	1647	183,2	6,04	104	MAP kinaseactivating death domain protein GN=MADD PE=1 SV=2 [MADD]
P80303	10	420	50,2	5,12	104	Nucleobindin2 GN=NUCB2 PE=1 SV=2 [NUCB2]
P12270	20	2363	267,1	5,02	103	Nucleoprotein TPR GN=TPR PE=1 SV=3 [TPR]
Q5TBA9	17	3013	338,7	5,99	103	Protein furry homolog GN=FRY PE=1 SV=1 [FRY]
Q4G0P3	23	5121	575,5	6,06	103	Hydrocephalusinducing protein homolog GN=HYDIN PE=1 SV=3 [HYDIN]
Q8IUD2	21	1116	128,0	5,97	103	ELKS/Rab6interacting/CAST family member 1 GN=ERC1 PE=1 SV=1 [RB6I2]
Q92817	58	2033	231,5	6,96	103	Envoplakin GN=EVPL PE=1 SV=3 [EVPL]
Q6UB99	39	2663	297,7	7,11	103	Ankyrin repeat domaincontaining protein 11 GN=ANKRD11 PE=1 SV=3 [ANR11]
Q6VAB6	15	950	107,6	8,69	103	Kinase suppressor of Ras 2 GN=KSR2 PE=1 SV=2 [KSR2]
O14686	10	5537	593,0	5,58	102	Histonelysine Nmethyltransferase MLL2 GN=MLL2 PE=1 SV=2 [MLL2]
Q8N3K9	1	4069	448,9	4,78	101	Cardiomyopathyassociated protein 5 GN=CMYA5 PE=1 SV=3 [CMYA5]
A6QL64	18	1941	217,3	8,66	101	Ankyrin repeat domaincontaining protein 36A GN=ANKRD36 PE=2 SV=3 [AN36A]
Q15413	68	4870	551,7	5,68	101	Ryanodine receptor 3 GN=RYR3 PE=1 SV=3 [RYR3]
Q5VYS8	13	1495	171,1	6,83	100	Terminal uridylyltransferase 7 GN=ZCCHC6 PE=1 SV=1 [TUT7]
P16591	10	822	94,6	7,14	100	Protooncogene tyrosineprotein kinase FER (EC 2.7.10.2) (p94FER) (cFER) (Tyrosine k
Q12756	39	1690	190,9	6,21	100	Kinesinlike protein KIF1A GN=KIF1A PE=2 SV=2 [KIF1A]
Q99880	21	126	13,9	10,32	99	Histone H2B type 1L GN=HIST1H2BL PE=1 SV=3 [H2B1L]

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Q02880	34	1626	183,2	8,00	97	DNA topoisomerase 2beta GN=TOP2B PE=1 SV=3 [TOP2B]
Q9HD33	13	250	29,4	10,37	97	39S ribosomal protein L47, mitochondrial GN=MRPL47 PE=1 SV=2 [RM47]
Q13098	384	491	55,5	6,74	97	COP9 signalosome complex subunit 1 GN=GPS1 PE=1 SV=4 [CSN1]
O75747	394	1445	165,6	6,93	97	Phosphatidylinositol4phosphate 3kinase C2 domaincontaining subunit gamma GN=
Q2VIQ3	68	1234	139,9	6,18	96	Chromosomeassociated kinesin KIF4B GN=KIF4B PE=1 SV=2 [KIF4B]
Q9UFH2	162	4485	511,5	5,77	96	Dynein heavy chain 17, axonemal GN=DNAH17 PE=2 SV=2 [DYH17]
Q96Q89	34	1820	210,5	5,67	96	Kinesinlike protein KIF20B GN=KIF20B PE=1 SV=3 [KI20B]
P07339	37	412	44,5	6,54	95	Cathepsin D GN=CTSD PE=1 SV=1 [CATD]
Q9UKN7	9	3530	395,0	9,17	95	MyosinXV GN=MYO15A PE=1 SV=2 [MYO15]
O14578	57	2027	231,3	6,57	95	Citron Rhointeracting kinase GN=CIT PE=1 SV=2 [CTRO]
Q9Y411	447	1855	215,3	8,48	95	MyosinVa GN=MYO5A PE=1 SV=2 [MYO5A]
Q9NYC9	151	4486	511,6	5,91	95	Dynein heavy chain 9, axonemal GN=DNAH9 PE=1 SV=3 [DYH9]
Q5VYK3	3	1845	204,2	7,12	94	Proteasomeassociated protein ECM29 homolog GN=ECM29 PE=1 SV=2 [ECM29]
Q7Z6Z7	14	4374	481,6	5,22	94	E3 ubiquitinprotein ligase HUWE1 GN=HUWE1 PE=1 SV=3 [HUWE1]
Q9UM54	17	1294	149,6	8,53	93	MyosinVI GN=MYO6 PE=1 SV=4 [MYO6]
P35749	13	1972	227,2	5,50	93	Myosin11 GN=MYH11 PE=1 SV=3 [MYH11]
Q14C86	38	1478	164,9	5,22	93	GTPaseactivating protein and VPS9 domaincontaining protein 1 GN=GAPVD1 PE=1 S
Q96M86	9	4753	533,3	6,71	93	Dynein heavy chain domaincontaining protein 1 GN=DNHD1 PE=1 SV=2 [DNHD1]
Q8WZ64	26	1704	193,3	7,39	93	ArfGAP with RhoGAP domain, ANK repeat and PH domaincontaining protein 2 GN=
Q96L93	3	1317	151,9	6,16	93	Kinesinlike protein KIF16B GN=KIF16B PE=1 SV=2 [KI16B]
Q8IZT6	1	3477	409,5	10,45	92	Abnormal spindlelike microcephalyassociated protein GN=ASPM PE=1 SV=2 [ASPM
Q5SZL2	26	805	91,8	6,33	92	Coiledcoil domaincontaining protein C6orf204 GN=C6orf204 PE=2 SV=1 [CF204]
A6NNM3	73	1639	180,8	6,77	92	RIMSBinding protein 3B GN=RIMBP3B PE=2 SV=3 [RIM3B]
Q8NCA5	69	519	55,4	9,03	92	Protein FAM98A GN=FAM98A PE=1 SV=1 [FA98A]
Q9NS87	24	1388	160,1	6,00	92	Kinesinlike protein KIF15 GN=KIF15 PE=1 SV=1 [KIF15]
Q63HN8	41	5207	591,0	6,48	92	E3 ubiquitinprotein ligase RNF213 GN=RNF213 PE=1 SV=3 [RN213]
O43290	4	800	90,2	6,13	91	U4/U6.U5 trisnRNPassociated protein 1 GN=SART1 PE=1 SV=1 [SNUT1]
O15083	18	957	110,5	6,99	91	ERC protein 2 GN=ERC2 PE=1 SV=3 [ERC2]
P01625	4	114	12,6	7,93	90	Ig kappa chain VIV region Len PE=1 SV=2 [KV402]
O15020	25	2390	271,2	6,11	90	Spectrin beta chain, brain 2 GN=SPTBN2 PE=1 SV=3 [SPTN2]
B2RTY4	1	2548	292,5	8,88	90	MyosinIXa GN=MYO9A PE=1 SV=2 [MYO9A]
A2VDJ0	18	1609	179,2	6,86	89	Transmembrane protein 131like GN=KIAA0922 PE=1 SV=2 [T131L]
P06576	10	529	56,5	5,40	89	ATP synthase subunit beta, mitochondrial GN=ATP5B PE=1 SV=3 [ATPB]
P31629	15	2446	268,9	6,96	88	Transcription factor HIVEP2 GN=HIVEP2 PE=1 SV=2 [ZEP2]
P13533	16	1939	223,6	5,73	88	Myosin6 GN=MYH6 PE=1 SV=5 [MYH6]
P05165	132	728	80,0	7,52	88	PropionylCoA carboxylase alpha chain, mitochondrial GN=PCCA PE=1 SV=4 [PCCA]
P01767	52	115	12,4	9,25	88	Ig heavy chain VIII region BUT PE=1 SV=1 [HV306]
Q9NQW8	15	809	92,1	7,93	88	Cyclic nucleotidedegated cation channel beta3 GN=CNGB3 PE=1 SV=2 [CNGB3]
P98161	178	4303	462,2	6,73	87	Polycystin1 GN=PKD1 PE=1 SV=3 [PKD1]
Q9BZF9	9	1416	162,4	7,03	87	Uveal autoantigen with coiledcoil domains and ankyrin repeats GN=UACA PE=1 SV=
O75122	28	1294	141,0	8,47	87	CLIPassociating protein 2 GN=CLASP2 PE=1 SV=2 [CLAP2]
O60293	116	1989	226,2	8,13	87	Zinc finger C3H1 domaincontaining protein GN=ZFC3H1 PE=1 SV=3 [ZC3H1]
Q96QB1	53	1528	170,5	6,40	87	Rho GTPaseactivating protein 7 GN=DLC1 PE=1 SV=4 [RHG07]
Q8NFC6	13	3051	330,3	5,08	87	Biorientation of chromosomes in cell division protein 1like GN=BOD1L PE=1 SV=2 [I
Q15652	17	2540	284,3	7,87	86	Probable JmjC domaincontaining histone demethylation protein 2C GN=JMJD1C PE
Q92614	28	2054	233,0	6,30	86	MyosinXVIIIa GN=MYO18A PE=1 SV=3 [MY18A]
Q9UQ35	19	2752	299,4	12,06	86	Serine/arginine repetitive matrix protein 2 GN=SRRM2 PE=1 SV=2 [SRRM2]
P49792	59	3224	358,0	6,20	85	E3 SUMOprotein ligase RanBP2 GN=RANBP2 PE=1 SV=2 [RBP2]
Q96DT5	69	4523	520,7	6,44	84	Dynein heavy chain 11, axonemal GN=DNAH11 PE=1 SV=3 [DYH11]
Q14679	27	1199	133,3	8,85	84	Tubulin polyglutamylase TTL4 GN=TLL4 PE=1 SV=2 [TLL4]
Q9UN79	23	622	69,2	6,71	84	Transcription factor SOX13 GN=SOX13 PE=1 SV=3 [SOX13]
P07951	8	284	32,8	4,70	84	Tropomyosin beta chain (Tropomyosin 2) (Betatropomyosin) [TPM2]
A2RUB6	91	948	109,3	8,25	84	Coiledcoil domaincontaining protein 66 GN=CCDC66 PE=1 SV=4 [CCD66]
Q9GT17	62	732	81,9	8,84	84	MAP7 domaincontaining protein 2 GN=MAP7D2 PE=1 SV=2 [MA7D2]
P30040	21	261	29,0	7,31	83	Endoplasmic reticulum resident protein 29 GN=ERP29 PE=1 SV=4 [ERP29]
Q9NR99	46	2828	312,0	8,32	83	Matrixremodelingassociated protein 5 GN=MXRA5 PE=2 SV=3 [MXRA5]
Q9Y283	17	1065	117,8	9,35	83	Inversin GN=INVS PE=1 SV=2 [INVS]
Q5VU43	24	2346	264,9	5,44	83	Myomegalin GN=PDE4DIP PE=1 SV=1 [MYOME]
Q0VDD8	5	3507	399,6	6,93	83	Dynein heavy chain 14, axonemal GN=DNAH14 PE=2 SV=3 [DYH14]
Q86VI3	13	1631	184,6	7,65	83	Ras GTPaseactivatinglike protein IQGAP3 GN=IQGAP3 PE=1 SV=2 [IQGA3]
Q96J81	69	4490	514,3	6,32	83	Dynein heavy chain 8, axonemal GN=DNAH8 PE=1 SV=2 [DYH8]
Q05D60	428	604	70,9	6,21	83	Coiledcoil domaincontaining protein 67 GN=CCDC67 PE=2 SV=2 [CCD67]
Q7Z7A1	7	2325	268,7	5,55	82	Centriolin GN=CEP110 PE=1 SV=2 [CE110]
P47895	17	512	56,1	7,25	82	Aldehyde dehydrogenase family 1 member A3 GN=ALDH1A3 PE=1 SV=2 [AL1A3]
Q9H1J1	8	476	54,7	9,03	82	Regulator of nonsense transcripts 3A GN=UPF3A PE=1 SV=1 [REN3A]

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Q7Z5Q5	76	900	100,2	8,29	82	DNA polymerase nu GN=POLN PE=1 SV=2 [DPOLN]
P01624	27	109	11,9	8,94	82	Ig kappa chain VIII region POM PE=1 SV=1 [KV306]
A4D1E1	1	1349	152,5	8,54	82	Zinc finger protein 804B GN=ZNF804B PE=1 SV=2 [Z804B]
P05787	4	483	53,7	5,59	82	Keratin, type II cytoskeletal 8 GN=KRT8 PE=1 SV=7 [K2C8]
Q07954	10	4544	504,3	5,39	81	Prolowdensity lipoprotein receptorrelated protein 1 GN=LRP1 PE=1 SV=2 [LRP1]
Q96Q15	1	3661	410,2	6,46	81	Serine/threonineprotein kinase SMG1 GN=SMG1 PE=1 SV=3 [SMG1]
P20742	11	1482	163,8	6,38	81	Pregnancy zone protein GN=PZP PE=1 SV=4 [PZP]
P57071	47	1507	169,2	8,24	81	PR domain zinc finger protein 15 GN=PRDM15 PE=2 SV=4 [PRD15]
Q87DN4	1	633	67,6	9,17	81	CDK5 and ABL1 enzyme substrate 1 GN=CABLES1 PE=1 SV=2 [CABL1]
Q86UK5	3	1308	147,9	6,96	81	Limbin GN=EVC2 PE=1 SV=1 [LBN]
Q15811	14	1721	195,3	7,77	80	Intersectin1 GN=ITSN1 PE=1 SV=3 [ITSN1]
P33151	19	784	87,5	5,43	79	Cadherin5 GN=CDH5 PE=1 SV=5 [CADH5]
Q9H3R1	31	872	100,7	7,55	79	Bifunctional heparan sulfate Ndeacetylase/Nsulfotransferase 4 GN=NDST4 PE=2 SV
Q96T21	70	854	95,4	8,12	79	Selenocysteine insertion sequencebinding protein 2 GN=SECISBP2 PE=1 SV=2 [SEBI
Q62T98	8	887	102,9	9,29	79	Tubulin polyglutamylase TTL7 GN=TTL7 PE=2 SV=2 [TLL7]
Q15154	28	2024	228,4	5,02	78	Pericentriolar material 1 protein GN=PCM1 PE=1 SV=4 [PCM1]
Q70CQ4	20	1352	146,6	9,22	78	Ubiquitin carboxylterminal hydrolase 31 GN=USP31 PE=1 SV=2 [UBP31]
Q9NRZ9	97	838	97,0	7,93	78	Lymphoidspecific helicase GN=HELLS PE=1 SV=1 [HELLS]
Q9NQ38	22	1064	120,6	8,06	78	Serine protease inhibitor Kazaltype 5 GN=SPINK5 PE=1 SV=2 [ISK5]
P48634	15	2157	228,7	9,45	77	Large prolinerich protein BAT2 GN=BAT2 PE=1 SV=3 [BAT2]
Q6ZU35	50	1233	136,7	5,60	77	Uncharacterized protein KIAA1211 GN=KIAA1211 PE=1 SV=3 [K1211]
P78559	13	2803	305,3	4,92	77	Microtubuleassociated protein 1A GN=MAP1A PE=1 SV=6 [MAP1A]
Q14722	10	419	46,5	8,94	77	Voltagegated potassium channel subunit beta1 GN=KCNAB1 PE=2 SV=1 [KCAB1]
P54652	6	639	70,0	5,74	77	Heat shockrelated 70 kDa protein 2 GN=HSPA2 PE=1 SV=1 [HSP72]
O15066	13	747	85,1	7,69	77	Kinesinlike protein KIF3B GN=KIF3B PE=1 SV=1 [KIF3B]
O95071	20	2799	309,2	5,85	76	E3 ubiquitinprotein ligase UBR5 GN=UBR5 PE=1 SV=2 [UBR5]
Q8TF72	21	1996	216,7	7,80	76	Protein Shroom3 GN=SHROOM3 PE=1 SV=2 [SHRM3]
Q14566	7	821	92,8	5,41	76	DNA replication licensing factor MCM6 GN=MCM6 PE=1 SV=1 [MCM6]
Q14966	10	1978	220,5	6,38	76	Zinc finger protein 638 GN=ZNF638 PE=1 SV=2 [ZNF638]
O43149	4	2961	330,9	5,95	76	Zinc finger ZZtype and EFhand domaincontaining protein 1 GN=ZZEF1 PE=1 SV=6 [Z
P12883	18	1935	223,0	5,80	76	Myosin7 GN=MYH7 PE=1 SV=5 [MYH7]
Q86UQ4	5	5058	575,8	6,46	75	ATPbinding cassette subfamily A member 13 GN=ABCA13 PE=2 SV=3 [ABCAD]
Q9UPZ9	442	632	71,4	9,77	75	Serine/threonineprotein kinase ICK GN=ICK PE=1 SV=1 [ICK]
Q5T200	11	1668	196,5	9,42	74	Zinc finger CCH domaincontaining protein 13 GN=ZC3H13 PE=1 SV=1 [ZC3HD]
Q8N2N9	9	1353	153,5	8,85	74	Ankyrin repeat domaincontaining protein 36B GN=ANKRD36B PE=2 SV=4 [AN36B]
P07195	2	334	36,6	6,05	74	Lactate dehydrogenase B chain (EC 1.1.1.27) (LDHB) (LDH heart subunit) (LDHH) (Re
Q5TAX3	28	1644	185,0	7,97	74	Terminal uridylyltransferase 4 GN=ZCCHC11 PE=1 SV=3 [TUT4]
Q6ZUT3	28	714	81,6	7,90	74	FERM domaincontaining protein 7 GN=FRMD7 PE=1 SV=1 [FRMD7]
Q5VZK9	28	1371	151,5	7,85	74	Leucinerich repeatcontaining protein 16A GN=LRRC16A PE=1 SV=1 [LR16A]
Q5TCS8	10	1911	221,3	5,01	74	Adenylate kinase domaincontaining protein 1 GN=AKD1 PE=1 SV=2 [AKD1]
Q5JR59	11	1369	150,1	6,68	74	Microtubuleassociated tumor suppressor candidate 2 GN=MTUS2 PE=1 SV=3 [MTUS
Q9UBC5	8	1043	118,3	9,31	73	Myosin1A GN=MYO1A PE=1 SV=1 [MYO1A]
Q9Y4D8	25	3996	439,1	6,19	73	Probable E3 ubiquitinprotein ligase C12orf51 GN=C12orf51 PE=1 SV=5 [K0614]
Q6IPM2	2	695	77,3	9,07	73	IQ domaincontaining protein E GN=IQCE PE=1 SV=2 [IQCE]
Q5JU85	24	1478	161,6	8,56	73	IQ motif and SEC7 domaincontaining protein 2 GN=IQSEC2 PE=1 SV=1 [IQEC2]
Q14571	58	2701	307,9	6,43	73	Inositol 1,4,5trisphosphate receptor type 2 GN=ITPR2 PE=1 SV=2 [ITPR2]
Q15147	63	1175	134,4	6,90	73	1phosphatidylinositol4,5bisphosphate phosphodiesterase beta4 GN=PLCB4 PE=1 S
Q07864	60	2286	261,4	6,39	73	DNA polymerase epsilon catalytic subunit A GN=POLE PE=1 SV=5 [DPOE1]
Q9Y4C0	22	1643	180,5	5,54	72	Neurexin3alpha GN=NRX3 PE=2 SV=4 [NRX3A]
O60449	6	1722	198,2	6,67	72	Lymphocyte antigen 75 GN=LY75 PE=1 SV=3 [LY75]
P11142	6	646	70,9	5,52	72	Heat shock cognate 71 kDa protein GN=HSPA8 PE=1 SV=1 [HSP7C]
P06313	26	133	14,6	6,58	72	Ig kappa chain VIV region JI PE=4 SV=1 [KV403]
Q86W11	79	4243	465,4	6,11	72	FibrocystinL GN=PKHD1L1 PE=2 SV=2 [PKHL1]
Q13023	11	2319	256,6	5,01	72	Akinase anchor protein 6 GN=AKAP6 PE=1 SV=3 [AKAP6]
Q81WZ3	220	2542	269,3	5,73	72	Ankyrin repeat and KH domaincontaining protein 1 GN=ANKHD1 PE=1 SV=1 [ANKH:
Q8N9T8	18	709	83,2	5,17	72	Protein KRI1 homolog GN=KRI1 PE=1 SV=2 [KRI1]
O60673	10	3130	352,6	8,47	72	DNA polymerase zeta catalytic subunit GN=REV3L PE=1 SV=2 [DPOLZ]
O75185	52	946	103,1	5,66	72	Calciumtransporting ATPase type 2C member 2 GN=ATP2C2 PE=1 SV=2 [AT2C2]
Q2LD37	1	5005	555,1	6,58	72	Uncharacterized protein KIAA1109 GN=KIAA1109 PE=1 SV=2 [K1109]
Q99490	244	1192	124,6	9,89	71	Centaurin gamma 1 (ARFGAP with GTPbinding proteinlike, ankyrin repeat and pleck
Q9NNX1	12	390	44,2	6,00	71	Tuftelin GN=TUFT1 PE=2 SV=1 [TUFT1]
O95359	9	2948	309,2	4,79	71	Transforming acidic coiledcoilcontaining protein 2 GN=TACC2 PE=1 SV=3 [TACC2]
Q7Z2W7	46	1104	127,6	7,24	71	Transient receptor potential cation channel subfamily M member 8 GN=TRPM8 PE=
Q8NCU4	12	936	110,5	9,60	71	Coiledcoil domaincontaining protein KIAA1407 GN=KIAA1407 PE=2 SV=1 [K1407]
Q70UQ0	19	350	39,3	9,17	71	Inhibitor of nuclear factor kappaB kinaseinteracting protein GN=IKBIP PE=1 SV=1 [I

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Q86UP2	1	1357	156,2	5,64	70	Kinectin GN=KTN1 PE=1 SV=1 [KTN1]
Q9UPV0	11	1460	164,2	5,36	70	Centrosomal protein of 164 kDa GN=CEP164 PE=1 SV=3 [CE164]
P61164	35	376	42,6	6,64	70	Alphacentractin GN=Actr1a PE=2 SV=1 [ACTZ]
Q86Y38	11	959	107,5	9,22	70	Xylosyltransferase 1 GN=XYLT1 PE=1 SV=1 [XYLT1]
Q9HCD6	10	1990	219,5	8,07	70	Protein TANC2 GN=TANC2 PE=1 SV=3 [TANC2]
Q8IUG5	21	2567	285,0	6,90	70	MyosinXVIIIb GN=MYO18B PE=1 SV=1 [MY18B]
Q9HCC0	11	563	61,3	7,68	70	MethylcrotonoylCoA carboxylase beta chain, mitochondrial GN=MCCC2 PE=1 SV=1
Q86UR5	1	1692	189,0	9,66	70	Regulating synaptic membrane exocytosis protein 1 GN=RIMS1 PE=1 SV=1 [RIMS1]
P07602	11	524	58,1	5,17	70	Proactivator polypeptide GN=PSAP PE=1 SV=2 [SAP]
Q6AWC2	11	1192	133,8	5,53	70	Protein WWC2 GN=WWC2 PE=1 SV=2 [WWC2]
Q9NZM3	9	1697	193,3	8,12	70	Intersectin2 GN=ITSN2 PE=1 SV=3 [ITSN2]
O60237	2	982	110,3	5,67	70	Protein phosphatase 1 regulatory subunit 12B GN=PPP1R12B PE=1 SV=2 [MYPT2]
Q8IV32	21	467	49,6	11,80	69	Coiledcoil domaincontaining protein 71 GN=CCDC71 PE=2 SV=3 [CCD71]
Q13427	41	754	88,6	10,29	69	Peptidylprolyl cistrans isomerase G GN=PPIG PE=1 SV=2 [PPIG]
Q14126	15	1118	122,2	5,24	69	Desmoglein2 GN=DSG2 PE=1 SV=2 [DSG2]
P35251	9	1148	128,2	9,36	69	Replication factor C subunit 1 GN=RFC1 PE=1 SV=4 [RFC1]
Q9H6T3	34	665	75,7	6,84	69	RNA polymerase II associated protein 3 GN=RPAP3 PE=1 SV=2 [RPAP3]
P07384	1	714	81,8	5,67	69	Calpain1 catalytic subunit GN=CAPN1 PE=1 SV=1 [CAN1]
Q6WRI0	65	2623	290,7	9,13	69	Immunoglobulin superfamily member 10 GN=IGSF10 PE=1 SV=1 [IGS10]
Q5VWV2	2	1013	112,8	7,65	68	GTPaseactivating Rap/RanGAP domainlike protein 3 GN=GARNL3 PE=1 SV=2 [GARL3]
Q6PGP7	15	1564	175,4	7,53	68	Tetratricopeptide repeat protein 37 GN=TTC37 PE=1 SV=1 [TTC37]
Q8NBS3	10	891	99,5	7,68	68	Sodium bicarbonate transporterlike protein 11 GN=SLC4A11 PE=1 SV=2 [S4A11]
P19237	61	187	21,7	9,58	68	Troponin I, slow skeletal muscle GN=TNNI1 PE=1 SV=3 [TNNI1]
Q3SY84	13	523	57,3	6,61	68	Keratin, type II cytoskeletal 71 GN=KRT71 PE=1 SV=3 [K2C71]
Q7Z7B0	5	1213	138,0	8,32	68	FilaminAinteracting protein 1 GN=FILIP1 PE=1 SV=1 [FLIP1]
Q9ULU4	40	1186	131,6	7,20	68	Protein kinase C binding protein 1 GN=ZMYND8 PE=1 SV=2 [PKCB1]
Q96L96	12	1907	201,1	7,58	67	Alphaprotein kinase 3 GN=ALPK3 PE=2 SV=2 [ALPK3]
P06737	3	847	97,1	7,17	67	Glycogen phosphorylase, liver form GN=PYGL PE=1 SV=4 [PYGL]
Q5VVJ2	435	828	95,0	5,53	67	Histone H2A deubiquitinase MYSM1 GN=MYSM1 PE=1 SV=1 [MYSM1]
Q96M83	10	486	55,7	7,78	67	Coiledcoil domaincontaining protein 7 GN=CCDC7 PE=2 SV=2 [CCDC7]
Q9NSB2	114	600	64,8	7,56	67	Keratin, type II cuticular Hb4 GN=KRT84 PE=1 SV=2 [KRT84]
Q95197	65	1032	112,5	4,96	66	Reticulon3 GN=RTN3 PE=1 SV=2 [RTN3]
Q96N67	357	2140	242,4	6,80	66	Dedicator of cytokinesis protein 7 GN=DOCK7 PE=1 SV=4 [DOCK7]
Q86SR1	44	603	68,9	8,59	66	Polypeptide Nacetylgalactosaminyltransferase 10 GN=GALNT10 PE=1 SV=2 [GLT10]
O60333	30	1816	204,3	5,60	66	Kinesinlike protein KIF1B GN=KIF1B PE=1 SV=5 [KIF1B]
P12955	13	493	54,5	6,00	66	XaaPro dipeptidase GN=PEPD PE=1 SV=3 [PEPD]
O15042	52	1029	118,2	8,47	66	U2associated protein SR140 GN=SR140 PE=1 SV=2 [SR140]
Q9BXX2	24	1392	157,9	6,35	66	Ankyrin repeat domaincontaining protein 30B GN=ANKRD30B PE=2 SV=3 [AN30B]
Q86VF7	70	1730	197,0	9,20	66	Nebulinrelatedanchoring protein GN=NRAP PE=2 SV=2 [NRAP]
Q08378	12	1498	167,3	5,44	66	Golgin subfamily A member 3 GN=GOLGA3 PE=1 SV=2 [GOGA3]
Q9UHN6	29	1383	154,3	8,15	66	Transmembrane protein 2 GN=TMEM2 PE=1 SV=1 [TMEM2]
Q9H4L7	122	1026	117,3	5,55	66	SWI/SNFrelated matrixassociated actindependent regulator of chromatin subfamily
Q8NDM7	8	1665	191,9	5,99	66	WD repeatcontaining protein C10orf79 GN=C10orf79 PE=2 SV=3 [CJ079]
O60281	17	2723	304,6	7,39	66	Zinc finger protein 292 GN=ZNF292 PE=1 SV=3 [ZN292]
Q5KS16	467	1271	141,7	5,53	66	Diacylglycerol kinase kappa GN=DGKK PE=1 SV=1 [DGKK]
Q99575	4	1024	114,6	9,22	65	Ribonucleases P/MRP protein subunit POP1 GN=POP1 PE=1 SV=2 [POP1]
A5PLK6	35	1076	125,6	9,01	65	Regulator of Gprotein signaling proteinlike GN=RGSL1 PE=2 SV=1 [RGSL]
Q9UPW8	5	1703	192,9	5,35	65	Protein unc13 homolog A GN=UNC13A PE=2 SV=4 [UN13A]
Q8TC20	1	777	90,2	5,29	65	Cancerassociated gene 1 protein GN=CAGE1 PE=2 SV=2 [CAGE1]
Q92973	150	898	102,3	4,98	65	Transportin1 GN=TNPO1 PE=1 SV=2 [TNPO1]
Q9BQ52	122	826	92,2	7,90	65	Zinc phosphodiesterase ELAC protein 2 GN=ELAC2 PE=1 SV=2 [RNZ2]
Q8N4C6	19	2090	243,1	5,03	65	Ninein GN=NIN PE=1 SV=4 [NIN]
P19174	66	1290	148,4	6,05	65	1phosphatidylinositol4,5bisphosphate phosphodiesterase gamma1 GN=PLCG1 PE=
Q5T2S8	28	1044	115,6	7,77	65	Armadillo repeatcontaining protein 4 GN=ARMC4 PE=2 SV=1 [ARMC4]
Q96S19	10	672	73,6	8,72	65	Spermatid perinuclear RNAbinding protein GN=STRBP PE=1 SV=1 [STRBP]
Q9BYJ4	19	488	56,8	7,44	65	Tripartite motifcontaining protein 34 GN=TRIM34 PE=1 SV=2 [TRI34]
Q86UP3	5	3567	393,5	6,37	65	Zinc finger homeobox protein 4 GN=ZFHX4 PE=1 SV=1 [ZFHX4]
P20591	14	662	75,5	5,83	65	Interferoninduced GTPbinding protein Mx1 GN=MX1 PE=1 SV=4 [MX1]
Q8NDI1	1	1231	139,9	5,35	65	EH domainbinding protein 1 GN=EHP1 PE=1 SV=3 [EHP1]
Q96C24	3	671	76,0	8,98	64	Synaptotagminlike protein 4 GN=SYTL4 PE=1 SV=2 [SYTL4]
P35712	22	828	91,9	7,78	64	Transcription factor SOX6 GN=SOX6 PE=1 SV=3 [SOX6]
A6NKT7	55	1758	197,4	6,33	64	RanBP2like and GRIP domaincontaining protein 3 GN=RGPD3 PE=2 SV=2 [RGPD3]
Q14678	12	1352	147,2	5,30	64	KN motif and ankyrin repeat domaincontaining protein 1 GN=KANK1 PE=1 SV=3 [KANK1]
Q6ZN28	17	852	96,6	6,90	64	Metastasisassociated in colon cancer protein 1 GN=MACC1 PE=1 SV=2 [MACC1]
P52907	99	286	32,9	5,69	64	Factincapping protein subunit alpha1 GN=CAPZA1 PE=1 SV=3 [CAZA1]

Appendices

Q9C099	8	1032	119,5	5,88	64	Leucinerich repeat and coiledcoil domaincontaining protein 1 GN=LRRCC1 PE=2 SV=
P19013	15	534	57,2	6,61	64	Keratin, type II cytoskeletal 4 GN=KRT4 PE=1 SV=4 [K2C4]
Q9HC62	72	589	67,8	9,48	64	Sentrinspecific protease 2 GN=SEN2 PE=1 SV=3 [SEN2]
Q86TE4	363	346	38,9	8,73	64	Leucine zipper protein 2 GN=LUZP2 PE=2 SV=2 [LUZP2]
Q8IYE0	24	955	112,7	8,48	64	Coiledcoil domaincontaining protein 146 GN=CCDC146 PE=2 SV=2 [CC146]
P30613	15	574	61,8	7,74	63	Pyruvate kinase isozymes R/L GN=PKLR PE=1 SV=2 [KPYR]
Q9P219	4	2028	228,1	6,23	63	Protein Daple GN=CCDC88C PE=1 SV=3 [DAPLE]
Q68CJ6	48	796	91,1	8,63	63	GTPase SLIPGC GN=C8orf80 PE=1 SV=3 [SLIP]
Q8IXZ2	1	948	101,9	10,95	63	Zinc finger CCH domaincontaining protein 3 GN=ZC3H3 PE=1 SV=3 [ZC3H3]
Q9BSK4	25	669	73,6	6,07	63	Protein fem1 homolog A GN=FEM1A PE=1 SV=1 [FEM1A]
A2RUR9	10	1427	165,0	5,36	63	Coiledcoil domaincontaining protein 144A GN=CCDC144A PE=1 SV=1 [C144A]
Q9COC9	7	1292	141,2	5,12	63	Ubiquitinconjugating enzyme E2 O GN=UBE2O PE=1 SV=3 [UBE2O]
Q8IWN7	29	2480	261,0	4,41	63	Retinitis pigmentosa 1like 1 protein GN=RP1L1 PE=1 SV=4 [RP1L1]
Q6ZU80	1	1094	127,9	6,52	63	Uncharacterized protein C14orf145 GN=C14orf145 PE=1 SV=2 [CN145]
Q15003	35	741	82,5	5,06	63	Condensin complex subunit 2 GN=NCAPH PE=1 SV=3 [CND2]
Q86XP1	8	1220	134,8	6,54	63	Diaclylglycerol kinase eta (EC 2.7.1.107) (Diglyceride kinase eta) (DGKeta) (DAG kina:
O75460	17	977	109,7	6,42	63	Serine/threonineprotein kinase/endoribonuclease IRE1 GN=ERN1 PE=1 SV=2 [ERN:
O75096	11	1905	211,9	5,27	63	Lowdensity lipoprotein receptorrelated protein 4 GN=LRP4 PE=1 SV=4 [LRP4]
Q99456	23	494	53,5	4,78	63	Keratin, type I cytoskeletal 12 GN=KRT12 PE=1 SV=1 [K1C12]
O15021	1	2626	284,2	8,62	63	Microtubuleassociated serine/threonineprotein kinase 4 GN=MAST4 PE=1 SV=3 [M
P35498	11	2009	228,8	5,81	62	Sodium channel protein type 1 subunit alpha GN=SCN1A PE=1 SV=2 [SCN1A]
Q86VQ6	438	682	74,8	8,22	62	Thioredoxin reductase 3 (Fragment) GN=TXNRD3 PE=1 SV=3 [TRXR3]
Q9P2D1	41	2997	335,7	6,34	62	ChromodomainhelicaseDNAbinding protein 7 GN=CHD7 PE=1 SV=3 [CHD7]
Q99698	1	3801	428,9	6,61	62	Lysosomaltrafficking regulator GN=LYST PE=1 SV=3 [LYST]
Q5THJ4	4	4387	491,5	6,58	62	Vacuolar protein sortingassociated protein 13D GN=VPS13D PE=1 SV=1 [VP13D]
Q15560	6	299	33,6	9,13	62	Transcription elongation factor A protein 2 GN=TCEA2 PE=1 SV=1 [TCEA2]
Q8IYW2	75	2715	303,3	7,36	62	Tetratricopeptide repeat protein 40 GN=TTC40 PE=2 SV=3 [TTC40]
Q15858	9	1988	226,2	6,93	62	Sodium channel protein type 9 subunit alpha GN=SCN9A PE=1 SV=3 [SCN9A]
P10632	38	490	55,8	8,50	62	Cytochrome P450 2C8 GN=CYP2C8 PE=1 SV=2 [CP2C8]
P52948	16	1817	197,5	6,40	62	Nuclear pore complex protein Nup98Nup96 GN=NUP98 PE=1 SV=4 [NUP98]
Q92667	95	903	97,3	4,94	62	Akinase anchor protein 1, mitochondrial GN=AKAP1 PE=1 SV=1 [AKAP1]
Q96NB3	13	372	42,0	5,31	62	Zinc finger protein 830 GN=ZNF830 PE=1 SV=2 [ZN830]
Q9H582	412	1327	149,5	8,16	62	Zinc finger protein 644 GN=ZNF644 PE=1 SV=2 [ZN644]
P30046	2	118	12,7	7,30	62	Ddopachrome decarboxylase GN=DDT PE=1 SV=3 [DOPD]
Q9BYB0	1	1741	186,2	9,03	62	SH3 and multiple ankyrin repeat domains protein 3 GN=SHANK3 PE=1 SV=2 [SHAN3
Q9ULD0	46	1010	114,4	6,65	62	2oxoglutarate dehydrogenaselike, mitochondrial GN=OGDHL PE=2 SV=3 [OGDHL]
Q8IY85	54	973	110,1	6,57	62	EFhand domaincontaining protein C17orf57 GN=C17orf57 PE=2 SV=2 [CQ057]
O94913	6	1555	172,9	8,48	62	PremRNA cleavage complex 2 protein Pcf11 GN=PCF11 PE=1 SV=3 [PCF11]
Q8IZF0	24	1738	200,2	8,68	61	Sodium leak channel nonselective protein GN=NALCN PE=1 SV=1 [NALCN]
Q9NRG4	40	433	49,7	6,71	61	SET and MYND domaincontaining protein 2 GN=SMYD2 PE=1 SV=2 [SMYD2]
B7ZAP0	8	253	29,0	5,33	61	Rab GTPaseactivating protein 1like, isoform 10 GN=RABGAP1L PE=2 SV=1 [RBG10]
Q5TCY1	34	1321	142,6	5,60	61	Tautubulin kinase 1 GN=TTBK1 PE=1 SV=2 [TTBK1]
Q5TB80	7	1403	161,8	5,47	61	Protein QN1 homolog GN=KIAA1009 PE=1 SV=2 [QN1]
P46063	3	649	73,4	7,88	61	ATPdependent DNA helicase Q1 GN=RECQL PE=1 SV=3 [RECQ1]
Q7RTX0	35	852	93,3	7,14	60	Taste receptor type 1 member 3 GN=TAS1R3 PE=1 SV=2 [TS1R3]
Q8IV76	4	773	87,4	5,07	60	PAS domaincontaining protein 1 GN=PASD1 PE=2 SV=1 [PASD1]
O60603	90	784	89,8	6,61	60	Tolllike receptor 2 GN=TLR2 PE=1 SV=1 [TLR2]
Q96C45	1	1275	142,4	6,29	60	Serine/threonineprotein kinase ULK4 GN=ULK4 PE=2 SV=2 [ULK4]
Q7Z478	7	1369	155,1	8,09	60	ATPdependent RNA helicase DHX29 GN=DHX29 PE=1 SV=2 [DHX29]
Q6ZS81	5	3184	353,4	6,32	60	WD repeat and FYVE domaincontaining protein 4 GN=WDFY4 PE=1 SV=3 [WDFY4]
Q6P3X8	18	592	68,0	8,54	60	PiggyBac transposable elementderived protein 2 GN=PGBD2 PE=2 SV=1 [PGBD2]
Q5T0Z8	73	1188	124,0	9,45	60	Uncharacterized protein C6orf132 GN=C6orf132 PE=1 SV=4 [CF132]
P15121	9	316	35,8	6,98	60	Aldose reductase GN=AKR1B1 PE=1 SV=3 [ALDR]
Q9UPX8	8	1470	158,7	6,90	60	SH3 and multiple ankyrin repeat domains protein 2 GN=SHANK2 PE=1 SV=3 [SHAN2
Q13075	24	1403	159,5	5,99	60	Baculoviral IAP repeatcontaining protein 1 GN=NAIP PE=1 SV=3 [BIRC1]
P01780	42	115	12,6	9,29	60	Ig heavy chain VIII region JON [HV319]
Q8IZD9	109	2030	233,0	6,98	60	Dedicator of cytokinesis protein 3 GN=DOCK3 PE=1 SV=1 [DOCK3]
P61161	23	394	44,7	6,74	60	Actinrelated protein 2 GN=Actr2 PE=1 SV=1 [ARP2]
Q9ULW6	14	460	52,5	4,49	59	Nucleosome assembly protein 1like 2 GN=NAP1L2 PE=2 SV=1 [NP1L2]
Q8N4A0	37	578	66,6	7,61	59	Polypeptide Nacetylglactosaminyltransferase 4 GN=GALNT4 PE=1 SV=2 [GALT4]
Q70CQ2	36	3546	404,0	5,82	59	Ubiquitin carboxylterminal hydrolase 34 GN=USP34 PE=1 SV=2 [UBP34]
Q9Y2R2	36	807	91,6	7,59	59	Tyrosineprotein phosphatase nonreceptor type 22 GN=PTPN22 PE=1 SV=2 [PTN22]
Q15911	4	3703	404,2	6,20	59	Zinc finger homeobox protein 3 GN=ZFHX3 PE=1 SV=2 [ZFHX3]
P35579	4	1960	226,4	5,60	59	Myosin9 GN=MYH9 PE=1 SV=4 [MYH9]
O94986	10	1654	189,0	5,44	59	Centrosomal protein of 152 kDa GN=CEP152 PE=1 SV=3 [CE152]

Appendices

Q9UUK3	177	1724	192,5	5,66	59	Poly [ADPribose] polymerase 4 GN=PARP4 PE=1 SV=3 [PARP4]
O75116	13	1388	160,8	6,02	59	Rhoassociated protein kinase 2 GN=ROCK2 PE=1 SV=4 [ROCK2]
Q9HAP6	3	207	22,9	8,66	58	Protein lin7 homolog B GN=LIN7B PE=1 SV=1 [LIN7B]
Q00013	1	466	52,3	7,37	58	55 kDa erythrocyte membrane protein GN=MPP1 PE=1 SV=2 [EM55]
Q9H0B6	16	622	68,9	7,15	58	Kinesin light chain 2 GN=KLC2 PE=1 SV=1 [KLC2]
Q9Y2H5	20	1048	117,1	9,10	58	Pleckstrin homology domaincontaining family A member 6 GN=PLEKHA6 PE=1 SV=4
Q7Z3V4	2	1068	123,0	8,19	58	Ubiquitinprotein ligase E3B GN=UBE3B PE=1 SV=3 [UBE3B]
O75145	14	1194	133,4	5,68	58	Liprinalpha3 GN=PPFIA3 PE=1 SV=3 [LIPA3]
Q01973	18	937	104,2	7,17	58	Tyrosineprotein kinase transmembrane receptor ROR1 GN=ROR1 PE=2 SV=2 [ROR1]
Q86UW7	7	1296	147,6	6,19	58	Calciumdependent secretion activator 2 GN=CADPS2 PE=1 SV=2 [CAPS2]
P62331	1	175	20,1	8,95	58	ADPriboseylation factor 6 GN=Arf6 PE=1 SV=2 [ARF6]
P46952	133	286	32,5	5,88	58	3hydroxyanthranilate 3,4dioxygenase GN=HAAO PE=1 SV=2 [3HAO]
Q8TE56	2	1095	121,0	8,06	58	A disintegrin and metalloproteinase with thrombospondin motifs 17 GN=ADAMTS1
Q1MSJ5	22	1256	145,4	6,80	58	Centrosome and spindle poleassociated protein 1 GN=CSPP1 PE=1 SV=4 [CSPP1]
Q969T7	80	292	33,5	6,68	58	Cytosolic 5'nucleotidase IIIlike protein GN=NT5C3L PE=1 SV=3 [5NT3L]
Q5T5U3	4	1957	217,2	7,80	58	Rho GTPaseactivating protein 21 GN=ARHGAP21 PE=1 SV=1 [RHG21]
Q2PPJ7	1	1873	210,6	6,07	57	Ral GTPaseactivating protein subunit alpha2 GN=RALGAP2 PE=1 SV=2 [RGPA2]
P08729	14	469	51,4	5,48	57	Keratin, type II cytoskeletal 7 GN=KRT7 PE=1 SV=5 [K2C7]
Q67726	63	1436	163,8	6,92	57	Nonstructural polyprotein 1AB astrovirus1 GN=ORF1 PE=3 SV=1 [NS1ABHASV1]
Q14289	16	1009	115,8	6,25	57	Proteintyrosine kinase 2beta GN=PTK2B PE=1 SV=2 [FAK2]
Q5TAH2	168	1124	129,0	6,92	57	Sodium/hydrogen exchanger 11 GN=SLC9A11 PE=2 SV=1 [S9A11]
Q8N9H9	190	656	69,7	5,43	57	Uncharacterized protein C1orf127 GN=C1orf127 PE=2 SV=2 [CA127]
O60330	22	932	100,9	5,05	57	Protocadherin gammaA12 GN=PCDHGA12 PE=2 SV=1 [PCDGC]
Q13535	177	2644	301,2	7,43	57	Serine/threonineprotein kinase ATR GN=ATR PE=1 SV=3 [ATR]
Q13561	33	401	44,2	5,21	57	Dynactin subunit 2 GN=DCTN2 PE=1 SV=4 [DCTN2]
Q9BQE9	1	202	22,2	4,75	56	Bcell CLL/lymphoma 7 protein family member B GN=BCL7B PE=1 SV=1 [BCL7B]
P42694	21	1942	218,8	7,42	56	Probable helicase with zinc finger domain GN=HELZ PE=1 SV=2 [HELZ]
Q711Q0	5	1435	156,4	6,38	56	Uncharacterized protein C10orf71 GN=C10orf71 PE=2 SV=2 [CJ071]
Q2M1K9	1	1284	144,5	6,89	56	Zinc finger protein 423 GN=ZNF423 PE=1 SV=1 [ZN423]
P20338	4	213	23,9	6,07	56	Rasrelated protein Rab4A GN=RAB4A PE=1 SV=2 [RAB4A]
P61018	6	213	23,6	6,06	56	Rasrelated protein Rab4B GN=RAB4B PE=1 SV=1 [RAB4B]
Q96SB3	19	815	89,1	4,97	56	Neurabin2 GN=PPP1R9B PE=1 SV=2 [NEB2]
O14744	15	637	72,6	6,29	56	Protein arginine Nmethyltransferase 5 GN=PRMT5 PE=1 SV=4 [ANM5]
Q9NRL2	12	1556	178,6	6,60	56	Bromodomain adjacent to zinc finger domain protein 1A GN=BAZ1A PE=1 SV=2 [BA
P35670	19	1465	157,2	6,70	56	Coppertransporting ATPase 2 GN=ATP7B PE=1 SV=4 [ATP7B]
Q96HP0	10	2047	229,4	6,74	56	Dedicator of cytokinesis protein 6 GN=DOCK6 PE=1 SV=3 [DOCK6]
Q9ULV0	440	1848	213,5	7,20	56	MyosinVb GN=MYO5B PE=1 SV=3 [MYO5B]
Q7Z6J4	436	655	74,8	6,93	56	FYVE, RhoGEF and PH domaincontaining protein 2 GN=FGD2 PE=2 SV=1 [FGD2]
Q3ZCX4	61	644	74,3	8,22	56	Zinc finger protein 568 GN=ZNF568 PE=2 SV=2 [ZN568]
Q9HAU0	12	1116	127,4	7,53	56	Pleckstrin homology domaincontaining family A member 5 GN=PLEKHA5 PE=1 SV=1
Q7RTW8	78	1153	128,5	5,82	56	Otoancorin GN=OTOA PE=1 SV=1 [OTOAN]
Q09666	11	5890	628,7	6,15	55	Neuroblast differentiationassociated protein AHNAC GN=AHNAC PE=1 SV=2 [AHN
Q00536	8	496	55,7	7,62	55	Cyclindependent kinase 16 GN=CDK16 PE=1 SV=1 [CDK16]
Q00526	5	305	35,0	8,79	55	Cyclindependent kinase 3 GN=CDK3 PE=1 SV=1 [CDK3]
Q8NFT6	108	615	67,2	8,34	55	Protein DBF4 homolog B GN=DBF4B PE=1 SV=1 [DBF4B]
Q7Z5J4	74	1906	203,2	8,79	55	Retinoic acidinduced protein 1 GN=RAI1 PE=1 SV=2 [RAI1]
O95425	10	2214	247,6	6,98	55	Supervillin GN=SVIL PE=1 SV=2 [SVIL]
P10412	9	219	21,9	11,03	55	Histone H1.4 GN=HIST1H1E PE=1 SV=2 [H14]
Q9Y6K9	290	419	48,2	5,71	55	NFkappaB essential modulator (NEMO) (NFkappaB essential modifier) (Inhibitor of
Q7Z6G8	21	1248	138,0	6,37	55	Ankyrin repeat and sterile alpha motif domaincontaining protein 1B GN=ANKS1B P
O95819	8	1239	142,0	7,46	55	Mitogenactivated protein kinase kinase kinase 4 GN=MAP4K4 PE=1 SV=2 [M
Q9BXJ9	445	866	101,2	7,42	55	Nalpaacetyltransferase 15, NatA auxiliary subunit GN=NAA15 PE=1 SV=1 [NAA15]
Q9UHA4	1	124	13,6	7,34	55	Ragulator complex protein LAMTOR3 GN=LAMTOR3 PE=1 SV=1 [LTOR3]
O75683	26	361	41,4	10,64	55	Surfeit locus protein 6 GN=SURF6 PE=1 SV=3 [SURF6]
Q2VWP7	52	1150	127,0	7,59	55	Protogenin GN=PRTG PE=2 SV=1 [PRTG]
Q9HD67	443	2058	237,2	6,21	54	MyosinX GN=MYO10 PE=1 SV=3 [MYO10]
Q6ULP2	19	937	102,1	4,54	54	Aftiphilin GN=AFTPH PE=1 SV=2 [AFTIN]
O00192	52	962	104,6	6,81	54	Armadillo repeat protein deleted in velocardiocardiac syndrome GN=ARVCF PE=1 SV
Q81WG1	1	891	102,9	5,74	54	WD repeatcontaining protein 63 GN=WDR63 PE=2 SV=1 [WDR63]
Q9BSV6	186	310	33,6	8,43	54	tRNAsplicing endonuclease subunit Sen34 GN=TSN34 PE=1 SV=1 [SEN34]
Q9BXT4	18	1180	131,9	6,34	54	Tudor domaincontaining protein 1 GN=TDRD1 PE=1 SV=2 [TDRD1]
Q81XQ6	1	854	96,3	7,91	54	Poly [ADPribose] polymerase 9 GN=PARP9 PE=1 SV=2 [PARP9]
Q7Z494	89	1330	150,8	6,76	54	Nephrocystin3 GN=NPHP3 PE=1 SV=1 [NPHP3]
Q14146	28	1524	170,4	7,31	54	Unhealthy ribosome biogenesis protein 2 homolog GN=URB2 PE=1 SV=2 [URB2]
O43795	5	1136	131,9	9,38	54	MyosinIb GN=MYO1B PE=1 SV=3 [MYO1B]

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Q9UPU5	42	2620	294,2	6,14	54	Ubiquitin carboxylterminal hydrolase 24 GN=USP24 PE=1 SV=3 [UBP24]
PODJ1	45	1756	197,2	6,20	54	RANBP2like and GRIP domaincontaining protein 2 GN=RGPD2 PE=2 SV=1 [RGPD2]
Q8TC71	9	538	61,1	8,63	54	Spermatogenesisassociated protein 18 GN=SPATA18 PE=1 SV=1 [SPT18]
POC221	10	828	97,4	5,99	54	Uncharacterized protein C14orf38 GN=C14orf38 PE=2 SV=1 [CN038]
Q8WVJ2	66	157	17,7	5,07	53	NudC domaincontaining protein 2 GN=NUDC2 PE=1 SV=1 [NUDC2]
Q9Y4E6	10	1490	163,7	6,92	53	WD repeatcontaining protein 7 GN=WDR7 PE=2 SV=2 [WDR7]
Q96A58	1	199	22,6	7,37	53	Rasrelated and estrogenregulated growth inhibitor GN=REG PE=1 SV=1 [REG]
P48547	2	511	57,9	6,58	53	Potassium voltagegated channel subfamily C member 1 GN=KCNK1 PE=1 SV=1 [KCNK1]
AOMZ66	9	631	71,6	5,33	53	Shoottin1 GN=KIAA1598 PE=1 SV=4 [SHOT1]
O94933	1	977	108,9	7,37	53	SLIT and NTRKlike protein 3 GN=SLITRK3 PE=2 SV=2 [SLIK3]
Q96PY6	8	1258	142,7	5,94	53	Serine/threonineprotein kinase Nek1 GN=NEK1 PE=1 SV=2 [NEK1]
Q13428	3	1488	152,0	9,04	53	Treacle protein GN=TCOF1 PE=1 SV=3 [TCOF]
Q727M9	36	940	106,2	9,47	53	Polypeptide Nacetylglactosaminyltransferase 5 GN=GALNT5 PE=1 SV=1 [GALNT5]
Q13395	60	1621	181,6	7,05	53	Probable methyltransferase TARBP1 GN=TARBP1 PE=1 SV=1 [TARB1]
Q9UF12	28	536	58,8	8,60	53	Probable proline dehydrogenase 2 GN=PRODH2 PE=1 SV=1 [PROD2]
Q6P996	4	788	86,7	5,38	53	Pyridoxaldependent decarboxylase domaincontaining protein 1 GN=PDXDC1 PE=1 SV=1 [PDXDC1]
Q5H9M0	16	696	79,0	4,97	53	PWWP domaincontaining protein MUM1L1 GN=MUM1L1 PE=2 SV=1 [MUM1L1]
Q96J65	8	1359	152,2	8,35	53	Multidrug resistanceassociated protein 9 GN=ABCC12 PE=1 SV=2 [MRP9]
Q8NHM5	70	1336	152,5	8,56	53	Lysinespecific demethylase 2B GN=KDM2B PE=1 SV=1 [KDM2B]
P49917	8	911	103,9	7,96	52	DNA ligase 4 GN=LIG4 PE=1 SV=2 [DNLI4]
Q8TBF8	3	368	42,4	8,92	52	Protein FAM81A GN=FAM81A PE=2 SV=3 [FA81A]
Q9ULD4	12	1205	135,7	6,58	52	Bromodomain and PHD fingercontaining protein 3 GN=BRPF3 PE=1 SV=2 [BRPF3]
Q9P2E9	21	1410	152,4	8,60	52	Ribosomebinding protein 1 GN=RRBP1 PE=1 SV=4 [RRBP1]
Q9BRK4	9	669	72,7	6,51	52	Leucine zipper putative tumor suppressor 2 GN=LZTS2 PE=1 SV=2 [LZTS2]
Q96S53	407	571	63,6	7,06	52	Dual specificity testisspecific protein kinase 2 GN=TESK2 PE=2 SV=1 [TESK2]
Q14315	38	2725	290,8	5,97	52	FilaminC GN=FLNC PE=1 SV=3 [FLNC]
Q03989	1	594	64,0	9,25	52	ATrich interactive domaincontaining protein 5A GN=ARID5A PE=2 SV=2 [ARISA]
Q13905	12	1077	120,5	5,92	52	Rap guanine nucleotide exchange factor 1 GN=RAPGEF1 PE=1 SV=3 [RPGF1]
Q5THR3	6	1501	172,8	8,40	52	EFhand calciumbinding domaincontaining protein 6 GN=EFCAB6 PE=1 SV=1 [EFCB6]
P57768	10	344	39,1	4,65	52	Sorting nexin16 GN=SNX16 PE=1 SV=2 [SNX16]
Q5W0A0	6	696	81,6	4,75	52	Protein FAM194B GN=FAM194B PE=2 SV=1 [F194B]
Q9NQG7	2	708	76,9	5,45	52	HermanskyPudlak syndrome 4 protein GN=HPS4 PE=1 SV=2 [HPS4]
Q726E9	23	1792	201,4	9,64	52	E3 ubiquitinprotein ligase RBBP6 GN=RBBP6 PE=1 SV=1 [RBBP6]
Q9ULE3	53	1009	113,8	8,95	52	DENN domaincontaining protein 2A GN=DENND2A PE=2 SV=4 [DEN2A]
Q14028	11	1251	139,6	4,81	52	Cyclic nucleotidedegated cation channel beta1 GN=CNGB1 PE=1 SV=2 [CNGB1]
Q4AC94	7	2353	260,2	7,12	52	C2 domaincontaining protein 3 GN=C2CD3 PE=1 SV=4 [C2CD3]
Q99613	36	913	105,3	5,68	52	Eukaryotic translation initiation factor 3 subunit C GN=EIF3C PE=1 SV=1 [EIF3C]
Q8NA56	13	475	55,0	5,71	52	Tetratricopeptide repeat protein 29 GN=TTC29 PE=2 SV=2 [TTC29]
Q96RV3	15	2341	258,5	7,21	52	Pecanexlike protein 1 GN=PCNX PE=2 SV=2 [PCX1]
Q01518	9	475	51,9	8,06	52	Adenyl cyclaseassociated protein 1 GN=CAP1 PE=1 SV=5 [CAP1]
Q9UKA4	8	1901	210,4	5,39	51	Akinase anchor protein 11 GN=AKAP11 PE=1 SV=1 [AKA11]
P13611	1	3396	372,6	4,51	51	Versican core protein GN=VCAN PE=1 SV=3 [CSPG2]
Q9HC36	79	420	47,0	8,73	51	RNA methyltransferaselike protein 1 GN=RNMTL1 PE=1 SV=2 [RMTL1]
P55786	5	919	103,2	5,72	51	Puromycinsensitive aminopeptidase GN=NPEPPS PE=1 SV=2 [PSA]
Q12965	8	1108	127,0	8,92	51	MyosinIe GN=MYO1E PE=1 SV=2 [MYO1E]
O43150	23	1006	111,6	6,68	51	ArfGAP with SH3 domain, ANK repeat and PH domaincontaining protein 2 GN=ASAP2 PE=1 SV=2 [ASAP2]
Q13206	9	875	100,8	8,63	51	Probable ATPdependent RNA helicase DDX10 GN=DDX10 PE=1 SV=2 [DDX10]
O14727	79	1248	141,7	6,40	51	Apoptotic proteaseactivating factor 1 GN=APAF1 PE=1 SV=2 [APAF]
Q86SQ0	23	1253	142,1	7,43	51	Pleckstrin homologylike domain family B member 2 GN=PHLB2 PE=1 SV=2 [PHLB2]
O43566	329	566	61,4	8,19	51	Regulator of Gprotein signaling 14 GN=RGS14 PE=1 SV=4 [RGS14]
P50995	60	505	54,4	7,65	51	Annexin A11 GN=ANXA11 PE=1 SV=1 [ANX11]
Q5QJE6	348	756	84,4	6,16	51	Deoxynucleotidyltransferase terminalinteracting protein 2 GN=DNITP2 PE=1 SV=2 [DNITP2]
Q9H3S7	7	1636	178,9	6,92	51	Tyrosineprotein phosphatase nonreceptor type 23 GN=PTPN23 PE=1 SV=1 [PTN23]
P01763	42	114	12,2	8,50	51	Ig heavy chain VIII region WEA PE=1 SV=1 [HV302]
Q96L58	29	329	37,1	9,66	51	Beta1,3galactosyltransferase 6 GN=B3GALT6 PE=2 SV=2 [B3GT6]
O95267	5	797	90,3	7,96	51	RAS guanylreleasing protein 1 GN=RASGRP1 PE=1 SV=2 [GRP1]
Q6N022	19	2769	307,8	6,55	51	Teneurin4 GN=ODZ4 PE=2 SV=2 [TEN4]
O94808	15	682	76,9	7,37	51	Glucosaminfructose6phosphate aminotransferase [isomerizing] 2 GN=GFPT2 PE=1 SV=2 [GFPT2]
Q58F21	6	947	107,9	8,95	51	Bromodomain testisspecific protein GN=BRDT PE=1 SV=4 [BRDT]
Q8N884	450	522	58,8	9,48	50	Uncharacterized protein C6orf150 GN=C6orf150 PE=1 SV=2 [CF150]
O15055	22	1255	136,5	6,47	50	Period circadian protein homolog 2 GN=PER2 PE=1 SV=2 [PER2]
Q9UBJ2	69	740	83,2	8,92	50	ATPbinding cassette subfamily D member 2 GN=ABCD2 PE=1 SV=1 [ABCD2]
Q96CP6	1	724	80,6	6,74	50	GRAM domaincontaining protein 1A GN=GRAMD1A PE=1 SV=2 [GRM1A]
Q8NEP3	5	725	80,0	4,67	50	Leucinerich repeatcontaining protein 50 GN=LRC50 PE=1 SV=5 [LRC50]
Q86YP4	14	633	68,0	9,94	50	Transcriptional repressor p66alpha GN=GATAD2A PE=1 SV=1 [P66A]

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O43424	69	1007	113,3	6,07	50	Glutamate receptor delta2 subunit GN=GRID2 PE=2 SV=2 [GRID2]
Q16236	56	605	67,8	4,78	50	Nuclear factor erythroid 2related factor 2 GN=NFE2L2 PE=1 SV=3 [NFE2L2]
P08237	20	780	85,1	7,99	50	6phosphofructokinase, muscle type GN=PFKM PE=1 SV=2 [K6PF]
P02786	1	760	84,8	6,61	50	Transferrin receptor protein 1 GN=TFRC PE=1 SV=2 [TFR1]
A6NI56	14	674	76,0	8,38	50	Coiledcoil domaincontaining protein 154 GN=CCDC154 PE=2 SV=4 [CC154]
P23634	12	1241	137,8	6,60	50	Plasma membrane calciumtransporting ATPase 4 GN=ATP2B4 PE=1 SV=2 [AT2B4]
O00159	7	1063	121,6	9,41	50	Myosinic GN=MYO1C PE=1 SV=4 [MYO1C]
Q96JD6	3	320	36,6	7,49	50	1,5anhydroDfructose reductase GN=AKR1E2 PE=1 SV=2 [AKCL2]
Q05397	18	1052	119,2	6,62	50	Focal adhesion kinase 1 GN=PTK2 PE=1 SV=2 [FAK1]
Q7Z745	5	1585	180,7	6,28	50	HEAT repeatcontaining protein 7B2 GN=HEATR7B2 PE=2 SV=3 [HTRB2]
Q15561	21	434	48,3	7,33	50	Transcriptional enhancer factor TEF3 GN=TEAD4 PE=1 SV=3 [TEAD4]
Q14203	16	1278	141,6	5,81	50	Dynactin subunit 1 GN=DCTN1 PE=1 SV=3 [DCTN1]
Q8NHH1	7	538	58,0	9,20	50	Tubulin polyglutamylase TTL11 GN=TLL11 PE=2 SV=1 [TLL11]
Q15021	8	1401	157,1	6,61	49	Condensin complex subunit 1 GN=NCAPD2 PE=1 SV=3 [CND1]
Q92797	367	1274	141,1	6,13	49	Symplekin GN=SYMPK PE=1 SV=2 [SYMPK]
B1AJ29	4	1412	161,8	6,95	49	Forkheadassociated domaincontaining protein 1 GN=FHAD1 PE=2 SV=2 [FHAD1]
Q9BXX0	4	1053	115,6	6,46	49	EMILIN2 GN=EMILIN2 PE=1 SV=3 [EMIL2]
Q8WXX1	8	588	65,8	5,83	49	Ankyrin repeat and SOCS box protein 15 GN=ASB15 PE=2 SV=3 [ASB15]
Q14511	10	834	92,8	6,70	49	Enhancer of filamentation 1 GN=NEDD9 PE=1 SV=1 [CASL]
Q86VF2	8	1251	137,7	7,56	49	Immunoglobulinlike and fibronectin type III domaincontaining protein 1 GN=IGFN1
P35236	21	360	40,5	6,79	49	Tyrosineprotein phosphatase nonreceptor type 7 GN=PTPN7 PE=1 SV=3 [PTN7]
Q02846	1	1103	120,0	7,44	49	Retinal guanylyl cyclase 1 GN=GUCY2D PE=1 SV=2 [GUC2D]
Q8IZJ3	20	1885	206,6	6,42	49	C3 and PZPIlike alpha2macroglobulin domaincontaining protein 8 GN=CPAMD8 PE=1
Q9H3U1	16	944	103,0	6,07	49	Protein unc45 homolog A GN=UNC45A PE=1 SV=1 [UN45A]
Q96JP5	14	570	63,4	7,36	49	E3 ubiquitinprotein ligase ZFP91 GN=ZFP91 PE=1 SV=1 [ZFP91]
Q9UH92	16	298	33,3	8,06	49	Maxlike protein X GN=MLX PE=1 SV=2 [MLX]
P51451	18	505	57,7	7,87	49	Tyrosineprotein kinase Blk GN=BLK PE=1 SV=3 [BLK]
Q8WYB5	13	2073	231,2	5,96	49	Histone acetyltransferase MYST4 GN=MYST4 PE=1 SV=3 [MYST4]
Q9H4A3	15	2382	250,6	6,34	49	Serine/threonineprotein kinase WNK1 GN=WNK1 PE=1 SV=2 [WNK1]
Q07866	14	573	65,3	6,20	49	Kinesin light chain 1 GN=KLC1 PE=1 SV=2 [KLC1]
Q7L592	11	441	49,2	8,34	49	Protein midA homolog, mitochondrial GN=C2orf56 PE=1 SV=1 [MIDA]
Q6UB98	1	2062	235,5	7,01	49	Ankyrin repeat domaincontaining protein 12 GN=ANKRD12 PE=1 SV=3 [ANR12]
Q8IXK2	36	581	66,9	6,80	49	Polypeptide Nacetylgalactosaminyltransferase 12 GN=GALNT12 PE=1 SV=3 [GLT12]
Q96L73	9	2696	296,5	8,03	49	Histone lysine Nmethyltransferase, H3 lysine36 and H4 lysine20 specific GN=NSD1F
P56945	11	870	93,3	5,67	48	Breast cancer antiestrogen resistance protein 1 GN=BCAR1 PE=1 SV=2 [BCAR1]
P13612	26	1032	114,8	6,48	48	Integrin alpha4 GN=ITGA4 PE=1 SV=3 [ITA4]
Q95460	16	341	39,3	6,30	48	Major histocompatibility complex class Irelated gene protein GN=MR1 PE=1 SV=1 [
A6NKG5	3	1359	155,1	5,20	48	Retrotransposonlike protein 1 GN=RTL1 PE=2 SV=2 [RTL1]
Q9NW08	181	1133	127,7	8,50	48	DNAdirected RNA polymerase III subunit RPC2 GN=POLR3B PE=1 SV=2 [RPC2]
Q8N371	339	416	47,2	5,80	48	Lysinespecific demethylase 8 GN=JMJD5 PE=1 SV=1 [KDM8]
Q86Z14	25	1044	119,7	9,22	48	Betaklotho GN=KLB PE=1 SV=1 [KLOTB]
P08582	363	738	80,2	5,94	48	Melanotransferrin GN=MFI2 PE=1 SV=2 [TRFM]
O43639	91	380	42,9	6,95	48	Cytoplasmic protein NCK2 GN=NCK2 PE=1 SV=2 [NCK2]
Q96N23	26	812	92,3	7,71	48	Uncharacterized protein C12orf55 GN=C12orf55 PE=2 SV=2 [CLO55]
O95185	34	931	103,1	6,10	48	Netrin receptor UNC5C GN=UNC5C PE=1 SV=2 [UNC5C]
Q9UMW8	8	372	43,0	7,80	48	Ubl carboxylterminal hydrolase 18 GN=USP18 PE=1 SV=1 [UBP18]
Q9BXT8	58	1623	184,5	5,40	48	RING finger protein 17 GN=RNF17 PE=1 SV=3 [RNF17]
Q8TEL6	70	797	90,8	7,61	48	Short transient receptor potential channel 4associated protein GN=TRPC4AP PE=1.S
Q9Y2I1	68	1504	166,5	5,14	48	Nischarin GN=NISCH PE=1 SV=3 [NISCH]
Q9UKE5	6	1360	154,8	7,17	48	TRAF2 and NCKinteracting protein kinase GN=TNIK PE=1 SV=1 [TNIK]
P23921	40	792	90,0	7,15	48	Ribonucleosidediphosphate reductase large subunit GN=RRM1 PE=1 SV=1 [RIR1]
Q9BUQ8	12	820	95,5	9,55	48	Probable ATPdependent RNA helicase DDX23 GN=DDX23 PE=1 SV=3 [DDX23]
P25705	11	553	59,7	9,13	47	ATP synthase subunit alpha, mitochondrial GN=ATP5A1 PE=1 SV=1 [ATPA]
Q16820	20	701	79,5	5,74	47	Meprip A subunit beta GN=MEP1B PE=1 SV=3 [MEP1B]
P01700	9	112	11,9	8,91	47	Ig lambda chain VI region HA PE=1 SV=1 [LV102]
Q8NEB9	11	887	101,5	6,81	47	Phosphatidylinositol 3kinase catalytic subunit type 3 GN=PIK3C3 PE=1 SV=1 [PK3C3]
Q6NT04	13	549	63,2	8,75	47	Tigger transposable elementderived protein 7 GN=TIGD7 PE=2 SV=1 [TIGD7]
O94769	7	699	79,7	5,41	47	Extracellular matrix protein 2 GN=ECM2 PE=2 SV=1 [ECM2]
Q4G0S7	14	254	30,0	9,06	47	Coiledcoil domaincontaining protein 152 GN=CCDC152 PE=2 SV=3 [CC152]
P09131	3	477	50,3	7,77	47	P3 protein GN=SLC10A3 PE=2 SV=1 [P3]
P13639	4	858	95,3	6,83	47	Elongation factor 2 GN=EEF2 PE=1 SV=4 [EF2]
O94927	18	633	71,6	8,51	47	HAUS augminlike complex subunit 5 GN=HAUS5 PE=1 SV=2 [HAUS5]
Q8NFP4	39	955	105,7	8,34	47	MAM domaincontaining glycosylphosphatidylinositol anchor protein 1 GN=MDGA1
Q9Y426	29	696	75,5	6,92	47	C2 domaincontaining protein 2 GN=C2CD2 PE=1 SV=2 [CU025]
Q86S16	8	1040	113,8	4,56	47	Desmoglein4 GN=DSG4 PE=1 SV=1 [DSG4]

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Q6PKG0	34	1096	123,4	8,82	47	Larelated protein 1 GN=LARP1 PE=1 SV=2 [LARP1]
Q99590	10	1463	164,6	8,41	47	Protein SCAF11 GN=SCAF11 PE=1 SV=2 [SCAFB]
Q13402	3	2215	254,2	8,56	47	MyosinVIIa GN=MYO7A PE=1 SV=1 [MYO7A]
Q9Y6N6	21	1575	171,1	6,58	47	Laminin subunit gamma3 GN=LAMC3 PE=2 SV=3 [LAMC3]
Q8NHV1	3	300	34,5	6,46	47	GTPase IMAP family member 7 GN=GIMAP7 PE=2 SV=1 [GIMA7]
P22455	7	802	87,9	6,81	47	Fibroblast growth factor receptor 4 GN=FGFR4 PE=1 SV=2 [FGFR4]
Q8IZU8	20	1212	139,1	8,32	47	Dermatansulfate epimeraselike protein GN=DSEL PE=2 SV=2 [DSEL]
Q14832	4	879	98,8	7,66	46	Metabotropic glutamate receptor 3 GN=GRM3 PE=2 SV=2 [GRM3]
Q9BV20	1	369	39,1	6,30	46	Methylthioribose1phosphate isomerase GN=MRI1 PE=1 SV=1 [MTNA]
P82932	157	125	14,2	9,26	46	Mitochondrial 28S ribosomal protein S6 (S6mt) (MRPS6) [RT06]
Q9NR16	20	1453	159,1	5,76	46	Scavenger receptor cysteine-rich type 1 protein M160 GN=CD163L1 PE=1 SV=2 [C163]
Q96BY6	9	2186	249,4	7,14	46	Dedicator of cytokinesis protein 10 GN=DOCK10 PE=1 SV=3 [DOC10]
Q9H9J4	11	1324	145,3	8,63	46	Ubiquitin carboxylterminal hydrolase 42 GN=USP42 PE=1 SV=3 [UBP42]
Q96AA8	56	810	94,9	6,16	46	Uncharacterized protein KIAA0555 [K0555]
O75197	16	1615	179,0	5,34	46	Lowdensity lipoprotein receptorrelated protein 5 GN=LRP5 PE=1 SV=2 [LRP5]
Q13424	29	505	53,9	6,80	46	Alpha1syntrophin GN=SNTA1 PE=1 SV=1 [SNTA1]
P59045	14	1033	117,7	7,71	46	NACHT, LRR and PYD domainscontaining protein 11 GN=NLRP11 PE=2 SV=2 [NAL11]
Q49AM1	14	385	44,4	8,97	46	mTERF domaincontaining protein 3, mitochondrial GN=MTERFD3 PE=1 SV=2 [MTERF3]
Q68C22	64	1445	155,2	6,81	46	Tensin3 GN=TNS3 PE=1 SV=2 [TENS3]
Q92558	2	559	61,6	6,46	46	WiskottAldrich syndrome protein family member 1 GN=WASF1 PE=1 SV=1 [WASF1]
P35711	19	763	84,0	6,60	46	Transcription factor SOX5 GN=SOX5 PE=2 SV=3 [SOX5]
Q13733	14	1029	114,1	6,64	46	Sodium/potassiumtransporting ATPase subunit alpha4 GN=ATP1A4 PE=1 SV=3 [AT1A4]
Q9Y4G8	14	1499	167,3	6,67	46	Rap guanine nucleotide exchange factor 2 (Neural RAP guanine nucleotide exchange factor 2) GN=RAPGEF2 PE=1 SV=2 [RAPGEF2]
P51160	1	858	99,1	5,72	46	Cone cGMPspecific 3',5'cyclic phosphodiesterase subunit alpha' GN=PDE6C PE=1 SV=2 [PDE6C]
Q12934	9	665	74,5	5,14	46	Filensin GN=BFSP1 PE=1 SV=3 [BFSP1]
Q9COB5	36	715	77,5	9,01	46	Probable palmitoyltransferase ZDHHC5 GN=ZDHHC5 PE=1 SV=2 [ZDHHC5]
Q9BYX2	48	928	105,3	6,58	46	TBC1 domain family member 2A GN=TBC1D2 PE=1 SV=3 [TBD2A]
Q9Y6D6	18	1849	208,6	5,85	46	Brefeldin A inhibited guanine nucleotideexchange protein 1 GN=ARFGEF1 PE=1 SV=2 [ARFGEF1]
P01604	9	108	12,1	8,81	46	Ig kappa chain VI region Kue PE=1 SV=1 [KV112]
Q9UEG4	42	869	96,6	7,93	46	Zinc finger protein 629 GN=ZNF629 PE=2 SV=2 [ZNF629]
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Q6L8Q7	11	609	67,3	6,57	46	2',5'phosphodiesterase 12 GN=PDE12 PE=1 SV=2 [PDE12]
Q9NV06	16	445	51,4	9,19	46	DDB1 and CUL4associated factor 13 GN=DCAF13 PE=1 SV=2 [DCAF13]
Q96AX9	1	1013	109,9	8,44	45	E3 ubiquitinprotein ligase MIB2 GN=MIB2 PE=1 SV=3 [MIB2]
O43299	4	807	88,5	7,01	45	Uncharacterized protein KIAA0415 GN=KIAA0415 PE=2 SV=2 [K0415]
Q9ULT8	20	2610	289,2	5,35	45	E3 ubiquitinprotein ligase HECTD1 GN=HECTD1 PE=1 SV=3 [HECTD1]
Q9BZJ0	7	848	100,4	8,00	45	Crooked necklike protein 1 GN=CRNKL1 PE=1 SV=4 [CRNL1]
Q38SD2	1	2015	225,2	6,68	45	Leucinerich repeat serine/threonineprotein kinase 1 GN=LRRK1 PE=1 SV=3 [LRRK1]
P00480	56	354	39,9	8,63	45	Ornithine carbamoyltransferase, mitochondrial precursor (EC 2.1.3.3) (OTCase) (Ornithine carbamoyltransferase) GN=ORNT1 PE=1 SV=2 [ORNT1]
Q12912	145	555	62,1	5,85	45	Lymphoidrestricted membrane protein GN=LRMP PE=1 SV=3 [LRMP]
Q13017	4	1502	172,4	6,62	45	Rho GTPaseactivating protein 5 GN=ARHGAP5 PE=1 SV=2 [RHG05]
P33992	10	734	82,2	8,37	45	DNA replication licensing factor MCM5 GN=MCM5 PE=1 SV=5 [MCM5]
Q2M1Z3	3	1444	156,9	5,76	45	Rho GTPaseactivating protein 31 GN=ARHGAP31 PE=1 SV=2 [RHG31]
Q8WXG9	9	6306	692,6	4,64	45	Gprotein coupled receptor 98 GN=GPR98 PE=1 SV=2 [GPR98]
Q9NVE4	3	849	96,3	8,59	45	Coiledcoil domaincontaining protein 87 GN=CCDC87 PE=2 SV=2 [CCD87]
Q9C0G6	15	4158	475,7	6,00	45	Dynein heavy chain 6, axonemal GN=DNAH6 PE=1 SV=3 [DYH6]
Q9H8K7	7	445	49,2	6,25	45	Uncharacterized protein C10orf88 GN=C10orf88 PE=2 SV=2 [C1088]
P78562	12	749	86,4	8,76	45	Phosphateregulating neutral endopeptidase GN=PHEX PE=1 SV=1 [PHEX]
Q13576	75	1575	180,5	5,64	45	Ras GTPaseactivatinglike protein IQGAP2 GN=IQGAP2 PE=1 SV=4 [IQGA2]
O60522	10	2096	236,4	5,25	45	Tudor domaincontaining protein 6 GN=TDRD6 PE=2 SV=2 [TDRD6]
P52888	14	689	78,8	6,05	45	Thimet oligopeptidase GN=THOP1 PE=1 SV=2 [THOP1]
Q9NWM0	9	555	61,8	5,45	45	Spermine oxidase GN=SMOX PE=1 SV=1 [SMOX]
Q9BYD6	24	325	36,9	8,78	45	39S ribosomal protein L1, mitochondrial GN=MRPL1 PE=1 SV=2 [RM01]
P14136	36	432	49,8	5,52	44	Glial fibrillary acidic protein GN=GFAP PE=1 SV=1 [GFAP]
Q86SG4	1	172	20,4	9,83	44	Dresden prostate carcinoma protein 2 GN=C15orf21 PE=2 SV=1 [DPCA2]
Q8WYN3	10	585	64,9	4,75	44	Cysteine/serinerich nuclear protein 3 GN=CSRN3 PE=1 SV=1 [CSRN3]
C9JTQ0	142	380	39,6	11,03	44	Ankyrin repeat domaincontaining protein 63 GN=ANKRD63 PE=4 SV=1 [ANR63]
P14616	1	1297	143,6	6,47	44	Insulin receptorrelated protein GN=INSRR PE=2 SV=2 [INSRR]
Q8WY54	16	764	84,9	5,03	44	Protein phosphatase 1E GN=PPM1E PE=1 SV=2 [PPM1E]
Q99684	12	422	45,3	9,00	44	Zinc finger protein Gfi1 GN=GFI1 PE=1 SV=2 [GFI1]
Q9H013	7	955	104,9	8,35	44	Disintegrin and metalloproteinase domaincontaining protein 19 GN=ADAM19 PE=1 SV=2 [ADAM19]
Q722Y8	18	2422	278,9	6,55	44	Interferoninduced very large GTPase 1 GN=GVIN1 PE=2 SV=2 [GVIN1]
Q96A08	11	127	14,2	10,32	44	Histone H2B type 1A GN=HIST1H2BA PE=1 SV=3 [H2B1A]
Q96CN5	6	670	75,9	6,23	44	Leucinerich repeatcontaining protein 45 GN=LRR45 PE=2 SV=1 [LRC45]
P55060	2	971	110,3	5,77	44	Exportin2 GN=CSE1L PE=1 SV=3 [XPO2]

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P52746	11	1687	187,8	7,91	44	Zinc finger protein 142 GN=ZNF142 PE=1 SV=4 [ZNF142]
Q8NH42	31	731	80,4	6,84	44	E3 ubiquitinprotein ligase RFW2 GN=RFWD2 PE=1 SV=1 [RFWD2]
Q9NVL8	12	296	34,7	9,28	44	Uncharacterized protein C14orf105 GN=C14orf105 PE=2 SV=2 [CN105]
Q8WUM0	43	1156	128,9	5,10	44	Nuclear pore complex protein Nup133 GN=NUP133 PE=1 SV=2 [NUP133]
Q9ULV3	197	898	100,0	6,11	44	Cip1interacting zinc finger protein GN=CIZ1 PE=1 SV=2 [CIZ1]
Q99608	13	321	36,1	8,78	44	Necdin GN=NDN PE=1 SV=1 [NECD]
Q9Y2G9	3	1366	150,2	6,52	44	Protein strawberry notch homolog 2 GN=SBNO2 PE=2 SV=3 [SBNO2]
Q92540	2	1137	127,2	8,72	44	Protein SMG7 GN=SMG7 PE=1 SV=2 [SMG7]
O95219	328	450	51,9	5,99	43	Sorting nexin4 GN=SNX4 PE=1 SV=1 [SNX4]
Q8N7X0	38	1667	189,6	8,31	43	Calpain7like protein GN=C6orf103 PE=2 SV=3 [CAN7L]
Q14435	26	633	72,6	7,99	43	Polypeptide Nacetylglucosaminyltransferase 3 GN=GALNT3 PE=2 SV=2 [GALNT3]
Q49MI3	93	558	62,6	8,27	43	Ceramide kinaselike protein GN=CERKL PE=1 SV=1 [CERKL]
Q9Y4F4	368	1720	189,2	8,50	43	Protein FAM179B GN=FAM179B PE=1 SV=4 [F179B]
Q8N6S4	1	541	60,8	6,93	43	Ankyrin repeat domaincontaining protein 13C GN=ANKRD13C PE=2 SV=2 [AN13C]
Q9Y2D5	21	859	94,6	5,11	43	Akinase anchor protein 2 GN=AKAP2 PE=1 SV=3 [AKAP2]
Q9V697	7	457	50,2	8,31	43	Cysteine desulfurase, mitochondrial GN=NFS1 PE=1 SV=3 [NFS1]
O76027	442	345	38,3	5,77	43	Annexin A9 GN=ANXA9 PE=1 SV=3 [ANXA9]
O75161	16	1426	157,5	8,13	43	Nephrocystin4 GN=NPHP4 PE=1 SV=2 [NPHP4]
P32926	8	999	107,5	5,00	43	Desmoglein3 GN=DSG3 PE=1 SV=2 [DSG3]
P47756	2	277	31,3	5,59	43	Factinocapping protein subunit beta GN=CAPZB PE=1 SV=4 [CAPZB]
Q9NPF5	28	467	53,0	9,50	43	DNA methyltransferase 1associated protein 1 GN=DMAP1 PE=1 SV=1 [DMAP1]
P56730	31	875	97,0	8,03	43	Neurotrypsin GN=PRSS12 PE=2 SV=2 [NETR]
O95447	7	670	76,5	9,48	42	Lebercilinlike protein GN=LCA5L PE=2 SV=1 [LCA5L]
Q96M91	9	514	61,8	8,90	42	Coiledcoil domaincontaining protein 11 GN=CCDC11 PE=2 SV=2 [CCDC11]
O75369	3	2602	278,0	5,73	42	FilaminB GN=FLNB PE=1 SV=2 [FLNB]
O43526	29	872	95,8	9,23	42	Potassium voltagegated channel subfamily KQT member 2 GN=KCNQ2 PE=1 SV=2 [KCNQ2]
Q02GT2	19	675	80,6	5,33	42	Nexilin GN=NEXN PE=1 SV=1 [NEXN]
O00160	6	1098	124,8	9,11	42	Myosinif GN=MYO1F PE=1 SV=3 [MYO1F]
P30281	17	292	32,5	7,06	42	G1/Sspecific cyclinD3 GN=CCND3 PE=1 SV=2 [CCND3]
P51523	43	738	85,4	8,78	42	Zinc finger protein 84 (Zinc finger protein HPF2) [ZNF84]
Q6PIF6	9	2116	241,4	8,57	42	MyosinVIIb GN=MYO7B PE=2 SV=2 [MYO7B]
Q9NZQ8	5	1165	131,4	6,77	42	Transient receptor potential cation channel subfamily M member 5 GN=TRPM5 PE=1 SV=1 [TRPM5]
P17480	4	764	89,4	5,81	42	Nucleolar transcription factor 1 GN=UBTF PE=1 SV=1 [UBTF]
Q9V277	42	283	30,6	8,66	42	Voltagegated anionselective channel protein 3 GN=VDAC3 PE=1 SV=1 [VDAC3]
Q9BZF1	432	889	101,1	6,96	42	Oxysterolbinding proteinrelated protein 8 GN=OSBPL8 PE=1 SV=3 [OSBPL8]
Q96DR5	434	249	27,0	5,59	42	Short palate, lung and nasal epithelium carcinomaassociated protein 2 GN=SPLUNC2 PE=1 SV=1 [SPLUNC2]
Q8TBK2	18	473	53,2	5,34	42	Nlysine methyltransferase SETD6 GN=SETD6 PE=1 SV=2 [SETD6]
P49910	1	485	55,7	7,17	42	Zinc finger protein 165 GN=ZNF165 PE=1 SV=1 [ZNF165]
Q8WVM7	1	1258	144,3	5,59	42	Cohesin subunit SA1 GN=STAG1 PE=1 SV=3 [STAG1]
Q72736	8	793	85,3	7,83	42	Pleckstrin homology domaincontaining family H member 3 GN=PLEKH3 PE=1 SV=2 [PLEKH3]
Q6UWX4	19	724	80,7	9,01	42	HHLIPlike protein 2 GN=HHLIP2 PE=2 SV=1 [HHLIP2]
P52272	1	730	77,5	8,70	42	Heterogeneous nuclear ribonucleoprotein M GN=HNRNPM PE=1 SV=3 [HNRNPM]
Q9UQN3	2	213	23,9	8,76	41	Charged multivesicular body protein 2b GN=CHMP2B PE=1 SV=1 [CHMP2B]
Q86X52	1	802	91,7	9,23	41	Chondroitin sulfate synthase 1 GN=CHSY1 PE=1 SV=3 [CHSY1]
P17643	25	537	60,7	5,99	41	5,6dihydroxyindole2carboxylic acid oxidase GN=TYRP1 PE=1 SV=2 [TYRP1]
Q9HAU5	25	1272	147,7	5,69	41	Regulator of nonsense transcripts 2 GN=UPF2 PE=1 SV=1 [RENT2]
Q96BQ5	5	260	30,8	9,20	41	Coiledcoil domaincontaining protein 127 [CC127]
Q86V15	454	1759	189,9	7,03	41	Zinc finger protein castor homolog 1 GN=CASZ1 PE=2 SV=4 [CASZ1]
Q99665	6	862	97,1	7,75	41	Interleukin12 receptor beta2 chain precursor (IL12 receptor beta2) (IL12Rbeta2) [IL12Rbeta2]
Q76NI1	23	1749	191,3	6,16	41	Protein very KIND GN=KNDC1 PE=2 SV=2 [VKIND]
Q96EF6	59	278	31,5	8,10	41	Fbox only protein 17 GN=FBXO17 PE=1 SV=1 [FBX17]
P23919	1	212	23,8	8,27	41	Thymidylate kinase GN=DTYMK PE=1 SV=4 [KTHY]
Q16394	20	746	86,2	9,04	41	Exostosin1 GN=EXT1 PE=1 SV=2 [EXT1]
Q70JA7	14	882	100,2	8,75	41	Chondroitin sulfate synthase 3 GN=CHSY3 PE=1 SV=3 [CHSY3]
Q9HCS4	1	588	62,6	8,94	41	Transcription factor 7like 1 GN=TCF7L1 PE=2 SV=1 [TF7L1]
Q96NH3	7	1257	144,7	6,74	41	Protein broadminded GN=BROMI PE=2 SV=4 [BROMI]
P10253	1	952	105,3	6,00	41	Lysosomal alphaglucosidase GN=GAA PE=1 SV=4 [LYAG]
P11678	8	715	81,0	10,29	41	Eosinophil peroxidase GN=EPX PE=1 SV=2 [PERE]
Q8N8U2	93	506	56,5	8,79	41	Chromodomain Ylike protein 2 GN=CDYL2 PE=2 SV=2 [CDYL2]
Q2Q1W2	12	868	93,3	7,61	41	Tripartite motifcontaining protein 71 GN=TRIM71 PE=2 SV=1 [LIN41]
Q9UPN9	19	1127	122,5	6,67	40	E3 ubiquitinprotein ligase TRIM33 GN=TRIM33 PE=1 SV=3 [TRI33]
Q14153	20	422	45,7	6,76	40	Protein FAM53B GN=FAM53B PE=1 SV=2 [FA53B]
Q8IW52	10	837	94,3	7,80	40	SLIT and NTRKlike protein 4 GN=SLITRK4 PE=2 SV=1 [SLIK4]
Q8NA47	119	563	66,2	9,06	40	Coiledcoil domaincontaining protein 63 GN=CCDC63 PE=1 SV=1 [CCDC63]
Q86TU7	17	594	67,2	5,96	40	SET domaincontaining protein 3 GN=SETD3 PE=1 SV=1 [SETD3]

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O75127	27	700	78,8	8,59	40	Pentatricopeptide repeatcontaining protein 1 GN=PTCD1 PE=1 SV=2 [PTCD1]
P17302	28	382	43,0	8,76	40	Gap junction alpha1 protein GN=GJA1 PE=1 SV=2 [CXA1]
Q6UQ28	26	207	23,4	5,12	40	Placentaexpressed transcript 1 protein GN=PLET1 PE=2 SV=2 [PLET1]
Q96CU9	1	486	53,8	7,78	40	FADdependent oxidoreductase domaincontaining protein 1 GN=FOXRED1 PE=1 SV=
Q96J02	23	903	102,7	6,30	40	E3 ubiquitinprotein ligase Itchy homolog GN=ITCH PE=1 SV=2 [ITCH]
P78316	25	857	97,6	7,58	40	Nucleolar protein 14 GN=NOP14 PE=1 SV=3 [NOP14]
Q8TBR7	5	257	29,4	9,41	40	Protein FAM57A GN=FAM57A PE=1 SV=2 [FA57A]
P16402	327	221	22,3	11,02	40	Histone H1.3 GN=HIST1H1D PE=1 SV=2 [H13]
Q9NT68	70	2774	307,6	6,68	40	Teneurin2 GN=ODZ2 PE=1 SV=3 [TEN2]
O43593	7	1189	127,4	7,08	40	Protein hairless GN=HR PE=1 SV=5 [HAIR]
Q92538	4	1859	206,3	5,73	40	Golgispecific brefeldin Aresistance guanine nucleotide exchange factor 1 GN=GBF1
Q8WWZ4	4	1543	175,7	6,65	40	ATPbinding cassette subfamily A member 10 GN=ABCA10 PE=1 SV=3 [ABCAA]
Q2M329	49	555	62,7	4,94	40	Coiledcoil domaincontaining protein 96 GN=CCDC96 PE=2 SV=2 [CCD96]
P54756	71	1037	114,7	6,93	40	Ephrin typeA receptor 5 GN=EPHA5 PE=1 SV=3 [EPHA5]
Q8NOX2	1	631	70,8	6,33	39	Spermassociated antigen 16 protein GN=SPAG16 PE=2 SV=2 [SPG16]
P22352	5	226	25,5	8,13	39	Glutathione peroxidase 3 GN=GPX3 PE=1 SV=2 [GPX3]
O15530	4	556	63,1	7,36	39	3phosphoinositidedependent protein kinase 1 GN=PDPK1 PE=1 SV=1 [PDPK1]
Q8NDZ2	1	872	96,8	6,62	39	Uncharacterized protein C5orf25 GN=C5orf25 PE=1 SV=2 [CE025]
Q35XZ7	41	439	51,4	8,76	39	Probable tubulin polyglutamylase TTL9 GN=TLL9 PE=2 SV=3 [TLL9]
Q96G75	288	393	44,4	6,61	39	Protein RMD5 homolog B GN=RMND5B PE=2 SV=1 [RMD5B]
Q96BN8	24	352	40,2	5,47	39	Protein FAM105B GN=FAM105B PE=1 SV=3 [F105B]
O60241	1	1585	172,5	7,42	39	Brainspecific angiogenesis inhibitor 2 GN=BAI2 PE=2 SV=2 [BAI2]
Q00537	4	523	59,5	9,03	39	Serine/threonineprotein kinase PCTAIRE2 (EC 2.7.11.22) (PCTAIREmotif protein kin
Q96QE3	16	1844	207,4	9,19	39	ATPase family AAA domaincontaining protein 5 GN=ATAD5 PE=1 SV=4 [ATAD5]
P15884	12	667	71,3	7,01	39	Transcription factor 4 GN=TCF4 PE=1 SV=3 [ITF2]
Q8WY91	27	577	62,9	9,28	39	THAP domaincontaining protein 4 GN=THAP4 PE=1 SV=2 [THAP4]
Q5VYM1	5	1079	117,6	7,36	39	Uncharacterized protein C9orf131 GN=C9orf131 PE=2 SV=3 [C131]
Q96GW9	3	593	66,5	8,09	39	MethionyltRNA synthetase, mitochondrial GN=MARS2 PE=1 SV=2 [SYMM]
Q6ZQW0	1	407	45,4	6,84	39	Indoleamine 2,3dioxygenase 2 GN=IDO2 PE=1 SV=3 [I23O2]
Q86YE8	123	645	75,8	9,14	39	Zinc finger protein 573 GN=ZNF573 PE=2 SV=3 [ZN573]
O00566	10	681	78,8	4,86	39	U3 small nucleolar ribonucleoprotein protein MPP10 GN=MPHOSPH10 PE=1 SV=2 [I
Q8N239	124	644	70,6	5,60	39	Kelchlike protein 34 GN=KLHL34 PE=2 SV=1 [KLH34]
Q9ULM3	13	1422	150,7	8,98	39	YEATS domaincontaining protein 2 GN=YEATS2 PE=1 SV=2 [YETS2]
Q9BXW9	8	1471	166,4	6,24	39	Fanconi anemia group D2 protein GN=FANCD2 PE=1 SV=1 [FACD2]
Q5TFE4	2	455	51,8	6,35	39	5'nucleotidase domaincontaining protein 1 GN=NT5DC1 PE=1 SV=1 [NT5D1]
Q17R98	8	1081	119,1	6,89	39	Zinc finger protein 827 GN=ZNF827 PE=2 SV=1 [ZN827]
P07741	3	180	19,6	6,02	39	Adenine phosphoribosyltransferase GN=APRT PE=1 SV=2 [APT]
Q9NYF0	17	836	90,1	8,69	38	Dapper homolog 1 GN=DACT1 PE=2 SV=2 [DACT1]
Q96LP6	27	360	39,7	9,60	38	Uncharacterized protein C12orf42 GN=C12orf42 PE=2 SV=2 [CLO42]
Q99700	3	1313	140,2	9,57	38	Ataxin2 GN=ATXN2 PE=1 SV=2 [ATX2]
Q9UKG1	15	709	79,6	5,41	38	DCCinteracting protein 13alpha GN=APPL1 PE=1 SV=1 [DP13A]
Q9Y5H2	22	935	101,5	4,96	38	Protocadherin gammaA11 GN=PCDHGA11 PE=1 SV=1 [PCDGB]
Q9BRD0	3	619	70,5	9,86	38	BUD13 homolog GN=BUD13 PE=1 SV=1 [BUD13]
Q13615	10	1198	133,5	5,80	38	Myotubularinrelated protein 3 GN=MTMR3 PE=1 SV=3 [MTMR3]
Q8N5C8	23	712	78,6	8,46	38	TGFbetaactivated kinase 1 and MAP3K7binding protein 3 GN=TAB3 PE=1 SV=2 [TAB
B3KS81	3	715	80,3	12,06	38	Serine/arginine repetitive matrix protein 5 GN=SRRM5 PE=2 SV=3 [SRRM5]
Q4W5G0	408	525	59,6	9,00	38	Tigger transposable elementderived protein 2 GN=TIGD2 PE=2 SV=1 [TIGD2]
P53004	4	296	33,4	6,44	38	Biliverdin reductase A GN=BLVRA PE=1 SV=2 [BIEA]
P19367	13	917	102,4	6,80	38	Hexokinase1 (EC 2.7.1.1) (Hexokinase type I) (HK I) (Brain form hexokinase) [HXK1
Q7Z7M0	19	2845	302,9	6,87	38	Multiple epidermal growth factorlike domains protein 8 GN=MEGF8 PE=1 SV=2 [ME
Q5D862	9	2391	247,9	8,31	38	Filaggrin2 GN=FLG2 PE=1 SV=1 [FLA2]
A6NJ16	4	123	13,6	9,60	38	Putative Vset and immunoglobulin domaincontaining protein 6 GN=VSIG6 PE=5 SV=
Q969N2	8	578	65,7	8,38	38	GPI transamidase component PIGT GN=PIGT PE=1 SV=1 [PIGT]
Q460N3	1	656	72,5	8,82	38	Poly [ADPribose] polymerase 15 GN=PARP15 PE=1 SV=1 [PAR15]
Q8I2Z0	1	524	56,9	8,69	38	Zinc finger protein 683 GN=ZNF683 PE=2 SV=3 [ZN683]
Q8NF86	9	280	29,8	9,94	38	Serine protease 33 GN=PRSS33 PE=1 SV=3 [PRS33]
Q8TCB7	26	284	33,2	6,19	38	Methyltransferaselike protein 6 GN=METTL6 PE=2 SV=2 [METL6]
Q9H825	17	291	33,4	6,95	38	Methyltransferaselike protein 8 GN=METTL8 PE=2 SV=2 [METL8]
Q68DN1	12	1984	224,2	10,08	38	Uncharacterized protein C2orf16 GN=C2orf16 PE=1 SV=3 [CB016]
Q07001	2	517	58,9	6,55	38	Acetylcholine receptor subunit delta GN=CHRND PE=1 SV=1 [ACHD]
Q7Z410	144	1059	113,9	8,07	38	Transmembrane protease serine 9 GN=TMPRSS9 PE=1 SV=2 [TMP59]
Q9BPX7	1	421	46,4	6,42	38	UPF0415 protein C7orf25 GN=C7orf25 PE=1 SV=1 [CG025]
Q9C030	36	488	56,4	7,55	38	Tripartite motifcontaining protein 6 GN=TRIM6 PE=1 SV=1 [TRIM6]
P11586	369	935	101,5	7,30	37	C1tetrahydrofolate synthase, cytoplasmic GN=MTHFD1 PE=1 SV=3 [C1TC]
Q8N4C8	5	1332	149,7	7,85	37	Missshapenlike kinase 1 GN=MINK1 PE=1 SV=2 [MINK1]

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P29590	5	882	97,5	6,21	37	Protein PML GN=PML PE=1 SV=3 [PML]
Q9NQ75	2	786	87,1	7,08	37	Cas scaffolding protein family member 4 GN=CASS4 PE=1 SV=2 [CASS4]
Q9P2D3	39	2071	224,2	7,17	37	HEAT repeatcontaining protein 5B GN=HEATR5B PE=1 SV=2 [HTR5B]
P22304	26	550	61,8	5,45	37	Iduronate 2sulfatase GN=IDS PE=1 SV=1 [IDS]
Q9BZE7	1	217	24,9	9,80	37	UPF0193 protein EVG1 [EVG1]
A2VCL2	4	907	103,8	7,90	37	Coiledcoil domaincontaining protein 162 GN=CCDC162P PE=2 SV=3 [CC162]
P20930	10	4061	434,9	9,25	37	Filaggrin GN=FLG PE=1 SV=3 [FLA]
Q9BRX2	3	385	43,3	6,34	37	Protein pelota homolog GN=PELO PE=1 SV=2 [PELO]
P21796	42	283	30,8	8,54	37	Voltagedependent anionselective channel protein 1 GN=VDAC1 PE=1 SV=2 [VDAC1]
Q6IBS0	6	349	39,5	6,84	37	Twinfilin2 GN=TWF2 PE=1 SV=2 [TWF2]
Q9BZ23	3	570	62,6	9,28	37	Pantothenate kinase 2, mitochondrial GN=PANK2 PE=1 SV=3 [PANK2]
Q9P2E3	16	1918	220,1	7,30	37	NFX1type zinc fingercontaining protein 1 GN=ZNF1 PE=1 SV=2 [ZNF1]
P41219	15	470	53,6	5,47	37	Peripherin GN=PRPH PE=1 SV=2 [PERI]
Q658Y4	86	838	93,8	6,39	37	Protein FAM91A1 GN=FAM91A1 PE=1 SV=3 [F91A1]
Q12840	1	1032	117,3	5,90	37	Kinesin heavy chain isoform 5A GN=KIF5A PE=1 SV=2 [KIF5A]
Q9BZ95	3	1437	161,5	8,21	37	Histoneylsine Nmethyltransferase NSD3 GN=WHSC1L1 PE=1 SV=1 [NSD3]
Q9BQE4	10	189	21,2	9,70	37	Selenoprotein S GN=SELS PE=1 SV=3 [SELS]
Q8IWW2	5	1026	113,4	7,47	37	Contactin4 GN=CNTN4 PE=1 SV=1 [CNTN4]
Q9NRP7	23	1315	143,9	5,90	37	Serine/threonineprotein kinase 36 GN=STK36 PE=1 SV=2 [STK36]
Q5JTD7	10	316	33,4	4,84	37	Uncharacterized protein C6orf154 GN=C6orf154 PE=2 SV=1 [CF154]
Q8N187	123	725	80,6	5,69	37	Amyotrophic lateral sclerosis 2 chromosomal region candidate gene 8 protein GN=
P55345	3	433	49,0	5,17	37	Protein arginine Nmethyltransferase 2 GN=PRMT2 PE=1 SV=1 [ANM2]
Q2VPB7	18	878	93,9	5,92	37	AP5 complex subunit beta1 GN=AP5B1 PE=1 SV=4 [AP5B1]
Q9UQP3	6	1299	143,9	5,63	37	TenascinN GN=TNN PE=1 SV=2 [TENN]
Q15291	24	538	59,1	5,10	36	Retinoblastomabinding protein 5 GN=RBBP5 PE=1 SV=2 [RBBP5]
Q9NUS5	13	200	22,5	6,87	36	Uncharacterized protein C20orf29 GN=C20orf29 PE=2 SV=1 [CT029]
Q92692	1	538	57,7	4,82	36	Poliovirus receptorrelated protein 2 GN=PVR2 PE=1 SV=1 [PVR2]
Q8N3C0	10	2202	251,3	7,09	36	Activating signal cointegrator 1 complex subunit 3 GN=ASCC3 PE=1 SV=3 [HELC1]
Q8TER5	3	1519	164,6	6,15	36	Rho guanine nucleotide exchange factor 40 GN=ARHGEF40 PE=1 SV=3 [ARH40]
Q99715	30	3063	332,9	5,53	36	Collagen alpha1(XII) chain GN=COL12A1 PE=1 SV=2 [COCA1]
Q96K76	1	1375	157,2	5,08	36	Ubiquitin carboxylterminal hydrolase 47 GN=USP47 PE=1 SV=3 [UBP47]
Q9H6A9	10	2034	221,9	6,64	36	Pecanexlike protein 3 GN=PCNXL3 PE=1 SV=2 [PCX3]
Q9NV72	25	465	53,8	9,22	36	Zinc finger protein 701 GN=ZNF701 PE=2 SV=2 [ZN701]
P56645	1	1201	131,8	6,89	36	Period circadian protein homolog 3 GN=PER3 PE=2 SV=4 [PER3]
O95271	9	1327	142,0	7,05	36	Tankyrase1 GN=TNKS PE=1 SV=2 [TNKS1]
Q5VT99	3	294	32,1	4,97	36	Leucinerich repeatcontaining protein 38 GN=LRR38 PE=2 SV=1 [LRC38]
Q9V2F9	7	522	58,4	7,46	36	BTB/POZ domaincontaining protein 3 GN=BTBD3 PE=2 SV=1 [BTBD3]
Q06210	26	699	78,8	7,11	36	Glucosaminfructose6phosphate aminotransferase [isomerizing] 1 GN=GFPT1 PE=1
Q9UDX4	40	400	46,0	6,11	36	SEC14like protein 3 GN=SEC14L3 PE=2 SV=1 [S14L3]
Q8N584	17	583	65,8	6,99	36	Tetratricopeptide repeat protein 39C GN=TTC39C PE=1 SV=2 [TT39C]
Q9ULI0	30	1458	164,8	6,80	36	ATPase family AAA domaincontaining protein 2B GN=ATAD2B PE=1 SV=3 [ATD2B]
Q8N201	28	2190	244,1	6,13	36	Integrator complex subunit 1 GN=INTS1 PE=1 SV=2 [INT1]
Q9C0I3	15	900	99,4	7,83	36	Protein FAM190A GN=FAM190A PE=2 SV=2 [F190A]
P05155	171	500	55,1	6,55	36	Plasma protease C1 inhibitor GN=SERPING1 PE=1 SV=2 [IC1]
Q96PP8	5	586	66,6	5,55	36	Guanylatebinding protein 5 GN=GBP5 PE=2 SV=1 [GBP5]
Q5W5X9	7	447	50,0	8,38	36	Tetratricopeptide repeat protein 23 GN=TTC23 PE=2 SV=1 [TTC23]
Q96C03	5	454	49,2	5,12	36	SmithMagenis syndrome chromosomal region candidate gene 7 protein GN=SMCR7
Q9P283	21	1151	125,8	7,65	36	Semaphorin5B GN=SEMA5B PE=2 SV=4 [SEM5B]
O00116	440	658	72,9	7,34	36	Alkyldihydroxyacetonephosphate synthase, peroxisomal GN=AGPS PE=1 SV=1 [AD
Q6DN14	26	999	111,6	8,15	36	Multiple C2 and transmembrane domaincontaining protein 1 GN=MCTP1 PE=2 SV=2
Q8NCI6	2	653	74,8	8,95	36	Betagalactosidase1like protein 3 GN=GLB1L3 PE=2 SV=3 [GLBL3]
Q15404	189	277	31,5	8,65	36	Ras suppressor protein 1 GN=RSU1 PE=1 SV=3 [RSU1]
Q9NPZ5	36	323	36,9	10,62	36	Galactosylgalactosylxylosylprotein 3betagalucuronosyltransferase 2 GN=B3GAT2 PE=
Q96FB5	1	475	52,9	7,97	36	UPF0431 protein C1orf66 GN=C1orf66 PE=2 SV=2 [CA066]
Q6ZU65	8	1347	146,0	9,19	36	Ubinuclein2 GN=UBN2 PE=1 SV=2 [UBN2]
P52735	8	878	101,2	7,08	36	Guanine nucleotide exchange factor VAV2 GN=VAV2 PE=1 SV=2 [VAV2]
A6NEL2	8	793	85,7	9,60	36	Ankyrin repeat domaincontaining protein 56 GN=ANKRD56 PE=2 SV=1 [ANR56]
Q15067	4	660	74,4	8,16	36	Peroxisomal acylcoenzyme A oxidase 1 GN=ACOX1 PE=1 SV=3 [ACOX1]
Q49A17	6	601	69,7	7,46	36	Polypeptide Nacetylglactosaminyltransferaselike 6 GN=GALNTL6 PE=2 SV=2 [GLTL
Q63HK5	10	1081	118,5	7,25	36	Teashirt homolog 3 GN=TSHZ3 PE=1 SV=2 [TSH3]
A8MZ36	32	301	33,9	6,20	36	Envoplakinlike protein GN=EVPLL PE=2 SV=1 [EVPLL]
Q8N6V9	3	391	44,8	6,60	36	Testisexpressed sequence 9 protein GN=TEX9 PE=2 SV=1 [TEX9]
Q9NQ87	440	328	35,1	10,68	36	Hairy/enhancerofsplit related with YRPW motiflike protein GN=HEYL PE=2 SV=2 [HI
P12081	12	509	57,4	5,88	36	HistidyltRNA synthetase, cytoplasmic GN=HARS PE=1 SV=2 [SYHC]
Q8IWR0	12	971	110,5	7,30	36	Zinc finger CCCH domaincontaining protein 7A GN=ZC3H7A PE=1 SV=1 [Z3H7A]

Appendices

P33261	28	490	55,9	7,39	36	Cytochrome P450 2C19 GN=CYP2C19 PE=1 SV=3 [CP2CJ]
Q93084	13	1043	113,9	5,57	36	Sarcoplasmic/endoplasmic reticulum calcium ATPase 3 (EC 3.6.3.8) (Calcium pump 3
Q11203	2	375	42,1	8,98	36	CMPNacetylneuraminatete1,4galactoside alpha2,3sialyltransferase GN=ST3GAL3
Q9NQ66	17	1216	138,5	6,23	35	1phosphatidylinositol4,5bisphosphate phosphodiesterase beta1 GN=PLCB1 PE=1 SV=1
P17544	53	494	52,9	8,65	35	Cyclic AMPdependent transcription factor ATF7 GN=ATF7 PE=1 SV=2 [ATF7]
Q92805	16	767	88,1	5,27	35	Golgin subfamily A member 1 GN=GOLGA1 PE=1 SV=3 [GOGA1]
Q5THK1	6	2151	237,2	6,33	35	Uncharacterized protein C22orf30 GN=C22orf30 PE=1 SV=1 [CV030]
P48553	173	1259	142,1	6,04	35	Trafficking protein particle complex subunit 10 GN=TRAPPC10 PE=1 SV=2 [TPC10]
P08069	1	1367	154,7	5,80	35	Insulinlike growth factor 1 receptor GN=IGF1R PE=1 SV=1 [IGF1R]
Q92674	10	756	86,7	8,76	35	Centromere protein I GN=CENPI PE=1 SV=2 [CENPI]
P16066	9	1061	118,8	6,64	35	Atrial natriuretic peptide receptor 1 GN=NPR1 PE=1 SV=1 [ANPRA]
Q01581	223	520	57,3	5,41	35	HydroxymethylglutarylCoA synthase, cytoplasmic, mitochondrial GN=PDHX PE=1 SV=3
O00330	1	501	54,1	8,66	35	Pyruvate dehydrogenase protein X component, mitochondrial GN=PDHX PE=1 SV=3
Q81VG5	6	1584	184,4	8,02	35	Sterile alpha motif domaincontaining protein 9like GN=SAMD9L PE=1 SV=2 [SAM9L
Q8NEM0	148	835	92,8	8,25	35	Microcephalin GN=MCPH1 PE=1 SV=3 [MCPH1]
Q5JPI3	311	329	37,5	6,47	35	Uncharacterized protein C3orf38 GN=C3orf38 PE=2 SV=1 [CC038]
Q9HC56	2	1237	136,0	5,47	35	Protocadherin9 GN=PCDH9 PE=1 SV=2 [PCDH9]
Q6ZTR7	4	304	34,8	8,66	35	Protein FAM92B GN=FAM92B PE=2 SV=1 [FA92B]
A7MCY6	2	615	67,7	5,86	35	TANKbinding kinase 1binding protein 1 GN=TBKBP1 PE=1 SV=1 [TBKB1]
Q08999	18	1139	128,3	7,44	35	Retinoblastomalike protein 2 GN=RBL2 PE=1 SV=3 [RBL2]
Q9H0I3	10	377	44,2	8,73	35	Coiledcoil domaincontaining protein 113 GN=CCDC113 PE=2 SV=1 [CC113]
P33908	21	653	72,9	6,47	35	Mannosyloligosaccharide 1,2alphamannosidase IA GN=MAN1A1 PE=1 SV=3 [MA1A
Q6ZRR7	2	1453	166,8	7,65	35	Leucinerich repeatcontaining protein 9 GN=LRR9 PE=2 SV=2 [LRR9]
Q8N323	12	547	63,1	8,56	35	Protein FAM55A GN=FAM55A PE=2 SV=2 [FA55A]
P14543	2	1247	136,3	5,29	35	Nidogen1 GN=NID1 PE=1 SV=3 [NID1]
P83105	3	476	50,9	8,02	35	Probable serine protease HTRA4 GN=HTRA4 PE=2 SV=1 [HTRA4]
Q9H307	163	717	81,6	7,14	35	Pinin GN=PNN PE=1 SV=4 [PININ]
O15440	2	1437	160,6	8,66	35	Multidrug resistanceassociated protein 5 GN=ABCC5 PE=1 SV=2 [MRP5]
Q8TCG2	56	481	54,7	5,97	35	Phosphatidylinositol 4kinase type 2beta GN=PI4K2B PE=1 SV=1 [P4K2B]
Q9V5P8	2	575	65,0	5,14	35	Serine/threonineprotein phosphatase 2A regulatory subunit B" subunit beta GN=P
Q7Z553	3	956	107,4	7,20	35	MAM domaincontaining glycosylphosphatidylinositol anchor protein 2 GN=MDGA2
Q9HAW4	8	1339	151,0	4,82	35	Claspin GN=CLSPN PE=1 SV=3 [CLSPN]
P61764	37	594	67,5	6,96	35	Syntaxinbinding protein 1 GN=STXB1 PE=1 SV=1 [STXB1]
O15068	239	1137	128,0	6,43	35	Guanine nucleotide exchange factor DBS GN=MCF2L PE=1 SV=2 [MCF2L]
Q9Y5B9	57	1047	119,8	5,66	35	FACT complex subunit SPT16 GN=SUPT16H PE=1 SV=1 [SP16H]
Q9ULH0	58	1771	196,4	6,62	35	Kinase Dinteracting substrate of 220 kDa GN=KIDINS220 PE=1 SV=3 [KDIS]
Q8IX95	2	158	18,0	4,92	35	Putative cutaneous Tcell lymphomaassociated antigen 3 GN=CTAGE3P PE=5 SV=1 [C
Q15008	7	389	45,5	5,62	35	26S proteasome nonATPase regulatory subunit 6 GN=PSMD6 PE=1 SV=1 [PSMD6]
Q9Y2Z2	2	717	79,9	8,31	35	Protein MTO1 homolog, mitochondrial GN=MTO1 PE=1 SV=2 [MTO1]
Q9H6W3	4	641	71,0	6,46	35	Lysinespecific demethylase NO66 GN=NO66 PE=1 SV=2 [NO66]
Q9H0I9	3	626	67,8	6,33	35	Transketolaselike protein 2 GN=TKTL2 PE=1 SV=1 [TKTL2]
Q96C10	14	678	76,6	7,37	34	Probable ATPdependent RNA helicase DHX58 GN=DHX58 PE=1 SV=1 [DHX58]
Q96ST2	7	819	91,9	4,69	34	Protein IWS1 homolog GN=IWS1 PE=1 SV=2 [IWS1]
O14595	8	271	30,6	5,53	34	Carboxyterminal domain RNA polymerase II polypeptide A small phosphatase 2 GN
Q9H3U7	18	446	49,6	8,46	34	SPARCrelated modular calciumbinding protein 2 GN=SMOC2 PE=2 SV=2 [SMOC2]
Q13042	6	620	71,6	5,85	34	Cell division cycle protein 16 homolog GN=CDC16 PE=1 SV=2 [CDC16]
P29475	5	1434	160,9	7,42	34	Nitric oxide synthase, brain GN=NOS1 PE=1 SV=2 [NOS1]
P51948	21	309	35,8	6,09	34	CDKactivating kinase assembly factor MAT1 GN=MNAT1 PE=1 SV=1 [MAT1]
Q15650	2	581	66,1	7,85	34	Activating signal cointegrator 1 GN=TRIP4 PE=1 SV=4 [TRIP4]
Q7Z5J8	3	1434	161,9	8,10	34	Ankyrin and armadillo repeatcontaining protein GN=ANKAR PE=2 SV=3 [ANKAR]
Q9H0A9	4	340	37,6	7,43	34	Uncharacterized protein C21orf56 GN=C21orf56 PE=2 SV=3 [CU056]
Q9GZY0	13	626	71,6	7,64	34	Nuclear RNA export factor 2 GN=NXF2 PE=1 SV=1 [NXF2]
Q8N556	23	730	80,7	8,68	34	Actin filamentassociated protein 1 GN=AFAP1 PE=1 SV=2 [AFAP1]
P78333	11	572	63,7	6,81	34	Glypican5 GN=GPC5 PE=1 SV=1 [GPC5]
Q6UWF7	10	544	62,2	8,85	34	Protein FAM55D GN=FAM55D PE=2 SV=1 [FA55D]
P42766	7	123	14,5	11,05	34	60S ribosomal protein L35 GN=RPL35 PE=1 SV=2 [RL35]
Q9H324	1	1103	120,8	7,94	34	A disintegrin and metalloproteinase with thrombospondin motifs 10 GN=ADAMTS1
Q16665	18	826	92,6	5,33	34	Hypoxiainducible factor 1alpha GN=HIF1A PE=1 SV=1 [HIF1A]
Q9H3G5	27	476	54,1	5,62	34	Probable serine carboxypeptidase CPVL GN=CPVL PE=1 SV=2 [CPVL]
Q6W4X9	1	2439	256,9	7,39	34	Mucin6 GN=MUC6 PE=1 SV=3 [MUC6]
Q9UPW6	211	733	82,5	6,90	34	DNAbinding protein SATB2 GN=SATB2 PE=1 SV=2 [SATB2]
Q9NRL3	1	753	80,5	5,40	34	Striatin4 GN=STRN4 PE=1 SV=2 [STRN4]
O75420	20	1035	114,5	5,39	34	PERQ amino acidrich with GYF domaincontaining protein 1 GN=GIGYF1 PE=1 SV=2 [I
P59046	8	1061	120,1	6,99	34	NACHT, LRR and PYD domainscontaining protein 12 GN=NLRP12 PE=2 SV=2 [NAL12]
Q9NRR4	13	1374	159,2	7,87	33	Ribonuclease 3 GN=DROSHA PE=1 SV=2 [RNC]

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Q08828	11	1119	123,4	8,43	33	Adenylate cyclase type 1 GN=ADCY1 PE=1 SV=2 [ADCY1]
Q96J6	17	628	65,8	6,81	33	Junctophilin4 GN=JPH4 PE=1 SV=2 [JPH4]
P35228	6	1153	131,0	7,96	33	Nitric oxide synthase, inducible GN=NOS2 PE=1 SV=2 [NOS2]
Q15262	9	1439	162,0	5,90	33	Receptortype tyrosineprotein phosphatase kappa GN=PTPRK PE=1 SV=2 [PTPRK]
O14490	101	977	108,8	7,08	33	Disks largeassociated protein 1 GN=DLGAP1 PE=1 SV=1 [DLGP1]
Q96QR8	4	312	33,2	5,43	33	Transcriptional activator protein Purbeta GN=PURB PE=1 SV=3 [PURB]
Q9HCM3	37	1950	210,6	6,11	33	UPF0606 protein KIAA1549 GN=KIAA1549 PE=1 SV=4 [K1549]
P29375	12	1690	192,0	6,49	33	Lysinespecific demethylase 5A GN=KDM5A PE=1 SV=3 [KDM5A]
Q8WXA9	24	508	59,3	10,39	33	Splicing regulatory glutamine/lysinerich protein 1 GN=SREK1 PE=1 SV=1 [SREK1]
O15117	2	783	85,3	6,48	33	FYNbinding protein GN=FYB PE=1 SV=2 [FYB]
P30954	11	320	35,9	9,07	33	Olfactory receptor 10J1 GN=OR10J1 PE=2 SV=2 [O10J1]
Q81WB9	9	1127	125,2	6,01	33	Testisexpressed sequence 2 protein GN=TEX2 PE=1 SV=2 [TEX2]
P20701	24	1170	128,7	5,63	33	Integrin alphaL GN=ITGAL PE=1 SV=3 [ITAL]
Q9NNZ3	2	241	27,6	10,55	33	Dnaj homolog subfamily C member 4 GN=DNAJC4 PE=2 SV=1 [DNJC4]
P42680	5	631	73,5	8,44	33	Tyrosineprotein kinase Tec GN=TEC PE=1 SV=2 [TEC]
Q9UBD0	6	423	46,7	7,11	33	Heat shock transcription factor, Xlinked GN=HSFX1 PE=1 SV=1 [HSFX1]
P00751	3	764	85,5	7,06	33	Complement factor B GN=CFB PE=1 SV=2 [CFAB]
Q494X3	2	552	65,4	9,14	33	Zinc finger protein 404 GN=ZNF404 PE=2 SV=1 [ZN404]
P11171	2	864	97,0	5,58	33	Protein 4.1 GN=EPB41 PE=1 SV=4 [41]
Q1X8D7	6	754	83,8	7,15	33	Leucinerich repeatcontaining protein 36 GN=LRR36 PE=2 SV=2 [LRC36]
Q9Y238	4	1755	195,6	6,33	32	Deleted in lung and esophageal cancer protein 1 GN=DLEC1 PE=2 SV=2 [DLEC1]
P22557	16	587	64,6	8,12	32	Saminolevulinate synthase, erythroidspecific, mitochondrial GN=ALAS2 PE=1 SV=2
Q96ER9	6	411	45,8	8,19	32	Coiledcoil domaincontaining protein 51 GN=CCDC51 PE=1 SV=2 [CCD51]
Q9GZQ6	4	430	47,8	9,41	32	Neuropeptide FF receptor 1 GN=NPFFR1 PE=1 SV=1 [NPFF1]
Q8WYP5	3	2266	252,3	6,60	32	Protein ELYS GN=AHCTF1 PE=1 SV=3 [ELYS]
Q9Y575	11	518	57,7	6,27	32	Ankyrin repeat and SOCS box protein 3 GN=ASB3 PE=1 SV=1 [ASB3]
Q16795	8	377	42,5	9,80	32	NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 9, mitochondrial p
Q10981	4	343	39,0	8,54	32	Galactoside 2alphafucosyltransferase 2 GN=FUT2 PE=2 SV=1 [FUT2]
P21580	62	790	89,6	8,22	32	Tumor necrosis factor alphainduced protein 3 GN=TNFAIP3 PE=1 SV=1 [TNAP3]
Q6WKZ4	5	1283	137,1	5,47	32	Rab11 familyinteracting protein 1 GN=RAB11FIP1 PE=1 SV=2 [RFIP1]
P49750	24	1951	219,8	6,57	32	YLP motifcontaining protein 1 GN=YLPM1 PE=1 SV=3 [YLPM1]
Q6NUQ1	22	792	90,6	5,45	32	RAD50interacting protein 1 GN=RINT1 PE=1 SV=1 [RINT1]
Q14191	3	1432	162,4	6,34	32	Werner syndrome ATPdependent helicase GN=WRN PE=1 SV=2 [WRN]
P17661	8	470	53,5	5,27	32	Desmin GN=DES PE=1 SV=3 [DESM]
O75146	4	1068	119,3	6,67	32	Huntingtininteracting protein 1related protein GN=HIP1R PE=1 SV=2 [HIP1R]
Q2WJG6	2	581	65,5	7,93	32	Kelchlike protein 38 GN=KLHL38 PE=2 SV=3 [KLH38]
O00294	7	542	60,6	9,47	32	Tubbyrelated protein 1 GN=TULP1 PE=1 SV=3 [TULP1]
O43148	29	476	54,8	6,61	32	mRNA cap guanineN7 methyltransferase GN=RNMT PE=1 SV=1 [MCES]
O76039	5	1030	115,5	9,54	32	Cyclindependent kinase5 GN=CDK5 PE=1 SV=1 [CDK5]
Q92974	23	986	111,5	7,27	32	Rho guanine nucleotide exchange factor 2 GN=ARHGEF2 PE=1 SV=4 [ARHG2]
Q9H0B3	3	1180	127,6	10,23	32	Uncharacterized protein KIAA1683 GN=KIAA1683 PE=1 SV=1 [K1683]
Q9Y2G1	108	1151	124,3	7,44	32	Myelin gene regulatory factor GN=MRF PE=1 SV=3 [MRF]
Q99714	12	261	26,9	7,78	32	3hydroxyacylCoA dehydrogenase type2 GN=HSD17B10 PE=1 SV=3 [HCD2]
O43283	7	966	108,2	6,49	32	Mitogenactivated protein kinase kinase kinase 13 GN=MAP3K13 PE=1 SV=1 [M3K13]
Q9H3R5	3	247	28,5	5,29	32	Centromere protein H (CENPH) (Interphase centromere complex protein 35) [CEN
Q9UJC3	11	728	84,6	5,15	32	Protein Hook homolog 1 GN=HOOK1 PE=1 SV=2 [HOOK1]
Q96T51	2	708	79,8	5,74	32	RUN and FYVE domaincontaining protein 1 GN=RUFY1 PE=1 SV=2 [RUFY1]
Q8N998	3	374	43,8	5,36	31	Coiledcoil domaincontaining protein 89 GN=CCDC89 PE=2 SV=1 [CCD89]
O60244	3	1454	160,5	8,73	31	Mediator of RNA polymerase II transcription subunit 14 GN=MED14 PE=1 SV=2 [MEI
Q99988	1	308	34,1	9,66	31	Growth/differentiation factor 15 GN=GDF15 PE=1 SV=3 [GDF15]
Q55RN2	4	563	61,6	9,25	31	Uncharacterized protein C6orf10 GN=C6orf10 PE=1 SV=3 [CF010]
P49760	3	499	60,1	9,66	31	Dual specificity protein kinase CLK2 GN=CLK2 PE=1 SV=1 [CLK2]
Q5FYB1	5	569	64,0	8,62	31	Arylsulfatase I GN=ARSI PE=1 SV=1 [ARSI]
Q96G42	6	594	63,3	9,00	31	Kelch domaincontaining protein 7B GN=KLHDC7B PE=2 SV=2 [KLD7B]
Q99729	6	332	36,2	8,21	31	Heterogeneous nuclear ribonucleoprotein A/B GN=HNRNPAB PE=1 SV=2 [ROAA]
Q9HCE9	17	1232	135,9	5,82	31	Anoctamin8 GN=ANO8 PE=2 SV=3 [ANO8]
Q5T7V8	2	394	45,0	7,08	31	RAB6interacting golgin GN=GORAB PE=1 SV=1 [GORAB]
P17213	3	487	53,9	9,38	31	Bactericidal permeabilityincreasing protein GN=BPI PE=1 SV=4 [BPI]
Q9UNN5	17	650	73,9	4,88	31	FASassociated factor 1 GN=FAF1 PE=1 SV=2 [FAF1]
Q9Y3E1	67	203	22,6	7,99	31	Hepatomaderived growth factorrelated protein 3 GN=HDGFRP3 PE=1 SV=1 [HDGR3]
A4FU01	4	709	79,5	7,03	31	Myotubularinrelated protein 11 GN=MTMR11 PE=2 SV=2 [MTMRB]
Q05195	24	221	25,2	8,53	31	Max dimerization protein 1 GN=MXD1 PE=1 SV=1 [MAD1]
Q494V2	198	611	71,1	7,11	30	Coiledcoil domaincontaining protein 37 GN=CCDC37 PE=1 SV=1 [CCD37]
P19823	5	946	106,4	6,86	30	Interalphatrypsin inhibitor heavy chain H2 GN=ITIH2 PE=1 SV=2 [ITIH2]
O43913	8	435	50,3	7,74	30	Origin recognition complex subunit 5 GN=ORC5 PE=1 SV=1 [ORC5]

Appendices

Appendix 4: LC ESI MS identified NHC-IOBA cell proteins.

Accession	#Peptides	# AAs	MW [kDa]	calc. pI	Score	Description
P07900	1111	732	84,6	5,02	17236	Heat shock protein HSP 90-alpha GN=HSP90AA1 PE=1 SV=5
A2Q0Z1	874	646	70,9	5,52	16909	Heat shock cognate 71 kDa protein GN=HSPA8 PE=2 SV=1
P08238	1102	724	83,2	5,03	16858	Heat shock protein HSP 90-beta GN=HSP90AB1 PE=1 SV=4
P60712	1063	375	41,7	5,48	15447	Actin, cytoplasmic 1 GN=ACTB PE=1 SV=1
P06733	904	434	47,1	7,39	15161	Alpha-enolase GN=ENO1 PE=1 SV=2
P14618	809	531	57,9	7,84	14932	Pyruvate kinase isozymes M1/M2 GN=PKM2 PE=1 SV=4
Q2KJDO	635	444	49,6	4,89	13736	Tubulin beta-5 chain GN=TUBB5 PE=2 SV=1
P31327	1057	1500	164,8	6,74	13385	Carbamoyl-phosphate synthase [ammonia], mitochondrial GN=CPS1 PE=1 SV=2
P11021	662	654	72,3	5,16	12923	78 kDa glucose-regulated protein GN=HSPA5 PE=1 SV=2
Q3MHM5	606	445	49,8	4,89	12793	Tubulin beta-4B chain GN=TUBB4B PE=2 SV=1
P07355	760	339	38,6	7,75	12539	Annexin A2 GN=ANXA2 PE=1 SV=2
P21333	1072	2647	280,6	6,06	12012	Filamin-A GN=FLNA PE=1 SV=4
P68103	898	462	50,1	9,01	11679	Elongation factor 1-alpha 1 GN=EEF1A1 PE=2 SV=1
P04406	695	335	36,0	8,46	11605	Glyceraldehyde-3-phosphate dehydrogenase GN=GAPDH PE=1 SV=3
Q3ZC07	823	377	42,0	5,39	11333	Actin, alpha cardiac muscle 1 GN=ACTC1 PE=2 SV=1
P62739	758	377	42,0	5,39	10443	Actin, aortic smooth muscle GN=ACTA2 PE=1 SV=1
Q13885	478	445	49,9	4,89	10178	Tubulin beta-2A chain GN=TUBB2A PE=1 SV=1
Q2T9S0	450	450	50,4	4,93	10153	Tubulin beta-3 chain GN=TUBB3 PE=2 SV=1
P81947	701	451	50,1	5,06	9968	Tubulin alpha-1B chain PE=1 SV=2
P63258	626	375	41,8	5,48	9606	Actin, cytoplasmic 2 GN=ACTG1 PE=1 SV=1
Q32PH8	650	463	50,4	9,03	9574	Elongation factor 1-alpha 2 GN=EEF1A2 PE=2 SV=1
Q06830	780	199	22,1	8,13	9544	Peroxisomal protein 1 GN=PRDX1 PE=1 SV=1
O75369	847	2602	278,0	5,73	9474	Filamin-B GN=FLNB PE=1 SV=2
P08107	500	641	70,0	5,66	9053	Heat shock 70 kDa protein 1A/1B GN=HSPA1A PE=1 SV=5
P04264	650	644	66,0	8,12	8972	Keratin, type II cytoskeletal 1 GN=KRT1 PE=1 SV=6
A6NMY6	567	339	38,6	6,95	8312	Putative annexin A2-like protein GN=ANXA2P2 PE=5 SV=2
P68362	648	451	50,1	5,06	8271	Tubulin alpha-1A chain OS=Cricetulus griseus GN=TUBA1A PE=2 SV=1
P54652	448	639	70,0	5,74	7852	Heat shock-related 70 kDa protein 2 GN=HSPA2 PE=1 SV=1
P00558	564	417	44,6	8,10	7648	Phosphoglycerate kinase 1 GN=PGK1 PE=1 SV=3
Q3ZBU7	429	444	49,6	4,88	7498	Tubulin beta-4A chain GN=TUBB4A PE=2 SV=1
P38646	386	679	73,6	6,16	7254	Stress-70 protein, mitochondrial GN=HSPA9 PE=1 SV=2
P81948	513	448	49,9	5,06	6852	Tubulin alpha-4A chain GN=TUBA4A PE=1 SV=2
P08670	412	466	53,6	5,12	6846	Vimentin GN=VIM PE=1 SV=4
P04083	282	346	38,7	7,02	6596	Annexin A1 GN=ANXA1 PE=1 SV=2
Q09666	1140	5890	628,7	6,15	6396	Neuroblast differentiation-associated protein AHNAK GN=AHNAK PE=1 SV=2
P22314	383	1058	117,8	5,76	6353	Ubiquitin-like modifier-activating enzyme 1 GN=UBA1 PE=1 SV=3
Q658J3	456	1075	121,3	6,20	6331	POTE ankyrin domain family member E GN=POTEE PE=1 SV=3
P34931	306	641	70,3	6,02	6284	Heat shock 70 kDa protein 1-like GN=HSPA1L PE=1 SV=2
ASA3E0	425	1075	121,4	6,20	6266	POTE ankyrin domain family member F GN=POTEF PE=1 SV=2
P07195	446	334	36,6	6,05	6193	L-lactate dehydrogenase B chain GN=LDHB PE=1 SV=2
P60174	293	286	30,8	5,92	5602	Triosephosphate isomerase GN=TPI1 PE=1 SV=3
P07737	264	140	15,0	8,27	5594	Profilin-1 GN=PFN1 PE=1 SV=2
P10809	405	573	61,0	5,87	5582	60 kDa heat shock protein, mitochondrial GN=HSPD1 PE=1 SV=2
Q6B856	219	445	49,9	4,89	5580	Tubulin beta-2B chain GN=TUBB2B PE=1 SV=2
P06744	331	558	63,1	8,32	5578	Glucose-6-phosphate isomerase GN=GPI PE=1 SV=4
P06576	269	529	56,5	5,40	5405	ATP synthase subunit beta, mitochondrial GN=ATP5B PE=1 SV=3
P08758	338	320	35,9	5,05	5291	Annexin A5 GN=ANXA5 PE=1 SV=2
P09211	179	210	23,3	5,64	5286	Glutathione S-transferase P GN=GSTP1 PE=1 SV=2
P46940	411	1657	189,1	6,48	5207	Ras GTPase-activating-like protein IQGAP1 GN=IQGAP1 PE=1 SV=1
P00338	439	332	36,7	8,27	5121	L-lactate dehydrogenase A chain GN=LDHA PE=1 SV=2
Q562R1	468	376	42,0	5,59	5088	Beta-actin-like protein 2 GN=ACTBL2 PE=1 SV=2
P17066	261	643	71,0	6,14	5043	Heat shock 70 kDa protein 6 GN=HSPA6 PE=1 SV=2
Q13748	460	450	49,9	5,10	5037	Tubulin alpha-3C/D chain GN=TUBA3C PE=1 SV=3
P32119	384	198	21,9	5,97	4946	Peroxisomal protein 2 GN=PRDX2 PE=1 SV=5
Q6PEY2	405	450	49,8	5,14	4876	Tubulin alpha-3E chain GN=TUBA3E PE=1 SV=2
P37802	260	199	22,4	8,25	4871	Transgelin-2 GN=TAGLN2 PE=1 SV=3
P62938	500	165	18,0	7,81	4726	Peptidyl-prolyl cis-trans isomerase A OS=Chlorocebus aethiops GN=PPIA PE=2 SV=2
P13639	409	858	95,3	6,83	4618	Elongation factor 2 GN=EEF2 PE=1 SV=4
P13645	319	584	58,8	5,21	4601	Keratin, type I cytoskeletal 10 GN=KRT10 PE=1 SV=6
Q58FF7	428	597	68,3	4,79	4411	Putative heat shock protein HSP 90-beta-3 GN=HSP90AB3P PE=5 SV=1
Q5E9B5	335	376	41,8	5,48	4399	Actin, gamma-enteric smooth muscle GN=ACTG2 PE=2 SV=1
P14625	353	803	92,4	4,84	4209	Endoplasmic reticulum chaperone GN=HSP90B1 PE=1 SV=1
Q9BUI5	271	446	49,8	4,88	4182	Tubulin beta-6 chain GN=TUBB6 PE=1 SV=1
Q9BYX7	256	375	42,0	6,33	4143	Putative beta-actin-like protein 3 GN=POTEKP PE=5 SV=1

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P23284	397	216	23,7	9,41	4139	Peptidyl-prolyl cis-trans isomerase B GN=PPIB PE=1 SV=2
Q9NY65	310	449	50,1	5,06	4114	Tubulin alpha-8 chain GN=TUBA8 PE=1 SV=1
P09104	163	434	47,2	5,03	4082	Gamma-enolase GN=ENO2 PE=1 SV=3
POCG38	387	1075	121,2	6,21	3941	POTE ankyrin domain family member I GN=POTEI PE=3 SV=1
Q58FF8	315	381	44,3	4,84	3906	Putative heat shock protein HSP 90-beta 2 GN=HSP90AB2P PE=1 SV=2
Q3SZ54	176	406	46,1	5,48	3837	Eukaryotic initiation factor 4A-I GN=EIF4A1 PE=2 SV=1
P62259	315	255	29,2	4,74	3577	14-3-3 protein epsilon GN=Ywhae PE=1 SV=1
Q12931	144	704	80,1	8,21	3460	Heat shock protein 75 kDa, mitochondrial GN=TRAP1 PE=1 SV=3
P49327	296	2511	273,3	6,44	3405	Fatty acid synthase GN=FASN PE=1 SV=3
Q00610	299	1675	191,5	5,69	3382	Clathrin heavy chain 1 GN=CLTC PE=1 SV=5
P07205	221	417	44,8	8,54	3317	Phosphoglycerate kinase 2 GN=PGK2 PE=1 SV=3
Q13162	305	271	30,5	6,29	3295	Peroxiredoxin-4 GN=PRDX4 PE=1 SV=1
Q14568	252	343	39,3	4,65	3240	Putative heat shock protein HSP 90-alpha A2 GN=HSP90AA2 PE=1 SV=2
P13929	112	434	46,9	7,71	3236	Beta-enolase GN=ENO3 PE=1 SV=4
P04075	276	364	39,4	8,09	3229	Fructose-bisphosphate aldolase A GN=ALDOA PE=1 SV=2
POCG39	328	1038	117,3	5,97	3152	POTE ankyrin domain family member J GN=POTEJ PE=3 SV=1
Q3ZCM7	237	444	49,7	4,89	3109	Tubulin beta-8 chain GN=TUBB8 PE=1 SV=2
Q3SZ65	129	407	46,4	5,48	3011	Eukaryotic initiation factor 4A-II GN=EIF4A2 PE=2 SV=1
P61603	209	102	10,9	8,92	2974	10 kDa heat shock protein, mitochondrial GN=HSPE1 PE=3 SV=2
P48741	136	367	40,2	7,87	2873	Putative heat shock 70 kDa protein 7 GN=HSPA7 PE=5 SV=2
P35908	228	639	65,4	8,00	2835	Keratin, type II cytoskeletal 2 epidermal GN=KRT2 PE=1 SV=2
P23528	233	166	18,5	8,09	2800	Cofilin-1 GN=CFL1 PE=1 SV=3
Q58FG1	129	418	47,7	5,19	2763	Putative heat shock protein HSP 90-alpha A4 GN=HSP90AA4P PE=5 SV=1
P26038	237	577	67,8	6,40	2740	Moesin GN=MSN PE=1 SV=3
P31946	304	246	28,1	4,83	2721	14-3-3 protein beta/alpha GN=YWHAB PE=1 SV=3
Q01105	194	290	33,5	4,32	2713	Protein SET GN=SET PE=1 SV=3
P30086	110	187	21,0	7,53	2673	Phosphatidylethanolamine-binding protein 1 GN=PEBP1 PE=1 SV=3
A6NNZ2	215	444	49,5	4,86	2663	Tubulin beta-8 chain B PE=1 SV=1
P63103	239	245	27,7	4,79	2645	14-3-3 protein zeta/delta GN=YWHAZ PE=1 SV=1
P30041	271	224	25,0	6,38	2630	Peroxiredoxin-6 GN=PRDX6 PE=1 SV=3
P29401	346	623	67,8	7,66	2602	Transketolase GN=TKT PE=1 SV=3
P40926	102	338	35,5	8,68	2571	Malate dehydrogenase, mitochondrial GN=MDH2 PE=1 SV=3
P27797	184	417	48,1	4,44	2541	Calreticulin GN=CALR PE=1 SV=1
P05141	194	298	32,8	9,69	2463	ADP/ATP translocase 2 GN=SLC25A5 PE=1 SV=7
P05387	104	115	11,7	4,54	2352	60S acidic ribosomal protein P2 GN=RPLP2 PE=1 SV=1
P50990	155	548	59,6	5,60	2339	T-complex protein 1 subunit theta GN=CCT8 PE=1 SV=4
Q99867	181	434	48,4	5,26	2329	Putative tubulin beta-4q chain GN=TUBB4Q PE=5 SV=1
P15531	241	152	17,1	6,19	2312	Nucleoside diphosphate kinase A GN=NME1 PE=1 SV=1
P27824	124	592	67,5	4,60	2272	Calnexin GN=CANX PE=1 SV=2
P25705	167	553	59,7	9,13	2257	ATP synthase subunit alpha, mitochondrial GN=ATP5A1 PE=1 SV=1
P16152	76	277	30,4	8,32	2237	Carbonyl reductase [NADPH] 1 GN=CBR1 PE=1 SV=3
P62984	211	128	14,7	9,83	2235	Ubiquitin-60S ribosomal protein L40 GN=Uba52 PE=1 SV=2
P08195	244	630	68,0	5,01	2226	4F2 cell-surface antigen heavy chain GN=SLC3A2 PE=1 SV=3
P10599	125	105	11,7	4,92	2225	Thioredoxin GN=TXN PE=1 SV=3
Q3SZ14	225	245	27,7	4,78	2147	14-3-3 protein theta GN=YWHAQ PE=2 SV=1
P29692	80	281	31,1	5,01	2144	Elongation factor 1-delta GN=EEF1D PE=1 SV=5
P78371	90	535	57,5	6,46	2144	T-complex protein 1 subunit beta GN=CCT2 PE=1 SV=4
P19338	221	710	76,6	4,70	2108	Nucleolin GN=NCL PE=1 SV=3
P26641	195	437	50,1	6,67	2076	Elongation factor 1-gamma GN=EEF1G PE=1 SV=3
Q148F1	129	166	18,7	7,88	2048	Cofilin-2 GN=CFL2 PE=2 SV=1
Q72794	100	578	61,9	5,99	1998	Keratin, type II cytoskeletal 1b GN=KRT77 PE=1 SV=3
P12956	154	609	69,8	6,64	1997	X-ray repair cross-complementing protein 6 GN=XRCC6 PE=1 SV=2
P30044	139	214	22,1	8,70	1995	Peroxiredoxin-5, mitochondrial GN=PRDX5 PE=1 SV=4
P55060	198	971	110,3	5,77	1993	Exportin-2 GN=CSE1L PE=1 SV=3
P12236	158	298	32,8	9,74	1991	ADP/ATP translocase 3 GN=SLC25A6 PE=1 SV=4
P24534	93	225	24,7	4,67	1980	Elongation factor 1-beta GN=EEF1B2 PE=1 SV=3
P12277	152	381	42,6	5,59	1978	Creatine kinase B-type GN=CKB PE=1 SV=1
P30101	164	505	56,7	6,35	1978	Protein disulfide-isomerase A3 GN=PDIA3 PE=1 SV=4
P49368	121	545	60,5	6,49	1962	T-complex protein 1 subunit gamma GN=CCT3 PE=1 SV=4
P31948	259	543	62,6	6,80	1959	Stress-induced-phosphoprotein 1 GN=STIP1 PE=1 SV=1
P67936	196	248	28,5	4,69	1921	Tropomyosin alpha-4 chain GN=TPM4 PE=1 SV=3
P13797	199	630	70,8	5,60	1910	Plastin-3 GN=PLS3 PE=1 SV=4
P02786	153	760	84,8	6,61	1908	Transferrin receptor protein 1 GN=TFRC PE=1 SV=2
Q9H853	82	241	27,5	7,83	1888	Putative tubulin-like protein alpha-4B GN=TUBA4B PE=5 SV=2
Q9H4B7	160	451	50,3	5,17	1883	Tubulin beta-1 chain GN=TUBB1 PE=1 SV=1

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P22392	242	152	17,3	8,41	1851	Nucleoside diphosphate kinase B GN=NME2 PE=1 SV=1
P35241	138	583	68,5	6,37	1850	Radixin GN=RDY PE=1 SV=1
Q14974	111	876	97,1	4,78	1838	Importin subunit beta-1 GN=KPNB1 PE=1 SV=2
P18669	161	254	28,8	7,18	1825	Phosphoglycerate mutase 1 GN=PGAM1 PE=1 SV=2
P78527	237	4128	468,8	7,12	1811	DNA-dependent protein kinase catalytic subunit GN=PRKDC PE=1 SV=3
P50395	124	445	50,6	6,47	1808	Rab GDP dissociation inhibitor beta GN=GDI2 PE=1 SV=2
P61982	221	247	28,3	4,89	1786	14-3-3 protein gamma GN=Ywhag PE=1 SV=2
P61205	141	181	20,6	7,43	1781	ADP-ribosylation factor 3 GN=Arf3 PE=2 SV=2
P84078	133	181	20,7	6,80	1780	ADP-ribosylation factor 1 GN=Arf1 PE=1 SV=2
P15311	140	586	69,4	6,27	1741	Ezrin GN=EZR PE=1 SV=4
P22234	111	425	47,0	7,23	1718	Multifunctional protein ADE2 GN=PAICS PE=1 SV=3
P52209	64	483	53,1	7,23	1715	6-phosphogluconate dehydrogenase, decarboxylating GN=PGD PE=1 SV=3
Q99832	88	543	59,3	7,65	1689	T-complex protein 1 subunit eta GN=CCT7 PE=1 SV=2
Q58FG0	150	334	38,7	6,57	1684	Putative heat shock protein HSP 90-alpha A5 GN=HSP90AA5P PE=1 SV=1
P05023	104	1023	112,8	5,49	1666	Sodium/potassium-transporting ATPase subunit alpha-1 GN=ATP1A1 PE=1 SV=1
P30048	93	256	27,7	7,78	1652	Thioredoxin-dependent peroxide reductase, mitochondrial GN=PRDX3 PE=1 SV=3
Q14204	219	4646	532,1	6,40	1651	Cytoplasmic dynein 1 heavy chain 1 GN=DYNC1H1 PE=1 SV=5
P07237	225	508	57,1	4,87	1629	Protein disulfide-isomerase GN=P4HB PE=1 SV=3
P12429	95	323	36,4	5,92	1622	Annexin A3 GN=ANXA3 PE=1 SV=3
P23526	116	432	47,7	6,34	1617	Adenosylhomocysteinase GN=AHCY PE=1 SV=4
P62827	180	216	24,4	7,49	1584	GTP-binding nuclear protein Ran GN=Ran PE=1 SV=3
P60661	105	151	16,9	4,65	1571	Myosin light polypeptide 6 GN=MYL6 PE=2 SV=2
O14818	94	248	27,9	8,46	1528	Proteasome subunit alpha type-7 GN=PSMA7 PE=1 SV=1
P31947	212	248	27,8	4,74	1518	14-3-3 protein sigma GN=SFN PE=1 SV=1
P53675	102	1640	186,9	5,85	1493	Clathrin heavy chain 2 GN=CLTCL1 PE=1 SV=2
P00441	112	154	15,9	6,13	1492	Superoxide dismutase [Cu-Zn] GN=SOD1 PE=1 SV=2
P02538	127	564	60,0	8,00	1481	Keratin, type II cytoskeletal 6A GN=KRT6A PE=1 SV=3
P08865	71	295	32,8	4,87	1454	40S ribosomal protein SA GN=RPSA PE=1 SV=4
O14737	54	125	14,3	6,04	1444	Programmed cell death protein 5 GN=PDCD5 PE=1 SV=3
O75828	37	277	30,8	6,18	1431	Carbonyl reductase [NADPH] 3 GN=CBR3 PE=1 SV=3
P05388	92	317	34,3	5,97	1415	60S acidic ribosomal protein P0 GN=RPLP0 PE=1 SV=1
Q07021	68	282	31,3	4,84	1406	Complement component 1 Q subcomponent-binding protein, mitochondrial GN=C1C
P63242	135	154	16,8	5,24	1400	Eukaryotic translation initiation factor 5A-1 GN=Elf5a PE=1 SV=2
P04259	122	564	60,0	8,00	1396	Keratin, type II cytoskeletal 6B GN=KRT6B PE=1 SV=5
Q370C7	169	149	17,3	5,97	1380	Stathmin GN=STMN1 PE=2 SV=3
P12235	106	298	33,0	9,76	1374	ADP/ATP translocase 1 GN=SLC25A4 PE=1 SV=4
Q99714	99	261	26,9	7,78	1366	3-hydroxyacyl-CoA dehydrogenase type-2 GN=HSD17B10 PE=1 SV=3
Q15084	67	440	48,1	5,08	1360	Protein disulfide-isomerase A6 GN=PDIA6 PE=1 SV=1
P26639	146	723	83,4	6,67	1352	Threonine--tRNA ligase, cytoplasmic GN=TARS PE=1 SV=3
P61979	106	463	50,9	5,54	1343	Heterogeneous nuclear ribonucleoprotein K GN=Hnrnpk PE=1 SV=1
P36952	115	375	42,1	6,05	1343	Serpin B5 GN=SERPINB5 PE=1 SV=2
P18206	172	1134	123,7	5,66	1327	Vinculin GN=VCL PE=1 SV=4
O60361	154	137	15,5	8,57	1318	Putative nucleoside diphosphate kinase GN=NME2P1 PE=5 SV=1
O43707	118	911	104,8	5,44	1315	Alpha-actinin-4 GN=ACTN4 PE=1 SV=2
Q3T169	105	243	26,7	9,66	1315	40S ribosomal protein S3 GN=RPS3 PE=2 SV=1
P30040	105	261	29,0	7,31	1314	Endoplasmic reticulum resident protein 29 GN=ERP29 PE=1 SV=4
P61284	88	165	17,8	9,42	1299	60S ribosomal protein L12 GN=RPL12 PE=2 SV=1
Q8BGY2	107	153	16,8	5,58	1295	Eukaryotic translation initiation factor 5A-2 GN=Elf5a2 PE=2 SV=3
Q13765	30	215	23,4	4,56	1291	Nascent polypeptide-associated complex subunit alpha GN=NACA PE=1 SV=1
Q61S14	104	154	16,8	5,00	1284	Eukaryotic translation initiation factor 5A-1-like GN=EIF5A1 PE=1 SV=2
Q9CXW4	82	178	20,2	9,60	1284	60S ribosomal protein L11 GN=Rpl11 PE=1 SV=4
Q8NHW5	75	317	34,3	5,55	1276	60S acidic ribosomal protein P0-like GN=RPLP0P6 PE=5 SV=1
P23921	74	792	90,0	7,15	1247	Ribonucleoside-diphosphate reductase large subunit GN=RRM1 PE=1 SV=1
P07741	100	180	19,6	6,02	1223	Adenine phosphoribosyltransferase GN=APRT PE=1 SV=2
O00299	71	241	26,9	5,17	1217	Chloride intracellular channel protein 1 GN=CLIC1 PE=1 SV=4
Q04917	182	246	28,2	4,84	1202	14-3-3 protein eta GN=YWHAH PE=1 SV=4
Q58FF6	168	505	58,2	4,73	1198	Putative heat shock protein HSP 90-beta 4 GN=HSP90AB4P PE=5 SV=1
P04843	116	607	68,5	6,38	1195	Dolichyl-diphosphooligosaccharide--protein glycosyltransferase subunit 1 GN=RPN1
P09382	100	135	14,7	5,50	1191	Galectin-1 GN=LGALS1 PE=1 SV=2
P62992	118	156	18,0	9,64	1191	Ubiquitin-40S ribosomal protein S27a GN=RPS27A PE=1 SV=2
P30613	115	574	61,8	7,74	1182	Pyruvate kinase isozymes R/L GN=PKLR PE=1 SV=2
P62204	71	149	16,8	4,22	1180	Calmodulin GN=Calm1 PE=1 SV=2
Q92688	97	251	28,8	4,06	1174	Acidic leucine-rich nuclear phosphoprotein 32 family member B GN=ANP32B PE=1 SV
P06748	136	294	32,6	4,78	1172	Nucleophosmin GN=NPM1 PE=1 SV=2
P11586	125	935	101,5	7,30	1166	C-1-tetrahydrofolate synthase, cytoplasmic GN=MTHFD1 PE=1 SV=3

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P31939	109	592	64,6	6,71	1152	Bifunctional purine biosynthesis protein PURH GN=ATIC PE=1 SV=3
O35129	114	299	33,3	9,83	1146	Prohibitin-2 GN=Phb2 PE=1 SV=1
P05787	94	483	53,7	5,59	1130	Keratin, type II cytoskeletal 8 GN=KRT8 PE=1 SV=7
O95373	105	1038	119,4	4,82	1129	Importin-7 GN=IPO7 PE=1 SV=1
Q14697	172	944	106,8	6,14	1126	Neutral alpha-glucosidase AB GN=GANAB PE=1 SV=3
Q9Z2U1	106	241	26,4	4,79	1119	Proteasome subunit alpha type-5 GN=Pma5 PE=1 SV=1
P14131	90	146	16,4	10,21	1112	40S ribosomal protein S16 GN=Rps16 PE=2 SV=4
P48643	98	541	59,6	5,66	1110	T-complex protein 1 subunit epsilon GN=CCT5 PE=1 SV=1
P07910	65	306	33,6	5,08	1110	Heterogeneous nuclear ribonucleoproteins C1/C2 GN=HNRNPC PE=1 SV=4
P34932	93	840	94,3	5,19	1108	Heat shock 70 kDa protein 4 GN=HSPA4 PE=1 SV=4
O60506	72	623	69,6	8,59	1106	Heterogeneous nuclear ribonucleoprotein Q GN=SYNCRIP PE=1 SV=2
Q14315	116	2725	290,8	5,97	1103	Filamin-C GN=FLNC PE=1 SV=3
P07951	116	284	32,8	4,70	1098	Tropomyosin beta chain GN=TPM2 PE=1 SV=1
P35579	119	1960	226,4	5,60	1094	Myosin-9 GN=MYH9 PE=1 SV=4
P17987	123	556	60,3	6,11	1085	T-complex protein 1 subunit alpha GN=TCP1 PE=1 SV=1
P13647	102	590	62,3	7,74	1083	Keratin, type II cytoskeletal 5 GN=KRT5 PE=1 SV=3
A6NL28	50	223	26,3	4,51	1080	Putative tropomyosin alpha-3 chain-like protein PE=5 SV=2
P31949	94	105	11,7	7,12	1071	Protein S100-A11 GN=S100A11 PE=1 SV=2
P04080	104	98	11,1	7,56	1056	Cystatin-B GN=CSTB PE=1 SV=2
P62821	94	205	22,7	6,21	1047	Ras-related protein Rab-1A GN=Rab1A PE=1 SV=3
P12268	72	514	55,8	6,90	1046	Inosine-5'-monophosphate dehydrogenase 2 GN=IMPDH2 PE=1 SV=2
Q9H0C2	46	315	35,0	9,89	1046	ADP/ATP translocase 4 GN=SLC25A31 PE=1 SV=1
P52565	51	204	23,2	5,11	1039	Rho GDP-dissociation inhibitor 1 GN=ARHGDI1 PE=1 SV=3
Q9Y2T3	78	454	51,0	5,68	1035	Guanine deaminase GN=GDA PE=1 SV=1
P49748	84	655	70,3	8,75	1016	Very long-chain specific acyl-CoA dehydrogenase, mitochondrial GN=ACADVL PE=1 SV=1
P55072	130	806	89,3	5,26	1002	Transitional endoplasmic reticulum ATPase GN=VCP PE=1 SV=4
P06703	175	90	10,2	5,48	988	Protein S100-A6 GN=S100A6 PE=1 SV=1
P53999	102	127	14,4	9,60	980	Activated RNA polymerase II transcriptional coactivator p15 GN=SUB1 PE=1 SV=3
P61007	79	207	23,7	9,07	980	Ras-related protein Rab-8A OS=Canis familiaris GN=RAB8A PE=2 SV=1
P27105	51	288	31,7	7,88	979	Erythrocyte band 7 integral membrane protein GN=STOM PE=1 SV=3
P50993	44	1020	112,2	5,66	978	Sodium/potassium-transporting ATPase subunit alpha-2 GN=ATP1A2 PE=1 SV=1
P13637	41	1013	111,7	5,38	975	Sodium/potassium-transporting ATPase subunit alpha-3 GN=ATP1A3 PE=1 SV=3
Q8TAA3	55	256	28,5	8,98	971	Proteasome subunit alpha type-7-like GN=PSMA8 PE=1 SV=3
P60900	52	246	27,4	6,76	971	Proteasome subunit alpha type-6 GN=PSMA6 PE=1 SV=1
Q5KR47	113	284	32,8	4,72	971	Tropomyosin alpha-3 chain GN=TPM3 PE=2 SV=1
P14314	56	528	59,4	4,41	963	Glucosidase 2 subunit beta GN=PRKCSH PE=1 SV=2
P12814	75	892	103,0	5,41	942	Alpha-actinin-1 GN=ACTN1 PE=1 SV=2
P25815	61	95	10,4	4,88	927	Protein S100-P GN=S100P PE=1 SV=2
Q32PD5	106	145	16,1	10,32	926	40S ribosomal protein S19 GN=RPS19 PE=2 SV=3
P00505	32	430	47,5	9,01	908	Aspartate aminotransferase, mitochondrial GN=GOT2 PE=1 SV=3
Q3ZCF3	53	163	18,6	4,54	904	S-phase kinase-associated protein 1 GN=SKP1 PE=2 SV=1
P18124	82	248	29,2	10,65	899	60S ribosomal protein L7 GN=RPL7 PE=1 SV=1
P40227	83	531	58,0	6,68	882	T-complex protein 1 subunit zeta GN=CCT6A PE=1 SV=3
Q15293	27	331	38,9	5,00	881	Reticulocalbin-1 GN=RCN1 PE=1 SV=1
P12004	76	261	28,8	4,69	881	Proliferating cell nuclear antigen GN=PCNA PE=1 SV=1
P09493	91	284	32,7	4,74	879	Tropomyosin alpha-1 chain GN=TPM1 PE=1 SV=2
P02533	122	472	51,5	5,16	878	Keratin, type I cytoskeletal 14 GN=KRT14 PE=1 SV=4
Q3T0F4	74	165	18,9	10,15	867	40S ribosomal protein S10 GN=RPS10 PE=2 SV=1
O43175	72	533	56,6	6,71	866	D-3-phosphoglycerate dehydrogenase GN=PHGDH PE=1 SV=4
P35268	56	128	14,8	9,19	855	60S ribosomal protein L22 GN=RPL22 PE=1 SV=2
P15259	99	253	28,7	8,88	852	Phosphoglycerate mutase 2 GN=PGAM2 PE=1 SV=3
P50991	91	539	57,9	7,83	850	T-complex protein 1 subunit delta GN=CCT4 PE=1 SV=4
Q9H0U4	82	201	22,2	5,73	850	Ras-related protein Rab-1B GN=RAB1B PE=1 SV=1
P51150	59	207	23,5	6,70	844	Ras-related protein Rab-7a GN=Rab7a PE=1 SV=2
Q5E958	56	208	24,2	10,32	832	40S ribosomal protein S8 GN=RPS8 PE=2 SV=3
O95678	72	551	59,5	7,74	831	Keratin, type II cytoskeletal 75 GN=KRT75 PE=1 SV=2
P33316	51	252	26,5	9,36	830	Deoxyuridine 5'-triphosphate nucleotidohydrolase, mitochondrial GN=DUT PE=1 SV=1
Q3T0F7	27	118	12,9	5,52	829	Myotrophin GN=MTPN PE=1 SV=3
P61286	104	636	70,6	9,50	829	Polyadenylate-binding protein 1 GN=PABPC1 PE=2 SV=1
P48668	63	564	60,0	8,00	817	Keratin, type II cytoskeletal 6C GN=KRT6C PE=1 SV=3
O00410	97	1097	123,5	4,94	815	Importin-5 GN=IPO5 PE=1 SV=4
P49736	69	904	101,8	5,52	808	DNA replication licensing factor MCM2 GN=MCM2 PE=1 SV=4
Q08211	94	1270	140,9	6,84	807	ATP-dependent RNA helicase A GN=DHX9 PE=1 SV=4
P68040	57	317	35,1	7,69	799	Guanine nucleotide-binding protein subunit beta-2-like 1 GN=Gnb2l1 PE=1 SV=3
P05783	52	430	48,0	5,45	788	Keratin, type I cytoskeletal 18 GN=KRT18 PE=1 SV=2

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P28074	46	263	28,5	6,92	788	Proteasome subunit beta type-5 GN=PSMB5 PE=1 SV=3
Q04837	34	148	17,2	9,60	772	Single-stranded DNA-binding protein, mitochondrial GN=SSBP1 PE=1 SV=1
P61027	56	200	22,5	8,38	767	Ras-related protein Rab-10 GN=Rab10 PE=1 SV=1
P00491	61	289	32,1	6,95	762	Purine nucleoside phosphorylase GN=PNP PE=1 SV=2
Q969H8	68	173	18,8	6,68	758	UPF0556 protein C19orf10 GN=C19orf10 PE=1 SV=1
Q07020	49	188	21,6	11,72	756	60S ribosomal protein L18 GN=RPL18 PE=1 SV=2
Q91V41	54	215	23,9	6,21	744	Ras-related protein Rab-14 GN=Rab14 PE=1 SV=3
O43776	61	548	62,9	6,25	740	Asparagine--tRNA ligase, cytoplasmic GN=NARS PE=1 SV=1
Q56K14	32	114	11,5	4,32	739	60S acidic ribosomal protein P1 GN=RPLP1 PE=3 SV=1
P78417	99	241	27,5	6,60	738	Glutathione S-transferase omega-1 GN=GSTO1 PE=1 SV=2
P38919	42	411	46,8	6,73	738	Eukaryotic initiation factor 4A-III GN=EIF4A3 PE=1 SV=4
P62264	45	151	16,3	10,05	737	40S ribosomal protein S14 GN=Rps14 PE=2 SV=3
Q92930	61	207	23,6	9,07	724	Ras-related protein Rab-8B GN=RAB8B PE=1 SV=2
Q02790	96	459	51,8	5,43	721	Peptidyl-prolyl cis-trans isomerase FKBP4 GN=FKBP4 PE=1 SV=3
P59190	53	212	24,4	5,71	719	Ras-related protein Rab-15 GN=RAB15 PE=1 SV=1
Q6PHN9	57	201	23,0	8,29	719	Ras-related protein Rab-35 GN=Rab35 PE=1 SV=1
Q5XKE5	64	535	57,8	7,20	716	Keratin, type II cytoskeletal 79 GN=KRT79 PE=1 SV=2
P08729	63	469	51,4	5,48	706	Keratin, type II cytoskeletal 7 GN=KRT7 PE=1 SV=5
Q3T165	57	272	29,8	5,76	703	Prohibitin GN=PHB PE=2 SV=1
P62852	70	125	13,7	10,11	703	40S ribosomal protein S25 GN=Rps25 PE=2 SV=1
P62702	87	263	29,6	10,15	703	40S ribosomal protein S4, X isoform GN=Rps4x PE=2 SV=2
P13693	41	172	19,6	4,93	698	Translationally-controlled tumor protein GN=TPT1 PE=1 SV=1
P16442	355	354	40,9	8,98	698	Histo-blood group ABO system transferase GN=ABO PE=1 SV=2
Q9NPC6	350	264	29,9	8,25	698	Myozenin-2 GN=MYOZ2 PE=1 SV=1
Q6DD88	39	541	60,5	5,66	688	Atlantin-3 GN=ATL3 PE=1 SV=1
P30049	23	168	17,5	5,49	681	ATP synthase subunit delta, mitochondrial GN=ATP5D PE=1 SV=2
P53396	89	1101	120,8	7,33	677	ATP-citrate synthase GN=ACLY PE=1 SV=3
P18085	77	180	20,5	7,14	673	ADP-ribosylation factor 4 GN=ARF4 PE=1 SV=3
P60902	89	97	11,2	7,37	673	Protein S100-A10 GN=S100A10 PE=1 SV=2
O43399	45	206	22,2	5,36	670	Tumor protein D54 GN=TPD52L2 PE=1 SV=2
P52292	39	529	57,8	5,40	669	Importin subunit alpha-2 GN=KPNA2 PE=1 SV=1
P56192	41	900	101,1	6,16	665	Methionine--tRNA ligase, cytoplasmic GN=MARS PE=1 SV=2
P62270	93	152	17,7	10,99	663	40S ribosomal protein S18 GN=Rps18 PE=1 SV=3
P30046	68	118	12,7	7,30	660	D-dopachrome decarboxylase GN=DDT PE=1 SV=3
P42704	86	1394	157,8	6,13	654	Leucine-rich PPR motif-containing protein, mitochondrial GN=LRPPRC PE=1 SV=3
Q60972	16	425	47,6	4,89	654	Histone-binding protein RBBP4 GN=Rbbp4 PE=1 SV=5
P00492	45	218	24,6	6,68	652	Hypoxanthine-guanine phosphoribosyltransferase GN=HPRT1 PE=1 SV=2
Q6PDM2	67	248	27,7	10,36	650	Serine/arginine-rich splicing factor 1 GN=Srsf1 PE=1 SV=3
P39687	75	249	28,6	4,09	649	Acidic leucine-rich nuclear phosphoprotein 32 family member A GN=ANP32A PE=1 SV=1
P37108	29	136	14,6	10,04	649	Signal recognition particle 14 kDa protein GN=SRP14 PE=1 SV=2
P07814	91	1512	170,5	7,33	644	Bifunctional glutamate/proline--tRNA ligase GN=EPRS PE=1 SV=5
Q92928	54	201	22,0	5,43	643	Putative Ras-related protein Rab-1C GN=RAB1C PE=5 SV=2
P21796	46	283	30,8	8,54	636	Voltage-dependent anion-selective channel protein 1 GN=VDAC1 PE=1 SV=2
Q99497	116	189	19,9	6,79	633	Protein DJ-1 GN=PARK7 PE=1 SV=2
P08779	96	473	51,2	5,05	633	Keratin, type I cytoskeletal 16 GN=KRT16 PE=1 SV=4
Q96FW1	25	271	31,3	4,94	632	Ubiquitin thioesterase OTUB1 GN=OTUB1 PE=1 SV=2
P84084	77	180	20,5	6,79	630	ADP-ribosylation factor 5 GN=Arf5 PE=2 SV=2
Q02878	58	288	32,7	10,58	620	60S ribosomal protein L6 GN=RPL6 PE=1 SV=3
P22102	48	1010	107,7	6,70	614	Trifunctional purine biosynthetic protein adenosine-3 GN=GART PE=1 SV=1
P61087	33	200	22,4	5,44	613	Ubiquitin-conjugating enzyme E2 K GN=Ube2k PE=1 SV=3
O14980	64	1071	123,3	6,06	612	Exportin-1 GN=XPO1 PE=1 SV=1
Q9UJK9	36	219	24,3	4,94	609	ADP-sugar pyrophosphatase GN=NUDT5 PE=1 SV=1
P17931	56	250	26,1	8,56	608	Galectin-3 GN=LGALS3 PE=1 SV=5
O14684	65	152	17,1	9,50	603	Prostaglandin E synthase GN=PTGES PE=1 SV=2
Q99733	41	375	42,8	4,69	597	Nucleosome assembly protein 1-like 4 GN=NAP1L4 PE=1 SV=1
Q9BSJ8	51	1104	122,8	5,83	595	Extended synaptotagmin-1 GN=ESYT1 PE=1 SV=1
Q9Y266	44	331	38,2	5,38	594	Nuclear migration protein nudC GN=NUDC PE=1 SV=1
P46777	52	297	34,3	9,72	585	60S ribosomal protein L5 GN=RPL5 PE=1 SV=3
P49755	36	219	25,0	7,44	581	Transmembrane emp24 domain-containing protein 10 GN=TMED10 PE=1 SV=2
P54819	48	239	26,5	7,81	574	Adenylate kinase 2, mitochondrial GN=AK2 PE=1 SV=2
P55854	29	103	11,6	5,49	568	Small ubiquitin-related modifier 3 GN=SUMO3 PE=1 SV=2
P57088	21	247	28,0	9,70	566	Transmembrane protein 33 GN=TMEM33 PE=1 SV=2
P62245	53	130	14,8	10,13	562	40S ribosomal protein S15a GN=Rps15a PE=2 SV=2
P42126	23	302	32,8	8,54	561	Enoyl-CoA delta isomerase 1, mitochondrial GN=EC11 PE=1 SV=1
P61018	27	213	23,6	6,06	557	Ras-related protein Rab-4B GN=RAB4B PE=1 SV=1

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P61294	29	208	23,4	5,53	557	Ras-related protein Rab-6B GN=Rab6b PE=1 SV=1
Q9H082	31	229	25,7	7,18	557	Ras-related protein Rab-33B GN=RAB33B PE=1 SV=1
Q8N0Y7	72	254	28,8	6,65	553	Probable phosphoglycerate mutase 4 GN=PGAM4 PE=1 SV=1
Q99829	62	537	59,0	5,83	549	Copine-1 GN=CPNE1 PE=1 SV=1
Q9Y230	23	463	51,1	5,64	546	RuvB-like 2 GN=RUVBL2 PE=1 SV=3
P61290	50	254	29,5	5,95	545	Proteasome activator complex subunit 3 GN=Psmc3 PE=1 SV=1
P49411	30	452	49,5	7,61	544	Elongation factor Tu, mitochondrial GN=TUFM PE=1 SV=2
P55209	34	391	45,3	4,46	538	Nucleosome assembly protein 1-like 1 GN=NAP1L1 PE=1 SV=1
P67809	21	324	35,9	9,88	538	Nuclease-sensitive element-binding protein 1 GN=YBX1 PE=1 SV=3
Q01650	33	507	55,0	7,72	535	Large neutral amino acids transporter small subunit 1 GN=SLC7A5 PE=1 SV=2
P14174	14	115	12,5	7,88	534	Macrophage migration inhibitory factor GN=MIF PE=1 SV=4
A7MBJ5	70	1230	136,3	5,78	532	Cullin-associated NEDD8-dissociated protein 1 GN=CAND1 PE=2 SV=1
P62751	52	156	17,7	10,45	531	60S ribosomal protein L23a GN=Rpl23a PE=1 SV=1
P55821	92	179	20,8	8,32	529	Stathmin-2 GN=Stmn2 PE=1 SV=1
Q13263	48	835	88,5	5,77	527	Transcription intermediary factor 1-beta GN=TRIM28 PE=1 SV=5
Q13011	60	328	35,8	8,00	526	Delta(3,5)-Delta(2,4)-dienoyl-CoA isomerase, mitochondrial GN=ECH1 PE=1 SV=2
P62301	322	151	17,2	10,54	523	40S ribosomal protein S13 GN=Rps13 PE=1 SV=2
B0CM99	93	215	24,9	5,74	522	High mobility group protein B1 OS=Callithrix jacchus GN=HMGB1 PE=3 SV=1
Q9BZK3	14	213	23,3	4,59	522	Putative nascent polypeptide-associated complex subunit alpha-like protein GN=NA
P37837	47	337	37,5	6,81	521	Transaldolase GN=TALDO1 PE=1 SV=2
P26640	38	1264	140,4	7,59	521	Valine--tRNA ligase GN=VARS PE=1 SV=4
P47755	26	286	32,9	5,85	515	F-actin-capping protein subunit alpha-2 GN=CAPZA2 PE=1 SV=3
P08559	30	390	43,3	8,06	514	Pyruvate dehydrogenase E1 component subunit alpha, somatic form, mitochondrial (
P80723	52	227	22,7	4,63	513	Brain acid soluble protein 1 GN=BASP1 PE=1 SV=2
Q14651	54	629	70,2	5,41	511	Plastin-1 GN=PLS1 PE=1 SV=2
P62858	27	69	7,8	10,70	511	40S ribosomal protein S28 GN=Rps28 PE=2 SV=1
P48047	40	213	23,3	9,96	509	ATP synthase subunit O, mitochondrial GN=ATP5O PE=1 SV=1
P27695	31	318	35,5	8,12	508	DNA-(apurinic or apyrimidinic site) lyase GN=APEX1 PE=1 SV=2
P40925	52	334	36,4	7,36	508	Malate dehydrogenase, cytoplasmic GN=MDH1 PE=1 SV=4
Q9NQ39	20	176	20,1	10,13	505	Putative 40S ribosomal protein S10-like GN=RPS10P5 PE=5 SV=1
A6H769	65	194	22,1	10,10	505	40S ribosomal protein S7 GN=RPS7 PE=2 SV=1
Q15758	48	541	56,6	5,48	504	Neutral amino acid transporter B(0) GN=SLC1A5 PE=1 SV=2
Q3T0Y5	30	234	25,9	7,43	502	Proteasome subunit alpha type-2 GN=PSMA2 PE=1 SV=3
P00966	59	412	46,5	8,02	502	Argininosuccinate synthase GN=ASS1 PE=1 SV=2
Q5E9E6	63	217	24,8	9,94	498	60S ribosomal protein L10a GN=RPL10A PE=2 SV=3
P62900	50	125	14,5	10,54	495	60S ribosomal protein L31 GN=Rpl31 PE=2 SV=1
P13010	68	732	82,7	5,81	491	X-ray repair cross-complementing protein 5 GN=XRCC5 PE=1 SV=3
P00568	42	194	21,6	8,63	490	Adenylate kinase isoenzyme 1 GN=AK1 PE=1 SV=3
A6NHG4	50	134	14,2	6,29	481	D-dopachrome decarboxylase-like protein GN=DDTL PE=2 SV=1
Q7Z3Y7	67	464	50,5	5,47	480	Keratin, type I cytoskeletal 28 GN=KRT28 PE=1 SV=2
P14649	23	208	22,7	5,73	477	Myosin light chain 6B GN=MYL6B PE=1 SV=1
Q96QK1	38	796	91,6	5,49	476	Vacuolar protein sorting-associated protein 35 GN=VPS35 PE=1 SV=2
P52597	44	415	45,6	5,58	467	Heterogeneous nuclear ribonucleoprotein F GN=HNRNPF PE=1 SV=3
P49721	41	201	22,8	7,02	462	Proteasome subunit beta type-2 GN=PSMB2 PE=1 SV=1
Q13310	33	644	70,7	9,26	462	Polyadenylate-binding protein 4 GN=PABPC4 PE=1 SV=1
Q14103	32	355	38,4	7,81	461	Heterogeneous nuclear ribonucleoprotein D0 GN=HNRNPD PE=1 SV=1
P05455	25	408	46,8	7,12	461	Lupus La protein GN=SSB PE=1 SV=2
Q75964	34	103	11,4	9,64	461	ATP synthase subunit g, mitochondrial GN=ATP5L PE=1 SV=3
A6NHL2	76	446	49,9	6,05	453	Tubulin alpha chain-like 3 GN=TUBAL3 PE=1 SV=2
P30153	73	589	65,3	5,11	453	Serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A alpha isoform
Q15181	60	289	32,6	5,86	451	Inorganic pyrophosphatase GN=PPA1 PE=1 SV=2
P20674	47	150	16,8	6,79	450	Cytochrome c oxidase subunit 5A, mitochondrial GN=COX5A PE=1 SV=2
O75396	17	215	24,6	6,92	449	Vesicle-trafficking protein SEC22b GN=SEC22B PE=1 SV=4
Q6ZWN5	93	194	22,6	10,65	449	40S ribosomal protein S9 GN=Rps9 PE=2 SV=3
P35609	30	894	103,8	5,45	446	Alpha-actinin-2 GN=ACTN2 PE=1 SV=1
P61956	26	95	10,8	5,50	445	Small ubiquitin-related modifier 2 GN=SUMO2 PE=1 SV=2
P24752	19	427	45,2	8,85	444	Acetyl-CoA acetyltransferase, mitochondrial GN=ACAT1 PE=1 SV=1
O00231	28	422	47,4	6,48	444	26S proteasome non-ATPase regulatory subunit 11 GN=PSMD11 PE=1 SV=3
P61105	33	212	23,5	6,54	443	Ras-related protein Rab-2A OS=Canis familiaris GN=RAB2A PE=1 SV=1
O75340	37	191	21,9	5,40	438	Programmed cell death protein 6 GN=PDCD6 PE=1 SV=1
Q7Z3Z0	52	450	49,3	5,08	437	Keratin, type I cytoskeletal 25 GN=KRT25 PE=1 SV=1
P63010	25	937	104,5	5,38	436	AP-2 complex subunit beta GN=AP2B1 PE=1 SV=1
Q10567	22	949	104,6	5,06	434	AP-1 complex subunit beta-1 GN=AP1B1 PE=1 SV=2
Q9BYC2	30	517	56,1	7,14	432	Succinyl-CoA:3-ketoacid coenzyme A transferase 2, mitochondrial GN=OXCT2 PE=2 SV
Q01813	31	784	85,5	7,55	432	6-phosphofructokinase type C GN=PFKP PE=1 SV=2

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P51148	38	216	23,5	8,41	431	Ras-related protein Rab-5C GN=RAB5C PE=1 SV=2
Q6WUX8	43	105	11,7	9,57	429	Cytochrome c OS=Gorilla gorilla gorilla GN=CYCS PE=3 SV=3
O43598	17	174	19,1	5,05	429	Deoxyribonucleoside 5'-monophosphate N-glycosidase GN=RCL PE=1 SV=1
Q9NR45	33	359	40,3	6,74	427	Sialic acid synthase GN=NANS PE=1 SV=2
P47897	44	775	87,7	7,15	425	Glutamine--tRNA ligase GN=QARS PE=1 SV=1
P29803	17	388	42,9	8,46	424	Pyruvate dehydrogenase E1 component subunit alpha, testis-specific form, mitochondrion
Q32P51	30	320	34,2	9,00	422	Heterogeneous nuclear ribonucleoprotein A1-like 2 GN=HNRNPA1L2 PE=2 SV=2
Q3TOE7	21	330	37,5	6,33	422	Serine/threonine-protein phosphatase PP1-alpha catalytic subunit GN=PPP1CA PE=2
Q9H3K6	21	86	10,1	6,52	419	Bola-like protein 2 GN=BOLA2 PE=1 SV=1
P07954	29	510	54,6	8,76	419	Fumarate hydratase, mitochondrial GN=FB PE=1 SV=3
P31040	38	664	72,6	7,39	419	Succinate dehydrogenase [ubiquinone] flavoprotein subunit, mitochondrial GN=SDH
P11177	26	359	39,2	6,65	417	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial GN=PDHB PE=1
P16989	12	372	40,1	9,77	414	DNA-binding protein A GN=CSDA PE=1 SV=4
Q9Y2T7	10	364	38,5	10,80	414	Y-box-binding protein 2 GN=YBX2 PE=1 SV=2
Q9NQP4	17	134	15,3	4,53	414	Prefoldin subunit 4 GN=PFDN4 PE=1 SV=1
O43143	29	795	90,9	7,46	414	Putative pre-mRNA-splicing factor ATP-dependent RNA helicase DHX15 GN=DHX15 P
P50213	18	366	39,6	6,92	414	Isocitrate dehydrogenase [NAD] subunit alpha, mitochondrial GN=IDH3A PE=1 SV=1
Q9BVC6	29	243	26,2	10,48	411	Transmembrane protein 109 GN=TMEM109 PE=1 SV=1
O43242	36	534	60,9	8,44	411	26S proteasome non-ATPase regulatory subunit 3 GN=PSMD3 PE=1 SV=2
P33991	46	863	96,5	6,74	407	DNA replication licensing factor MCM4 GN=MCM4 PE=1 SV=5
P55884	38	814	92,4	5,00	406	Eukaryotic translation initiation factor 3 subunit B GN=EIF3B PE=1 SV=3
Q16658	14	493	54,5	7,24	406	Fascin GN=FSCN1 PE=1 SV=3
P60867	38	119	13,4	9,94	406	40S ribosomal protein S20 GN=Rps20 PE=1 SV=1
Q15561	64	434	48,3	7,33	402	Transcriptional enhancer factor TEF-3 GN=TEAD4 PE=1 SV=3
Q08043	38	901	103,2	5,52	400	Alpha-actinin-3 GN=ACTN3 PE=1 SV=2
P49720	35	205	22,9	6,55	397	Proteasome subunit beta type-3 GN=PSMB3 PE=1 SV=2
P60766	40	191	21,2	6,55	397	Cell division control protein 42 homolog GN=Cdc42 PE=1 SV=2
P35749	34	1972	227,2	5,50	397	Myosin-11 GN=MYH11 PE=1 SV=3
Q99584	46	98	11,5	6,16	396	Protein S100-A13 GN=S100A13 PE=1 SV=1
Q9CXY6	14	390	43,0	5,26	395	Interleukin enhancer-binding factor 2 GN=Ilf2 PE=1 SV=1
P31943	38	449	49,2	6,30	394	Heterogeneous nuclear ribonucleoprotein H GN=HNRNPH1 PE=1 SV=4
P54709	18	279	31,5	8,35	392	Sodium/potassium-transporting ATPase subunit beta-3 GN=ATP1B3 PE=1 SV=1
P62492	40	216	24,4	6,57	389	Ras-related protein Rab-11A GN=Rab11a PE=1 SV=3
O60812	25	293	32,1	5,06	389	Heterogeneous nuclear ribonucleoprotein C-like 1 GN=HNRNPCL1 PE=1 SV=1
P17661	83	470	53,5	5,27	388	Desmin GN=DES PE=1 SV=3
Q13620	33	913	103,9	7,37	387	Cullin-4B GN=CUL4B PE=1 SV=4
P62424	38	266	30,0	10,61	386	60S ribosomal protein L7a GN=RPL7A PE=1 SV=2
O60888	21	179	19,1	5,50	385	Protein CutA GN=CUTA PE=1 SV=2
P36578	67	427	47,7	11,06	384	60S ribosomal protein L4 GN=RPL4 PE=1 SV=5
Q9Y265	29	456	50,2	6,42	382	RuvB-like 1 GN=RUVBL1 PE=1 SV=1
P25685	40	340	38,0	8,63	382	DnaJ homolog subfamily B member 1 GN=DNAJB1 PE=1 SV=4
Q9Y490	72	2541	269,6	6,07	379	Talin-1 GN=TLN1 PE=1 SV=3
Q13185	19	183	20,8	5,33	378	Chromobox protein homolog 3 GN=CBX3 PE=1 SV=4
Q9H361	61	631	70,0	9,67	375	Polyadenylate-binding protein 3 GN=PABPC3 PE=1 SV=2
Q9UNM6	19	376	42,9	5,81	375	26S proteasome non-ATPase regulatory subunit 13 GN=PSMD13 PE=1 SV=2
Q9Y262	37	564	66,7	6,34	374	Eukaryotic translation initiation factor 3 subunit L GN=EIF3L PE=1 SV=1
P14927	21	111	13,5	8,78	373	Cytochrome b-c1 complex subunit 7 GN=UQCRCB PE=1 SV=2
P13798	11	732	81,2	5,48	372	Acylamino-acid-releasing enzyme GN=APEH PE=1 SV=4
P23381	8	471	53,1	6,23	370	Tryptophan--tRNA ligase, cytoplasmic GN=WARS PE=1 SV=2
P38117	63	255	27,8	8,10	368	Electron transfer flavoprotein subunit beta GN=ETFB PE=1 SV=3
P50914	21	215	23,4	10,93	366	60S ribosomal protein L14 GN=RPL14 PE=1 SV=4
P13073	44	169	19,6	9,51	366	Cytochrome c oxidase subunit 4 isoform 1, mitochondrial GN=COX4I1 PE=1 SV=1
Q00688	37	224	25,2	9,28	365	Peptidyl-prolyl cis-trans isomerase FKBP3 GN=FKBP3 PE=1 SV=1
Q3THE2	15	172	19,8	4,84	364	Myosin regulatory light chain 12B GN=Myl12b PE=1 SV=2
Q04760	35	184	20,8	5,31	359	Lactoylglutathione lyase GN=GLO1 PE=1 SV=4
Q9BY22	21	381	41,9	8,65	358	L-lactate dehydrogenase A-like 6B GN=LDHAL6B PE=1 SV=3
Q9H254	49	2564	288,8	6,01	358	Spectrin beta chain, brain 3 GN=SPTBN4 PE=1 SV=2
Q96TA1	40	746	84,1	6,19	357	Niban-like protein 1 GN=FAM129B PE=1 SV=3
P62849	17	133	15,4	10,78	356	40S ribosomal protein S24 GN=Rps24 PE=1 SV=1
B2RPKO	43	211	24,2	6,21	356	Putative high mobility group protein B1-like 1 GN=HMGB1P1 PE=5 SV=1
Q9UI30	16	125	14,2	5,26	355	tRNA methyltransferase 112 homolog GN=TRMT112 PE=1 SV=1
Q06323	23	249	28,7	6,02	355	Proteasome activator complex subunit 1 GN=PSME1 PE=1 SV=1
P17655	34	700	79,9	4,98	354	Calpain-2 catalytic subunit GN=CAPN2 PE=1 SV=6
Q9NZ01	38	308	36,0	9,45	351	Trans-2,3-enoyl-CoA reductase GN=TECR PE=1 SV=1
P28347	32	426	47,9	8,15	351	Transcriptional enhancer factor TEF-1 GN=TEAD1 PE=1 SV=2

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Q76I81	49	132	14,5	7,21	350	40S ribosomal protein S12 GN=RPS12 PE=2 SV=1
P13804	28	333	35,1	8,38	349	Electron transfer flavoprotein subunit alpha, mitochondrial GN=ETFA PE=1 SV=1
Q13283	25	466	52,1	5,52	349	Ras GTPase-activating protein-binding protein 1 GN=G3BP1 PE=1 SV=1
P32969	46	192	21,8	9,95	347	60S ribosomal protein L9 GN=RPL9 PE=1 SV=1
Q04695	70	432	48,1	5,02	345	Keratin, type I cytoskeletal 17 GN=KRT17 PE=1 SV=2
Q9UI15	10	199	22,5	7,33	345	Transgelin-3 GN=TAGLN3 PE=1 SV=2
O75874	16	414	46,6	7,01	343	Isocitrate dehydrogenase [NADP] cytoplasmic GN=IDH1 PE=1 SV=2
Q8BMJ3	44	144	16,5	5,24	343	Eukaryotic translation initiation factor 1A, X-chromosomal GN=EIF1ax PE=2 SV=3
Q16719	30	465	52,3	7,03	338	Kynureninase GN=KYNU PE=1 SV=1
P35237	21	376	42,6	5,27	338	Serpin B6 GN=SERPINB6 PE=1 SV=3
P06454	18	111	12,2	3,78	337	Prothymosin alpha GN=PTMA PE=1 SV=2
Q02413	29	1049	113,7	5,03	336	Desmoglein-1 GN=DSG1 PE=1 SV=2
Q09019	31	674	70,4	7,24	336	Dystrophia myotonica WD repeat-containing protein GN=DMWD PE=1 SV=3
Q04941	29	152	16,7	7,24	335	Proteolipid protein 2 GN=PLP2 PE=1 SV=1
P10620	10	155	17,6	9,39	333	Microsomal glutathione S-transferase 1 GN=MGST1 PE=1 SV=1
P19012	62	456	49,2	4,77	333	Keratin, type I cytoskeletal 15 GN=KRT15 PE=1 SV=3
Q9V536	112	164	18,2	9,22	332	Peptidyl-prolyl cis-trans isomerase A-like 4A/B/C GN=PPIAL4A PE=1 SV=1
P35527	85	623	62,0	5,24	331	Keratin, type I cytoskeletal 9 GN=KRT9 PE=1 SV=3
P36871	19	562	61,4	6,76	331	Phosphoglucomutase-1 GN=PGM1 PE=1 SV=3
P51571	17	173	19,0	6,15	331	Translocon-associated protein subunit delta GN=SSR4 PE=1 SV=1
P61222	34	599	67,3	8,34	330	ATP-binding cassette sub-family E member 1 GN=Abce1 PE=2 SV=1
O95831	36	613	66,9	8,95	327	Apoptosis-inducing factor 1, mitochondrial GN=AIFM1 PE=1 SV=1
Q58FF3	47	399	45,8	5,26	326	Putative endoplasmin-like protein GN=HSP90B2P PE=5 SV=1
P30084	42	290	31,4	8,07	326	Enoyl-CoA hydratase, mitochondrial GN=ECHS1 PE=1 SV=4
P09874	24	1014	113,0	8,88	326	Poly [ADP-ribose] polymerase 1 GN=PARP1 PE=1 SV=4
P25788	47	255	28,4	5,33	325	Proteasome subunit alpha type-3 GN=PSMA3 PE=1 SV=2
Q0P5D0	85	177	19,2	8,07	323	Peptidyl-prolyl cis-trans isomerase H GN=PPIH PE=2 SV=1
P40429	61	203	23,6	10,93	322	60S ribosomal protein L13a GN=RPL13A PE=1 SV=2
P27708	69	2225	242,8	6,46	321	CAD protein GN=CAD PE=1 SV=3
P51858	34	240	26,8	4,73	321	Hepatoma-derived growth factor GN=HDGF PE=1 SV=1
Q13451	21	457	51,2	5,90	320	Peptidyl-prolyl cis-trans isomerase FKBP5 GN=FKBP5 PE=1 SV=2
P62267	20	143	15,8	10,49	320	40S ribosomal protein S23 GN=Rps23 PE=2 SV=3
P45877	76	212	22,7	8,40	319	Peptidyl-prolyl cis-trans isomerase C GN=PPIC PE=1 SV=1
O95747	12	527	58,0	6,43	318	Serine/threonine-protein kinase OSR1 GN=OSR1 PE=1 SV=1
P20648	18	1035	114,0	5,81	318	Potassium-transporting ATPase alpha chain 1 GN=ATP4A PE=2 SV=5
P13796	39	627	70,2	5,43	317	Plastin-2 GN=LCP1 PE=1 SV=6
Q5VT79	16	327	36,8	5,63	314	Annexin A8-like protein 2 GN=ANXA8L2 PE=2 SV=1
Q92973	51	898	102,3	4,98	313	Transportin-1 GN=TNPO1 PE=1 SV=2
O00116	17	658	72,9	7,34	311	Alkylidihydroxyacetonephosphate synthase, peroxisomal GN=AGPS PE=1 SV=1
Q9NTK5	10	396	44,7	7,81	310	Obg-like ATPase 1 GN=OLA1 PE=1 SV=2
Q9BQ67	8	446	49,4	4,92	310	Glutamate-rich WD repeat-containing protein 1 GN=GRWD1 PE=1 SV=1
Q9UQ80	34	394	43,8	6,55	309	Proliferation-associated protein 2G4 GN=PA2G4 PE=1 SV=3
Q96A49	12	352	39,9	4,53	309	Synapse-associated protein 1 GN=SYAP1 PE=1 SV=1
P15880	76	293	31,3	10,24	308	40S ribosomal protein S2 GN=RPS2 PE=1 SV=2
P13928	23	327	36,9	5,78	308	Annexin A8 GN=ANXA8 PE=1 SV=3
P49589	14	748	85,4	6,76	306	Cysteine--tRNA ligase, cytoplasmic GN=CARS PE=1 SV=3
P05198	28	315	36,1	5,08	306	Eukaryotic translation initiation factor 2 subunit 1 GN=EIF2S1 PE=1 SV=3
Q15645	20	432	48,5	6,09	306	Pachytene checkpoint protein 2 homolog GN=TRIP13 PE=1 SV=2
Q15366	25	365	38,6	6,79	306	Poly(rC)-binding protein 2 GN=PCBP2 PE=1 SV=1
Q72406	35	1995	227,7	5,60	303	Myosin-14 GN=MYH14 PE=1 SV=2
Q00325	37	362	40,1	9,38	301	Phosphate carrier protein, mitochondrial GN=SLC25A3 PE=1 SV=2
P41252	59	1262	144,4	6,15	301	Isoleucine--tRNA ligase, cytoplasmic GN=IARS PE=1 SV=2
Q14964	19	217	25,0	7,65	300	Ras-related protein Rab-39A GN=RAB39A PE=2 SV=2
P51153	25	203	22,8	9,19	300	Ras-related protein Rab-13 GN=RAB13 PE=1 SV=1
Q6EEV6	17	95	10,7	7,18	298	Small ubiquitin-related modifier 4 GN=SUMO4 PE=1 SV=2
O60763	19	962	107,8	4,91	298	General vesicular transport factor p115 GN=USO1 PE=1 SV=2
P17174	32	413	46,2	7,01	296	Aspartate aminotransferase, cytoplasmic GN=GOT1 PE=1 SV=3
Q14126	37	1118	122,2	5,24	295	Desmoglein-2 GN=DSG2 PE=1 SV=2
Q9BRA2	30	123	13,9	5,52	292	Thioredoxin domain-containing protein 17 GN=TXNDC17 PE=1 SV=1
Q6NZI2	11	390	43,4	5,60	291	Polymerase I and transcript release factor GN=PTRF PE=1 SV=1
O00429	20	736	81,8	6,81	288	Dynamin-1-like protein GN=DNM1L PE=1 SV=2
Q96AE4	18	644	67,5	7,61	288	Far upstream element-binding protein 1 GN=FUBP1 PE=1 SV=3
Q5E9D5	32	165	18,5	7,85	288	Destrin GN=DSTN PE=2 SV=3
P20962	24	102	11,5	4,16	287	Parathymosin GN=PTMS PE=1 SV=2
P09972	58	364	39,4	6,87	287	Fructose-bisphosphate aldolase C GN=ALDOC PE=1 SV=2

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P25786	27	263	29,5	6,61	286	Proteasome subunit alpha type-1 GN=PSMA1 PE=1 SV=1
Q8NCW5	29	288	31,7	7,66	286	Apolipoprotein A-I-binding protein GN=APOA1BP PE=1 SV=2
Q9CZM2	27	204	24,1	11,62	283	60S ribosomal protein L15 GN=Rpl15 PE=2 SV=4
Q3T0W9	18	196	23,5	11,47	281	60S ribosomal protein L19 GN=RPL19 PE=2 SV=1
Q14116	32	193	22,3	4,67	280	Interleukin-18 GN=IL18 PE=1 SV=1
O14595	25	271	30,6	5,53	280	Carboxy-terminal domain RNA polymerase II polypeptide A small phosphatase 2 GN=
P63221	17	83	9,1	8,50	279	40S ribosomal protein S21 OS=Sus scrofa GN=RPS21 PE=3 SV=1
P24666	21	158	18,0	6,74	279	Low molecular weight phosphotyrosine protein phosphatase GN=ACP1 PE=1 SV=3
P43487	21	201	23,3	5,29	278	Ran-specific GTPase-activating protein GN=RANBP1 PE=1 SV=1
P38606	16	617	68,3	5,52	277	V-type proton ATPase catalytic subunit A GN=ATP6V1A PE=1 SV=2
O00159	28	1063	121,6	9,41	277	Unconventional myosin-Ic GN=MYO1C PE=1 SV=4
Q3T035	15	178	20,5	8,59	276	Actin-related protein 2/3 complex subunit 3 GN=ARPC3 PE=1 SV=3
P02794	15	183	21,2	5,55	275	Ferritin heavy chain GN=FTH1 PE=1 SV=2
P61358	33	136	15,8	10,56	274	60S ribosomal protein L27 GN=Rpl27 PE=2 SV=2
Q9H4A4	11	650	72,5	5,74	273	Aminopeptidase B GN=RNPEP PE=1 SV=2
P04908	24	130	14,1	11,05	273	Histone H2A type 1-B/E GN=HIST1H2AB PE=1 SV=2
Q96CT7	12	223	25,8	9,54	272	Coiled-coil domain-containing protein 124 GN=CCDC124 PE=1 SV=1
Q9NZ45	10	108	12,2	9,09	271	CDGSH iron-sulfur domain-containing protein 1 GN=CISD1 PE=1 SV=1
P55809	22	520	56,1	7,46	271	Succinyl-CoA:3-ketoacid coenzyme A transferase 1, mitochondrial GN=OXCT1 PE=1 SV=
P17812	9	591	66,6	6,46	271	CTP synthase 1 GN=CTPS1 PE=1 SV=2
P09525	36	319	35,9	6,13	269	Annexin A4 GN=ANXA4 PE=1 SV=4
P07108	21	87	10,0	6,57	269	Acyl-CoA-binding protein GN=DBI PE=1 SV=2
Q9Y617	57	370	40,4	7,66	269	Phosphoserine aminotransferase GN=PSAT1 PE=1 SV=2
Q9Y2B0	17	182	20,6	4,92	269	Protein canopy homolog 2 GN=CNPY2 PE=1 SV=1
Q13619	18	759	87,6	8,13	268	Cullin-4A GN=CUL4A PE=1 SV=3
Q3ZCK9	15	261	29,5	7,72	268	Proteasome subunit alpha type-4 GN=PSMA4 PE=1 SV=1
P06737	39	847	97,1	7,17	266	Glycogen phosphorylase, liver form GN=PYGL PE=1 SV=4
O00764	6	312	35,1	6,13	266	Pyridoxal kinase GN=PDXK PE=1 SV=1
Q9NQG5	18	326	36,9	5,97	266	Regulation of nuclear pre-mRNA domain-containing protein 1B GN=RPRD1B PE=1 SV=
Q15785	16	309	34,5	8,98	266	Mitochondrial import receptor subunit TOM34 GN=TOMM34 PE=1 SV=2
P26373	62	211	24,2	11,65	266	60S ribosomal protein L13 GN=RPL13 PE=1 SV=4
P33993	33	719	81,3	6,46	265	DNA replication licensing factor MCM7 GN=MCM7 PE=1 SV=4
Q9UNL2	10	185	21,1	9,61	265	Translocon-associated protein subunit gamma GN=SSR3 PE=1 SV=1
O95292	20	243	27,2	7,30	264	Vesicle-associated membrane protein-associated protein B/C GN=VAPB PE=1 SV=3
P55327	20	224	24,3	4,83	263	Tumor protein D52 GN=TPD52 PE=1 SV=2
Q08J23	31	767	86,4	6,77	263	tRNA (cytosine(34)-C(5))-methyltransferase GN=NSUN2 PE=1 SV=2
P21266	23	225	26,5	5,54	261	Glutathione S-transferase Mu 3 GN=GSTM3 PE=1 SV=3
P14324	41	419	48,2	6,15	261	Farnesyl pyrophosphate synthase GN=FDPS PE=1 SV=4
P11413	27	515	59,2	6,84	261	Glucose-6-phosphate 1-dehydrogenase GN=G6PD PE=1 SV=4
Q9Y6C9	16	303	33,3	7,97	259	Mitochondrial carrier homolog 2 GN=MTCH2 PE=1 SV=1
Q16576	9	425	47,8	5,05	258	Histone-binding protein RBBP7 GN=RBBP7 PE=1 SV=1
P20290	20	206	22,2	9,38	257	Transcription factor BTF3 GN=BTF3 PE=1 SV=1
P83917	10	185	21,4	4,93	257	Chromobox protein homolog 1 GN=Cbx1 PE=1 SV=1
P41250	24	739	83,1	7,03	257	Glycyl-tRNA synthetase GN=GARS PE=1 SV=3
P39748	16	380	42,6	8,62	256	Flap endonuclease 1 GN=FEN1 PE=1 SV=1
Q13838	25	428	49,0	5,67	256	Spliceosome RNA helicase DDX39B GN=DDX39B PE=1 SV=1
Q16836	30	314	34,3	8,85	256	Hydroxyacyl-coenzyme A dehydrogenase, mitochondrial GN=HADH PE=1 SV=3
P04181	30	439	48,5	7,03	255	Ornithine aminotransferase, mitochondrial GN=OAT PE=1 SV=1
P61021	17	215	23,7	8,13	255	Ras-related protein Rab-5B GN=Rab5b PE=1 SV=1
P17980	30	439	49,2	5,24	255	26S protease regulatory subunit 6A GN=PSMC3 PE=1 SV=3
O75347	32	108	12,8	5,29	255	Tubulin-specific chaperone A GN=TBCA PE=1 SV=3
Q96IX5	7	58	6,5	9,76	254	Up-regulated during skeletal muscle growth protein 5 GN=USMG5 PE=1 SV=1
Q3S252	39	147	16,5	7,93	254	Ubiquitin-conjugating enzyme E2 variant 1 GN=UBE2V1 PE=2 SV=1
Q9UHV9	16	154	16,6	6,58	253	Prefoldin subunit 2 GN=PFDN2 PE=1 SV=1
P17858	13	780	85,0	7,50	253	6-phosphofructokinase, liver type GN=PFKL PE=1 SV=6
O76021	19	490	54,9	10,13	252	Ribosomal L1 domain-containing protein 1 GN=RSL1D1 PE=1 SV=3
Q3S243	29	145	16,4	8,09	251	Ubiquitin-conjugating enzyme E2 variant 2 GN=UBE2V2 PE=2 SV=3
P33992	24	734	82,2	8,37	250	DNA replication licensing factor MCM5 GN=MCM5 PE=1 SV=5
P56134	18	94	10,9	9,67	250	ATP synthase subunit f, mitochondrial GN=ATP5J2 PE=1 SV=3
Q16543	13	378	44,4	5,25	247	Hsp90 co-chaperone Cdc37 GN=CDC37 PE=1 SV=1
O95716	17	219	24,3	4,93	246	Ras-related protein Rab-3D GN=RAB3D PE=1 SV=1
Q13423	35	1086	113,8	8,09	246	NAD(P) transhydrogenase, mitochondrial GN=NNT PE=1 SV=3
P41091	39	472	51,1	8,40	246	Eukaryotic translation initiation factor 2 subunit 3 GN=EIF2S3 PE=1 SV=3
Q6ZVH7	32	1005	108,1	6,47	245	Espin-like protein GN=ESPNL PE=1 SV=3
Q9UN86	15	482	54,1	5,55	244	Ras GTPase-activating protein-binding protein 2 GN=G3BP2 PE=1 SV=2

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P13646	31	458	49,6	4,96	244	Keratin, type I cytoskeletal 13 GN=KRT13 PE=1 SV=4
Q8WUD1	16	216	24,2	7,83	243	Ras-related protein Rab-2B GN=RAB2B PE=1 SV=1
P20339	22	215	23,6	8,15	243	Ras-related protein Rab-5A GN=RAB5A PE=1 SV=2
Q9HC84	46	5762	596,0	6,64	242	Mucin-5B GN=MUC5B PE=1 SV=3
Q9UL46	19	239	27,4	5,73	240	Proteasome activator complex subunit 2 GN=PSME2 PE=1 SV=4
P60335	16	356	37,5	7,09	240	Poly(rC)-binding protein 1 GN=Pcbp1 PE=1 SV=1
Q5XTY7	17	184	21,4	10,17	238	60S ribosomal protein L17 OS=Felis catus GN=RPL17 PE=2 SV=3
Q96FQ6	12	103	11,8	6,79	237	Protein S100-A16 GN=S100A16 PE=1 SV=1
Q9HAV0	20	340	37,5	6,00	235	Guanine nucleotide-binding protein subunit beta-4 GN=GNB4 PE=1 SV=3
O14949	11	82	9,9	10,08	235	Cytochrome b-c1 complex subunit 8 GN=UQCRCQ PE=1 SV=4
Q6NVV1	17	102	12,1	10,76	233	Putative 60S ribosomal protein L13a-like MGC87657 PE=5 SV=1
Q12965	12	1108	127,0	8,92	233	Unconventional myosin-Ie GN=MYO1E PE=1 SV=2
P61164	35	376	42,6	6,64	233	Alpha-centractin GN=Actr1a PE=2 SV=1
P68433	60	136	15,4	11,12	232	Histone H3.1 GN=Hist1h3a PE=1 SV=2
Q86WQ0	16	139	15,9	6,16	229	Nuclear receptor 2C2-associated protein GN=NR2C2AP PE=1 SV=1
P49419	18	539	58,5	7,99	229	Alpha-aminoadipic semialdehyde dehydrogenase GN=ALDH7A1 PE=1 SV=5
Q9Y2Q3	9	226	25,5	8,41	227	Glutathione S-transferase kappa 1 GN=GSTK1 PE=1 SV=3
P61009	192	180	20,3	8,62	227	Signal peptidase complex subunit 3 GN=SPCS3 PE=1 SV=1
P55010	7	431	49,2	5,58	226	Eukaryotic translation initiation factor 5 GN=EIF5 PE=1 SV=2
Q92598	21	858	96,8	5,39	225	Heat shock protein 105 kDa GN=HSPH1 PE=1 SV=1
P54577	21	528	59,1	7,05	224	Tyrosyl-tRNA synthetase, cytoplasmic GN=YARS PE=1 SV=4
Q15562	19	447	49,2	6,47	224	Transcriptional enhancer factor TEF-4 GN=TEAD2 PE=1 SV=2
Q7L2H7	15	374	42,5	5,63	223	Eukaryotic translation initiation factor 3 subunit M GN=EIF3M PE=1 SV=1
Q9Y5L4	11	95	10,5	8,18	223	Mitochondrial import inner membrane translocase subunit Tim13 GN=TIMM13 PE=1 SV=1
P13667	23	645	72,9	5,07	223	Protein disulfide-isomerase A4 GN=PDIA4 PE=1 SV=2
Q13347	44	325	36,5	5,64	223	Eukaryotic translation initiation factor 3 subunit I GN=EIF3I PE=1 SV=1
P30085	18	196	22,2	5,57	222	UMP-CMP kinase GN=CMPK1 PE=1 SV=3
P39023	16	403	46,1	10,18	221	60S ribosomal protein L3 GN=RPL3 PE=1 SV=2
Q9H2P0	33	1102	123,5	7,34	221	Activity-dependent neuroprotector homeobox protein GN=ADNP PE=1 SV=1
O14787	28	897	101,3	5,01	221	Transportin-2 GN=TNPO2 PE=1 SV=3
P62281	41	158	18,4	10,30	220	40S ribosomal protein S11 GN=Rps11 PE=2 SV=3
P36957	28	453	48,7	8,95	220	Dihydrolipoyllysine-residue succinyltransferase component of 2-oxoglutarate dehydrogenase GN=DLGAP1 PE=1 SV=1
Q15008	17	389	45,5	5,62	220	26S proteasome non-ATPase regulatory subunit 6 GN=PSMD6 PE=1 SV=1
P05062	27	364	39,4	7,87	219	Fructose-bisphosphate aldolase B GN=ALDOB PE=1 SV=2
O15357	31	1258	138,5	6,54	219	Phosphatidylinositol 3,4,5-trisphosphate 5-phosphatase 2 GN=INPPL1 PE=1 SV=2
O76064	20	485	55,5	7,33	219	E3 ubiquitin-protein ligase RNF8 GN=RNF8 PE=1 SV=1
P35557	23	465	52,2	5,20	219	Glucokinase GN=GCK PE=1 SV=1
Q12829	20	278	30,9	9,61	219	Ras-related protein Rab-40B GN=RAB40B PE=2 SV=1
Q8IUM7	20	802	87,1	4,64	219	Neuronal PAS domain-containing protein 4 GN=NPAS4 PE=1 SV=1
Q8TE56	19	1095	121,0	8,06	219	A disintegrin and metalloproteinase with thrombospondin motifs 17 GN=ADAMTS17
Q8WXH6	21	277	31,1	9,32	219	Ras-related protein Rab-40A GN=RAB40A PE=2 SV=2
Q99571	25	388	43,3	7,99	219	P2X purinoceptor 4 GN=P2RX4 PE=1 SV=2
Q61Q22	11	244	27,2	8,41	219	Ras-related protein Rab-12 GN=RAB12 PE=1 SV=3
P53621	28	1224	138,3	7,66	219	Coatamer subunit alpha GN=COPA PE=1 SV=2
Q9Y3F4	10	350	38,4	5,12	218	Serine-threonine kinase receptor-associated protein GN=STRAP PE=1 SV=1
P52907	13	286	32,9	5,69	218	F-actin-capping protein subunit alpha-1 GN=CAPZA1 PE=1 SV=3
P28663	13	298	33,5	5,47	217	Beta-soluble NSF attachment protein GN=Napb PE=1 SV=2
Q8CGP0	53	126	13,9	10,32	217	Histone H2B type 3-B GN=Hist3h2bb PE=1 SV=3
P57053	57	126	13,9	10,37	217	Histone H2B type F-S GN=H2BFS PE=1 SV=2
O43423	22	234	26,7	4,21	216	Acidic leucine-rich nuclear phosphoprotein 32 family member C GN=ANP32C PE=2 SV=2
P35580	41	1976	228,9	5,54	214	Myosin-10 GN=MYH10 PE=1 SV=3
P35754	12	106	11,8	8,09	214	Glutaredoxin-1 GN=GLRX PE=1 SV=2
Q00839	22	825	90,5	6,00	214	Heterogeneous nuclear ribonucleoprotein U GN=HNRNPU PE=1 SV=6
O14979	37	420	46,4	9,57	213	Heterogeneous nuclear ribonucleoprotein D-like GN=HNRPDL PE=1 SV=3
P62942	18	108	11,9	8,16	212	Peptidyl-prolyl cis-trans isomerase FKBP1A GN=FKBP1A PE=1 SV=2
P48444	9	511	57,2	6,21	212	Coatamer subunit delta GN=ARCN1 PE=1 SV=1
P62996	15	288	33,6	11,25	212	Transformer-2 protein homolog beta GN=Tra2b PE=1 SV=1
O43324	11	174	19,8	8,54	211	Eukaryotic translation elongation factor 1 epsilon-1 GN=EEF1E1 PE=1 SV=1
P49588	18	968	106,7	5,53	211	Alanine--tRNA ligase, cytoplasmic GN=AARS PE=1 SV=2
P21291	11	193	20,6	8,57	211	Cysteine and glycine-rich protein 1 GN=CSRP1 PE=1 SV=3
P15121	22	316	35,8	6,98	210	Aldose reductase GN=AKR1B1 PE=1 SV=3
P11216	31	843	96,6	6,86	210	Glycogen phosphorylase, brain form GN=PYGB PE=1 SV=5
O14579	24	308	34,5	5,12	210	Coatamer subunit epsilon GN=COPE PE=1 SV=3
Q99536	10	393	41,9	6,29	209	Synaptic vesicle membrane protein VAT-1 homolog GN=VAT1 PE=1 SV=2
P43307	13	286	32,2	4,49	209	Translocon-associated protein subunit alpha GN=SSR1 PE=1 SV=3

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P62806	48	103	11,4	11,36	209	Histone H4 GN=Hist1h4a PE=1 SV=2
P42771	20	156	16,5	5,81	208	Cyclin-dependent kinase inhibitor 2A, isoforms 1/2/3 GN=CDKN2A PE=1 SV=2
O00148	13	427	49,1	5,68	208	ATP-dependent RNA helicase DDX39A GN=DDX39A PE=1 SV=2
P31942	7	346	36,9	6,87	208	Heterogeneous nuclear ribonucleoprotein H3 GN=HNRNP3 PE=1 SV=2
Q8N9W4	52	650	79,3	5,54	208	Golgin subfamily A member 6-like protein 2 GN=GOLGA6L2 PE=2 SV=2
P16402	317	221	22,3	11,02	208	Histone H1.3 GN=HIST1H1D PE=1 SV=2
Q14257	23	317	36,9	4,40	206	Reticulocalbin-2 GN=RCN2 PE=1 SV=1
Q2VIR3	34	472	51,2	8,40	206	Putative eukaryotic translation initiation factor 2 subunit 3-like protein GN=EIF2S3L P
Q01546	33	638	65,8	8,12	205	Keratin, type II cytoskeletal 2 oral GN=KRT76 PE=1 SV=2
P00367	22	558	61,4	7,80	205	Glutamate dehydrogenase 1, mitochondrial GN=GLUD1 PE=1 SV=2
P14550	21	325	36,5	6,79	204	Alcohol dehydrogenase [NADP(+)] GN=AKR1A1 PE=1 SV=3
P26599	33	531	57,2	9,17	204	Polypyrimidine tract-binding protein 1 GN=PTBP1 PE=1 SV=1
P59999	13	168	19,7	8,43	204	Actin-related protein 2/3 complex subunit 4 GN=Arpc4 PE=1 SV=3
P62192	22	440	49,2	6,21	204	26S protease regulatory subunit 4 GN=Psmc1 PE=1 SV=1
Q02818	30	461	53,8	5,25	203	Nucleobindin-1 GN=NUCB1 PE=1 SV=4
P21980	20	687	77,3	5,22	201	Protein-glutamine gamma-glutamyltransferase 2 GN=TGM2 PE=1 SV=2
P50502	9	369	41,3	5,27	200	Hsc70-interacting protein GN=ST13 PE=1 SV=2
Q13057	12	564	62,3	6,99	198	Bifunctional coenzyme A synthase GN=COASY PE=1 SV=4
Q04446	14	702	80,4	6,32	198	1,4-alpha-glucan-branching enzyme GN=GBE1 PE=1 SV=3
Q8NEB9	44	887	101,5	6,81	197	Phosphatidylinositol 3-kinase catalytic subunit type 3 GN=PIK3C3 PE=1 SV=1
P46778	13	160	18,6	10,49	197	60S ribosomal protein L21 GN=RPL21 PE=1 SV=2
O75122	186	1294	141,0	8,47	197	CLIP-associating protein 2 GN=CLASP2 PE=1 SV=2
Q86YS6	10	212	23,3	5,64	197	Ras-related protein Rab-43 GN=RAB43 PE=1 SV=1
A6NDJ8	12	181	20,2	6,54	197	Putative Rab-43-like protein ENSP00000330714 PE=5 SV=3
P20340	10	208	23,6	5,54	197	Ras-related protein Rab-6A GN=RAB6A PE=1 SV=3
Q96JK9	29	1134	121,7	7,52	197	Mastermind-like protein 3 GN=MAML3 PE=1 SV=3
O00160	7	1098	124,8	9,11	196	Unconventional myosin-Ib GN=MYO1F PE=1 SV=3
Q86211	25	157	17,8	11,25	196	60S ribosomal protein L24 GN=RPL24 PE=2 SV=2
O15020	71	2390	271,2	6,11	196	Spectrin beta chain, brain 2 GN=SPTBN2 PE=1 SV=3
P84104	17	164	19,3	11,65	195	Serine/arginine-rich splicing factor 3 GN=Srsf3 PE=1 SV=1
Q3MIP1	43	535	58,4	9,42	195	Inositol 1,4,5-trisphosphate receptor-interacting protein-like 2 GN=ITPRIPL2 PE=2 SV=
Q96DG6	25	245	28,0	7,18	195	Carboxymethylglutaminase homolog GN=CMBL PE=1 SV=1
P56537	27	245	26,6	4,68	194	Eukaryotic translation initiation factor 6 GN=EIF6 PE=1 SV=1
P15927	13	270	29,2	6,15	194	Replication protein A 32 kDa subunit GN=RPA2 PE=1 SV=1
O00264	14	195	21,7	4,70	193	Membrane-associated progesterone receptor component 1 GN=PGRMC1 PE=1 SV=3
P61247	29	264	29,9	9,73	193	40S ribosomal protein S3a GN=RPS3A PE=1 SV=2
A6PW82	21	633	72,0	6,09	193	Putative uncharacterized protein CXorf30 GN=CXorf30 PE=2 SV=2
Q86WR0	292	208	24,5	6,80	193	Coiled-coil domain-containing protein 25 GN=CCDC25 PE=1 SV=2
P31150	35	447	50,6	5,14	192	Rab GDP dissociation inhibitor alpha GN=GDI1 PE=1 SV=2
Q9H7Z7	6	377	41,9	9,16	192	Prostaglandin E synthase 2 GN=PTGES2 PE=1 SV=1
P08174	23	381	41,4	7,59	192	Complement decay-accelerating factor GN=CD55 PE=1 SV=4
Q16629	18	238	27,4	11,82	192	Serine/arginine-rich splicing factor 7 GN=SRSF7 PE=1 SV=1
Q8WZ42	323	34350	3813,8	6,34	192	Titin GN=TTN PE=1 SV=3
Q15691	37	268	30,0	5,14	191	Microtubule-associated protein RP/EB family member 1 GN=MAPRE1 PE=1 SV=3
Q09161	29	790	91,8	6,43	191	Nuclear cap-binding protein subunit 1 GN=NCBP1 PE=1 SV=1
O15173	19	223	23,8	4,88	191	Membrane-associated progesterone receptor component 2 GN=PGRMC2 PE=1 SV=1
Q32LA7	53	128	13,5	10,58	190	Histone H2A.V GN=H2AFV PE=2 SV=3
Q92945	17	711	73,1	7,30	190	Far upstream element-binding protein 2 GN=KHSRP PE=1 SV=4
P20618	20	241	26,5	8,13	189	Proteasome subunit beta type-1 GN=PSMB1 PE=1 SV=2
P08727	45	400	44,1	5,14	189	Keratin, type I cytoskeletal 19 GN=KRT19 PE=1 SV=4
Q99460	22	953	105,8	5,39	189	26S proteasome non-ATPase regulatory subunit 1 GN=PSMD1 PE=1 SV=2
Q9CQC6	59	419	48,0	5,92	188	Basic leucine zipper and W2 domain-containing protein 1 GN=Bzw1 PE=1 SV=1
A6NI21	12	184	20,9	5,48	187	Ras-related protein Rap-1b-like protein PE=2 SV=1
P62835	14	184	21,0	6,67	187	Ras-related protein Rap-1A GN=Rap1a PE=2 SV=1
O15067	41	1338	144,6	5,76	187	Phosphoribosylformylglycinamide synthase GN=PFAS PE=1 SV=4
P40673	43	209	24,0	7,81	187	High mobility group protein B2 GN=HMGB2 PE=1 SV=3
Q9NYA3	39	693	79,9	5,41	186	Golgin subfamily A member 6A GN=GOLGA6A PE=1 SV=3
Q01082	45	2364	274,4	5,57	186	Spectrin beta chain, brain 1 GN=SPTBN1 PE=1 SV=2
P07196	27	543	61,5	4,65	186	Neurofilament light polypeptide GN=NEFL PE=1 SV=3
P07197	36	916	102,4	4,91	186	Neurofilament medium polypeptide GN=NEFM PE=1 SV=3
P12036	45	1026	112,4	6,18	186	Neurofilament heavy polypeptide GN=NEFH PE=1 SV=4
Q16352	27	499	55,4	5,40	186	Alpha-internexin GN=INA PE=1 SV=2
P80303	32	420	50,2	5,12	186	Nucleobindin-2 GN=NUCB2 PE=1 SV=2
Q9H009	5	215	23,2	4,73	185	Nascent polypeptide-associated complex subunit alpha-2 GN=NACA2 PE=1 SV=1
Q15019	9	361	41,5	6,60	185	Septin-2 GN=SEPT2 PE=1 SV=1

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A2RTX5	37	802	92,6	6,05	185	Probable threonine--tRNA ligase 2, cytoplasmic GN=TARSL2 PE=1 SV=1
P27482	16	149	16,9	4,42	184	Calmodulin-like protein 3 GN=CALML3 PE=1 SV=2
P22695	12	453	48,4	8,63	184	Cytochrome b-c1 complex subunit 2, mitochondrial GN=UQCRC2 PE=1 SV=3
Q96L21	18	214	24,5	10,01	183	60S ribosomal protein L10-like GN=RPL10L PE=1 SV=3
Q9H444	6	224	24,9	4,82	183	Charged multivesicular body protein 4b GN=CHMP4B PE=1 SV=1
Q92526	29	530	57,8	7,24	183	T-complex protein 1 subunit zeta-2 GN=CCT6B PE=1 SV=5
Q7Z3Y8	29	459	49,8	5,05	183	Keratin, type I cytoskeletal 27 GN=KRT27 PE=1 SV=2
P68401	17	229	25,6	5,92	183	Platelet-activating factor acetylhydrolase IB subunit beta GN=PFAFH1B2 PE=1 SV=1
Q9NP97	12	96	10,9	7,25	182	Dynein light chain roadblock-type 1 GN=DYNLRB1 PE=1 SV=3
Q8NF37	7	534	59,1	6,02	182	Lysophosphatidylcholine acyltransferase 1 GN=LPCAT1 PE=1 SV=2
P62276	25	56	6,7	10,13	181	40S ribosomal protein S29 GN=RPS29 PE=3 SV=2
Q12791	36	1236	137,5	7,06	181	Calcium-activated potassium channel subunit alpha-1 GN=KCNMA1 PE=1 SV=2
O00154	16	380	41,8	8,54	181	Cytosolic acyl coenzyme A thioester hydrolase GN=ACOT7 PE=1 SV=3
Q15843	22	81	9,1	8,43	180	NEDD8 GN=NEDD8 PE=1 SV=1
Q99798	193	780	85,4	7,61	180	Aconitate hydratase, mitochondrial GN=ACO2 PE=1 SV=2
O75947	27	161	18,5	5,30	179	ATP synthase subunit d, mitochondrial GN=ATP5H PE=1 SV=3
Q9NR31	24	198	22,4	6,68	179	GTP-binding protein SAR1a GN=SAR1A PE=1 SV=1
Q6ZMW3	35	1958	217,8	7,44	179	Echinoderm microtubule-associated protein-like 6 GN=EML6 PE=2 SV=2
Q9UMY4	8	172	19,7	7,87	178	Sorting nexin-12 GN=SNX12 PE=1 SV=3
P54105	6	237	26,2	4,11	178	Methylosome subunit pICln GN=CLNS1A PE=1 SV=1
P54136	25	660	75,3	6,68	177	Arginyl-tRNA synthetase, cytoplasmic GN=RARS PE=1 SV=2
P55795	9	449	49,2	6,30	177	Heterogeneous nuclear ribonucleoprotein H2 GN=HNRNPH2 PE=1 SV=1
P30043	6	206	22,1	7,65	176	Flavin reductase (NADPH) GN=BLVRB PE=1 SV=3
Q9Y5M8	6	271	29,7	9,04	176	Signal recognition particle receptor subunit beta GN=SRPRB PE=1 SV=3
Q13061	27	729	81,5	9,42	176	Triadin GN=TRDN PE=1 SV=4
Q9H169	20	189	22,1	5,99	175	Stathmin-4 GN=STMN4 PE=2 SV=1
O43390	12	633	70,9	8,13	175	Heterogeneous nuclear ribonucleoprotein R GN=HNRNPR PE=1 SV=1
Q96RP9	16	751	83,4	7,01	175	Elongation factor G, mitochondrial GN=GFM1 PE=1 SV=2
Q6P1F6	13	447	51,7	6,20	175	Serine/threonine-protein phosphatase 2A 55 kDa regulatory subunit B alpha isoform
Q9UHY7	15	261	28,9	4,78	175	Enolase-phosphatase E1 GN=ENOPH1 PE=1 SV=1
O75694	45	1391	155,1	6,16	174	Nuclear pore complex protein Nup155 GN=NUP155 PE=1 SV=1
Q9HC38	19	313	34,8	5,60	174	Glyoxalase domain-containing protein 4 GN=GLOD4 PE=1 SV=1
Q02218	35	1023	115,9	6,86	174	2-oxoglutarate dehydrogenase, mitochondrial GN=OGDH PE=1 SV=3
Q96EK6	11	184	20,7	7,99	174	Glucosamine 6-phosphate N-acetyltransferase GN=GPNAT1 PE=1 SV=1
Q9BTP7	14	215	23,9	9,19	174	Fanconi anemia-associated protein of 24 kDa GN=FAAP24 PE=1 SV=2
Q16643	7	649	71,4	4,45	173	Drebrin GN=DBN1 PE=1 SV=4
O60762	34	260	29,6	9,57	173	Dolichol-phosphate mannosyltransferase GN=DPM1 PE=1 SV=1
O60664	8	434	47,0	5,44	172	Perilipin-3 GN=PLIN3 PE=1 SV=3
Q8NF91	125	8797	1010,4	5,53	172	Nesprin-1 GN=SYNE1 PE=1 SV=3
P52815	17	198	21,3	8,87	171	39S ribosomal protein L12, mitochondrial GN=MRPL12 PE=1 SV=2
Q7Z3Y9	21	468	51,9	4,92	171	Keratin, type I cytoskeletal 26 GN=KRT26 PE=1 SV=2
Q9NUJ1	16	306	33,9	8,57	171	Abhydrolase domain-containing protein 10, mitochondrial GN=ABHD10 PE=1 SV=1
P18077	26	110	12,5	11,06	170	60S ribosomal protein L35a GN=RPL35A PE=1 SV=2
P06493	11	297	34,1	8,40	169	Cyclin-dependent kinase 1 GN=CDK1 PE=1 SV=3
Q99613	30	913	105,3	5,68	168	Eukaryotic translation initiation factor 3 subunit C GN=EIF3C PE=1 SV=1
P21281	6	511	56,5	5,81	168	V-type proton ATPase subunit B, brain isoform GN=ATP6V1B2 PE=1 SV=3
Q9Y224	12	244	28,1	6,65	168	UPF0568 protein C14orf166 GN=C14orf166 PE=1 SV=1
P61161	31	394	44,7	6,74	168	Actin-related protein 2 GN=Actr2 PE=1 SV=1
Q16401	6	504	56,2	5,48	168	26S proteasome non-ATPase regulatory subunit 5 GN=PSMD5 PE=1 SV=3
P62715	10	309	35,6	5,43	167	Serine/threonine-protein phosphatase 2A catalytic subunit beta isoform GN=Ppp2cb
P54886	9	795	87,2	7,12	167	Delta-1-pyrroline-5-carboxylate synthase GN=ALDH18A1 PE=1 SV=2
P11279	10	417	44,9	8,75	167	Lysosome-associated membrane glycoprotein 1 GN=LAMP1 PE=1 SV=3
Q7Z3V4	39	1068	123,0	8,19	166	Ubiquitin-protein ligase E3B GN=UBE3B PE=1 SV=3
P14616	36	1297	143,6	6,47	166	Insulin receptor-related protein GN=INSRR PE=1 SV=2
Q921L3	14	188	21,2	9,74	166	Transmembrane and coiled-coil domain-containing protein 1 GN=Tmco1 PE=2 SV=1
P24539	17	256	28,9	9,36	166	ATP synthase subunit b, mitochondrial GN=ATP5F1 PE=1 SV=2
P21397	16	527	59,6	7,85	166	Amine oxidase [flavin-containing] A GN=MAOA PE=1 SV=1
P04792	14	205	22,8	6,40	166	Heat shock protein beta-1 GN=HSPB1 PE=1 SV=2
Q9CXU9	9	113	12,8	7,37	165	Eukaryotic translation initiation factor 1b GN=EIF1b PE=2 SV=2
O75167	34	634	69,7	8,16	165	Phosphatase and actin regulator 2 GN=PHACTR2 PE=1 SV=2
P19623	13	302	33,8	5,49	165	Spermidine synthase GN=SRM PE=1 SV=1
O95571	10	254	27,9	6,83	164	Protein ETHE1, mitochondrial GN=ETHE1 PE=1 SV=2
Q15149	168	4684	531,5	5,96	164	Plectin GN=PLEC PE=1 SV=3
Q13907	9	227	26,3	6,34	163	Isopentenyl-diphosphate Delta-isomerase 1 GN=IDI1 PE=1 SV=2
Q6ZMR3	19	332	36,5	6,99	163	L-lactate dehydrogenase A-like 6A GN=LDHAL6A PE=2 SV=1

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Q13442	25	181	20,6	8,87	163	28 kDa heat- and acid-stable phosphoprotein GN=PDAP1 PE=1 SV=1
O35593	44	310	34,6	6,52	163	26S proteasome non-ATPase regulatory subunit 14 GN=Psm14 PE=1 SV=2
Q15046	20	597	68,0	6,35	162	Lysyl-tRNA synthetase GN=KARS PE=1 SV=3
O60701	13	494	55,0	7,12	162	UDP-glucose 6-dehydrogenase GN=UGDH PE=1 SV=1
Q3MHP2	16	218	24,5	5,94	162	Ras-related protein Rab-11B GN=RAB11B PE=2 SV=3
P14866	11	589	64,1	8,22	161	Heterogeneous nuclear ribonucleoprotein L GN=HNRNPL PE=1 SV=2
P50995	24	505	54,4	7,65	161	Annexin A11 GN=ANXA11 PE=1 SV=1
O95336	18	258	27,5	6,05	160	6-phosphogluconolactonase GN=PGLS PE=1 SV=2
P34897	13	504	56,0	8,53	160	Serine hydroxymethyltransferase, mitochondrial GN=SHMT2 PE=1 SV=3
P12035	25	628	64,4	6,48	160	Keratin, type II cytoskeletal 3 GN=KRT3 PE=1 SV=3
P30154	23	601	66,2	4,94	159	Serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A beta isoform GN=PPP2R1B PE=1 SV=1
P34896	6	483	53,0	7,71	159	Serine hydroxymethyltransferase, cytosolic GN=SHMT1 PE=1 SV=1
Q62WV3	10	214	24,6	10,08	159	60S ribosomal protein L10 GN=Rpl10 PE=2 SV=3
Q12912	358	555	62,1	5,85	159	Lymphoid-restricted membrane protein GN=LRMP PE=1 SV=3
Q8NC51	10	408	44,9	8,65	158	Plasminogen activator inhibitor 1 RNA-binding protein GN=SERBP1 PE=1 SV=2
P04818	19	313	35,7	7,01	158	Thymidylate synthase GN=TYMS PE=1 SV=3
P61255	32	145	17,2	10,55	158	60S ribosomal protein L26 GN=Rpl26 PE=2 SV=1
Q9NQ38	78	1064	120,6	8,06	157	Serine protease inhibitor Kazal-type 5 GN=SPINK5 PE=1 SV=2
Q3ZCL8	18	93	10,4	4,93	157	SH3 domain-binding glutamic acid-rich-like protein 3 GN=SH3BGL3 PE=3 SV=1
Q14914	16	329	35,8	8,29	156	Prostaglandin reductase 1 GN=PTGR1 PE=1 SV=2
Q68C16	41	796	91,1	8,63	155	GTPase SLIP-GC GN=C8orf80 PE=1 SV=3
O75531	8	89	10,1	6,09	155	Barrier-to-autointegration factor GN=BANF1 PE=1 SV=1
Q8NBS9	28	432	47,6	5,97	155	Thioredoxin domain-containing protein 5 GN=TXNDC5 PE=1 SV=2
Q07812	6	192	21,2	5,22	155	Apoptosis regulator BAX GN=BAX PE=1 SV=1
P22061	14	227	24,6	7,21	155	Protein-L-isoaspartate(D-aspartate) O-methyltransferase GN=PCMT1 PE=1 SV=4
P62880	14	340	37,3	6,00	154	Guanine nucleotide-binding protein G(I)/G(S)/G(T) subunit beta-2 GN=Gnb2 PE=1 SV=2
P40121	9	348	38,5	6,19	154	Macrophage-capping protein GN=CAPG PE=1 SV=2
P22090	13	263	29,4	10,24	154	40S ribosomal protein S4, Y isoform 1 GN=RPS4Y1 PE=2 SV=2
Q8TD47	22	263	29,3	10,08	154	40S ribosomal protein S4, Y isoform 2 GN=RPS4Y2 PE=1 SV=3
Q96P16	6	312	35,7	7,55	154	Regulation of nuclear pre-mRNA domain-containing protein 1A GN=RPRD1A PE=1 SV=2
Q37003	21	176	20,7	10,71	153	60S ribosomal protein L18a GN=RPL18A PE=2 SV=1
Q15029	37	972	109,4	5,00	153	116 kDa U5 small nuclear ribonucleoprotein component GN=EFTUD2 PE=1 SV=1
Q53GQ0	14	312	34,3	9,32	152	Estradiol 17-beta-dehydrogenase 12 GN=HSD17B12 PE=1 SV=2
P17844	7	614	69,1	8,92	151	Probable ATP-dependent RNA helicase DDX5 GN=DDX5 PE=1 SV=1
P41219	56	470	53,6	5,47	151	Peripherin GN=PRPH PE=1 SV=2
Q5TID7	70	509	60,1	5,91	151	Uncharacterized protein C1orf114 GN=C1orf114 PE=2 SV=1
Q9BYJ4	59	488	56,8	7,44	151	Tripartite motif-containing protein 34 GN=TRIM34 PE=1 SV=2
Q96QV6	31	131	14,2	10,86	151	Histone H2A type 1-A GN=HIST1H2AA PE=1 SV=3
Q99877	42	126	13,9	10,32	151	Histone H2B type 1-N GN=HIST1H2BN PE=1 SV=3
O94986	49	1654	189,0	5,44	151	Centrosomal protein of 152 kDa GN=CEP152 PE=1 SV=3
Q96J02	25	903	102,7	6,30	151	E3 ubiquitin-protein ligase Itchy homolog GN=ITCH PE=1 SV=2
Q92841	10	650	72,3	8,59	150	Nicotinamide phosphoribosyltransferase GN=DDX17 PE=1 SV=1
P43490	4	491	55,5	7,15	150	Nicotinamide phosphoribosyltransferase GN=NAMPT PE=1 SV=1
O95340	10	614	69,5	8,03	150	Bifunctional 3'-phosphoadenosine 5'-phosphosulfate synthase 2 GN=PAPSS2 PE=1 SV=1
Q5THR3	21	1501	172,8	8,40	150	EF-hand calcium-binding domain-containing protein 6 GN=EFCAB6 PE=1 SV=1
Q92540	28	1137	127,2	8,72	150	Protein SMG7 GN=SMG7 PE=1 SV=2
P48552	30	1158	126,9	8,18	150	Nuclear receptor-interacting protein 1 GN=NRIP1 PE=1 SV=2
Q86XW9	15	330	36,8	4,89	149	Thioredoxin domain-containing protein 6 GN=NME9 PE=1 SV=1
P46779	12	137	15,7	12,02	149	60S ribosomal protein L28 GN=RPL28 PE=1 SV=3
Q96MM6	18	686	75,6	8,53	149	Heat shock 70 kDa protein 12B GN=HSPA12B PE=1 SV=2
P00540	53	346	37,8	8,91	149	Proto-oncogene serine/threonine-protein kinase mos GN=MOS PE=2 SV=1
Q9HCB6	10	807	90,9	6,11	149	Spondin-1 GN=SPON1 PE=1 SV=2
P06899	37	126	13,9	10,32	149	Histone H2B type 1-J GN=HIST1H2BJ PE=1 SV=3
P28070	33	264	29,2	5,97	149	Proteasome subunit beta type-4 GN=PSMB4 PE=1 SV=4
Q07960	24	439	50,4	6,29	148	Rho GTPase-activating protein 1 GN=ARHGAP1 PE=1 SV=1
Q3ZBF7	22	160	18,7	4,54	148	Prostaglandin E synthase 3 GN=PTGES3 PE=1 SV=1
P36542	13	298	33,0	9,22	147	ATP synthase subunit gamma, mitochondrial GN=ATP5C1 PE=1 SV=1
P45880	13	294	31,5	7,56	147	Voltage-dependent anion-selective channel protein 2 GN=VDAC2 PE=1 SV=2
Q4LE39	59	1312	147,7	5,12	147	AT-rich interactive domain-containing protein 4B GN=ARID4B PE=1 SV=2
P15374	23	230	26,2	4,92	147	Ubiquitin carboxyl-terminal hydrolase isozyme L3 GN=UCHL3 PE=1 SV=1
Q66LE6	18	453	52,0	6,39	147	Serine/threonine-protein phosphatase 2A 55 kDa regulatory subunit B delta isoform GN=PPP2R1D PE=1 SV=1
A7E2Y1	46	1941	221,3	6,01	147	Myosin-7B GN=MYH7B PE=2 SV=3
O60518	23	1105	124,6	5,01	147	Ran-binding protein 6 GN=RANBP6 PE=1 SV=2
P16070	8	742	81,5	5,33	146	CD44 antigen GN=CD44 PE=1 SV=3
Q8IWB6	41	1497	167,8	5,15	146	Inactive serine/threonine-protein kinase TEX14 GN=TEX14 PE=1 SV=2

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Q8WXX0	62	4024	460,9	6,00	146	Dynein heavy chain 7, axonemal GN=DNAH7 PE=1 SV=2
Q10471	14	571	64,7	8,35	145	Polypeptide N-acetylgalactosaminyltransferase 2 GN=GALNT2 PE=1 SV=1
P62141	9	327	37,2	6,19	145	Serine/threonine-protein phosphatase PP1-beta catalytic subunit GN=Ppp1cb PE=1 S
O00232	7	456	52,9	7,65	144	26S proteasome non-ATPase regulatory subunit 12 GN=PSMD12 PE=1 SV=3
Q92882	8	214	23,8	5,68	144	Osteoclast-stimulating factor 1 GN=OSTF1 PE=1 SV=2
O75489	21	264	30,2	7,50	144	NADH dehydrogenase [ubiquinone] iron-sulfur protein 3, mitochondrial GN=NDUFS3
P10412	26	219	21,9	11,03	144	Histone H1.4 GN=HIST1H1E PE=1 SV=2
Q14232	12	305	33,7	7,33	144	Translation initiation factor eIF-2B subunit alpha GN=EIF2B1 PE=1 SV=1
Q9BZZ5	9	524	59,0	7,34	144	Apoptosis inhibitor 5 GN=API5 PE=1 SV=3
P42766	38	123	14,5	11,05	144	60S ribosomal protein L35 GN=RPL35 PE=1 SV=2
P32926	15	999	107,5	5,00	143	Desmoglein-3 GN=DSG3 PE=1 SV=2
Q9Y6B6	14	198	22,4	6,11	143	GTP-binding protein SAR1b GN=SAR1B PE=1 SV=1
P15313	3	513	56,8	5,66	143	V-type proton ATPase subunit B, kidney isoform GN=ATP6V1B1 PE=1 SV=3
Q9H871	14	391	44,0	6,06	143	Protein RMD5 homolog A GN=RMND5A PE=1 SV=1
A2A2Y4	21	597	68,7	6,39	143	FERM domain-containing protein 3 GN=FRMD3 PE=2 SV=1
P52888	11	689	78,8	6,05	143	Thimet oligopeptidase GN=THOP1 PE=1 SV=2
Q14258	8	630	70,9	8,09	143	E3 ubiquitin/ISG15 ligase TRIM25 GN=TRIM25 PE=1 SV=2
P08243	10	561	64,3	6,86	142	Asparagine synthetase [glutamine-hydrolyzing] GN=ASNS PE=1 SV=4
Q8N531	21	539	58,6	9,29	142	F-box/LRR-repeat protein 6 GN=FBXL6 PE=2 SV=1
Q9HCE9	30	1232	135,9	5,82	142	Anoctamin-8 GN=ANO8 PE=2 SV=3
Q96KB5	9	322	36,1	5,12	142	Lymphokine-activated killer T-cell-originated protein kinase GN=PBK PE=1 SV=3
Q9NSE4	32	1012	113,7	7,20	142	Isoleucyl-tRNA synthetase, mitochondrial GN=IARS2 PE=1 SV=2
Q02880	28	1626	183,2	8,00	142	DNA topoisomerase 2-beta GN=TOP2B PE=1 SV=3
Q370V7	11	148	16,4	9,95	142	Endothelial differentiation-related factor 1 GN=EDF1 PE=2 SV=1
Q16555	18	572	62,3	6,38	141	Dihydropyrimidinase-related protein 2 GN=DPYSL2 PE=1 SV=1
P09651	8	372	38,7	9,13	141	Heterogeneous nuclear ribonucleoprotein A1 GN=HNRNPA1 PE=1 SV=5
P46776	20	148	16,6	11,00	141	60S ribosomal protein L27a GN=RPL27A PE=1 SV=2
P62331	21	175	20,1	8,95	140	ADP-ribosylation factor 6 GN=Arf6 PE=1 SV=2
Q9P2J5	16	1176	134,4	7,30	140	Leucine--tRNA ligase, cytoplasmic GN=LARS PE=1 SV=2
Q9UN52	31	423	47,8	6,65	140	COP9 signalosome complex subunit 3 GN=COPS3 PE=1 SV=3
O95197	10	1032	112,5	4,96	140	Reticulon-3 GN=RTN3 PE=1 SV=2
P68399	7	391	45,1	7,74	140	Casein kinase II subunit alpha GN=CSNK2A1 PE=1 SV=1
Q4VXU2	28	614	68,3	8,87	140	Polyadenylate-binding protein 1-like GN=PABPC1L PE=2 SV=1
P28072	14	239	25,3	4,92	139	Proteasome subunit beta type-6 GN=PSMB6 PE=1 SV=4
P63163	26	240	24,6	11,19	139	Small nuclear ribonucleoprotein-associated protein N GN=Snrpn PE=2 SV=1
Q8NE71	14	845	95,9	6,80	139	ATP-binding cassette sub-family F member 1 GN=ABCF1 PE=1 SV=2
P11498	37	1178	129,6	6,84	139	Pyruvate carboxylase, mitochondrial GN=PC PE=1 SV=2
P30203	18	668	71,8	4,92	139	T-cell differentiation antigen CD6 GN=CD6 PE=1 SV=3
P62320	9	126	13,9	10,32	139	Small nuclear ribonucleoprotein Sm D3 GN=Snrpd3 PE=1 SV=1
Q15370	12	118	13,1	4,88	138	Transcription elongation factor B polypeptide 2 GN=TCEB2 PE=1 SV=1
Q7KZF4	18	910	101,9	7,17	137	Staphylococcal nuclease domain-containing protein 1 GN=SDN1 PE=1 SV=1
Q9Y678	10	874	97,7	5,47	137	Coatmer subunit gamma-1 GN=COPG1 PE=1 SV=1
Q16795	16	377	42,5	9,80	137	NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 9, mitochondrial GN
Q4G0N4	18	442	49,4	8,18	137	NAD kinase domain-containing protein 1 GN=NADKD1 PE=1 SV=2
Q6BDS2	19	1440	159,4	6,14	137	UHRF1-binding protein 1 GN=UHRF1BP1 PE=1 SV=1
Q8IX03	22	1113	125,2	5,85	137	Protein KIBRA GN=WWC1 PE=1 SV=1
O15381	21	856	95,0	6,48	137	Nuclear valosin-containing protein-like GN=NVL PE=1 SV=1
P43003	24	542	59,5	8,41	137	Excitatory amino acid transporter 1 GN=SLC1A3 PE=1 SV=1
Q13608	24	980	104,0	6,34	137	Peroxisome assembly factor 2 GN=PEX6 PE=1 SV=2
Q15056	8	248	27,4	7,23	136	Eukaryotic translation initiation factor 4H GN=EIF4H PE=1 SV=5
Q9UMX0	4	589	62,5	5,11	136	Ubiquilin-1 GN=UBQLN1 PE=1 SV=2
Q9Y285	10	508	57,5	7,80	136	Phenylalanine--tRNA ligase alpha subunit GN=FARSA PE=1 SV=3
P84244	34	136	15,3	11,27	135	Histone H3.3 GN=H3f3a PE=1 SV=2
Q92616	24	2671	292,6	7,47	135	Translational activator GCN1 GN=GCN1L1 PE=1 SV=6
A8MW92	7	1017	114,9	6,83	135	PHD finger protein 20-like protein 1 GN=PHF20L1 PE=1 SV=2
Q14152	23	1382	166,5	6,79	135	Eukaryotic translation initiation factor 3 subunit A GN=EIF3A PE=1 SV=1
P19013	27	534	57,2	6,61	135	Keratin, type II cytoskeletal 4 GN=KRT4 PE=1 SV=4
Q3MHP5	9	367	40,5	8,90	135	Developmentally-regulated GTP-binding protein 1 GN=DRG1 PE=2 SV=1
Q9BTT0	7	268	30,7	3,85	134	Acidic leucine-rich nuclear phosphoprotein 32 family member E GN=ANP32E PE=1 SV
Q16531	23	1140	126,9	5,26	134	DNA damage-binding protein 1 GN=DDB1 PE=1 SV=1
P46926	7	289	32,6	6,92	134	Glucosamine-6-phosphate isomerase 1 GN=GNPDA1 PE=1 SV=1
Q99627	7	209	23,2	5,38	133	COP9 signalosome complex subunit 8 GN=COPS8 PE=1 SV=1
Q96A08	27	127	14,2	10,32	133	Histone H2B type 1-A GN=HIST1H2BA PE=1 SV=3
Q08050	227	763	84,2	7,91	133	Forkhead box protein M1 GN=FOXM1 PE=1 SV=3
Q9UHX3	226	823	90,4	6,87	133	EGF-like module-containing mucin-like hormone receptor-like 2 GN=EMR2 PE=1 SV=2

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Q9UBW8	12	275	30,3	8,22	133	COP9 signalosome complex subunit 7a GN=COPS7A PE=1 SV=1
Q9NRC6	67	3674	416,6	6,67	132	Spectrin beta chain, brain 4 GN=SPTBN5 PE=1 SV=1
Q80X95	38	313	36,5	7,72	132	Ras-related GTP-binding protein A GN=Rraga PE=2 SV=1
O76003	10	335	37,4	5,39	131	Glutaredoxin-3 GN=GLRX3 PE=1 SV=2
Q96P70	9	1041	115,9	4,81	131	Importin-9 GN=IPO9 PE=1 SV=3
O14744	12	637	72,6	6,29	131	Protein arginine N-methyltransferase 5 GN=PRMT5 PE=1 SV=4
O95235	41	890	100,2	6,92	131	Kinesin-like protein KIF20A GN=KIF20A PE=1 SV=1
P62855	18	115	13,0	11,00	131	40S ribosomal protein S26 GN=Rps26 PE=2 SV=3
Q8NBJ5	8	622	71,6	7,31	130	Procollagen galactosyltransferase 1 GN=GLT25D1 PE=1 SV=1
P63210	12	74	8,5	4,82	130	Guanine nucleotide-binding protein G(T) subunit gamma-T1 OS=Canis familiaris GN=C
Q4VNC0	14	1218	137,2	7,90	130	Probable cation-transporting ATPase 13A5 GN=ATP13A5 PE=2 SV=1
Q9UPN3	99	7388	837,8	5,39	130	Microtubule-actin cross-linking factor 1, isoforms 1/2/3/5 GN=MACF1 PE=1 SV=4
P78386	19	507	55,8	6,55	130	Keratin, type II cuticular Hb5 GN=KRT85 PE=1 SV=1
Q72627	20	4374	481,6	5,22	130	E3 ubiquitin-protein ligase HUWE1 GN=HUWE1 PE=1 SV=3
Q8TF09	6	96	10,8	7,50	129	Dynein light chain roadblock-type 2 GN=DYNLRB2 PE=1 SV=1
P27338	23	520	58,7	7,50	129	Amine oxidase [flavin-containing] B GN=MAOB PE=1 SV=3
Q8WUM4	18	868	96,0	6,52	129	Programmed cell death 6-interacting protein GN=PDCD6IP PE=1 SV=1
Q15652	68	2540	284,3	7,87	129	Probable JmjC domain-containing histone demethylation protein 2C GN=JMJD1C PE=
Q8IZP2	4	240	27,4	5,08	129	Putative protein FAM10A4 GN=ST13P4 PE=5 SV=1
Q8IWW8	20	1755	200,4	6,24	128	E3 ubiquitin-protein ligase UBR2 GN=UBR2 PE=1 SV=1
Q9HAV7	10	217	24,3	8,12	128	GrpE protein homolog 1, mitochondrial GN=GRPEL1 PE=1 SV=2
Q9Y2V2	8	147	15,9	8,21	128	Calcium-regulated heat stable protein 1 GN=CARHSP1 PE=1 SV=2
O95757	13	839	94,5	5,88	128	Heat shock 70 kDa protein 4L GN=HSPA4L PE=1 SV=3
P51812	16	740	83,7	6,89	128	Ribosomal protein S6 kinase alpha-3 GN=RPS6KA3 PE=1 SV=1
Q9BRP0	23	275	30,4	8,75	128	Transcription factor Ovo-like 2 GN=OVOL2 PE=2 SV=1
Q92905	14	334	37,6	6,54	127	COP9 signalosome complex subunit 5 GN=COP5 PE=1 SV=4
Q04637	19	1599	175,4	5,33	127	Eukaryotic translation initiation factor 4 gamma 1 GN=EIF4G1 PE=1 SV=4
Q96QB1	36	1528	170,5	6,40	126	Rho GTPase-activating protein 7 GN=DLC1 PE=1 SV=4
Q12792	19	350	40,3	6,96	126	Twinfilin-1 GN=TWF1 PE=1 SV=3
P36405	13	182	20,4	7,24	126	ADP-ribosylation factor-like protein 3 GN=ARL3 PE=1 SV=2
Q14847	20	261	29,7	7,05	126	LIM and SH3 domain protein 1 GN=LASP1 PE=1 SV=2
Q14135	26	290	30,9	8,22	126	Transcription cofactor vestigial-like protein 4 GN=VGLL4 PE=1 SV=4
Q8IYT4	23	538	61,2	7,56	126	Katanin p60 ATPase-containing subunit A-like 2 GN=KATNAL2 PE=2 SV=3
P54727	30	409	43,1	4,84	125	UV excision repair protein RAD23 homolog B GN=RAD23B PE=1 SV=1
P23497	25	879	100,4	8,22	125	Nuclear autoantigen Sp-100 GN=SP100 PE=1 SV=3
Q9BVV6	214	1533	169,2	5,54	125	Uncharacterized protein KIAA0586 GN=KIAA0586 PE=1 SV=4
O43313	14	823	88,3	5,16	125	ATM interactor GN=ATMIN PE=1 SV=2
Q9UIJ7	10	227	25,5	9,16	124	GTP:AMP phosphotransferase, mitochondrial GN=AK3 PE=1 SV=4
Q62WY9	17	126	13,9	10,32	124	Histone H2B type 1-C/E/G GN=Hist1h2bc PE=1 SV=3
Q9Y277	11	283	30,6	8,66	124	Voltage-dependent anion-selective channel protein 3 GN=VDAC3 PE=1 SV=1
P10515	8	647	69,0	7,84	123	Dihydrolipoyllysine-residue acetyltransferase component of pyruvate dehydrogenas
Q72222	16	1120	125,3	5,91	123	Elongation factor Tu GTP-binding domain-containing protein 1 GN=EFTUD1 PE=1 SV=2
P31689	12	397	44,8	7,08	123	DnaJ homolog subfamily A member 1 GN=DNAJA1 PE=1 SV=2
Q9HAV4	14	1204	136,2	5,80	123	Exportin-5 GN=XPO5 PE=1 SV=1
Q02539	11	215	21,8	10,99	123	Histone H1.1 GN=HIST1H1A PE=1 SV=3
Q9H2U2	24	334	37,9	7,39	123	Inorganic pyrophosphatase 2, mitochondrial GN=PPA2 PE=1 SV=2
P30419	11	496	56,8	7,80	123	Glycylpeptide N-tetradecanoyltransferase 1 GN=NMMT1 PE=1 SV=2
P07384	10	714	81,8	5,67	122	Calpain-1 catalytic subunit GN=CAPN1 PE=1 SV=1
O95996	28	2303	243,8	8,82	122	Adenomatous polyposis coli protein 2 GN=APC2 PE=1 SV=1
P10619	11	480	54,4	6,61	122	Lysosomal protective protein GN=CTSA PE=1 SV=2
P55786	8	919	103,2	5,72	122	Puromycin-sensitive aminopeptidase GN=NPEPPS PE=1 SV=2
O14793	44	375	42,7	6,76	122	Growth/differentiation factor 8 GN=MSTN PE=1 SV=1
Q9BPX3	61	1015	114,3	5,59	122	Condensin complex subunit 3 GN=NCAPG PE=1 SV=1
O95470	38	568	63,5	9,16	122	Sphingosine-1-phosphate lyase 1 GN=SGPL1 PE=1 SV=3
Q14566	296	821	92,8	5,41	121	DNA replication licensing factor MCM6 GN=MCM6 PE=1 SV=1
Q92900	18	1129	124,3	6,61	121	Regulator of nonsense transcripts 1 GN=UPF1 PE=1 SV=2
Q16881	5	649	70,9	7,39	121	Thioredoxin reductase 1, cytoplasmic GN=TXNRD1 PE=1 SV=3
P50416	15	773	88,3	8,65	120	Carnitine O-palmitoyltransferase 1, liver isoform GN=CPT1A PE=1 SV=2
P47756	16	277	31,3	5,59	120	F-actin-capping protein subunit beta GN=CAPZB PE=1 SV=4
P43243	11	847	94,6	6,25	120	Matrin-3 GN=MATR3 PE=1 SV=2
Q01518	24	475	51,9	8,06	120	Adenylyl cyclase-associated protein 1 GN=CAP1 PE=1 SV=5
P49902	7	561	64,9	6,14	120	Cytosolic purine 5'-nucleotidase GN=NT5C2 PE=1 SV=1
Q5VST9	57	7968	867,9	5,99	117	Obscurin GN=OBSCN PE=1 SV=3
Q9Y2L1	19	958	108,9	7,14	117	Exosome complex exonuclease RRP44 GN=DIS3 PE=1 SV=2
P26885	9	142	15,6	9,13	116	Peptidyl-prolyl cis-trans isomerase FKBP2 GN=FKBP2 PE=1 SV=2

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P62830	13	140	14,9	10,51	116	60S ribosomal protein L23 GN=Rpl23 PE=1 SV=1
Q9P2D7	70	4330	493,6	5,94	116	Dynein heavy chain 1, axonemal GN=DNAH1 PE=1 SV=4
Q9HB71	36	228	26,2	8,25	116	Calcyclin-binding protein GN=CACYBP PE=1 SV=2
O14907	4	124	13,7	8,48	115	Tax1-binding protein 3 GN=TAX1BP3 PE=1 SV=2
Q86TD4	21	932	100,7	4,40	115	Sarcolumenin GN=SRL PE=2 SV=2
Q86W28	12	1048	119,4	7,96	115	NACHT, LRR and PYD domains-containing protein 8 GN=NLRP8 PE=2 SV=2
Q96CX6	14	371	40,6	6,84	115	Leucine-rich repeat-containing protein 58 GN=LRRC58 PE=1 SV=2
Q9D5T0	19	361	40,7	6,90	115	ATPase family AAA domain-containing protein 1 GN=Atad1 PE=1 SV=1
Q9Y2H5	20	1048	117,1	9,10	115	Pleckstrin homology domain-containing family A member 6 GN=PLEKHA6 PE=1 SV=4
Q9Y4W6	21	797	88,5	8,66	115	AFG3-like protein 2 GN=AFG3L2 PE=1 SV=2
P15170	10	499	55,7	5,62	114	Eukaryotic peptide chain release factor GTP-binding subunit ERF3A GN=GSPT1 PE=1 SV=1
Q15436	7	765	86,1	7,08	114	Protein transport protein Sec23A GN=SEC23A PE=1 SV=2
Q9UPY8	12	281	32,0	5,54	114	Microtubule-associated protein RP/EB family member 3 GN=MAPRE3 PE=1 SV=1
Q9UBF2	7	871	97,6	5,81	114	Coatomer subunit gamma-2 GN=COPG2 PE=1 SV=1
P04632	31	268	28,3	5,20	114	Calpain small subunit 1 GN=CAPNS1 PE=1 SV=1
O75569	32	313	34,4	8,41	114	Interferon-inducible double stranded RNA-dependent protein kinase activator A GN=
P61089	12	152	17,1	6,57	114	Ubiquitin-conjugating enzyme E2 N GN=Ube2n PE=1 SV=1
O00625	7	290	32,1	6,92	113	Pirin GN=PIR PE=1 SV=1
P58107	18	5090	555,3	5,60	113	Epiplakin GN=EPPK1 PE=1 SV=2
Q6NXT2	39	135	15,2	11,11	112	Histone H3.3C GN=H3F3C PE=1 SV=3
P54920	7	295	33,2	5,36	111	Alpha-soluble NSF attachment protein GN=NAPA PE=1 SV=3
P61971	21	127	14,5	5,38	111	Nuclear transport factor 2 GN=Nutf2 PE=2 SV=1
O95456	4	288	32,8	7,17	111	Proteasome assembly chaperone 1 GN=PSMG1 PE=1 SV=1
Q86U38	7	636	69,4	7,28	111	Pumilio domain-containing protein C14orf21 GN=C14orf21 PE=1 SV=1
Q96AC1	15	680	77,8	6,70	110	Fermitin family homolog 2 GN=FERMT2 PE=1 SV=1
Q86UD5	15	537	57,5	6,71	110	Mitochondrial sodium/hydrogen exchanger 9B2 GN=SLC9B2 PE=1 SV=2
P49257	16	510	57,5	6,77	110	Protein ERGIC-53 GN=LMAN1 PE=1 SV=2
Q13155	33	320	35,3	8,22	110	Aminoacyl tRNA synthase complex-interacting multifunctional protein 2 GN=AIMP2 F
P20336	5	220	25,0	5,03	109	Ras-related protein Rab-3A GN=RAB3A PE=1 SV=1
P20338	7	213	23,9	6,07	109	Ras-related protein Rab-4A GN=RAB4A PE=1 SV=2
Q923S9	6	203	23,0	4,97	109	Ras-related protein Rab-30 GN=Rab30 PE=2 SV=1
Q96AX2	6	223	24,8	6,35	109	Ras-related protein Rab-37 GN=RAB37 PE=1 SV=3
Q9HCK1	43	2354	265,5	6,16	109	DBF4-type zinc finger-containing protein 2 GN=ZDBF2 PE=1 SV=3
Q9NWW4	31	346	39,4	6,80	109	UPF0609 protein C4orf27 GN=C4orf27 PE=1 SV=2
Q9Y483	61	593	67,1	8,72	109	Metal-response element-binding transcription factor 2 GN=MTF2 PE=1 SV=2
Q96M27	122	445	46,7	5,83	109	Protein PRRC1 GN=PRRC1 PE=1 SV=1
Q5VTH9	12	848	94,5	5,76	109	WD repeat-containing protein 78 GN=WDR78 PE=2 SV=1
Q8N3K9	32	4069	448,9	4,78	109	Cardiomyopathy-associated protein 5 GN=CMYA5 PE=1 SV=3
A2BFH1	30	164	18,2	9,06	108	Peptidyl-prolyl cis-trans isomerase A-like 4G GN=PPIAL4G PE=2 SV=1
Q9COA0	46	1308	145,2	6,68	108	Contactin-associated protein-like 4 GN=CNTNAP4 PE=1 SV=3
P67810	7	179	20,6	9,48	108	Signal peptidase complex catalytic subunit SEC11A GN=SEC11A PE=2 SV=1
P22626	8	353	37,4	8,95	108	Heterogeneous nuclear ribonucleoproteins A2/B1 GN=HNRNPA2B1 PE=1 SV=2
O15247	46	247	28,3	5,59	108	Chloride intracellular channel protein 2 GN=CLIC2 PE=1 SV=3
P48637	3	474	52,4	5,92	108	Glutathione synthetase GN=GSS PE=1 SV=1
Q8TDQ7	6	276	31,1	6,95	107	Glucosamine-6-phosphate isomerase 2 GN=GNPDA2 PE=1 SV=1
P33176	22	963	109,6	6,51	107	Kinesin-1 heavy chain GN=KIF5B PE=1 SV=1
Q12840	6	1032	117,3	5,90	107	Kinesin heavy chain isoform 5A GN=KIF5A PE=1 SV=2
Q8NEZ5	9	403	44,5	7,03	107	F-box only protein 22 GN=FBXO22 PE=1 SV=1
Q969N2	15	578	65,7	8,38	107	GPI transamidase component PIG-T GN=PIGT PE=1 SV=1
Q9NX40	7	245	27,6	7,49	107	OCIA domain-containing protein 1 GN=OCIAD1 PE=1 SV=1
Q9UPU5	38	2620	294,2	6,14	107	Ubiquitin carboxyl-terminal hydrolase 24 GN=USP24 PE=1 SV=3
Q86XS8	27	419	46,4	8,87	107	E3 ubiquitin-protein ligase RNF130 GN=RNF130 PE=1 SV=1
Q9NTG1	51	2253	255,3	9,11	107	Polycystic kidney disease and receptor for egg jelly-related protein GN=PKDREJ PE=2
Q9NXE8	48	425	49,6	10,18	107	Pre-mRNA-splicing factor CWC25 homolog GN=CWC25 PE=1 SV=1
Q9Y603	36	341	39,0	8,03	107	Transcription factor ETV7 GN=ETV7 PE=1 SV=1
P28482	30	360	41,4	6,98	107	Mitogen-activated protein kinase 1 GN=MAPK1 PE=1 SV=3
P46108	4	304	33,8	5,55	107	Adapter molecule crk GN=CRK PE=1 SV=2
P49458	8	86	10,1	7,97	107	Signal recognition particle 9 kDa protein GN=SRP9 PE=1 SV=2
O14879	12	490	55,9	5,20	107	Interferon-induced protein with tetratricopeptide repeats 3 GN=IFIT3 PE=1 SV=1
P20073	18	488	52,7	5,68	106	Annexin A7 GN=ANXA7 PE=1 SV=3
P53618	6	953	107,1	6,05	106	Coatomer subunit beta GN=COPB1 PE=1 SV=3
Q92888	29	912	102,4	5,66	105	Rho guanine nucleotide exchange factor 1 GN=ARHGEF1 PE=1 SV=2
P08574	24	325	35,4	9,00	105	Cytochrome c1, heme protein, mitochondrial GN=CYC1 PE=1 SV=3
P10635	16	497	55,7	7,25	105	Cytochrome P450 2D6 GN=CYP2D6 PE=1 SV=2
P16403	17	213	21,4	10,93	105	Histone H1.2 GN=HIST1H1C PE=1 SV=2

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Q9Y6K8	10	562	63,3	5,07	105	Adenylate kinase isoenzyme 5 GN=AK5 PE=1 SV=2
Q86Y46	12	540	58,9	7,23	105	Keratin, type II cytoskeletal 73 GN=KRT73 PE=1 SV=1
Q9NSB2	12	600	64,8	7,56	105	Keratin, type II cuticular Hb4 GN=KRT84 PE=1 SV=2
P69905	21	142	15,2	8,68	104	Hemoglobin subunit alpha GN=HBA1 PE=1 SV=2
Q08380	16	585	65,3	5,27	104	Galectin-3-binding protein GN=LGALS3BP PE=1 SV=1
Q8WXH0	94	6885	795,9	5,36	103	Nesprin-2 GN=SYNE2 PE=1 SV=3
O15371	31	548	63,9	6,05	103	Eukaryotic translation initiation factor 3 subunit D GN=EIF3D PE=1 SV=1
Q8WXI9	26	593	65,2	9,70	103	Transcriptional repressor p66-beta GN=GATAD2B PE=1 SV=1
Q02952	9	1782	191,4	4,41	102	A-kinase anchor protein 12 GN=AKAP12 PE=1 SV=4
P14406	3	83	9,4	9,76	102	Cytochrome c oxidase subunit 7A2, mitochondrial GN=COX7A2 PE=1 SV=1
P31153	24	395	43,6	6,48	102	S-adenosylmethionine synthase isoform type-2 GN=MAT2A PE=1 SV=1
P04899	13	355	40,4	5,54	102	Guanine nucleotide-binding protein G(i) subunit alpha-2 GN=GNAI2 PE=1 SV=3
Q9UG01	27	1749	197,5	6,13	102	Intraflagellar transport protein 172 homolog GN=IFT172 PE=1 SV=2
Q14201	28	252	29,1	8,95	102	Protein BTG3 GN=BTG3 PE=2 SV=3
A0AVT1	12	1052	117,9	6,14	102	Ubiquitin-like modifier-activating enzyme 6 GN=UBA6 PE=1 SV=1
O75665	29	1012	116,6	6,10	102	Oral-facial-digital syndrome 1 protein GN=OFD1 PE=1 SV=1
P10854	15	126	13,9	10,32	101	Histone H2B type 1-M GN=Hist1h2bm PE=1 SV=2
O75312	3	459	50,9	4,73	101	Zinc finger protein ZPR1 GN=ZNF259 PE=1 SV=1
P50454	7	418	46,4	8,69	101	Serpin H1 GN=SERPINH1 PE=1 SV=2
P42765	14	397	41,9	8,09	101	3-ketoacyl-CoA thiolase, mitochondrial GN=ACAA2 PE=1 SV=2
O60831	3	178	19,2	9,19	100	PRA1 family protein 2 GN=PRAF2 PE=1 SV=1
Q96AG4	17	307	34,9	9,57	100	Leucine-rich repeat-containing protein 59 GN=LRRC59 PE=1 SV=1
O75096	65	1905	211,9	5,27	100	Low-density lipoprotein receptor-related protein 4 GN=LRP4 PE=1 SV=4
Q8N1G4	3	583	63,4	8,28	99	Leucine-rich repeat-containing protein 47 GN=LRRC47 PE=1 SV=1
P62334	7	389	44,1	7,49	99	26S protease regulatory subunit 10B GN=Psmc6 PE=1 SV=1
P46379	7	1132	119,3	5,60	99	Large proline-rich protein BAG6 GN=BAG6 PE=1 SV=2
O00193	3	183	20,3	4,72	99	Small acidic protein GN=SMAP PE=1 SV=1
P07332	24	822	93,4	6,73	99	Tyrosine-protein kinase Fes/Fps GN=FES PE=1 SV=3
P09110	9	424	44,3	8,44	99	3-ketoacyl-CoA thiolase, peroxisomal GN=ACAA1 PE=1 SV=2
Q9Y547	6	144	16,3	5,03	99	Heat shock protein beta-11 GN=HSPB11 PE=1 SV=1
Q15555	10	327	37,0	5,57	99	Microtubule-associated protein RP/EB family member 2 GN=MAPRE2 PE=1 SV=1
Q05B71	9	135	15,3	9,61	99	CDGSH iron-sulfur domain-containing protein 2 GN=CISD2 PE=2 SV=1
Q969U7	13	264	29,4	6,98	99	Proteasome assembly chaperone 2 GN=PSMG2 PE=1 SV=1
P61769	13	119	13,7	6,52	98	Beta-2-microglobulin GN=B2M PE=1 SV=1
O60678	10	531	59,8	5,35	98	Protein arginine N-methyltransferase 3 GN=PRMT3 PE=1 SV=3
Q9H611	15	641	69,8	9,72	98	ATP-dependent DNA helicase PIF1 GN=PIF1 PE=1 SV=2
P54578	5	494	56,0	5,30	98	Ubiquitin carboxyl-terminal hydrolase 14 GN=USP14 PE=1 SV=3
O43488	3	359	39,6	7,17	98	Aflatoxin B1 aldehyde reductase member 2 GN=AKR7A2 PE=1 SV=3
Q8TF17	8	1288	144,7	6,37	98	SH3 domain and tetratricopeptide repeat-containing protein 2 GN=SH3TC2 PE=1 SV=2
Q9UPP2	11	1182	127,5	6,51	98	IQ motif and SEC7 domain-containing protein 3 GN=IQSEC3 PE=2 SV=3
Q15233	23	471	54,2	8,95	98	Non-POU domain-containing octamer-binding protein GN=NONO PE=1 SV=4
Q8TDN4	22	633	67,6	9,17	98	CDK5 and ABL1 enzyme substrate 1 GN=CABLES1 PE=1 SV=2
Q9NR09	55	4857	529,9	6,05	97	Baculoviral IAP repeat-containing protein 6 GN=BIRC6 PE=1 SV=2
P08912	21	532	60,0	9,28	97	Muscarinic acetylcholine receptor M5 GN=CHRM5 PE=2 SV=2
O95881	16	172	19,2	5,40	97	Thioredoxin domain-containing protein 12 GN=TXNDC12 PE=1 SV=1
P35270	2	261	28,0	8,05	97	Sepiapterin reductase GN=SPR PE=1 SV=1
Q08945	42	709	81,0	6,87	97	FACT complex subunit SSRP1 GN=SSRP1 PE=1 SV=1
P50552	2	380	39,8	8,94	96	Vasodilator-stimulated phosphoprotein GN=VASP PE=1 SV=3
P49915	29	693	76,7	6,87	96	GMP synthase [glutamine-hydrolyzing] GN=GMPS PE=1 SV=1
Q9BTF0	45	503	56,4	7,81	96	THUMP domain-containing protein 2 GN=THUMP2 PE=2 SV=2
P31350	7	389	44,8	5,38	96	Ribonucleoside-diphosphate reductase subunit M2 GN=RRM2 PE=1 SV=1
Q7Z4S6	44	1674	187,1	6,42	96	Kinesin-like protein KIF21A GN=KIF21A PE=1 SV=2
Q2M2I5	15	525	55,1	4,96	96	Keratin, type I cytoskeletal 24 GN=KRT24 PE=1 SV=1
Q8N196	15	739	74,5	4,96	95	Homeobox protein SIX5 GN=SIX5 PE=1 SV=3
P51686	12	369	42,0	8,19	95	C-C chemokine receptor type 9 GN=CCR9 PE=1 SV=2
Q15772	48	3267	354,1	8,51	95	Striated muscle preferentially expressed protein kinase GN=SPEG PE=1 SV=4
Q96IY1	32	281	32,1	6,79	95	Kinetochore-associated protein NSL1 homolog GN=NSL1 PE=1 SV=3
P36776	19	959	106,4	6,39	95	Lon protease homolog, mitochondrial GN=LONP1 PE=1 SV=2
P10244	13	700	78,7	6,87	94	Myb-related protein B GN=MYBL2 PE=1 SV=1
Q8NI35	23	1801	196,2	4,94	94	InaD-like protein GN=INADL PE=1 SV=3
P62889	13	115	12,8	9,63	94	60S ribosomal protein L30 GN=Rpl30 PE=2 SV=2
O75390	11	466	51,7	8,32	94	Citrate synthase, mitochondrial GN=CS PE=1 SV=2
Q6TFL3	29	1326	152,7	6,81	94	Coiled-coil domain-containing protein 171 GN=CCDC171 PE=2 SV=1
Q15631	12	228	26,2	6,44	94	Translin GN=TSN PE=1 SV=1
O75083	2	606	66,2	6,65	94	WD repeat-containing protein 1 GN=WDR1 PE=1 SV=4

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O14967	12	610	70,0	4,69	94	Calmegin GN=CLGN PE=1 SV=1
Q5VWN6	99	2430	268,7	5,90	94	Protein FAM208B GN=FAM208B PE=1 SV=1
Q15393	8	1217	135,5	5,26	93	Splicing factor 3B subunit 3 GN=SF3B3 PE=1 SV=4
Q01831	11	940	105,9	8,90	93	DNA repair protein complementing XP-C cells GN=XPC PE=1 SV=4
Q62093	3	221	25,5	11,85	93	Serine/arginine-rich splicing factor 2 GN=Srsf2 PE=1 SV=4
O60551	5	498	56,9	7,58	93	Glycylpeptide N-tetradecanoyltransferase 2 GN=NM2 PE=1 SV=1
Q9NRE1	9	261	29,7	6,47	93	Matrix metalloproteinase-26 GN=MMP26 PE=2 SV=2
Q5T013	22	277	30,4	5,50	92	Putative hydroxypyruvate isomerase GN=HYI PE=2 SV=2
Q9UBU9	30	619	70,1	8,51	92	Nuclear RNA export factor 1 GN=NXF1 PE=1 SV=1
Q9D7G0	11	318	34,8	6,98	92	Ribose-phosphate pyrophosphokinase 1 GN=Prps1 PE=1 SV=4
O75449	13	491	55,9	6,90	92	Katanin p60 ATPase-containing subunit A1 GN=KATNA1 PE=1 SV=1
Q96G03	7	612	68,2	6,73	92	Phosphoglucomutase-2 GN=PGM2 PE=1 SV=4
Q13393	27	1074	124,1	8,78	92	Phospholipase D1 GN=PLD1 PE=1 SV=1
Q14203	30	1278	141,6	5,81	91	Dynactin subunit 1 GN=DCTN1 PE=1 SV=3
Q9Y4C1	32	1321	147,2	8,07	91	Lysine-specific demethylase 3A GN=KDM3A PE=1 SV=4
Q9UK76	6	154	16,0	5,60	91	Hematological and neurological expressed 1 protein GN=HN1 PE=1 SV=3
P62754	17	249	28,7	10,84	91	40S ribosomal protein S6 GN=Rps6 PE=1 SV=1
P13521	19	617	70,9	4,75	91	Secretogranin-2 GN=SCG2 PE=1 SV=2
O14556	18	408	44,5	8,19	91	Glyceraldehyde-3-phosphate dehydrogenase, testis-specific GN=GAPDHs PE=1 SV=2
P13611	17	3396	372,6	4,51	91	Versican core protein GN=VCAN PE=1 SV=3
Q03001	55	7570	860,1	5,25	91	Dystonin GN=DST PE=1 SV=4
A4FU69	28	1503	173,3	5,85	90	EF-hand calcium-binding domain-containing protein 5 GN=EFCAB5 PE=1 SV=2
Q9NY33	5	737	82,5	5,10	90	Dipeptidyl peptidase 3 GN=DPP3 PE=1 SV=2
P63001	24	192	21,4	8,50	90	Ras-related C3 botulinum toxin substrate 1 GN=Rac1 PE=1 SV=1
O43169	3	146	16,3	4,97	90	Cytochrome b5 type B GN=CYB5B PE=1 SV=2
O60239	8	455	50,4	4,97	90	SH3 domain-binding protein 5 GN=SH3BP5 PE=1 SV=2
Q70HW3	8	274	29,4	9,35	90	S-adenosylmethionine mitochondrial carrier protein GN=SLC25A26 PE=2 SV=1
Q8IZ58	8	1091	122,9	5,78	90	Voltage-dependent calcium channel subunit alpha-2/delta-3 GN=CACNA2D3 PE=1 SV=1
Q15437	8	767	86,4	6,89	90	Protein transport protein Sec23B GN=SEC23B PE=1 SV=2
P98161	29	4303	462,2	6,73	90	Polycystin-1 GN=PKD1 PE=1 SV=3
Q5TDP6	8	509	57,2	6,35	90	Lengsin GN=LGSN PE=1 SV=1
Q7RTS7	15	529	57,8	7,71	90	Keratin, type II cytoskeletal 74 GN=KRT74 PE=1 SV=2
Q8IZ56	3	198	23,2	9,33	90	Tctex1 domain-containing protein 3 GN=TCTE3 PE=1 SV=1
Q96EX2	3	444	48,9	7,84	90	RING finger and transmembrane domain-containing protein 2 GN=RNFT2 PE=2 SV=2
O95568	18	372	42,1	6,76	90	Histidine protein methyltransferase 1 homolog GN=METTL8 PE=1 SV=1
P56470	18	323	35,9	9,16	90	Galectin-4 GN=LGALS4 PE=1 SV=1
Q8N5S1	20	370	40,8	9,22	90	Solute carrier family 25 member 41 GN=SLC25A41 PE=2 SV=2
Q8WW11	32	1683	192,6	8,09	90	LIM domain only protein 7 GN=LMO7 PE=1 SV=3
Q9BQL6	10	677	77,4	6,28	89	Fermitin family homolog 1 GN=FERMT1 PE=1 SV=1
P12955	2	493	54,5	6,00	89	Xaa-Pro dipeptidase GN=PEPD PE=1 SV=3
Q13126	7	283	31,2	7,18	89	S-methyl-5'-thioadenosine phosphorylase GN=MTAP PE=1 SV=2
Q56UQ5	9	140	15,8	6,19	89	TPT1-like protein PE=1 SV=1
Q3T030	3	418	47,3	5,21	89	26S protease regulatory subunit 6B GN=PSMC4 PE=2 SV=1
Q7Z5L2	27	792	87,8	5,12	88	Growth inhibition and differentiation-related protein 88 GN=GIDRP88 PE=2 SV=2
Q5K651	27	1589	184,2	7,83	88	Sterile alpha motif domain-containing protein 9 GN=SAMD9 PE=1 SV=1
Q6RI45	34	1802	203,5	7,84	88	Bromodomain and WD repeat-containing protein 3 GN=BRWD3 PE=1 SV=2
Q8WWQ0	25	1821	206,6	8,85	88	PH-interacting protein GN=PHIP PE=1 SV=2
Q9Y696	26	253	28,8	5,59	88	Chloride intracellular channel protein 4 GN=CLIC4 PE=1 SV=4
O14983	30	1001	110,2	5,16	88	Sarcoplasmic/endoplasmic reticulum calcium ATPase 1 GN=ATP2A1 PE=1 SV=1
O75955	31	427	47,3	7,49	88	Flotillin-1 GN=FLOT1 PE=1 SV=3
P30679	27	374	43,5	8,51	88	Guanine nucleotide-binding protein subunit alpha-15 GN=GNA15 PE=2 SV=2
P60879	43	206	23,3	4,77	88	Synaptosomal-associated protein 25 GN=Snap25 PE=1 SV=1
Q14254	31	428	47,0	5,25	88	Flotillin-2 GN=FLOT2 PE=1 SV=2
Q2TAC6	29	998	111,3	8,69	88	Kinesin-like protein KIF19 GN=KIF19 PE=2 SV=2
O00151	7	329	36,0	7,02	88	PDZ and LIM domain protein 1 GN=PDLM1 PE=1 SV=4
O14529	33	1486	161,6	5,48	88	Homeobox protein cut-like 2 GN=CUX2 PE=1 SV=4
Q8NGY6	17	317	35,7	8,82	88	Olfactory receptor 6N2 GN=OR6N2 PE=1 SV=1
Q8BWW3	4	437	49,0	5,71	88	Eukaryotic peptide chain release factor subunit 1 GN=Etf1 PE=1 SV=4
Q8NDA8	32	1641	181,1	6,89	88	HEAT repeat-containing protein 7A GN=HEATR7A PE=1 SV=3
Q8NGT2	11	312	34,7	8,51	87	Olfactory receptor 13J1 GN=OR13J1 PE=2 SV=1
Q9NVE4	245	849	96,3	8,59	87	Coiled-coil domain-containing protein 87 GN=CCDC87 PE=2 SV=2
P15559	14	274	30,8	8,88	87	NAD(P)H dehydrogenase [quinone] 1 GN=NQO1 PE=1 SV=1
O00443	39	1686	190,6	8,02	87	Phosphatidylinositol-4-phosphate 3-kinase C2 domain-containing subunit alpha GN=
P26368	9	475	53,5	9,09	87	Splicing factor U2AF 65 kDa subunit GN=U2AF2 PE=1 SV=4
P13674	21	534	61,0	6,01	86	Prolyl 4-hydroxylase subunit alpha-1 GN=P4HA1 PE=1 SV=2

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Q6AWC2	60	1192	133,8	5,53	86	Protein WWC2 GN=WWC2 PE=1 SV=2
Q9NZM1	14	2061	234,6	6,18	86	Myoferlin GN=MYOF PE=1 SV=1
Q96JM2	47	2506	284,5	7,58	86	Zinc finger protein 462 GN=ZNF462 PE=1 SV=3
P07602	41	524	58,1	5,17	86	Proactivator polypeptide GN=PSAP PE=1 SV=2
Q15323	13	416	47,2	4,88	85	Keratin, type I cuticular Ha1 GN=KRT31 PE=2 SV=3
O95816	8	211	23,8	6,70	85	BAG family molecular chaperone regulator 2 GN=BAG2 PE=1 SV=1
O14645	29	258	29,6	8,50	85	Axonemal dynein light intermediate polypeptide 1 GN=DNALI1 PE=2 SV=2
Q96F24	29	287	32,4	5,87	85	Nuclear receptor-binding factor 2 GN=NRBF2 PE=1 SV=1
Q9NZQ7	30	290	33,3	7,23	85	Programmed cell death 1 ligand 1 GN=CD274 PE=1 SV=1
Q9Y6Y8	10	1000	111,0	5,54	85	SEC23-interacting protein GN=SEC23IP PE=1 SV=1
P16401	6	226	22,6	10,92	85	Histone H1.5 GN=HIST1H1B PE=1 SV=3
P46063	8	649	73,4	7,88	85	ATP-dependent DNA helicase Q1 GN=RECQL PE=1 SV=3
P61082	18	183	20,9	7,69	85	NEDD8-conjugating enzyme Ubc12 GN=Ubc2m PE=2 SV=1
Q61411	3	189	21,3	5,31	85	GTPase HRas GN=Hras1 PE=1 SV=2
Q5VYK3	12	1845	204,2	7,12	84	Proteasome-associated protein ECM29 homolog GN=ECM29 PE=1 SV=2
Q14019	4	142	15,9	5,67	84	Coactosin-like protein GN=COTL1 PE=1 SV=3
O43847	9	1150	131,5	5,00	84	Nardilysin GN=NRD1 PE=1 SV=2
Q99615	7	494	56,4	6,96	84	DnaJ homolog subfamily C member 7 GN=DNAJC7 PE=1 SV=2
P62317	16	118	13,5	9,91	84	Small nuclear ribonucleoprotein Sm D2 GN=Snrpd2 PE=2 SV=1
P22492	8	207	22,0	11,71	84	Histone H1t GN=HIST1H1T PE=1 SV=4
Q9Y237	3	131	13,8	9,77	84	Peptidyl-prolyl cis-trans isomerase NIMA-interacting 4 GN=PIN4 PE=1 SV=1
P20042	3	333	38,4	5,80	83	Eukaryotic translation initiation factor 2 subunit 2 GN=EIF2S2 PE=1 SV=2
Q9BPU6	15	564	61,4	7,20	83	Dihydropyrimidinase-related protein 5 GN=DPYSL5 PE=1 SV=1
Q96N18	27	536	62,3	8,28	83	Zinc finger protein 570 GN=ZNF570 PE=2 SV=1
Q9NQH7	27	507	57,0	6,83	83	Probable Xaa-Pro aminopeptidase 3 GN=XPNPEP3 PE=1 SV=1
Q2VPB7	14	821	87,8	5,99	83	Uncharacterized protein DKFzp761E198 GN=PP1030 PE=1 SV=3
P35613	17	385	42,2	5,66	83	Basigin GN=BSG PE=1 SV=2
Q29RF7	10	1337	150,7	7,91	83	Sister chromatid cohesion protein PDS5 homolog A GN=PDS5A PE=1 SV=1
Q9UJZ1	22	356	38,5	7,39	83	Stomatin-like protein 2 GN=STOML2 PE=1 SV=1
Q9UMD9	20	1497	150,3	8,79	82	Collagen alpha-1(XVII) chain GN=COL17A1 PE=1 SV=3
Q9NPJ3	7	140	15,0	9,14	82	Acyl-coenzyme A thioesterase 13 GN=ACOT13 PE=1 SV=1
P47914	5	159	17,7	11,66	82	60S ribosomal protein L29 GN=RPL29 PE=1 SV=2
Q9NVD7	17	372	42,2	5,95	82	Alpha-parvin GN=PARVA PE=1 SV=1
Q0MQA1	4	116	13,5	5,99	82	NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 5 OS=Gorilla gorilla g
Q9BRL6	9	282	32,3	11,72	82	Serine/arginine-rich splicing factor 8 GN=SRSF8 PE=1 SV=1
Q8WUI4	29	952	102,9	7,58	82	Histone deacetylase 7 GN=HDAC7 PE=1 SV=2
Q99JI6	6	184	20,8	5,78	82	Ras-related protein Rap-1b GN=Rap1b PE=2 SV=2
P09960	11	611	69,2	6,18	82	Leukotriene A-4 hydrolase GN=LTA4H PE=1 SV=2
Q01581	2	520	57,3	5,41	82	Hydroxymethylglutaryl-CoA synthase, cytoplasmic GN=HMGCS1 PE=1 SV=2
Q96QD8	3	506	56,0	8,00	82	Sodium-coupled neutral amino acid transporter 2 GN=SLC38A2 PE=1 SV=2
O95394	9	542	59,8	6,25	82	Phosphoacetylglucosamine mutase GN=PGM3 PE=1 SV=1
Q9BY32	4	194	21,4	5,66	81	Inosine triphosphate pyrophosphatase GN=ITPA PE=1 SV=2
Q92508	30	2521	286,6	7,47	81	Piezo-type mechanosensitive ion channel component 1 GN=PIEZO1 PE=1 SV=4
Q13561	10	401	44,2	5,21	81	Dynactin subunit 2 GN=DCTN2 PE=1 SV=4
Q7LBE3	6	791	86,9	8,22	81	Solute carrier family 26 member 9 GN=SLC26A9 PE=2 SV=1
Q72221	32	1910	210,7	8,78	81	Treslin GN=TICRR PE=1 SV=2
Q8TF05	11	950	106,9	4,77	81	Serine/threonine-protein phosphatase 4 regulatory subunit 1 GN=PPP4R1 PE=1 SV=1
Q96BR1	13	496	57,1	6,93	81	Serine/threonine-protein kinase Sgk3 GN=SGK3 PE=1 SV=1
Q9HBY8	8	427	47,6	7,46	81	Serine/threonine-protein kinase Sgk2 GN=SGK2 PE=1 SV=1
Q9P1A2	14	438	49,5	5,44	81	Putative serine/threonine-protein phosphatase 4 regulatory subunit 1-like GN=PPP4
O75844	14	475	54,8	7,49	81	CAAX prenyl protease 1 homolog GN=ZMPSTE24 PE=1 SV=2
P58743	36	744	81,2	6,29	81	Prestin GN=SLC26A5 PE=2 SV=1
P07339	4	412	44,5	6,54	81	Cathepsin D GN=CTSD PE=1 SV=1
P83882	15	106	12,4	10,58	80	60S ribosomal protein L36a GN=Rpl36a PE=3 SV=2
P48059	10	325	37,2	8,05	80	LIM and senescent cell antigen-like-containing domain protein 1 GN=LIMS1 PE=1 SV=
A4D1E1	43	1349	152,5	8,54	80	Zinc finger protein 804B GN=ZNF804B PE=1 SV=2
Q2TVT4	101	97	11,1	8,76	79	Putative keratinocyte growth factor-like protein 1 GN=KGLFP1 PE=5 SV=1
O00303	4	357	37,5	5,45	79	Eukaryotic translation initiation factor 3 subunit F GN=EIF3F PE=1 SV=1
P39656	5	456	50,8	6,55	79	Dolichyl-diphosphooligosaccharide--protein glycosyltransferase 48 kDa subunit GN=
Q15125	7	230	26,3	7,90	79	3-beta-hydroxysteroid-Delta(8),Delta(7)-isomerase GN=EBP PE=1 SV=3
Q96L46	22	248	27,6	5,73	79	Calpain small subunit 2 GN=CAPNS2 PE=2 SV=2
Q86S19	16	138	15,1	11,41	79	Protein CEI GN=C5orf38 PE=2 SV=1
Q9Y3E0	8	138	15,4	10,36	79	Vesicle transport protein GOT1B GN=GOLT1B PE=1 SV=1
P29966	3	332	31,5	4,45	79	Myristoylated alanine-rich C-kinase substrate GN=MARCKS PE=1 SV=4
Q9BXJ9	8	866	101,2	7,42	79	N-alpha-acetyltransferase 15, NatA auxiliary subunit GN=NAA15 PE=1 SV=1

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P33778	10	126	13,9	10,32	79	Histone H2B type 1-B GN=HIST1H2BB PE=1 SV=2
Q9H845	7	621	68,7	7,96	79	Acyl-CoA dehydrogenase family member 9, mitochondrial GN=ACAD9 PE=1 SV=1
Q9BV73	48	2442	281,0	5,02	78	Centrosome-associated protein CEP250 GN=CEP250 PE=1 SV=2
Q6Z2NQ3	17	347	39,6	8,57	78	Leucine-rich repeat-containing protein 69 GN=LRRC69 PE=2 SV=2
Q9Y2K7	21	1162	132,7	7,58	78	Lysine-specific demethylase 2A GN=KDM2A PE=1 SV=3
Q16740	6	277	30,2	8,09	78	Putative ATP-dependent Clp protease proteolytic subunit, mitochondrial GN=CLPP P
Q86TM3	9	631	71,1	9,07	78	Probable ATP-dependent RNA helicase DDX53 GN=DDX53 PE=1 SV=3
P21580	25	790	89,6	8,22	77	Tumor necrosis factor alpha-induced protein 3 GN=TNFAIP3 PE=1 SV=1
Q13200	11	908	100,1	5,20	77	26S proteasome non-ATPase regulatory subunit 2 GN=PSMD2 PE=1 SV=3
Q2PPJ7	34	1873	210,6	6,07	77	Ral GTPase-activating protein subunit alpha-2 GN=RALGAPA2 PE=1 SV=2
Q8TDM6	32	1919	213,7	7,42	77	Disks large homolog 5 GN=DLG5 PE=1 SV=4
P15328	27	257	29,8	7,97	77	Folate receptor alpha GN=FOLR1 PE=1 SV=3
O15511	12	151	16,3	5,67	77	Actin-related protein 2/3 complex subunit 5 GN=ARPC5 PE=1 SV=3
O75822	5	258	29,0	4,83	77	Eukaryotic translation initiation factor 3 subunit J GN=EIF3J PE=1 SV=2
Q6NSJ2	19	640	71,9	6,57	77	Pleckstrin homology-like domain family B member 3 GN=PHLDB3 PE=2 SV=3
Q9Y566	15	2161	224,8	8,15	77	SH3 and multiple ankyrin repeat domains protein 1 GN=SHANK1 PE=1 SV=2
Q9Y6E2	27	419	48,1	6,68	77	Basic leucine zipper and W2 domain-containing protein 2 GN=BZW2 PE=1 SV=1
Q96M86	44	4753	533,3	6,71	77	Dynein heavy chain domain-containing protein 1 GN=DNHD1 PE=1 SV=2
Q99959	24	881	97,4	9,33	77	Plakophilin-2 GN=PKP2 PE=1 SV=2
Q9ULW6	22	460	52,5	4,49	77	Nucleosome assembly protein 1-like 2 GN=NAP1L2 PE=2 SV=1
Q86GK5	39	1308	147,9	6,96	77	Limbin GN=EVC2 PE=1 SV=1
Q9NXH8	9	423	46,9	9,94	76	Torsin family protein C9orf167 GN=C9orf167 PE=1 SV=2
P49454	61	3210	367,5	5,07	76	Centromere protein F GN=CENPF PE=1 SV=2
O15034	23	1052	116,0	5,30	76	RIMS-binding protein 2 GN=RIMBP2 PE=1 SV=3
P10072	22	659	75,1	9,25	76	Kruppel-related zinc finger protein 1 GN=HKR1 PE=2 SV=4
P53350	23	603	68,2	8,91	76	Serine/threonine-protein kinase PLK1 GN=PLK1 PE=1 SV=1
Q9P055	12	326	36,7	7,93	75	JNK1/MAPK8-associated membrane protein GN=JKAMP PE=2 SV=2
Q5E988	8	204	22,9	9,72	75	40S ribosomal protein S5 GN=RPS5 PE=2 SV=3
Q9H4Y5	12	243	28,2	7,56	75	Glutathione S-transferase omega-2 GN=GSTO2 PE=2 SV=1
Q6PGP7	40	1564	175,4	7,53	75	Tetratricopeptide repeat protein 37 GN=TTC37 PE=1 SV=1
Q0MQC2	12	156	17,4	4,93	75	Acyl carrier protein, mitochondrial OS=Gorilla gorilla gorilla GN=NDUFAB1 PE=2 SV=1
A4GXA9	26	379	41,2	6,89	75	Probable crossover junction endonuclease EME2 GN=EME2 PE=1 SV=2
P78344	33	907	102,3	7,14	75	Eukaryotic translation initiation factor 4 gamma 2 GN=EIF4G2 PE=1 SV=1
P02768	5	609	69,3	6,28	75	Serum albumin GN=ALB PE=1 SV=2
O60244	18	1454	160,5	8,73	75	Mediator of RNA polymerase II transcription subunit 14 GN=MED14 PE=1 SV=2
O94888	11	489	54,8	5,16	75	UBX domain-containing protein 7 GN=UBXN7 PE=1 SV=2
Q04323	3	297	33,3	5,25	75	UBX domain-containing protein 1 GN=UBXN1 PE=1 SV=2
Q9Y3A6	2	229	26,0	4,84	75	Transmembrane emp24 domain-containing protein 5 GN=TMED5 PE=1 SV=1
Q8WW22	15	397	44,8	7,59	74	DnaI homolog subfamily A member 4 GN=DNAJA4 PE=1 SV=1
P30414	22	1462	165,6	9,99	74	NK-tumor recognition protein GN=NKTR PE=1 SV=2
P62918	16	257	28,0	11,03	74	60S ribosomal protein L8 GN=Rpl8 PE=2 SV=2
O60281	40	2723	304,6	7,39	74	Zinc finger protein 292 GN=ZNF292 PE=1 SV=3
Q8IVL1	31	2488	268,0	9,03	74	Neuron navigator 2 GN=NAV2 PE=1 SV=3
O60673	43	3130	352,6	8,47	74	DNA polymerase zeta catalytic subunit GN=REV3L PE=1 SV=2
O95382	31	1288	142,5	7,11	74	Mitogen-activated protein kinase kinase kinase 6 GN=MAP3K6 PE=1 SV=3
O95905	16	644	72,7	4,87	74	Protein SGT1 GN=ECD PE=1 SV=1
P33764	14	101	11,7	4,78	74	Protein S100-A3 GN=S100A3 PE=1 SV=1
P51654	15	580	65,5	6,37	74	Glypican-3 GN=GPC3 PE=1 SV=1
Q7Z478	25	1369	155,1	8,09	74	ATP-dependent RNA helicase DHX29 GN=DHX29 PE=1 SV=2
Q8NFT6	16	615	67,2	8,34	74	Protein DBF4 homolog B GN=DBF4B PE=1 SV=1
Q8TBF8	23	368	42,4	8,92	74	Protein FAM81A GN=FAM81A PE=2 SV=3
Q9Y2R2	15	807	91,6	7,59	74	Tyrosine-protein phosphatase non-receptor type 22 GN=PTPN22 PE=1 SV=2
P10155	3	538	60,6	8,03	74	60 kDa SS-A/Ro ribonucleoprotein GN=TROVE2 PE=1 SV=2
O43196	20	834	92,8	6,37	74	MutS protein homolog 5 GN=MSH5 PE=1 SV=1
O75037	34	1637	182,5	7,08	74	Kinesin-like protein KIF21B GN=KIF21B PE=1 SV=2
Q96NT1	15	182	19,6	4,21	74	Nucleosome assembly protein 1-like 5 GN=NAP1L5 PE=2 SV=1
Q9NYQ8	18	4349	479,0	5,16	74	Protocadherin Fat 2 GN=FAT2 PE=1 SV=2
P51659	2	736	79,6	8,84	74	Peroxisomal multifunctional enzyme type 2 GN=HSD17B4 PE=1 SV=3
O00391	16	747	82,5	8,92	73	Sulfhydryl oxidase 1 GN=QSOX1 PE=1 SV=3
O94921	24	469	53,0	8,92	73	Cyclin-dependent kinase 14 GN=CDK14 PE=1 SV=3
Q13232	12	169	19,0	7,84	73	Nucleoside diphosphate kinase 3 GN=NME3 PE=1 SV=2
Q16671	13	573	62,7	5,81	73	Anti-Muellerian hormone type-2 receptor GN=AMHR2 PE=1 SV=1
Q59H18	28	835	92,8	6,74	73	Serine/threonine-protein kinase TNNI3K GN=TNNI3K PE=1 SV=3
Q92772	14	493	56,0	8,21	73	Cyclin-dependent kinase-like 2 GN=CDKL2 PE=2 SV=1
Q9BXT8	19	1623	184,5	5,40	73	RING finger protein 17 GN=RNF17 PE=1 SV=3

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Q9H093	16	628	69,6	8,87	73	NUAK family SNF1-like kinase 2 GN=NUAK2 PE=1 SV=1
O94913	45	1555	172,9	8,48	73	Pre-mRNA cleavage complex 2 protein Pcf11 GN=PCF11 PE=1 SV=3
P19367	3	917	102,4	6,80	73	Hexokinase-1 GN=HK1 PE=1 SV=3
Q9P209	15	647	71,7	6,52	73	Centrosomal protein of 72 kDa GN=CEP72 PE=1 SV=2
P16520	17	340	37,2	5,67	73	Guanine nucleotide-binding protein G(I)/G(S)/G(T) subunit beta-3 GN=GNB3 PE=1 SV
Q12907	12	356	40,2	6,95	73	Vesicular integral-membrane protein VIP36 GN=LMAN2 PE=1 SV=1
Q13535	29	2644	301,2	7,43	73	Serine/threonine-protein kinase ATR GN=ATR PE=1 SV=3
Q8NFI4	3	369	41,4	5,05	73	Putative protein FAM10A5 GN=ST13P5 PE=5 SV=1
Q96DM3	5	657	74,9	7,83	73	Uncharacterized protein C18orf8 GN=C18orf8 PE=1 SV=2
Q9Y3R5	37	2298	258,1	6,29	73	Protein dopey-2 GN=DOPEY2 PE=1 SV=5
A6NCI4	28	1184	133,9	8,46	73	von Willebrand factor A domain-containing protein 3A GN=VWA3A PE=2 SV=3
Q8IY92	18	1834	199,9	6,06	73	Structure-specific endonuclease subunit SLX4 GN=SLX4 PE=1 SV=3
Q07889	16	1333	152,4	6,84	73	Son of sevenless homolog 1 GN=SOS1 PE=1 SV=1
Q14571	51	2701	307,9	6,43	73	Inositol 1,4,5-trisphosphate receptor type 2 GN=ITPR2 PE=1 SV=2
Q9P2N5	22	1060	118,6	9,19	72	RNA-binding protein 27 GN=RBM27 PE=1 SV=2
Q8TAT6	2	608	68,1	6,38	72	Nuclear protein localization protein 4 homolog GN=NPLOC4 PE=1 SV=3
A7E2F4	23	631	70,1	6,24	72	Golgin subfamily A member 8A GN=GOLGA8A PE=1 SV=3
Q02978	6	314	34,0	9,91	72	Mitochondrial 2-oxoglutarate/malate carrier protein GN=SLC25A11 PE=1 SV=3
P78345	27	283	31,8	9,92	72	Ribonuclease P protein subunit p38 GN=RPP38 PE=1 SV=2
Q6P587	5	224	24,8	7,39	72	Acylpyruvase FAHD1, mitochondrial GN=FAHD1 PE=1 SV=2
Q96C19	11	240	26,7	5,20	72	EF-hand domain-containing protein D2 GN=EFHD2 PE=1 SV=1
Q9BUP0	8	239	26,9	5,39	72	EF-hand domain-containing protein D1 GN=EFHD1 PE=1 SV=1
P24941	13	298	33,9	8,68	72	Cyclin-dependent kinase 2 GN=CDK2 PE=1 SV=2
Q96DF8	17	476	52,5	7,56	72	Protein DGCR14 GN=DGCR14 PE=1 SV=1
Q13576	34	1575	180,5	5,64	72	Ras GTPase-activating-like protein IQGAP2 GN=IQGAP2 PE=1 SV=4
O14986	10	540	61,0	6,86	72	Phosphatidylinositol-4-phosphate 5-kinase type-1 beta GN=PIP5K1B PE=1 SV=2
O15455	12	904	103,8	7,18	72	Toll-like receptor 3 GN=TLR3 PE=1 SV=1
P05412	9	331	35,7	8,76	72	Transcription factor AP-1 GN=JUN PE=1 SV=2
P28340	14	1107	123,6	7,03	72	DNA polymerase delta catalytic subunit GN=POLD1 PE=1 SV=2
P43268	11	484	53,9	5,52	72	ETS translocation variant 4 GN=ETV4 PE=1 SV=3
P60508	8	538	59,5	8,85	72	HERV-FRD_6p24.1 provirus ancestral Env polyprotein GN=ERVFRD-1 PE=1 SV=1
Q14142	15	442	49,7	7,87	72	Tripartite motif-containing protein 14 GN=TRIM14 PE=2 SV=2
Q75QN2	11	995	113,0	7,05	72	Integrator complex subunit 8 GN=INTS8 PE=1 SV=1
Q86Y91	12	864	94,2	8,79	72	Kinesin-like protein KIF18B GN=KIF18B PE=1 SV=3
Q9NTI5	31	1447	164,6	8,47	72	Sister chromatid cohesion protein PDS5 homolog B GN=PDS5B PE=1 SV=1
Q9Y261	10	457	48,3	8,69	72	Hepatocyte nuclear factor 3-beta GN=FOXA2 PE=1 SV=1
Q8IWE2	4	563	60,7	4,68	72	Protein NOXP20 GN=FAM114A1 PE=1 SV=2
Q00526	22	305	35,0	8,79	72	Cyclin-dependent kinase 3 GN=CDK3 PE=1 SV=1
P49773	6	126	13,8	6,95	71	Histidine triad nucleotide-binding protein 1 GN=HINT1 PE=1 SV=2
O95684	13	399	43,0	4,81	71	FGFR1 oncogene partner GN=FGFR1OP PE=1 SV=1
Q8NA56	13	475	55,0	5,71	71	Tetratricopeptide repeat protein 29 GN=TTC29 PE=2 SV=2
Q8NEY4	9	427	48,7	6,14	71	V-type proton ATPase subunit C 2 GN=ATP6V1C2 PE=2 SV=2
Q96BY7	22	2078	232,6	5,76	71	Autophagy-related protein 2 homolog B GN=ATG2B PE=1 SV=5
O60282	12	957	109,4	6,19	71	Kinesin heavy chain isoform 5C GN=KIF5C PE=1 SV=1
P16870	4	476	53,1	5,14	71	Carboxypeptidase E GN=CPE PE=1 SV=1
P55265	40	1226	136,0	8,65	71	Double-stranded RNA-specific adenosine deaminase GN=ADAR PE=1 SV=4
P25205	4	808	90,9	5,77	71	DNA replication licensing factor MCM3 GN=MCM3 PE=1 SV=3
Q7Z6P3	26	723	77,6	4,92	71	Ras-related protein Rab-44 GN=RAB44 PE=3 SV=3
Q9H3S7	29	1636	178,9	6,92	71	Tyrosine-protein phosphatase non-receptor type 23 GN=PTPN23 PE=1 SV=1
Q9ULH7	21	1088	118,1	6,28	71	MKL/myocardin-like protein 2 GN=MKL2 PE=1 SV=3
Q9BZG1	11	259	29,0	7,88	71	Ras-related protein Rab-34 GN=RAB34 PE=1 SV=1
Q9NYY8	21	710	81,4	8,05	71	FAST kinase domain-containing protein 2 GN=FASTKD2 PE=1 SV=1
Q9POL0	2	249	27,9	8,62	71	Vesicle-associated membrane protein-associated protein A GN=VAPA PE=1 SV=3
Q6YN16	6	418	45,4	7,99	70	Hydroxysteroid dehydrogenase-like protein 2 GN=HSDL2 PE=1 SV=1
P61157	3	418	47,3	5,88	70	Actin-related protein 3 GN=ACTR3 PE=1 SV=3
O14827	21	1237	140,7	7,53	70	Ras-specific guanine nucleotide-releasing factor 2 GN=RASGRF2 PE=1 SV=2
O95279	8	499	55,1	6,74	70	Potassium channel subfamily K member 5 GN=KCNK5 PE=1 SV=1
Q2TB10	7	664	75,2	9,47	70	Zinc finger protein 800 GN=ZNF800 PE=1 SV=1
Q8WYA1	9	636	70,8	7,37	70	Aryl hydrocarbon receptor nuclear translocator-like protein 2 GN=ARNTL2 PE=1 SV=2
Q96NY7	10	704	73,0	4,37	70	Chloride intracellular channel protein 6 GN=CLIC6 PE=2 SV=3
Q9BX26	23	1530	175,5	8,85	70	Synaptonemal complex protein 2 GN=SYCP2 PE=2 SV=2
Q9H6X5	7	657	71,3	5,78	70	Uncharacterized protein C19orf44 GN=C19orf44 PE=2 SV=1
Q9UJW0	5	460	52,3	7,34	70	Dynactin subunit 4 GN=DCTN4 PE=1 SV=1
Q15102	11	231	25,7	6,84	70	Platelet-activating factor acetylhydrolase IB subunit gamma GN=PAFAH1B3 PE=1 SV=
Q969F9	10	1004	113,7	6,43	70	Hermansky-Pudlak syndrome 3 protein GN=HPS3 PE=1 SV=1

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Q13641	8	420	46,0	6,83	70	Trophoblast glycoprotein GN=TPBG PE=1 SV=1
P11766	6	374	39,7	7,49	70	Alcohol dehydrogenase class-3 GN=ADH5 PE=1 SV=4
O95714	52	4834	526,9	6,28	70	E3 ubiquitin-protein ligase HERC2 GN=HERC2 PE=1 SV=2
P36543	5	226	26,1	8,00	70	V-type proton ATPase subunit E 1 GN=ATP6V1E1 PE=1 SV=1
P36404	4	184	20,9	6,34	70	ADP-ribosylation factor-like protein 2 GN=ARL2 PE=1 SV=4
Q92995	2	863	97,3	5,53	70	Ubiquitin carboxyl-terminal hydrolase 13 GN=USP13 PE=1 SV=2
P40939	7	763	82,9	9,04	70	Trifunctional enzyme subunit alpha, mitochondrial GN=HADHA PE=1 SV=2
Q99873	3	361	41,5	5,43	70	Protein arginine N-methyltransferase 1 GN=PRMT1 PE=1 SV=2
Q9N9G9	8	546	59,5	7,50	70	Aladin GN=AAAS PE=1 SV=1
Q62R08	28	3092	356,7	6,19	69	Dynein heavy chain 12, axonemal GN=DNAH12 PE=1 SV=2
P04183	3	234	25,5	8,51	69	Thymidine kinase, cytosolic GN=TK1 PE=1 SV=2
Q29R16	28	341	39,8	6,89	69	Calcium-binding protein 39 GN=CAB39 PE=2 SV=1
O60610	6	1272	141,3	5,41	69	Protein diaphanous homolog 1 GN=DIAPH1 PE=1 SV=2
Q9Y3U8	3	105	12,2	11,59	69	60S ribosomal protein L36 GN=RPL36 PE=1 SV=3
P00403	12	227	25,5	4,82	69	Cytochrome c oxidase subunit 2 GN=MT-CO2 PE=1 SV=1
P54845	22	237	25,9	7,88	69	Neural retina-specific leucine zipper protein GN=NRL PE=1 SV=1
Q9UNX3	18	145	17,2	10,55	69	60S ribosomal protein L26-like 1 GN=RPL26L1 PE=1 SV=1
Q13642	7	323	36,2	8,97	69	Four and a half LIM domains protein 1 GN=FHL1 PE=1 SV=4
P05109	8	93	10,8	7,03	69	Protein S100-A8 GN=S100A8 PE=1 SV=1
Q8NGA1	14	313	34,8	8,91	69	Olfactory receptor 1M1 GN=OR1M1 PE=2 SV=1
P09622	3	509	54,1	7,85	69	Dihydropyridyl dehydrogenase, mitochondrial GN=DLD PE=1 SV=2
P0C221	27	828	97,4	5,99	69	Uncharacterized protein C14orf38 GN=C14orf38 PE=2 SV=1
Q96PK2	123	5938	669,7	5,30	68	Microtubule-actin cross-linking factor 1, isoform 4 GN=MACF1 PE=1 SV=2
Q6PKX4	353	331	38,3	8,48	68	Docking protein 6 GN=DOK6 PE=1 SV=1
Q5T6S3	16	580	65,5	8,84	68	PHD finger protein 19 GN=PHF19 PE=1 SV=1
P85298	14	464	53,5	9,41	68	Rho GTPase-activating protein 8 GN=ARHGAP8 PE=1 SV=1
Q8IUE6	19	130	14,0	10,89	68	Histone H2A type 2-B GN=HIST2H2AB PE=1 SV=3
P29144	2	1249	138,3	6,32	68	Tripeptidyl-peptidase 2 GN=TPP2 PE=1 SV=4
Q5T1N1	41	836	92,8	6,81	68	Protein AKNAD1 GN=AKNAD1 PE=2 SV=3
Q8NGX9	25	317	35,8	8,44	68	Olfactory receptor 6P1 GN=OR6P1 PE=2 SV=1
Q9P2P1	31	1898	208,2	8,02	68	Protein NYNRIN GN=NYNRIN PE=1 SV=3
Q8TCT9	7	377	41,5	6,43	68	Minor histocompatibility antigen H13 GN=HM13 PE=1 SV=1
Q9UKV3	24	1341	151,8	6,43	68	Apoptotic chromatin condensation inducer in the nucleus GN=ACIN1 PE=1 SV=2
P49366	7	369	40,9	5,36	68	Deoxyhypusine synthase GN=DHPS PE=1 SV=1
P20839	19	514	55,4	6,90	68	Inosine-5'-monophosphate dehydrogenase 1 GN=IMPDH1 PE=1 SV=2
O75368	3	114	12,8	5,25	67	SH3 domain-binding glutamic acid-rich-like protein GN=SH3BGL PE=1 SV=1
O94817	16	140	15,1	5,10	67	Ubiquitin-like protein ATG12 GN=ATG12 PE=1 SV=1
Q8TDB6	20	740	83,5	8,06	67	E3 ubiquitin-protein ligase DTX3L GN=DTX3L PE=1 SV=1
P11532	70	3685	426,5	5,88	67	Dystrophin GN=DMD PE=1 SV=3
Q15717	7	326	36,1	9,17	67	ELAV-like protein 1 GN=ELAVL1 PE=1 SV=2
Q99575	58	1024	114,6	9,22	67	Ribonucleases P/MRP protein subunit POP1 GN=POP1 PE=1 SV=2
Q3LXA3	2	575	58,9	7,49	67	Bifunctional ATP-dependent dihydroxyacetone kinase/FAD-AMP lyase (cyclizing) GN=
Q725Q5	45	900	100,2	8,29	67	DNA polymerase nu GN=POLN PE=1 SV=2
Q5JWR5	34	2465	277,2	6,29	67	Protein dopey-1 GN=DOPEY1 PE=1 SV=1
Q68CQ4	34	756	87,0	5,88	67	Digestive organ expansion factor homolog GN=DIEXF PE=1 SV=2
Q9Y5K8	17	247	28,2	9,36	67	V-type proton ATPase subunit D GN=ATP6V1D PE=1 SV=1
Q2TAZ0	20	1938	212,7	5,88	67	Autophagy-related protein 2 homolog A GN=ATG2A PE=1 SV=3
Q96Q15	56	3661	410,2	6,46	67	Serine/threonine-protein kinase SMG1 GN=SMG1 PE=1 SV=3
Q15011	13	391	43,7	5,25	67	Homocysteine-responsive endoplasmic reticulum-resident ubiquitin-like domain me
Q8IVF5	32	1701	190,0	7,21	67	T-lymphoma invasion and metastasis-inducing protein 2 GN=TIAM2 PE=2 SV=4
P04070	3	461	52,0	6,28	66	Vitamin K-dependent protein C GN=PROC PE=1 SV=1
Q14832	13	879	98,8	7,66	66	Metabotropic glutamate receptor 3 GN=GRM3 PE=1 SV=2
Q68DA7	9	1419	157,5	8,44	66	Formin-1 GN=FMN1 PE=1 SV=3
P45974	7	858	95,7	5,03	66	Ubiquitin carboxyl-terminal hydrolase 5 GN=USP5 PE=1 SV=2
Q9BPX5	3	153	16,9	6,60	66	Actin-related protein 2/3 complex subunit 5-like protein GN=ARPC5L PE=1 SV=1
Q86SJ6	6	1040	113,8	4,56	66	Desmoglein-4 GN=DSG4 PE=1 SV=1
A6NC57	36	917	106,4	6,67	66	Ankyrin repeat domain-containing protein 62 GN=ANKRD62 PE=2 SV=4
Q07864	10	2286	261,4	6,39	66	DNA polymerase epsilon catalytic subunit A GN=POLE PE=1 SV=5
Q9H3U1	33	944	103,0	6,07	66	Protein unc-45 homolog A GN=UNC45A PE=1 SV=1
A6NKT7	61	1758	197,4	6,33	66	RanBP2-like and GRIP domain-containing protein 3 GN=RGPD3 PE=2 SV=2
O14715	37	1765	198,9	6,49	66	RANBP2-like and GRIP domain-containing protein 8 GN=RGPD8 PE=1 SV=2
P49792	49	3224	358,0	6,20	66	E3 SUMO-protein ligase RanBP2 GN=RANBP2 PE=1 SV=2
Q68DN6	48	1748	196,5	6,10	66	RANBP2-like and GRIP domain-containing protein 1/2 GN=RGPD1 PE=1 SV=2
Q7Z3J3	58	1758	197,2	6,27	66	RanBP2-like and GRIP domain-containing protein 4 GN=RGPD4 PE=2 SV=3
Q99666	36	1765	198,8	6,42	66	RANBP2-like and GRIP domain-containing protein 5/6 GN=RGPD5 PE=1 SV=3

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Q86XX4	34	4007	442,6	5,59	66	Extracellular matrix protein FRAS1 GN=FRAS1 PE=2 SV=1
Q1KMD3	5	747	85,1	4,91	66	Heterogeneous nuclear ribonucleoprotein U-like protein 2 GN=HNRNPUL2 PE=1 SV=1
Q9UBQ0	3	182	20,5	6,79	66	Vacuolar protein sorting-associated protein 29 GN=VPS29 PE=1 SV=1
P51956	14	506	57,7	7,17	66	Serine/threonine-protein kinase Nek3 GN=NEK3 PE=1 SV=2
Q9H0K1	32	926	103,8	6,02	66	Serine/threonine-protein kinase SIK2 GN=SIK2 PE=1 SV=1
Q13637	10	225	25,0	6,54	66	Ras-related protein Rab-32 GN=RAB32 PE=1 SV=3
P81605	12	110	11,3	6,54	65	Dermcidin GN=DCD PE=1 SV=2
P61758	3	197	22,6	7,11	65	Prefoldin subunit 3 GN=VBP1 PE=1 SV=3
P43246	3	934	104,7	5,77	65	DNA mismatch repair protein Msh2 GN=MSH2 PE=1 SV=1
Q9UQ90	8	795	88,2	8,69	65	Paraplegin GN=SPG7 PE=1 SV=2
Q08257	9	329	35,2	8,44	65	Quinone oxidoreductase GN=CRYZ PE=1 SV=1
P48751	23	1232	135,7	6,44	65	Anion exchange protein 3 GN=SLC4A3 PE=1 SV=2
Q93009	17	1102	128,2	5,55	65	Ubiquitin carboxyl-terminal hydrolase 7 GN=USP7 PE=1 SV=2
Q9BSL1	26	405	45,3	4,92	65	Ubiquitin-associated domain-containing protein 1 GN=UBAC1 PE=1 SV=1
Q9N2K0	9	584	64,3	8,25	65	HERV-H_2q24.3 provirus ancestral Env polyprotein PE=2 SV=1
Q5VV41	14	709	80,1	7,36	65	Rho guanine nucleotide exchange factor 16 GN=ARHGEF16 PE=1 SV=1
Q9C0B7	12	1094	120,7	6,11	65	Transmembrane and coiled-coil domain-containing protein 7 GN=TMCO7 PE=2 SV=2
P16591	13	822	94,6	7,14	65	Tyrosine-protein kinase Fer GN=FER PE=1 SV=2
Q9ULM3	10	1422	150,7	8,98	65	YEATS domain-containing protein 2 GN=YEATS2 PE=1 SV=2
Q9NS73	22	344	39,3	7,24	64	MAP3K12-binding inhibitory protein 1 GN=MBIP PE=1 SV=2
P60229	11	445	52,2	6,04	64	Eukaryotic translation initiation factor 3 subunit E GN=EIF3e PE=1 SV=1
Q9HCM7	40	1045	110,8	9,67	64	Fibrosin-1-like protein GN=FBRSL1 PE=1 SV=4
Q9H9A6	29	602	68,2	6,43	64	Leucine-rich repeat-containing protein 40 GN=LRRC40 PE=1 SV=1
P53384	7	320	34,5	5,33	64	Cytosolic Fe-S cluster assembly factor NUBP1 GN=NUBP1 PE=1 SV=2
O43615	2	452	51,3	8,32	64	Mitochondrial import inner membrane translocase subunit TIM44 GN=TIMM44 PE=1 SV=1
Q9UBQ7	2	328	35,6	7,39	63	Glyoxylate reductase/hydroxypyruvate reductase GN=GRHPR PE=1 SV=1
Q8IVV2	11	1947	221,8	5,38	63	Lipoxygenase homology domain-containing protein 1 GN=LOXHD1 PE=2 SV=3
Q15024	5	291	31,8	5,19	63	Exosome complex component RRP42 GN=EXOSC7 PE=1 SV=3
P49895	8	249	28,9	8,69	63	Type I iodothyronine deiodinase GN=DIO1 PE=2 SV=3
Q9UBV8	4	284	30,4	6,54	63	Peflin GN=PEF1 PE=1 SV=1
Q13867	3	455	52,5	6,27	63	Bleomycin hydrolase GN=BLMH PE=1 SV=1
Q9UIA9	11	1087	123,8	6,32	63	Exportin-7 GN=XPO7 PE=1 SV=3
Q9H582	23	1327	149,5	8,16	63	Zinc finger protein 644 GN=ZNF644 PE=1 SV=2
Q13526	2	163	18,2	8,82	62	Peptidyl-prolyl cis-trans isomerase NIMA-interacting 1 GN=PIN1 PE=1 SV=1
Q9HC10	22	1997	226,6	5,69	62	Otoferrin GN=OTOF PE=1 SV=3
Q3SY84	12	523	57,3	6,61	62	Keratin, type II cytoskeletal 71 GN=KRT71 PE=1 SV=3
P46060	2	587	63,5	4,68	62	Ran GTPase-activating protein 1 GN=RANGAP1 PE=1 SV=1
P22307	6	547	59,0	6,89	62	Non-specific lipid-transfer protein GN=SCP2 PE=1 SV=2
Q8WVX9	27	515	59,3	9,17	61	Fatty acyl-CoA reductase 1 GN=FAR1 PE=1 SV=1
Q9H936	14	323	34,4	9,29	61	Mitochondrial glutamate carrier 1 GN=SLC25A22 PE=1 SV=1
O00571	2	662	73,2	7,18	61	ATP-dependent RNA helicase DDX3X GN=DDX3X PE=1 SV=3
P56385	2	69	7,9	9,35	61	ATP synthase subunit e, mitochondrial GN=ATP5I PE=1 SV=2
Q3ZCA2	5	227	26,2	8,82	61	Cleavage and polyadenylation specificity factor subunit 5 GN=NUDT21 PE=2 SV=1
Q9NZL4	2	362	39,4	5,21	61	Hsp70-binding protein 1 GN=HSPBP1 PE=1 SV=1
P62196	4	406	45,6	7,55	60	26S protease regulatory subunit 8 GN=Psmc5 PE=1 SV=1
Q9NQR4	4	276	30,6	7,21	60	Omega-amidase NIT2 GN=NIT2 PE=1 SV=1
Q15797	38	465	52,2	7,31	60	Mothers against decapentaplegic homolog 1 GN=SMAD1 PE=1 SV=1
Q8NEJ9	22	315	35,9	9,57	60	Neuroguidin GN=NGDN PE=1 SV=1
B2RSH2	3	354	40,3	5,97	60	Guanine nucleotide-binding protein G(i) subunit alpha-1 GN=Gnai1 PE=2 SV=1
P09471	5	354	40,0	5,53	60	Guanine nucleotide-binding protein G(o) subunit alpha GN=GNAO1 PE=1 SV=4
Q03113	4	381	44,3	9,83	60	Guanine nucleotide-binding protein subunit alpha-12 GN=GNA12 PE=1 SV=4
Q14344	9	377	44,0	8,00	60	Guanine nucleotide-binding protein subunit alpha-13 GN=GNA13 PE=1 SV=2
O95782	3	977	107,5	7,03	60	AP-2 complex subunit alpha-1 GN=AP2A1 PE=1 SV=3
Q38SD2	22	2015	225,2	6,68	59	Leucine-rich repeat serine/threonine-protein kinase 1 GN=LRRK1 PE=1 SV=3
Q9Y2Z9	19	468	50,8	7,30	59	Ubiquinone biosynthesis monooxygenase COQ6 GN=COQ6 PE=1 SV=2
Q99729	5	332	36,2	8,21	59	Heterogeneous nuclear ribonucleoprotein A/B GN=HNRNPAB PE=1 SV=2
Q9NQC3	5	1192	129,9	4,50	59	Reticulon-4 GN=RTN4 PE=1 SV=2
Q5JQF8	3	200	22,8	9,10	59	Polyadenylate-binding protein 1-like 2 GN=PABPC1L2A PE=1 SV=1
O95239	109	1232	139,8	6,27	59	Chromosome-associated kinesin KIF4A GN=KIF4A PE=1 SV=3
Q2VIQ3	107	1234	139,9	6,18	59	Chromosome-associated kinesin KIF4B GN=KIF4B PE=1 SV=2
P10301	9	218	23,5	6,93	58	Ras-related protein R-Ras GN=RRAS PE=1 SV=1
P04844	8	631	69,2	5,69	58	Dolichyl-diphosphooligosaccharide--protein glycosyltransferase subunit 2 GN=RPN2
Q13523	31	1007	116,9	10,26	58	Serine/threonine-protein kinase PRP4 homolog GN=PRPF4B PE=1 SV=3
P10606	4	129	13,7	8,81	57	Cytochrome c oxidase subunit 5B, mitochondrial GN=COX5B PE=1 SV=2
Q9UBT2	3	640	71,2	5,29	57	SUMO-activating enzyme subunit 2 GN=UBA2 PE=1 SV=2

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P24844	18	172	19,8	4,92	57	Myosin regulatory light polypeptide 9 GN=MYL9 PE=1 SV=4
Q9Y5L0	5	923	104,1	5,57	57	Transportin-3 GN=TNPO3 PE=1 SV=3
Q96K30	6	269	28,6	11,08	56	RBPJ-interacting and tubulin-associated protein GN=RITA PE=1 SV=1
Q1RMH8	1	162	18,8	8,66	56	Sorting nexin-3 GN=SNX3 PE=2 SV=3
P30520	8	456	50,1	6,55	55	Adenylosuccinate synthetase isozyme 2 GN=ADSS PE=1 SV=3
Q8IYD1	14	628	68,8	5,43	54	Eukaryotic peptide chain release factor GTP-binding subunit ERF3B GN=GSPT2 PE=1 SV=1
Q9Y4L1	10	999	111,3	5,22	54	Hypoxia up-regulated protein 1 GN=HYOU1 PE=1 SV=1
P52788	5	366	41,2	5,02	54	Spermine synthase GN=SMS PE=1 SV=2
Q9P0S9	1	112	11,6	9,88	54	Transmembrane protein 14C GN=TMEM14C PE=1 SV=1
P25325	2	297	33,2	6,60	54	3-mercaptopyruvate sulfurtransferase GN=MPST PE=1 SV=3
O95399	4	124	14,3	7,80	53	Urotensin-2 GN=UTS2 PE=1 SV=1
Q99456	24	494	53,5	4,78	52	Keratin, type I cytoskeletal 12 GN=KRT12 PE=1 SV=1
O43432	10	1585	176,5	5,38	52	Eukaryotic translation initiation factor 4 gamma 3 GN=EIF4G3 PE=1 SV=2
P21399	5	889	98,3	6,68	52	Cytoplasmic aconitate hydratase GN=ACO1 PE=1 SV=3
Q96M42	8	142	15,2	8,00	52	Putative uncharacterized protein encoded by LINC00479 GN=LINC00479 PE=5 SV=2
P30566	3	484	54,9	7,11	52	Adenylosuccinate lyase GN=ADSL PE=1 SV=2
P21912	26	280	31,6	8,76	51	Succinate dehydrogenase [ubiquinone] iron-sulfur subunit, mitochondrial GN=SDHB
O60437	44	1756	204,6	5,60	51	Periplakin GN=PPL PE=1 SV=4
P63280	3	158	18,0	8,66	51	SUMO-conjugating enzyme UBC9 GN=Ube2i PE=1 SV=1
P11217	10	842	97,0	7,03	51	Glycogen phosphorylase, muscle form GN=PYGM PE=1 SV=6
P02489	3	173	19,9	6,20	50	Alpha-crystallin A chain GN=CRYAA PE=1 SV=2
P43320	1	205	23,4	7,01	50	Beta-crystallin B2 GN=CRYBB2 PE=1 SV=2
O60925	4	122	14,2	6,81	49	Prefoldin subunit 1 GN=PFDN1 PE=1 SV=2
P11172	3	480	52,2	7,24	49	Uridine 5'-monophosphate synthase GN=UMPS PE=1 SV=1
Q9UBQ5	6	218	25,0	4,93	49	Eukaryotic translation initiation factor 3 subunit K GN=EIF3K PE=1 SV=1
Q62WH5	9	1172	133,1	6,80	48	Serine/threonine-protein kinase Nek10 GN=NEK10 PE=2 SV=2
P16615	4	1042	114,7	5,34	48	Sarcoplasmic/endoplasmic reticulum calcium ATPase 2 GN=ATP2A2 PE=1 SV=1
P02545	1	664	74,1	7,02	48	Prelamin-A/C GN=LMNA PE=1 SV=1
Q9UNH7	4	406	46,6	6,16	48	Sorting nexin-6 GN=SNX6 PE=1 SV=1
P10768	5	282	31,4	7,02	48	S-formylglutathione hydrolase GN=ESD PE=1 SV=2
P16083	1	231	25,9	6,29	48	Ribosyl-dihydroxynicotinamide dehydrogenase [quinone] GN=NQO2 PE=1 SV=5
Q9Y5F7	2	938	101,2	5,35	48	Protocadherin gamma-C4 GN=PCDHG4 PE=1 SV=1
P49591	9	514	58,7	6,43	48	Seryl-tRNA synthetase, cytoplasmic GN=SARS PE=1 SV=3
Q9H4M9	4	534	60,6	6,83	47	EH domain-containing protein 1 GN=EHD1 PE=1 SV=2
Q9NTN9	3	838	91,4	7,84	47	Semaphorin-4G GN=SEMA4G PE=2 SV=1
P68037	4	154	17,9	8,51	47	Ubiquitin-conjugating enzyme E2 L3 GN=Ube2l3 PE=2 SV=1
O60884	2	412	45,7	6,48	47	DnaJ homolog subfamily A member 2 GN=DNAJA2 PE=1 SV=1
O94979	50	1220	132,9	6,89	47	Protein transport protein Sec31A GN=SEC31A PE=1 SV=3
Q14CN4	9	511	55,8	6,89	47	Keratin, type II cytoskeletal 72 GN=KRT72 PE=1 SV=2
Q9BU89	2	302	32,9	4,83	47	Deoxyhypusine hydroxylase GN=DOHH PE=1 SV=1
P52272	2	730	77,5	8,70	46	Heterogeneous nuclear ribonucleoprotein M GN=HNRNPM PE=1 SV=3
O14966	17	203	23,1	7,18	45	Ras-related protein Rab-7L1 GN=RAB7L1 PE=1 SV=1
P35462	5	400	44,2	8,94	45	D(3) dopamine receptor GN=DRD3 PE=1 SV=2
P04179	2	222	24,7	8,25	45	Superoxide dismutase [Mn], mitochondrial GN=SOD2 PE=1 SV=2
Q16222	2	522	58,7	6,33	45	UDP-N-acetylhexosamine pyrophosphorylase GN=UAP1 PE=1 SV=3
Q7L014	25	1031	117,3	9,29	45	Probable ATP-dependent RNA helicase DDX46 GN=DDX46 PE=1 SV=2
Q13257	2	205	23,5	5,08	45	Mitotic spindle assembly checkpoint protein MAD2A GN=MAD2L1 PE=1 SV=1
Q16650	14	682	74,0	7,33	44	T-box brain protein 1 GN=TBR1 PE=1 SV=1
P51665	1	324	37,0	6,77	44	26S proteasome non-ATPase regulatory subunit 7 GN=PSMD7 PE=1 SV=2
Q96PD4	4	163	18,0	8,90	44	Interleukin-17F GN=IL17F PE=1 SV=3
P62874	1	340	37,4	6,00	44	Guanine nucleotide-binding protein G(I)/G(S)/G(T) subunit beta-1 GN=Gnb1 PE=1 SV=1
Q9C0K3	1	210	23,7	5,58	44	Actin-related protein 3C GN=ACTR3C PE=2 SV=1
A6NNC1	15	897	94,0	10,26	44	Putative POM121-like protein 1-like PE=5 SV=3
P51531	9	1590	181,2	7,20	44	Probable global transcription activator SNF2L2 GN=SMARCA2 PE=1 SV=2
O15523	1	660	73,1	7,55	44	ATP-dependent RNA helicase DDX3Y GN=DDX3Y PE=1 SV=2
Q13148	3	414	44,7	6,19	43	TAR DNA-binding protein 43 GN=TARDBP PE=1 SV=1
P47929	3	136	15,1	7,62	43	Galectin-7 GN=LGALS7 PE=1 SV=2
Q70J99	8	1090	123,2	6,65	42	Protein unc-13 homolog D GN=UNC13D PE=1 SV=1
P57729	2	211	23,7	7,77	42	Ras-related protein Rab-38 GN=RAB38 PE=1 SV=1
Q6P1M9	13	558	62,3	8,73	42	Armadillo repeat-containing X-linked protein 5 GN=ARMCX5 PE=2 SV=1
P00387	1	301	34,2	7,59	42	NADH-cytochrome b5 reductase 3 GN=CYB5R3 PE=1 SV=3
Q9UHD9	4	624	65,7	5,22	41	Ubiquilin-2 GN=UBQLN2 PE=1 SV=2
Q86TJ2	12	420	48,4	7,83	41	Transcriptional adapter 2-beta GN=TADA2B PE=1 SV=2
Q9H7B2	1	306	35,6	9,99	41	Ribosome production factor 2 homolog GN=RPF2 PE=1 SV=2
P51451	9	505	57,7	7,87	41	Tyrosine-protein kinase Blk GN=BLK PE=1 SV=3

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P61924	4	177	20,2	4,81	41	Coatomer subunit zeta-1 GN=Copz1 PE=2 SV=1
Q9BWW9	4	433	47,0	9,31	41	Apolipoprotein L5 GN=APOL5 PE=1 SV=1
P23786	3	658	73,7	8,18	41	Carnitine O-palmitoyltransferase 2, mitochondrial GN=CPT2 PE=1 SV=2
Q8TDN6	3	353	41,4	9,92	40	Ribosome biogenesis protein BRX1 homolog GN=BRX1 PE=1 SV=2
Q8TEX9	2	1081	118,6	4,96	40	Importin-4 GN=IPO4 PE=1 SV=2
Q16822	3	640	70,7	7,62	40	Phosphoenolpyruvate carboxykinase [GTP], mitochondrial GN=PCK2 PE=1 SV=3
Q53558	10	311	33,7	9,61	40	Transmembrane protein 177 GN=TMEM177 PE=2 SV=1
Q9POU1	4	55	6,2	10,29	40	Mitochondrial import receptor subunit TOM7 homolog GN=TOMM7 PE=1 SV=1
Q8NBT2	3	197	22,4	4,70	40	Kinetochore protein Spc24 GN=SPC24 PE=1 SV=1
P06702	6	114	13,2	6,13	40	Protein S100-A9 GN=S100A9 PE=1 SV=1
Q96RT7	6	1819	200,4	6,32	40	Gamma-tubulin complex component 6 GN=TUBGCP6 PE=1 SV=3
Q14980	15	2115	238,1	5,78	40	Nuclear mitotic apparatus protein 1 GN=NUMA1 PE=1 SV=2
P29992	12	359	42,1	5,69	40	Guanine nucleotide-binding protein subunit alpha-11 GN=GNA11 PE=1 SV=2
P35251	19	1148	128,2	9,36	40	Replication factor C subunit 1 GN=RFC1 PE=1 SV=4
P11908	5	318	34,7	6,61	39	Ribose-phosphate pyrophosphokinase 2 GN=PRPS2 PE=1 SV=2
P21108	5	318	34,8	6,35	39	Ribose-phosphate pyrophosphokinase 3 GN=PRPS1L1 PE=1 SV=2
O60218	5	316	36,0	7,84	39	Aldo-keto reductase family 1 member B10 GN=AKR1B10 PE=1 SV=2
Q8WYA6	38	563	65,1	5,05	39	Beta-catenin-like protein 1 GN=CTNBL1 PE=1 SV=1
Q9NXG2	6	353	39,3	7,88	39	THUMP domain-containing protein 1 GN=THUMP1 PE=1 SV=2
P49448	3	558	61,4	8,46	39	Glutamate dehydrogenase 2, mitochondrial GN=GLUD2 PE=1 SV=2
Q14558	3	356	39,4	7,20	39	Phosphoribosyl pyrophosphate synthase-associated protein 1 GN=PRPSAP1 PE=1 SV=
O00273	18	331	36,5	4,79	39	DNA fragmentation factor subunit alpha GN=DFFA PE=1 SV=1
Q7L155	8	443	52,0	9,38	39	Carbohydrate sulfotransferase 9 GN=CHST9 PE=2 SV=2
P33947	5	212	24,4	8,72	39	ER lumen protein retaining receptor 2 GN=KDEL2 PE=1 SV=1
P12524	1	364	40,3	5,66	39	Protein L-Myc-1 GN=MYCL1 PE=1 SV=2
O14764	8	452	50,7	8,54	39	Gamma-aminobutyric acid receptor subunit delta GN=GABRD PE=1 SV=2
Q5D0E6	6	543	59,3	7,42	39	DALR anticodon-binding domain-containing protein 3 GN=DALRD3 PE=2 SV=2
Q8IVF2	48	5795	616,2	5,36	39	Protein AHNAK2 GN=AHNAK2 PE=1 SV=2
P62315	2	119	13,3	11,56	39	Small nuclear ribonucleoprotein Sm D1 GN=Snrdp1 PE=2 SV=1
Q14C86	11	1478	164,9	5,22	39	GTPase-activating protein and VPS9 domain-containing protein 1 GN=GAPVD1 PE=1 S
Q9NP72	11	206	23,0	5,24	39	Ras-related protein Rab-18 GN=RAB18 PE=1 SV=1
Q5TZA2	6	2017	228,4	5,50	39	Rootletin GN=CROCC PE=1 SV=1
Q9HD20	26	1204	132,9	8,13	39	Probable cation-transporting ATPase 13A1 GN=ATP13A1 PE=1 SV=2
C9J158	8	452	52,5	8,37	39	Tripartite motif-containing protein 49-like protein 1 GN=TRIM49L1 PE=2 SV=1
POCI26	3	452	52,9	7,46	39	Tripartite motif-containing protein 49-like protein 2 GN=TRIM49L2 PE=2 SV=1
P40145	7	1251	140,0	6,99	39	Adenylate cyclase type 8 GN=ADCY8 PE=1 SV=1
Q15035	2	370	43,3	9,22	39	Translocating chain-associated membrane protein 2 GN=TRAM2 PE=1 SV=1
Q63HK5	17	1081	118,5	7,25	39	Teashirt homolog 3 GN=TSHZ3 PE=1 SV=2
Q62RF7	2	136	15,5	10,23	39	Putative zinc finger protein 818 GN=ZNF818P PE=5 SV=1
Q6ZS26	10	1077	117,8	7,06	39	Teashirt homolog 1 GN=TSHZ1 PE=2 SV=2
Q86YA3	18	1062	119,3	5,02	39	Uncharacterized protein C4orf21 GN=C4orf21 PE=1 SV=2
Q9NRE2	3	1034	114,9	7,83	39	Teashirt homolog 2 GN=TSHZ2 PE=1 SV=3
P35637	8	526	53,4	9,36	39	RNA-binding protein FUS GN=FUS PE=1 SV=1
Q86SQ0	12	1253	142,1	7,43	39	Pleckstrin homology-like domain family B member 2 GN=PHLDB2 PE=1 SV=2
Q13085	6	2346	265,4	6,37	39	Acetyl-CoA carboxylase 1 GN=ACACA PE=1 SV=2
Q9NUQ9	5	324	36,7	6,06	39	Protein FAM49B GN=FAM49B PE=1 SV=1
Q86SE5	14	291	32,3	7,93	39	RNA-binding Raly-like protein GN=RALYL PE=2 SV=2
Q9UKM9	7	306	32,4	9,17	39	RNA-binding protein Raly GN=RALY PE=1 SV=1
Q9P1Q0	4	977	110,5	6,55	38	Vacuolar protein sorting-associated protein 54 GN=VPS54 PE=1 SV=2
P40261	1	264	29,6	5,74	38	Nicotinamide N-methyltransferase GN=NNMT PE=1 SV=1
Q9NZ52	3	723	78,3	5,58	38	ADP-ribosylation factor-binding protein GGA3 GN=GGA3 PE=1 SV=1
Q9UJY5	1	639	70,3	5,29	38	ADP-ribosylation factor-binding protein GGA1 GN=GGA1 PE=1 SV=1
Q68DK2	6	2539	284,4	6,39	38	Zinc finger FYVE domain-containing protein 26 GN=ZFYVE26 PE=1 SV=3
Q96NW7	7	1537	172,5	6,81	38	Leucine-rich repeat-containing protein 7 GN=LRRC7 PE=1 SV=1
Q5U649	22	245	27,6	7,96	38	Uncharacterized protein C12orf60 GN=C12orf60 PE=2 SV=2
Q15418	7	735	82,7	7,83	38	Ribosomal protein S6 kinase alpha-1 GN=RPS6KA1 PE=1 SV=2
Q5JT29	17	985	107,3	6,27	38	Alanine--tRNA ligase, mitochondrial GN=AARS2 PE=1 SV=1
Q13144	2	721	80,3	5,08	38	Translation initiation factor eIF-2B subunit epsilon GN=EIF2B5 PE=1 SV=3
Q8IUZ0	19	686	78,8	7,87	38	Leucine-rich repeat-containing protein 49 GN=LRRC49 PE=2 SV=2
Q5HYI8	1	236	26,4	7,11	38	Rab-like protein 3 GN=RABL3 PE=1 SV=1
P49643	1	509	58,8	7,91	38	DNA primase large subunit GN=PRIM2 PE=1 SV=2
Q8ND04	8	991	109,6	7,68	38	Protein SMG8 GN=SMG8 PE=1 SV=1
Q9NVF7	4	368	41,1	9,55	38	F-box only protein 28 GN=FBXO28 PE=1 SV=1
Q9Y2G5	31	429	49,9	6,60	38	GDP-fucose protein O-fucosyltransferase 2 GN=POFUT2 PE=2 SV=3
Q9NRA8	33	985	108,1	8,32	38	Eukaryotic translation initiation factor 4E transporter GN=EIF4ENIF1 PE=1 SV=2

Appendices

Q9Y5C1	8	460	53,6	6,70	38	Angiopoietin-related protein 3 GN=ANGPTL3 PE=1 SV=1
P36639	12	197	22,5	5,27	38	7,8-dihydro-8-oxoguanine triphosphatase GN=NUDT1 PE=1 SV=3
Q9H4A6	5	298	33,8	6,44	37	Golgi phosphoprotein 3 GN=GOLPH3 PE=1 SV=1
P61202	4	443	51,6	5,53	37	COP9 signalosome complex subunit 2 GN=Cops2 PE=1 SV=1
Q63HQ2	11	1017	111,2	7,40	37	Pikachurin GN=EGFLAM PE=1 SV=2
Q93033	8	1021	115,0	6,96	37	Immunoglobulin superfamily member 2 GN=CD101 PE=1 SV=2
Q02809	1	727	83,5	6,95	37	Procollagen-lysine,2-oxoglutarate 5-dioxygenase 1 GN=PLOD1 PE=1 SV=2
Q9POX4	8	2223	244,9	6,54	37	Voltage-dependent T-type calcium channel subunit alpha-1l GN=CACNA1l PE=1 SV=1
Q5E9F9	2	433	48,6	5,95	37	26S protease regulatory subunit 7 GN=PSMC2 PE=2 SV=3
Q14498	2	530	59,3	10,10	37	RNA-binding protein 39 GN=RBM39 PE=1 SV=2
P83105	23	476	50,9	8,02	37	Serine protease HTRA4 GN=HTRA4 PE=2 SV=1
Q00266	16	395	43,6	6,29	37	S-adenosylmethionine synthase isoform type-1 GN=MAT1A PE=1 SV=2
Q674X7	21	775	86,3	7,03	37	Kazrin GN=KAZN PE=1 SV=2
Q8WUY3	38	3088	340,4	4,45	37	Protein prune homolog 2 GN=PRUNE2 PE=1 SV=3
O60344	14	883	99,7	5,10	37	Endothelin-converting enzyme 2 GN=ECE2 PE=1 SV=4
P05937	25	261	30,0	4,83	37	Calbindin GN=CALB1 PE=1 SV=2
P23508	23	829	93,0	5,52	37	Colorectal mutant cancer protein GN=MCC PE=1 SV=2
Q05655	21	676	77,5	7,75	37	Protein kinase C delta type GN=PRKCD PE=1 SV=2
Q6JEL2	16	608	68,9	5,68	37	Kelch-like protein 10 GN=KLHL10 PE=2 SV=1
Q9Y274	15	331	38,2	9,00	37	Type 2 lactosamine alpha-2,3-sialyltransferase GN=ST3GAL6 PE=1 SV=1
Q14012	10	370	41,3	5,29	37	Calcium/calmodulin-dependent protein kinase type 1 GN=CAMK1 PE=1 SV=1
O95626	5	131	14,8	5,02	37	Acidic leucine-rich nuclear phosphoprotein 32 family member D GN=ANP32D PE=1 SV=1
P00846	2	226	24,8	10,10	37	ATP synthase subunit a GN=MT-ATP6 PE=1 SV=1
O43868	2	658	71,9	7,94	37	Sodium/nucleoside cotransporter 2 GN=SLC28A2 PE=2 SV=2
Q08AD1	12	1489	168,0	6,80	37	Calmodulin-regulated spectrin-associated protein 2 GN=CAMSAP2 PE=1 SV=3
Q10713	20	525	58,2	6,92	37	Mitochondrial-processing peptidase subunit alpha GN=PMPCA PE=1 SV=2
P35244	2	121	13,6	5,08	37	Replication protein A 14 kDa subunit GN=RPA3 PE=1 SV=1
Q5VT25	29	1732	197,2	6,58	37	Serine/threonine-protein kinase MRCK alpha GN=CDC42BPA PE=1 SV=1
Q86SQ7	13	713	82,6	5,81	37	Serologically defined colon cancer antigen 8 GN=SDCCAG8 PE=1 SV=1
Q8IV03	11	231	24,6	5,14	37	Leucine rich adaptor protein 1-like GN=LURAP1L PE=2 SV=2
Q9P2R7	22	463	50,3	7,42	37	Succinyl-CoA ligase [ADP-forming] subunit beta, mitochondrial GN=SUCLA2 PE=1 SV=1
A6NDN3	19	693	79,9	5,50	37	Putative golgin subfamily A member 6B GN=GOLGA6B PE=5 SV=3
Q9BT78	9	406	46,2	5,83	37	COP9 signalosome complex subunit 4 GN=COPS4 PE=1 SV=1
P13473	3	410	44,9	5,63	37	Lysosome-associated membrane glycoprotein 2 GN=LAMP2 PE=1 SV=2
Q8WVT3	8	735	79,3	4,91	37	Trafficking protein particle complex subunit 12 GN=TRAPP12 PE=1 SV=3
O14530	5	226	26,5	5,88	36	Thioredoxin domain-containing protein 9 GN=TXNDC9 PE=1 SV=2
Q9Y310	21	505	55,2	7,23	36	tRNA-splicing ligase RtcB homolog GN=C22orf28 PE=1 SV=1
P19525	4	551	62,1	8,40	36	Interferon-induced, double-stranded RNA-activated protein kinase GN=EIF2AK2 PE=1 SV=1
Q9NQW7	2	623	69,9	5,67	36	Xaa-Pro aminopeptidase 1 GN=XPNPEP1 PE=1 SV=3
Q13243	19	272	31,2	11,59	36	Serine/arginine-rich splicing factor 5 GN=SRSF5 PE=1 SV=1
Q16533	24	368	43,0	9,51	36	snRNA-activating protein complex subunit 1 GN=SNAPC1 PE=1 SV=1
Q6P5Q4	4	547	61,6	5,78	36	Leiomodin-2 GN=LMOD2 PE=2 SV=2
O94760	5	285	31,1	5,81	36	N(G),N(G)-dimethylarginine dimethylaminohydrolase 1 GN=DDAH1 PE=1 SV=3
O00483	2	81	9,4	9,38	36	NADH dehydrogenase [ubiquinone] 1 alpha subcomplex subunit 4 GN=NDUFA4 PE=1 SV=1
A8K979	9	691	77,4	8,90	36	ERI1 exoribonuclease 2 GN=ERI2 PE=2 SV=2
A8MTJ3	2	354	40,3	6,01	36	Guanine nucleotide-binding protein G(t) subunit alpha-3 GN=GNAT3 PE=2 SV=2
P08754	2	354	40,5	5,69	36	Guanine nucleotide-binding protein G(k) subunit alpha GN=GNAI3 PE=1 SV=3
P11488	3	350	40,0	5,62	36	Guanine nucleotide-binding protein G(t) subunit alpha-1 GN=GNAT1 PE=1 SV=5
P38405	6	381	44,3	6,65	36	Guanine nucleotide-binding protein G(olf) subunit alpha GN=GNAL PE=1 SV=1
Q5JWF2	7	1037	111,0	5,03	36	Guanine nucleotide-binding protein G(s) subunit alpha isoforms XLas GN=GNAS PE=1 SV=1
P12273	1	146	16,6	8,05	36	Prolactin-inducible protein GN=PIP PE=1 SV=1
P56199	4	1179	130,8	6,29	36	Integrin alpha-1 GN=ITGA1 PE=1 SV=2
Q9NYL9	1	352	39,6	5,19	36	Tropomodulin-3 GN=TMOD3 PE=1 SV=1
Q7L576	6	1253	145,1	6,90	36	Cytoplasmic FMR1-interacting protein 1 GN=CYFIP1 PE=1 SV=1
Q12797	5	758	85,8	5,01	36	Aspartyl/asparaginyl beta-hydroxylase GN=ASPH PE=1 SV=3
Q96CN4	38	794	91,3	5,34	36	EVI5-like protein GN=EVI5L PE=1 SV=1
P62911	2	135	15,8	11,33	36	60S ribosomal protein L32 GN=Rpl32 PE=2 SV=2
Q96HE7	1	468	54,4	5,68	36	ERO1-like protein alpha GN=ERO1L PE=1 SV=2
O35900	1	95	10,8	6,52	36	U6 snRNA-associated Sm-like protein LSm2 GN=Lsm2 PE=2 SV=1
Q96AY3	3	582	64,2	5,62	36	Peptidyl-prolyl cis-trans isomerase FKBP10 GN=FKBP10 PE=1 SV=1
O95758	8	552	59,7	9,04	35	Regulator of differentiation 1 GN=ROD1 PE=1 SV=2
Q9UKA9	11	531	57,5	8,66	35	Polypyrimidine tract-binding protein 2 GN=PTBP2 PE=1 SV=1
O43345	21	1167	134,3	9,07	35	Zinc finger protein 208 GN=ZNF208 PE=2 SV=1
Q5JUK3	2	1230	138,3	7,52	35	Potassium channel subfamily T member 1 GN=KCNT1 PE=2 SV=2
Q7L5N1	3	327	36,1	5,73	35	COP9 signalosome complex subunit 6 GN=COPS6 PE=1 SV=1

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Q32MQ0	8	723	77,3	8,18	35	Zinc finger protein 750 GN=ZNF750 PE=1 SV=1
Q9UPQ0	12	1083	121,8	6,47	35	LIM and calponin homology domains-containing protein 1 GN=LIMCH1 PE=1 SV=4
P52926	1	109	11,8	10,62	35	High mobility group protein HMGI-C GN=HMGA2 PE=1 SV=1
Q8NG31	84	2342	265,2	5,47	35	Protein CASC5 GN=CASC5 PE=1 SV=3
P62862	3	59	6,6	12,15	35	40S ribosomal protein S30 GN=Fau PE=3 SV=1
O75208	1	318	35,5	5,94	35	Ubiquinone biosynthesis protein COQ9, mitochondrial GN=COQ9 PE=1 SV=1
O75131	13	537	60,1	5,85	35	Copine-3 GN=CPNE3 PE=1 SV=1
Q96FN4	8	548	61,2	6,07	35	Copine-2 GN=CPNE2 PE=1 SV=3

14 Eidesstattliche Erklärung

„Hiermit versichere ich, dass ich die vorliegende Arbeit selbstständig verfasst habe. Zur Anfertigung wurden keine anderen als die angegebenen Hilfsmittel verwendet. Die Stellen, die aus anderen Werken wörtlich oder sinngemäß entnommen wurden, sind als solche kenntlich gemacht. Ich versichere weiterhin, dass diese Arbeit in gleicher oder ähnlicher Form noch keiner anderen Prüfungsbehörde vorgelegen hat.“

Mainz, den 15.09.2014

Eidesstattliche Erklärung

Eidesstattliche Erklärung